osteoarthritis: a systematic review

Anna Boada-Pladellorens 🕩, Mercè Avellanet, Esther Pages-Bolibar and Anna Veiga

Abstract

Background: Regenerative cell therapies, such as adipose-derived stromal vascular fraction (SVF), have been postulated as potential treatments for knee osteoarthritis (KOA). **Objectives:** To assess the efficacy and safety of SVF treatment against placebo and other standard therapies for treating KOA in adult patients. **Design:** A systematic review.

Data sources and methods: We searched the following databases: MEDLINE via PubMed, Epistemonikos, PEDro, DynaMed, TripDatabase, Elsevier via Clinicalkey and Cochrane Controlled Trials Register. We included prospective interventional studies where treatment with SVF in adults with KOA was compared against placebo or other standard therapies, and results were objectively measured with at least one widely recognised osteoarthritis scale. **Results:** Among 266 studies published until May 2021, nine met our inclusion criteria. A total of 239 patients (274 knees) were included in our study. The follow-up ranged from 6 to 24 months. Six studies had a control group (only one being placebo). All studies showed that SVF improved pain and functionality measured, in most cases, with the visual analogue scale and the Western Ontario and McMaster Universities Osteoarthritis Index. In addition, five studies reported an improvement in anatomical structures, as detected in MR images. However, the number of cells contained in SVF varied substantially between different studies, which could induce a comparison bias.

Conclusion: Although based on a small number of dissimilar studies, SVF was considered a safe treatment for KOA and could be promising in terms of pain, functionality and anatomical structure improvement. However, SVF products need to be standardised, the number of cells homogenised and the use of concomitant treatments reduced to establish proper comparisons.

Registration: PROSPERO registration number: CRD42021284187.

Keywords: knee osteoarthritis, mesenchymal stem cells, regenerative medicine, stromal vascular fraction

Received: 15 February 2022; revised manuscript accepted: 19 July 2022.

Introduction

Osteoarthritis is a chronic degenerative joint condition characterised by the progressive destruction of the articular cartilage, leading to pain and functional loss. This disease is among the main causes of disability in adults, and the knee is the most frequent joint affected.^{1,2} Knee osteoarthritis (KOA) is estimated to affect 265 million people worldwide and its prevalence has risen to approximately 9% over 28 years (from 1990 to 2017).^{3–5} Besides, osteoarthritis prevalence is expected to continue increasing from 22% in 2003 to 25% by 2030 in the United States and from 26.6% in 2012 to 29.5% by 2032 in Sweden.⁶

Several treatments have historically been explored for KOA, including pharmacological and surgical approaches, but most provide only symptomatic relief. Intra-articular treatments, such as Ther Adv Musculoskelet Dis

2022, Vol. 14: 1–15

1759720X221117879

© The Author(s), 2022. Article reuse guidelines: sagepub.com/journalspermissions

Correspondence to: Anna Boada-Pladellorens Physical Medicine and

Physical Medicine and Rehabilitation Department, Hospital Nostra Senyora de Meritxell, Carrer dels Escalls, AD700 Escaldes-Engordany, Andorra.

Celular Clinic, Escaldes-Engordany, Andorra. Research Group in Health Sciences and Health Services, University of Andorra, Sant Julia de Loria, Andorra. annaboadal@gmail.com

annaboadaidgmail.con

Mercè Avellanet Physical Medicine and Rehabilitation Department, Hospital Nostra Senyora de Meritxell, Escaldes-Engordany, Andorra

Celular Clinic, Escaldes-Engordany, Andorra

Research Group in Health Sciences and Health Services, University of Andorra, Sant Julia de Loria, Andorra

Esther Pages-Bolibar

Physical Medicine and Rehabilitation Department, Hospital Nostra Senyora de Meritxell, Escaldes-Engordany, Andorra

Research Group in Health Sciences and Health Services, University of Andorra, Sant Julia de Loria, Andorra

