**CLINICAL RESEARCH** 

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Received: 2016.08.25 Class III  $\beta$ -Tubulin in Colorectal Cancer: Tissue Accepted: 2016.09.12 **Distribution and Clinical Analysis of Chinese** Published: 2016.10.23 Patients ABCDEF Xiaoli Zhao Authors' Contribution: Department of Pathology, Beijing Tongren Hospital, Capital Medical University and Study Design A Beijing Key Laboratory of Head and Neck Molecular Diagnostic Pathology, Beijing, ABCDEF **Changli Yue** Data Collection B P.R. China **Jiamin Chen** BCDEF Statistical Analysis C BCDEF Cheng Tian Data Interpretation D Manuscript Preparation E BCDEF Dongmei Yang Literature Search E Li Xing BEF Funds Collection G Honggang Liu ACDE Yulan Jin ABCDEFG **Corresponding Author:** Yulan Jin, e-mail: yulanjin24@163.com Source of support: This work was supported by the National Natural Science Foundation of China (Grant no. 81360361) Background: Class III β-tubulin (βIII-tubulin) has been reported to express at the invasive margin of colorectal cancer. The present study aimed to investigate the clinical implication of ßIII-tubulin expression at the invasive margin of colorectal cancer. We recruited 111 patients with surgically resected colorectal carcinoma for ßIII-tubulin expression analysis. Material/Methods: The cases with ßIII-tubulin-positive tumor cells found only in the invasive front tumor area were assigned to the invasive front group, while the remaining cases were all assigned to the non-invasive front group. Clinical analysis of BIII-tubulin and other clinical data was performed. The positive staining rates and staining intensity of  $\beta$ III-tubulin were significantly different between the inva-Results: sive and non-invasive front groups (p=0.001 and p=0.006), and there was a significant difference in tumor differentiation between the 2 groups (p=0.032). In the non-invasive front group, staining intensity of ßIII-tubulin was significantly associated with positive staining rates and lymphatic metastasis (p<0.001 and p=0.048). **Conclusions:** Our data showed the tissue distribution of ßIII-tubulin expression at invasive margin or diffuse distribution. Expression of BIII-tubulin was correlated with tumor differentiation and lymphatic metastasis, suggesting a potential role of BIII-tubulin in tumor differentiation and metastasis. This study may shed light on BIII-tubulin as a novel potential molecular target for a new anti-cancer drug. **MeSH Keywords: Colorectal Neoplasms • Lymphatic Metastasis • Tubulin** Full-text PDF: http://www.medscimonit.com/abstract/index/idArt/901252 2 2 28 **5** 2 2041



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## Background

As the third most common malignancy, colorectal carcinoma is the fourth leading cause of cancer-related death worldwide [1]. In China, colorectal carcinoma has been ranked as the fifth most common human malignant disease, accounting for 6.51% of mortality in urban areas and 4.64% in rural areas [2]. The etiology of colorectal cancers is known to be heterogeneous and various mechanisms have been associated with the increased risk for development and progression of colorectal carcinoma, as well as the aggressive behaviors of the tumors [3–5]. Tumors at the invasive front appear morphologically more poorly differentiated and show different molecular characteristics [6]. Previous studies of colorectal carcinoma have suggested the potential role of β-tubulin expression in rectal cancer development [7,8]. A study by Portyanko et al. found expression of class III  $\beta$ -tubulin ( $\beta$ III-tubulin, also known as TUBB3) at the invasive margin of colorectal cancer [9]. However, the authors mainly focused on the immunoreactivity of βIII-tubulin in colorectal cancer patients, and the clinical implication of their findings has not been well investigated. Recently, Mariani et al. investigated the role of  $\beta$ III-tubulin as a predictive biomarker [10]. The distribution of tubulin has also been reported in human leukemia cell line HL-60 [11]. However, the potential effect of ßIII-tubulin expression at the invasive margin of colorectal cancer was not taken into consideration. Therefore, the present investigated the clinical implication of βIII-tubulin expression at the invasive margin of colorectal cancer.