Anna Veiga

Barcelona Stem Cell Bank, Regenerative Medicine Programme, Institut d'Investigació Biomèdica de Bellvitge (IDIBELL), Hospital Duran i Reynals, Barcelona, Spain

journals.sagepub.com/home/tab



Creative Commons Non Commercial CC BY-NC: This article is distributed under the terms of the Creative Commons Attribution-NonCommercial 4.0 License (https://creativecommons.org/licenses/by-nc/4.0/) which permits non-commercial use, reproduction and distribution of the work without further permission provided the original work is attributed as specified on the SAGE and Open Access pages (https://us.sagepub.com/en-us/nam/open-access-at-sage).

corticoids, hyaluronic acid (HA) and platelet-rich plasma (PRP), are the most widely used drugs to treat KOA. Interestingly, recent studies have suggested that corticoids could be chondrotoxic, inducing early KOA.7 On the other hand, HA and PRP are therapies whose effectiveness is still debated, and a unanimous consensus about their appropriate indication has not vet been reached. Hence, no widespread criteria for prescribing the abovementioned intra-articular drugs are available.8-10 At present, arthroplasty for severe cases is the only curative treatment for KOA. However, this surgery entails non-negligible complications, including infection, residual pain and stiffness.¹¹ Treatment with mesenchymal stem cells (MSCs), namely, multipotent stromal cells that can differentiate into osteoblast, adipocytes and chondrocytes, is among the new strategies to treat KOA. MSCs have high plasticity, self-renewal capabilities, and immune-suppressive and anti-inflammatory properties.^{12,13}

Among the available sources of MSCs, two have received the greatest scientific attention: adiposederived mesenchymal stem cells (ADSCs) and marrow mesenchymal stem cells.14 bone Belonging to ADSCs, the stromal vascular fraction (SVF) isolated from adipose tissue is increasingly used to treat KOA.15 SVF is a heterogeneous product that contains ADSCs, macrophages, blood cells, pericytes, fibroblasts, endothelial cells and their progenitors. This cellular heterogeneity entails a high therapeutic potential because of their complementary mechanism of action. Up to 60% of the cells within SVF are CD34+, a marker present on cells from the vascular microenvironment. Basic characterization of SVF has been provided by the International Society for Cellular Therapy, but the exact phenotype of ADSCs is still unknown, as biomarkers differ in vivo and in vitro.16 SVF can be easily obtained in large amounts from autologous adipose tissue by liposuction and used without culture or differentiation. The interest in SVF stems from its extensively described immunomodulatory, anti-inflammatory, proangiogenic, antiapoptotic and antifibrotic properties.^{17,18} Some of these actions may be attributed to the presence of ADSC (ranging from 2% to 10%), while others are associated with the paracrine effect of cell phenotypes present in SVF.16

Although several studies have reported the superiority of using SVF over other therapies for KOA, the evidence remains limited by the lack of randomised placebo-controlled clinical trials.¹⁹ The most recently published systematic reviews and meta-analyses have pooled together, in the same analysis, different MSC therapy products, including SVF.^{20–22} The comparison of different types of cell therapies could be misleading and induce biased conclusions, as has already been warned by some authors.²³

Therefore, the aim of this systematic review was to assess the efficacy and safety of SVF against placebo and other standard therapies for treating KOA in adult patients.

Methods

Protocol and registration

We identified and examined the available literature but could not undertake a meta-analysis because of the heterogeneity of available data and study outcomes. We report our findings as suggested by the Preferred Reporting Items for Systematic Reviews and Meta-Analyses statement (PRISMA). The protocol for this systematic review was registered in the Prospective Register for Systematic Reviews (PROSPERO; registration number: CRD42021284187).

Population, intervention, comparison and outcomes

We created inclusion and exclusion criteria using the PICO (population, intervention, comparison and outcomes) model. To be eligible, a study had to meet the following PICO criteria:

- P: Adults over 18 years old suffering from KOA.
- I: Treatment with SVF.
- C: Placebo or other therapies, including arthroscopic microfracture (AM), ADSCs, HA, or PRP.
- O: Objective measure with at least one widely recognised osteoarthritis scale [e.g. WOMAC (Western Ontario and McMaster Universities Osteoarthritis Index), KOOS (Knee Injury and Osteoarthritis Outcome Score)] or a magnetic resonance imaging (MRI) scan and reported adverse effects.

Search strategy

In May 2021, we conducted a search in seven electronic databases: MEDLINE via PubMed, Epistemonikos, PEDro, DynaMed, TripDatabase,

Elsevier via Clinicalkev and Cochrane Controlled Trials Register. In addition, we manually searched references from all available reviews15,20-29 on SVF to verify that none were missing from our initial search. The search included the terms 'stromal vascular fraction', 'knee osteoarthritis' or 'osteoarthritis', 'cartilage', 'adipose-derived stem cells' or 'adipose-derived stromal cells', and 'mesenchymal stem cells' or 'mesenchymal stromal cells' in the title and abstract of all trial registers and databases. Date restriction was applied to only include studies published within the last 10 vears (when SVF treatments have increasingly been used). An example of a full-string search can be found in Appendix 1. We did not impose a language restriction in our search, but all articles meeting our inclusion criteria were written in English.

Study selection

We included only prospective interventional studies, either randomised or non-randomised, that met our PICO criteria. We only included published articles. We excluded articles not meeting our PICO criteria, narrative reviews, systematic reviews, meta-analysis and clinical trial registers. One reviewer (A.B.-P.) performed the eligibility assessment and two other reviewers (M.A. and E.P.-B.) revised it independently in an unblinded, standardised manner. Disagreements between reviewers were resolved by consensus. We used a designed electronic spreadsheet (Excel, Microsoft Corporation 2021) to enhance the consistency of data collected by the reviewers. First, we reviewed the titles to check for relevance and removed duplicates. Then, we screened abstracts to verify whether the article met our inclusion criteria. Finally, we retrieved and analysed full-text manuscripts.

Data extraction

Two independent reviewers (A.B.-P. and M.A.) collected the following characteristics from eligible studies: study registration, number of participants, study design, intervention, control, number of cells used, follow-up period, outcomes and adverse effects. Extracted data were summarised in an Excel spreadsheet and any disagreement between reviewers was discussed until reaching a consensus. A third reviewer (E.P.-B.) checked the extracted data to verify that the process was performed correctly. Unreported data were not considered, but reviewers asked for additional

details from the authors of the article if some relevant data were missing.

Risk of bias assessment

Three independent reviewers (A.B.-P., M.A. and E.P.-B.) critically appraised each study to ensure relevance using Risk of Bias 2 (RoB2) for randomised studies and ROBINS-1 for non-randomised ones. For randomised studies, we assessed the following characteristics: randomisation process, deviations from intended interventions, missing outcome data, measurement of the outcome selection of the reported result and overall bias. For non-randomised studies, we considered the following aspects: bias due to confounding, bias in selection of participants into the study, bias in the classification of interventions, bias due to deviations from intended interventions, bias due to missing data, bias in measurement of outcomes and bias in the selection of the reported results. We classified the risk of bias as 'low risk', 'some concerns' or 'high risk'. Any uncertainty was solved through discussion between the reviewers.

Data analysis

We performed a qualitative analysis based on extracted data by study type and population, and then by quality and specific results for each study. We also evaluated qualitatively the heterogeneity of the included studies. Furthermore, we listed the adverse effects reported to assess the safety of SVF.

Results

Study selection

The initial literature search resulted in 266 entries. Once duplicates were removed, 218 studies were screened by title and abstract. A total of 201 articles were excluded at this stage, mainly because they were not interventional trials, they studied different pathologies or joints from those of interest here, they were ongoing clinical trials or they were considered basic research articles (Figure 1). Eight of the 17 studies assessed in detail for eligibility were excluded, and the reasons can be found in Supplementary Table 1. A total of nine articles were finally analysed in this systematic review: two randomised clinical trials (RCT), two non-randomised clinical trials, three cohort studies, and two case series.