### **Material and Methods**

### Patients and tissue samples

This was a retrospective cohort study. A total of 111 patients with colorectal carcinoma who underwent radical colorectal resection were recruited at Beijing Tongren Hospital from February 2012 to February 2015. We excluded patients with tumor history, familial polyposis colon cancer, Crohn's disease, malignant transformation of ulcerative colitis, simultaneous multiple primary tumors, or incomplete clinical data. The formalin-fixed and paraffin-embedded tissue samples containing the invasive part of the tumors were obtained and stained with hematoxylin and eosin (HE).

### Immunohistochemical assay

Immunohistochemical staining of βIII-tubulin was performed with the method of immunohistochemistry envision-two steps using MaxVision™2 (Fuzhou Maixin Biotech. Co., Ltd., Fujian, China) according to the manufacturer's instructions. Briefly, the tissue specimen was cut into 4-µm-thick sections, and deparaffinized in xylene and later rehydrated in graded alcohol. Antigen was retrieved using citrate buffer (pH 6.0) in a microwave oven. After washing 3 times with Tris-buffered saline (TBS), the sections were processed with 3% hydrogen peroxide for 5-10 min to block endogenous peroxidase activity. Sections were then incubated with mouse anti-human antibody against βIII-tubulin (Clone TuJ-1, MAB-0636, Fuzhou Maixin Biotechnology Development Co., Ltd., Fuzhou, China) or cytokeratin (AE1/AE3, KIT-0009, Fuzhou Maixin Biotech Co., Ltd., Fujian, China) overnight at 4°C, followed by horseradish peroxidase-linked sheep anti-mouse secondary antibody (KIT-5930-1, Fuzhou Maixin Biotech Co., Ltd., Fujian, China) for 60 min. The reactions were visualized by 3, 3-diaminobenzidine (DAB). The slides were then counterstained with hematoxylin. The same procedure with the primary antibody omitted was used as a negative control. Sections of peripheral nerve tissue were used as a positive control.

#### **Staining evaluation**

Expression of  $\beta$ III-tubulin was analyzed under an Olympus BX51 microscope (Olympus, Tokyo, Japan) by 2 senior professional physicians who were blinded to the clinical data of patients. The proportion of positive tumor cells was scored from 5 randomly selected slides and the results were expressed as the average of data of the 2 physicians. The cytoplasmatic staining intensity was graded as: negative (–, no staining), weak positive (+, pale yellow), moderate positive (++, claybank), and strong positive (+++, sepia).

### Study group and follow-up

According to the distribution of  $\beta$ III-tubulin in the tumor tissue, patients were assigned to either the invasive front group or the non-invasive front group. The invasive front group consisted of cases with  $\beta$ III-tubulin-positive tumor cells found only in the invasive front tumor area, while the non-invasive front group consisted of the remaining cases. The invasive front was defined as the advancing edge of the tumor. We recorded clinical data on age, sex, tumor size, differentiation status, TNM stage, and lymphatic metastasis. Enrolled patients were followed up from the time of disease diagnosis to the time of death of patients or study deadline (May 2015). This study was approved by the Ethics Committee of Beijing Tongren Hospital of the Capital Medical University and informed consent was obtained from all patients.

### Statistical analysis

Qualitative data are expressed as proportion and analyzed by  $\chi^2$  or Fisher's exact test. Normally distributed quantitative data are expressed as mean  $\pm$ SD and analyzed by ANOVA. Data with abnormal distribution are expressed as interquartile range and were analyzed by Kruskal-Wallis test. Correlations analysis was

performed by Spearman's or Pearson's test. P <0.05 was considered statistically significant.