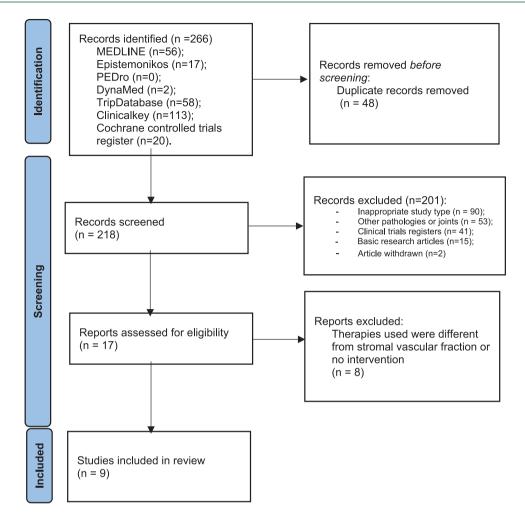


Figure 1. PRISMA flow diagram of article retrieval.

Study characteristics

The characteristics of all included studies are summarised in Table 1. The duration of the follow-up ranged from 6 to 24 months (6 months in one study,³⁰ 12 months in six studies,³¹⁻³⁶ 18 months in one study³⁷ and 24 months in one study).38 The included studies involved a total of 239 participants and the equivalent of 274 knees. The main inclusion criteria were adults with unilateral or bilateral KOA, according to the Kellgren–Lawrence (KL) grading scale, and who had undergone an ineffective conservative treatment. Exclusion criteria included the following: secondary arthritis, medical conditions that precluded an anaesthetic procedure, psychiatric disorders, history of cancer, pregnancy, coagulopathy, signs of infection or syphilis- or HIV-positive serological results, knee joint surgery and intraarticular injection of any drug within the previous 3 months.

Intervention

All studies were performed in a single centre. The studies were performed in Vietnam,^{37,38} Japan,^{30,34} the United States,^{32,33} China,³¹ Germany³⁶ and Australia.³⁵ In all studies, the intervention was a treatment with SVF. However, although the authors described the therapy as SVF, different methodological procedures and cell types were used in each study. Some of them used commercially manufactured kits,^{33,34,36,37} while others used a manual procedure^{31,32,35,38} to obtain the SVF. Three studies used concomitantly other therapies, such as AM^{37,38} or PRP.³⁶ All analysed studies employed autologous treatments.

Control

Only one study had a placebo control,³² five studies used an active control^{30,31,36–38} and three were non-controlled.^{33–35} The active controls in the

Study	Participants	Intervention (SVF kit used, if any)	Control	Outcome	Adverse effects, <i>n</i> (%)
Hong <i>et al.</i> ³¹ RCT	16 patients (32 knees; 16 knees in each group)	SVF	НА	VAS, WOMAC, ROM, WORMS, MOCART	Pain in the abdomen 1 week after liposuction [4 (25]] and pain and swelling in bilateral knee joints [6 (37.5]]
Garza <i>et al.</i> ³² RCT	39 patients (13 in each group)	High-dose SVF; low- dose SVF (GID SVF-2, Louisville, CO)	Placebo	WOMAC, modified Outerbridge classification	None
Tran <i>et al.</i> ³⁸ Non-RCT	33 patients	Microfracture + SVF (Geneworld Co. Ltd., Ho Chi Minh City, Vietnam)	Microfracture	VAS, Lysholm, WOMAC, modified Outerbridge classification	N/A
Fodor and Paulseth ³³ Cohort study	6 patients (8 knees)	SVF (GID SVF-1, Louisville, CO, USA)	None	VAS, WOMAC, ROM, TUG, MRI	Minimal discomfort, oedema and ecchymosis after liposuction. No adverse effects related to the knee injection
Tsubosaka <i>et al.</i> ³⁴ Cohort study	57 patients	SVF (SVF Celution® 800/ CRS System)	None	VAS, ROM, muscle force, WOMAC, JKOM, KOOS, radiographic imaging, MRI	None
Simunec <i>et al.</i> ³⁶ Case series	12 patients	SVF (SVF Q-graft®)	SVF + PRP	KOOS, MRI	Some pain-free swelling on the injection site and hematomas and muscle soreness-like pain in the tissue harvesting site that resolved without further interventions
Gibbs <i>et al.</i> 3 ⁵ Case series	4 patients (7 knees)	SVF + PRP	None	KOOS (TUG, Stair Climbing Test)	N/A
Yokota <i>et al.</i> ³⁰ Cohort	42 patients [59 knees]	SVF	ADSCs	VAS, KOOS, ROM	Mild knee effusion [3 (7.14)-one needed a knee aspiration], abdominal pain [6 (14.2]], internal bleeding [5 (11.9]] at the incision site, subcutaneous induration at the abdominal area fat harvest site [12 (28.5]]. All resolved without intervention
Nguyen <i>et al.³⁷</i> Non-RCT	30 patients	AM + SVF [GeneWorld] + PRP	AM	VAS, WOMAC, Lysholm, radiographic imaging, MRI	None

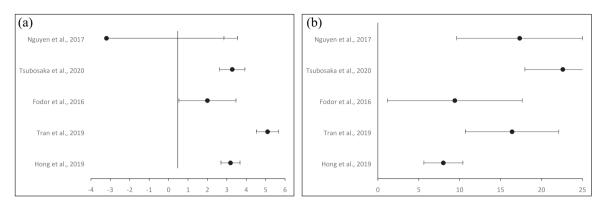


Figure 2. Forest plots showing changes in main outcomes 12 months after treatment: (a) mean VAS (Visual Analogue Scale) and (b) mean WOMAC (Western Ontario and McMaster Universities Osteoarthritis Index.

analysed studies included HA,³¹ AM^{37,38} (both studies treated the intervention group with micro-fracture and SVF), PRP mixed with SVF³⁶ and intra-articular injection of ADSCs.³⁰

Outcome

In all studies, the primary outcomes were knee pain and functionality assessed with either WOMAC or KOOS scales. Radiographic imaging or MRI was also evaluated in most studies (seven out of nine).^{31–34,36–38}

Adverse effects

Of the seven studies that assessed safety, three reported no adverse effects.^{32,34,37} After the lipo-suction, four studies described minimal discomfort, oedema and ecchymosis, which resolved without intervention, in a few cases.^{30,31,33,36} Two studies reported knee pain and swelling.^{30,31} All were described as minor adverse effects.

Results for included studies

The majority of the studies recorded two types of outcomes: clinical (concerning pain and/or functionality) and radiological outcomes. The most widely employed clinical scoring systems were the visual analogue scale (VAS)^{30,31,33,34,37,38} and the WOMAC.^{31-34,37,38} Four studies used the KOOS scale.^{30,34-36} Range of motion assessment,^{31,34} Lysholm score,^{37,38} Timed Up-and-Go test,³⁵ Stair Climbing Test³⁵ and the Japanese Knee Osteoarthritis Measure³⁰ were secondary clinical outcomes recorded in six articles. Functional outcomes improved in the intervention group in all studies. Seven studies assessed radiological outcomes using MR images.^{31–34,36–38} However, only four studies used standardised radiological scoring systems. The Whole-Organ MRI Score and the Magnetic Resonance Observation of Cartilage Repair Tissue were used in one study³¹ and the Outerbridge Classification System in three other studies.^{32,37,38} Among these seven studies, two did not observe significant differences in MR images. One performed the control MRI 3 months after intervention³³ and the other 6 and 12 months after treatment in a total of 26 patients.³² The five remaining studies found significant improvement in MR images between baseline and a 12- or 24-month control.^{31,34,36–38}

A pooled analysis of mean VAS and WOMAC scores 12 months after treatment, for studies with available data, can be found in Figure 2. Notably, two of these studies (Nguyen *et al.*³⁷ and Tran *et al.*³⁸) used AM as a coadjutant treatment in the intervention group, which may devalue the analysis. Figure 3 describes the pooled analysis of studies exclusively using SVF in the intervention group.