## Results

### Clinical data

A total of 111 patients who underwent radical colorectal resection were included in this study. There were 55 males and 56 females. Mean age of the patients was 66.41±12.64 years (median: 68.00). Clinical data are summarized in Table 1. βIIItubulin was positively stained in 0.31±0.27% of the tumor cells. The staining intensity was negative in 6 cases (5.41%), weakly positive in 10 cases (9.01%), moderately positive in 65 cases (58.56%), and strongly positive in 30 cases (27.03%). The invasive front group had 72 patients with ßIII-tubulin expressed only in the invasive front of tumor tissue, and the non-invasive front group had the remaining 39 patients. HE staining results of colorectal cancer in the non-invasive front group and the invasive front group are shown in Figure 1A and 1B. Immunohistochemical analysis showed positive staining of cytokeratin (AE1/AE3) in the non-invasive front group (Figure 1C) and the invasive front group (Figure 1D). Immunohistochemical analysis of BIII-tubulin showed heterogeneous labeling in both the non-invasive front group and invasive front group, with weakly, moderately, and strongly stained areas observed (Figure 1E-1J).

# Positive staining and staining intensity of $\beta\mbox{III-tubulin}$ in colorectal carcinoma patients

The rate of positive staining of  $\beta$ III-tubulin in colorectal carcinoma patients was significantly correlated with staining intensity and tissue distribution of  $\beta$ III-tubulin expression (p=0.018 and p=0.001, respectively, Table 2). The positive staining rate of  $\beta$ III-tubulin was significantly higher in tumor tissues with higher staining intensity (p<0.05). The rate of  $\beta$ III-tubulin positive staining in the non-invasive front group was also significantly higher when compared with the invasive front group. The other data (age, sex, lymphatic metastasis, TNM stage, local recurrence, distant metastasis, and mortality) were not found to be significantly associated with positive staining of  $\beta$ III-tubulin. There was no significant difference in the clinical data among the groups with different  $\beta$ III-tubulin staining intensity (Table 3)

# Tissue distribution of $\beta \mbox{III-tubulin}$ in colorectal carcinoma patients

The patients were further divided into invasive front and noninvasive front groups according to the tissue distribution of  $\beta$ IIItubulin in colorectal carcinoma. The rate of positive staining 
 Table 1. Clinical data of patients with colorectal carcinoma.

	Colorecta patien	al carcinoma ts (n=111)			
Age (years)	66.4	66.41±12.64			
Gender n (%)					
Male		55			
Female		56			
Lymphatic metastasis n (%)					
No	54	(48.65)			
Yes	57	(51.35)			
Tumor differentiation n (%)					
Well	18	(16.22)			
Moderate	68	(61.26)			
Poor	25	(22.52)			
TNM stage n (%)					
1	18	(16.22)			
II	35	(31.53)			
Ш	46	(41.44)			
IV	12	(10.81)			
Local recurrence n (%)					
No	109	(98.20%)			
Yes	2	(1.80)			
Distant metastasis n (%)					
No	97	(87.39%)			
Yes	14	(12.61)			
Mortality n (%)					
No	102	(91.89)			
Yes	9	(8.11)			
Positive staining of $\beta$ III-tubulin (%)	0.3	1±0.27			
Staining intensity of $\beta$ III-tubulin n (%)	)				
-	6	(5.41)			
+	10	(9.01)			
++	65	(58.56)			
+++	30	(27.03)			
Tissue distribution of $\beta$ III-tubulin n (%)	)				
Invasive front group	72	(64.87)			
Non-invasive front group	39	(35,13)			



Figure 1. Histological and immunohistochemical analysis of colorectal cancer. HE staining of tumor tissue in the non-invasive front group (A, ×50) and invasive front group (B, ×50). Immunohistochemical staining of cytokeratin AE1/AE3 in the non-invasive front group (C, ×50) and invasive front group (D, ×50). Immunohistochemical analysis of βIII-tubulin with different positive intensities in the non-invasive front group (E: +; G: ++; I: +++; E, ×100, G and I, ×50) and invasive front group (F: +; H: ++; J: +++) (E, F, ×100, strong positivity of βIII-tubulin in nervous tissue as internal control; G–J, ×50).

Table 2. Clinical analysis of positive staining of  $\beta$ III-tubulin in colorectal carcinoma patients.