Results for individual studies

The main results of included studies are summarised in Table 2. Hong *et al.*,³¹ in a RCT, showed a significant improvement in the SVF group for all clinical scores including VAS [mean: 3.19; standard deviation (SD): 0.98; p < 0.001], WOMAC pain (mean: 8; SD: 4.77; p < 0.001), WOMAC stiffness (mean: 2.25; SD: 2.11; p < 0.001) and knee range of motion (mean: 19.06; SD: 7.76; p < 0.001) 12 months after intervention. In addition, the study showed positive radiologic results:

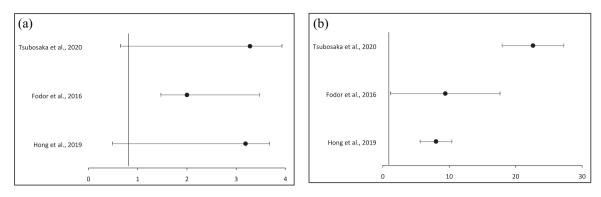


Figure 3. Forest plots showing changes in main outcomes 12 months after treatment in studies using exclusively SVF in the intervention group: (a) mean VAS (Visual Analogue Scale) and (b) mean WOMAC (Western Ontario and McMaster Universities Osteoarthritis Index).

a decrease in the Whole-Organ MRI Score (mean: 15.44; SD: 21.95; p < 0.05) and a significant Magnetic Resonance Observation of Cartilage Repair Tissue score improvement (mean: 62.81; SD: 8.16; p < 0.001) compared with the control group (HA). Similarly, Tran et al.³⁸ found that SVF treatment improved pain and functionality since a reduction in the mean VAS (from 5.1; SD: 1.2 at 12 months to 3.4; SD: 1.8 at 24 months) and mean WOMAC (from 16.4; SD: 12.1 at 12 months to 11.1; SD: 11.9 at 24 months; p < 0.05) scores was observed. Besides, radiological results also improved 24 months after treatment as assessed by the Outerbridge score (mean: 3.0; SD: 0.8 versus mean: 2.0; SD: 0.7; p<0.05). Characteristically, these authors observed that KL3-grade KOA patients showed a higher decrease in pain and improvement in functionality than those with a KL2 grade. On the contrary, Simunec et al.36 suggested that, although both KL3 and KL4 grade showed an improvement in KOOS scores 12 months after treatment, the effect was milder in KL4-grade patients (7.7%) than in those with a KL3 grade (34.5%).

On the other hand, Tsubosaka *et al.*³⁴ demonstrated a statistically significant clinical improvement with several functional scores (WOMAC, KOOS and Japanese Knee Osteoarthritis Measure) 12 months after SVF treatment. They also observed recovery in MR images 12 months after treatment, albeit no validated scoring system was used. Similarly, Nguyen *et al.*³⁷ found better Outerbridge scores in the MR images of the treatment group than in those of the control group (AM) 12 months after the intervention. In the study of Gibbs *et al.*,³⁵ improvement was only reported in knee-related quality-of-life KOOS scores. In addition, both Fodor and Paulseth's³³ and Garza *et al.*'s³² studies described better WOMAC results in the treatment group. However, Garza *et al.*³² could not find differences between the MR images of the SVF and the placebo group assessed at the 3-, 6-, and 12-month visits, and neither did Fodor and Paulseth³³ between MR images of their cohort taken at baseline and 3 months after treatment.

The number of cells contained in SVF varied substantially between the different studies and within participants of each study, which could induce a comparison bias. For example, Garza *et al.*,³² in an RCT, suggested that a high dose of SVF $(3.0 \times 10^7 \text{ SVF cells})$ provided an additional therapeutic relief of KOA symptoms than a low dose $(1.5 \times 10^7 \text{ SVF cells})$. Surprisingly, in the study of Yokota *et al.*,³⁰ the authors stated that the number of cells injected was unknown.