	Case (n)	Positive staining of bIII-tubulin (%)	t	Р
Age	111	_	-0.02	0.836
Gender				
Male	55	0.32±0.27	0.47	
Female	56	0.31±0.27	0.17	0.869
Lymphatic metastasis				
No	54	0.32±0.25	0.15	0.070
Yes	57	0.31±0.29	0.15	0.878
Tumor differentiation				
Well	18	0.40±0.30		
Moderate	68	0.28±0.25	1.76	0.177
Poor	25	0.34±0.29		
TNM stage				
I	18	0.44 <u>±</u> 0.31a		
11	35	0.28±0.21a	1.(2)	0.189
111	46	0.30±0.29a	1.02	
IV	12	0.28±0.26a		
Local recurrence				
No	109	0.31±0.27	0.25	0 7 9 2
Yes	2	0.47±0.62	-0.55	0.783
Distant metastasis				
No	97	0.32±0.27	0.16	0 975
Yes	14	0.30±0.30	0.10	0.075
Mortality				
No	102	0.31±0.27	0 58	0.566
Yes	9	0.36±0.33	-0.58	
Staining intensity of $\beta$ III-tubulin				
	6	0.00±0.00b		
+	10	0.24±0.31a	3 49	0.018
++	65	0.34±0.26a	J. <del>T</del> J	
+++	30	0.34±0.28a		
Tissue distribution of $\beta$ III-tubulin				
Non-invasive front group	39	0.44±0.33	3 5 2	0.001
Invasive front group	72	0.24±0.20	5.52	0.001

and staining intensity of  $\beta$ III-tubulin were significantly different between the invasive and non-invasive front groups (p=0.001 and p=0.006, respectively, Table 4, Figure 2A, 2B), and there was a significant difference in tumor differentiation between the 2 groups (p=0.032, Figure 2C). However, the differences in other data between the invasive and non-invasive front groups did not reach statistical significance.

# Clinical analysis of $\beta \text{III-tubulin}$ staining intensity in invasive and non-invasive front groups

Staining intensity of  $\beta$ III-tubulin in patients in the invasive and non-invasive front groups was further analyzed. As shown in Table 5, in the non-invasive front group, staining intensity of  $\beta$ III-tubulin was significantly associated with the rates of  $\beta$ III-tubulin-positive staining and lymphatic metastasis of tumors

#### **Table 3.** Clinical analysis of staining intensity of βIII-tubulin in colorectal carcinoma patients.

	– (n=6)	+ (n=10)	++ (n=65)	+++ (n=30)	χ²	Р
Age (years)	59.83±21.64	66.90±12.01	66.65±11.37	67.03±13.62	0.57*	0.635
Gender n (%)						
Male	3 (50.00)	6 (60.00)	33 (50.77)	13 (43.33)		0.829
Female	3 (50.00)	4 (40.00)	32 (49.23)	17 (56.67)	0.01	
Lymphatic metastasis						
No	5 (83.33)	4 (40.00)	31 (47.69)	14 (46.67)	0.00	0 202
Yes	1 (16.67)	6 (60.00)	34 (52.31)	16 (53.33)	0.00	0.565
Tumor differentiation n (%)						
Well	3 (50.00)	1 (10.00)	12 (18.46)	2 (6.67)		0.078
Moderate	1 (16.67)	6 (60.00)	42 (64.62)	19 (63.33)	0.00	
Poor	2 (33.33)	3 (30.00)	11 (16.92)	9 (30.00)		
TNM stage						
1	1 (16.67)	3 (30.00)	10 (15.38)	4 (13.33)		0.597
ll	4 (66.67)	1 (10.00)	22 (33.85)	8 (26.67)	0.00	
III	1 (16.67)	5 (50.00)	26 (40.00)	14 (46.67)	0.00	
IV	0 (0.00)	1 (10.00)	7 (10.77)	4 (13.33)		
Local recurrence						
No	6 (100.0)	9 (90.00)	65 (100.0)	29 (96.67)	0.05	0.100
Yes	0 (0.00)	1 (10.00)	0 (0.00)	1 (3.33)	0.05	
Distant metastasis						
No	5 (83.33)	9 (90.00)	58 (89.23)	25 (83.33)		0.785
Yes	1 (16.67)	1 (10.00)	7 (10.77)	5 (16.67)	0.03	
Mortality						
No	6 (100.0)	9 (90.00)	60 (92.31)	27 (90.00)	0.07	0.932
Yes	0 (0.00)	1 (10.00)	5 (7.69)	3 (10.00)	0.07	
Positive staining of $\beta \mbox{III-tubulin}$ (%)	0.00±0.00	0.24±0.31*	0.34±0.26*	0.34±0.28*	3.49	0.018

\* p<0.05 vs. the  $\beta$ III-tubulin negative (–) group.

(p<0.001 and p=0.048, respectively, Figure 2D), but there were no significant differences in any of the clinical data with respect to the  $\beta$ III-tubulin staining intensity in patients in the invasive front group.

### Discussion

 $\beta$ III-tubulin is one of 9  $\beta$ -isoforms that participate in formation of microtubules; it is known to play a critical role in cell growth, division, motility, signaling development, and cell shape maintenance [12]. Overexpression of  $\beta$ III-tubulin was originally identified to be a prominent factor contributing to

Table 4. Clinical analysis of patients with  $\beta$ III-tubulin distributed in invasive front and non-invasive front area.

	Non-invasive front group (n=39)		Invasive front group		χ2	Р	
Age (years)	64.	64.79±14.40		67.28±11.59		0.325	
Gender n (%)							
Male	22	(56.41)	33	(45.83)	1 1 2	0.207	
Female	17	(43.59)	39	(54.17)	1.13	0.287	
Lymphatic metastasis							
No	19	(48.72)	35	(48.61)	0.00	0.001	
Yes	20	(51.28)	37	(51.39)	0.00	0.991	
Tumor differentiation n (%)							
Well	11	(28.21)	7	(9.72)			
Moderate	19	(48.72)	49	(68.06)	6.88	0.032	
Poor	9	(23.08)	16	(22.22)			
TNM stage							
I	8	(20.51)	10	(13.89)			
II	11	(28.21)	24	(33.33)	2.01	0.390	
III	18	(46.15)	28	(38.89)	3.01		
IV	2	(5.13)	10	(13.89)			
Local recurrence							
No	38	(97.44)	71	(98.61)	0.46	1 000	
Yes	1	(2.56)	1	(1.39)	0.46	1.000	
Distant metastasis							
No	35	(89.74)	62	(86.11)	0.21	0.767	
Yes	4	(10.26)	10	(13.89)	0.21		
Mortality							
No	35	(89.74)	67	(93.06)	0.22	0 717	
Yes	4	(10.26)	5	(6.94)	0.23	0.717	
Staining intensity of $\beta$ III-tubulin							
-	6	(15.38)	0	(0.00)			
+	2	(5.13)	8	(11.11)	0.00	0.006	
++	23	(58.97)	42	(58.33)	0.00		
+++	8	(20.51)	22	(30.56)			
Positive staining of βIII-tubulin (%)	0.	0.44±0.33		0.24±0.20		0.001	

resistance to taxane drugs [13–18]. Down-regulation of  $\beta$ IIItubulin expression has been reported to contribute to favorable clinical outcome following anti-tubulin therapy [14]. Recently, however, increasing studies have shown that  $\beta$ III-tubulin is involved more in tumor development and progression than as a predictor of response to chemotherapy in various



Figure 2. Positive staining (A) and staining intensity of βIII-tubulin (B) and tumor differentiation (C) were significantly different in patients in the invasive and non-invasive front groups. Staining intensity of βIII-tubulin was significantly associated with lymphatic metastasis in the non-invasive front group (D). \*\* p<0.01 vs. the non-invasive front group.</p>

types of tumors [18–23].  $\beta$ III-tubulin has been reported to be expressed in a variety of tumors, especially in those with aggressive behavior that are likely to metastasize [24–26]. In the present study, the tissue distribution of class III  $\beta$ -tubulin expression at the invasive margin or diffuse distribution was investigated in colorectal cancer. Our study showed that  $\beta$ III-tubulin was expressed at the invasive margin in 64.78% of our colorectal cancer cases. The expression of  $\beta$ III-tubulin may be involved with tumor differentiation and lymphatic metastasis, suggesting a potential role of  $\beta$ III-tubulin in tumor differentiation and metastasis.