Risk of bias within studies

The overall risk of bias in all included studies was considered high, except for the two RCTs (Figure 4). The allocation sequence was randomised and concealed until the enrolment and intervention assignment in two studies.^{31,32} One study was double-blinded,³¹ whereas in others, the blinding process was unknown or non-existent. Data for main outcomes (pain and functionality) were available for nearly all included participants in all studies. In most studies, the measuring of the outcomes was appropriate, except for the prematurity on the MRI analysis (less than 12 months after the intervention) in two studies^{32,33} or the THERAPEUTIC ADVANCES in Musculoskeletal Disease

Study	Number of cells in SVF (mean \pm SD)	Pain (mean \pm SD)	Functionality (mean \pm SD)	Imaging (mean \pm SD)
Hong <i>et al.</i> ³¹	$29.8 \pm 3.73 \times 10^{6}$	VAS improved by 3.19 ± 0.98 ($p < 0.001$) at 12 months	Improved WOMAC pain (8 \pm 4.77), WOMAC stiffness (2.25 \pm 2.11), and ROM (19.06 \pm 7.76), (p < 0.001) at 12 months	Mean WORMS decreased by 15.44 ± 21.95 ($p < 0.05$) at 12 months Better MOCART score improvement in test group than in control group, both at 6 and at 12 months ($p < 0.001$)
Garza <i>et al.</i> ³²	High dose: 3.0×10^7 Low dose: 1.5×10^7 Placebo: no cells	N/A	At 6 months, the median percentage change in WOMAC score for high- and low-dose groups was greater than MCID and than that of the placebo group. At 12 months, high- and low-dose groups continued improving. whereas the placebo group returned towards baseline	No MRI changes from baseline or any evidence of disease progression at 6 months
Tran <i>et al.</i> ³⁸	9-12×10 ⁷	VAS reduced from 5.1 \pm 1.2 (at 12 months) to 3.4 \pm 1.8 (at 24 months) (p < 0.05). VAS in placebo group increased from 4.9 \pm 2 to 5.9 \pm 2.47 at 24 months	WOMAC score decreased at 12 months (44.7 \pm 15.4 versus 16.4 \pm 12.1, $p < 0.05$) and further at 24 months (11.1 \pm 11.9 versus 16.4 \pm 12.1, p < 0.05). Compared with placebo, the decreasing trend in the treatment group was larger	Bone marrow oedema length was larger before treatment (2.4 ± 0.34) than after 24 months (0.9 ± 0.73) , (p < 0.05). In the placebo group, the oedema increased $(1.9 \pm 0.74 \text{ versus } 2.1 \pm 0.64,$ p < 0.05). At 24 months, the Outerbridge score decreased in the treatment group $(3.0 \pm 0.8 \text{ versus } 2.0 \pm 0.7, p < 0.05)$ but remained unchanged in the placebo group
Fodor and Paulseth ³³	$14.1 \pm 11.8 \times 10^{6}$	VAS decreased (5.9 to 2.1, <i>p</i> < 0.05) at 12 months	WOMAC score decreased (32.9 to 9.4, $p = 0.05$) at 12 months	No MRI changes from baseline 3 months after treatment
Tsubosaka et al. ³⁴	$7.6 \pm 2.5 imes 10^7$	VAS improved (46.5 \pm 23.5 to 32.8 \pm 24.7, $p < 0.01$) at 12 months Both ROM and muscle force of knee extension and flexion improved at 12 months	At 12 months, improved total WOMAC score (33.4 ± 18.2 to 22.6 ± 17.5, $p < 0.01$); JKOM (34.9 ± 18.2 to 26.8 ± 19.7, $p = 0.04$); and KOOS (48.7 ± 15.8 to 58.6 ± 16.8 , $p < 0.01$)	T2 mapping values of anterior and posterior lateral and anterior medial compartment were lower at 12 months than at baseline
Simunec <i>et al</i> . ³⁶	7.56×10 ⁶	N/A	In grade 3 KOA patients, KOOS improved 34.5% <i>versus</i> 9.7% in control group at 12 months. In grade 4 KOA patients, the average KOOS score dropped by 7.7%, whereas it improved in control group to 28.8% at 12 months	MR images showed restructuration of the cartilage at 16 months and increase in joint space at 14 months