In colorectal cancer, aberrant  $\beta$ III-tubulin expression has been suggested to be involved with tumor development and poor survival of the patients [8,10]. However, a previous study of  $\beta$ III-tubulin expression in colorectal cancer indicated that expression of  $\beta$ III-tubulin was relatively uncommon in colorectal carcinomas [7]. A study by Portyanko et al. showed the preferential localization of  $\beta$ III-tubulin at the invasive margin in colorectal cancer, suggesting a potential role of  $\beta$ III-tubulin in modulation of the invading activity of cancer cells [9]. However, those studies had relatively small sample sizes and the clinical implications of  $\beta$ III-tubulin at the invasive margin in colorectal cancer had not yet been defined. Therefore, in the present study, 111 patients with surgically resected colorectal cancer were recruited, and the expression of  $\beta$ III-tubulin at the invasive margin was investigated. The findings of our

study showed that BIII-tubulin was expressed at the invasive margin in 72 of 111 cases of colorectal cancer, accounting for 64.87% of all cases. These data dramatically differed from the results reported by Portyanko et al., which showed BIIItubulin expression at the invasive margin in 28 of 29 cases of colorectal cancer [9]. These conflicting results can be partially explained by differences in race and number of patients recruited, as well as the lack of consistency in study criteria. However, despite these differences, our study showed a significant difference in tumor differentiation between the invasive front and non-invasive front groups, with many more poorly and moderately differentiated tumors identified in the invasive front group. This finding was partially corroborated with the results of a previous study reporting more ßIII-tubulin immunoreactivity in poorly differentiated colorectal carcinomas [7]. Sex has been recently suggested to play a role in the ability of βIII-tubulin to predict poor outcome in colorectal cancer [10]. In our study, however, no such sex-related difference was observed. The upregulated expression of βIII-tubulin has been reported in several types of cancers and was associated with aggressive behavior. A previous papillary thyroid carcinoma study showed strong staining of ßIII-tubulin in widely infiltrating PTCs, particularly at the invasive margin, or in moderately differentiated PTCs, and most cases with lymphatic metastasis showed strong ßIII-tubulin immunoreactivity [27]. In an esophageal squamous cell carcinoma study, Yu et al. found that BIII-tubulin expression was associated with

	Colorectal carcinoma patients										
	Non-invasive front group					Invasive front group					
	-	+	++	+++	χ²	Р	+	++	+++	χ²	P
	(n=6)	(n=2)	(n=23)	(n=8)			(n=8)	(n=42)	(n=22)		
Age (years)	59.83± 21.64	63.50± 7.78	64.74± 11.63	69.00± 17.78	0.45	0.719	67.75± 13.14	67.69± 11.23	66.32± 12.19	0.11	0.9
Gender n (%)											
Male	3 (50.00)	1 (50.00)	13 (56.52)	5 (62.50)	0.05	1	5 (62.50)	20 (47.62)	8 (36.36)	0.03 (	0 4 2 2
Female	3 (50.00)	1 (50.00)	10 (43.48)	3 (37.50)			3 (37.50)	22 (52.38)	14 (63.64)		0.422
Lymphatic metastasis											
No	5 (83.33)	1 (50.00)	12 (52.17)	1 (12.50)	0	0.048	3 (37.50)	19 (45.24)	13 (59.09)	0.03	0.515
Yes	1 (16.67)	1 (50.00)	11 (47.83)	7 (87.50)	U	0.040	5 (62.50)	23 (54.76)	9 (40.91)		
Tumor differentiation n (%	%)										
Well	3 (50.00)	1 (50.00)	7 (30.43)	0 (0.00)			0 (0.00)	5 (11.90)	2 (9.09)	0	0.633
Moderate	1 (16.67)	1 (50.00)	12 (52.17)	5 (62.50)	0	0.185	5 (62.50)	30 (71.43)	14 (63.64)		
Poor	2 (33.33)	0 (0.00)	4 (17.39)	3 (37.50)			3 (37.50)	7 (16.67)	6 (27.27)		
TNM stage											
I	1 (16.67)	1 (50.00)	5 (21.74)	1 (12.50)			2 (25.00)	5 (11.90)	3 (13.64)	0	0.876
II	4 (66.67)	0 (0.00)	7 (30.43)	0 (0.00)	0	0 160	1 (12.50)	15 (35.71)	8 (36.36)		
III	1 (16.67)	1 (50.00)	10 (43.48)	6 (75.00)	0	0.109	4 (50.00)	16 (38.10)	8 (36.36)		
IV	0 (0.00)	0 (0.00)	1 (4.35)	1 (12.50)			1 (12.50)	6 (14.29)	3 (13.64)		
Local recurrence											
No	6 (100.0)	2 (100.0)	23 (100.0)	7 (87.50)	0.21	0.414	7 (87.50)	42 (100.0)	22 (100.0)	0.11 (	0.110
Yes	0 (0.00)	0 (0.00)	0 (0.00)	1 (12.50)	0.21	0.414	1 (12.50)	0 (0.00)	0 (0.00)		
Distant metastasis											
No	5 (83.33)	2 (100.0)	22 (95.65)	6 (75.00)	0.05	0 202	7 (87.50)	36 (85.71)	19 (86.36)	0.12	1 000
Yes	1 (16.67)	0 (0.00)	1 (4.35)	2 (25.00)	0.05	0.302	1 (12.50)	6 (14.29)	3 (13.64)		1.000
Mortality											
No	6 (100.0)	2 (100.0)	21 (91.30)	6 (75.00)	0.00	0.440	7 (87.50)	39 (92.86)	21 (95.45)	0.14 (	0.640
Yes	0 (0.00)	0 (0.00)	2 (8.70)	2 (25.00)	0.09	0.449	1 (12.50)	3 (7.14)	1 (4.55)		
Positive staining of βIII-tubulin (%)	0.00± 0.00	0.80± 0.00*	0.46± 0.28*	0.64± 0.27*	8.91	<0.001	0.10± 0.10	0.27± 0.21	0.24± 0.20	2.47	0.092

### **Table 5.** clinical analysis of βIII-tubulin staining intensity in patients of invasive and non-invasive front groups.

\* p<0.05 vs. the βIII-tubulin negative (–) group.

the lymphatic metastasis of tumors [28]. The results of the present study showed that the staining intensity of  $\beta$ III-tubulin was associated with the lymphatic metastasis in the non-invasive front group, suggesting the potential role of  $\beta$ III-tubulin in tumor invasion or metastasis. However, our study showed no significant association of  $\beta$ III-tubulin with tumor recurrence, metastasis, or survival. Further research is needed to clarify the potential role of  $\beta$ III-tubulin in colorectal cancer and patient prognosis.

This study has some limitations. The number of patients recruited in was comparatively small. Therefore, only the patients with  $\beta$ III-tubulin expressed in the invasive front of tumor tissue were included in the invasive front group, and all remaining patients were included in the non-invasive front group. However, despite these limitations, our study did show the association of  $\beta$ III-tubulin expression with tumor differentiation and lymphatic metastasis in colorectal cancer. Studies with larger samples are needed to further clarify the clinical implications of  $\beta$ III-tubulin expression at the invasive margin in colorectal cancer.

### Conclusions

Our data showed the tissue distribution of class III  $\beta$ -tubulin expression at the invasive margin or diffuse distribution, and that  $\beta$ III-tubulin expression may be involved in tumor differentiation and metastasis. This study may shed light on  $\beta$ III-tubulin as a novel potential molecular target for new anti-cancer drugs.

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#### **Conflict of Interest**

None.

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