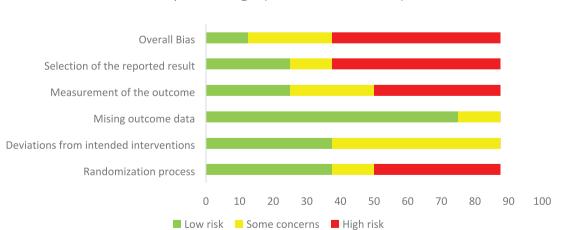
Table 2. Main results of the included studies.

(Continued)

Table 2. (Continued)

Study	Number of cells in SVF (mean \pm SD)	Pain (mean \pm SD)	Functionality (mean \pm SD)	Imaging (mean \pm SD)
Gibbs <i>et al.</i> ³⁵	$5.0 \pm 1.15 imes 10^{7}$	N/A	All 7 joints improved to >94 in knee-related quality of life. All patients improved in all five KOOS subscales >8–10 points	N/A
Yokota <i>et al.</i> ³⁰	Unknown	VAS improved in both groups at 6 months regardless of KL- grade KOA. Greater improvement in ADSC group $(54.6 \pm 21.7\%)$ than in SVF group $(44 \pm 26.1\%, p < 0.05)$	KOOS symptoms occurred earlier in the ADSC group, with significant improvement detected at 3 months ($p < 0.05$). Other KOOS domains were similar for both groups	N/A
Nguyen <i>et al</i> . ³⁷	1 × 10 ⁷	VAS scores in the treatment group increased gradually post-treatment. In the placebo group, they increased after 6 months and gradually decreased at 12 and 18 months	WOMAC score decreased at 6 (19.27 \pm 14.87), 12 (17.33 \pm 14.91) and 18 months (12.40 \pm 13.44) and was significantly different from placebo ($p < 0.05$) compared with baseline (42.87 \pm 16.29). In the placebo group, the WOMAC score increased from 25.60 \pm 19.69 at 12 months to 37.08 \pm 21.45 at 18 months (not significantly different from pretreatment scores)	At 12 months, the Outerbridge score in the treatment group (2.93 ± 0.88) decreased from baseline (3.33 ± 0.97) but was not statistically significant. In the placebo group, Outerbridge score clearly increased

AADSCs, adipose-derived tissue stem cells; JKOM; Japanese Knee Osteoarthritis Measure; KL, Kellgren–Lawrence; KOA, knee osteoarthritis; KOOS, Knee Injury and Osteoarthritis Outcome Score; MCID, minimal clinically important difference; MOCART, Magnetic Resonance Observation of Cartilage Repair Tissue; MRI, magnetic resonance imaging; N/A, not available; ROM, range of motion; SVF, stromal vascular fraction; VAS, Visual analogue scale; WOMAC, Western Ontario and McMaster Universities Osteoarthritis Index; WORMS, Whole-Organ Magnetic Resonance Imaging Score.



As percentage (intention-to-treat)

Figure 4. Overall risk of bias for all included studies in an intention-to-treat analysis. Visualised with the ROBVIS tool.

assessment of a unique outcome in another.³⁵ However, in several studies, outcome judgement assessors might have been influenced by the awareness of the intervention received. Finally, in most studies, results had been assessed based on multiple eligible outcome measurements within the outcome domain, but few analyses had been performed (Table 3).

Discussion

To the best of our knowledge, this is the first report to review the efficacy and safety of SVF to treat KOA in adult patients against placebo and other therapies. According to the analysed studies, SVF is safe to treat KOA, entailing only a few minor adverse effects. Although low-quality studies have also been included here, all studies concluded that SVF treatment improves the symptoms and functionality of KOA patients. Inconsistencies in its effect on anatomical structures have been found, maybe due to different follow-up periods, but the majority of studies showed an improvement in MRI scans 12 months after treatment.

We have reviewed the best available evidence on SVF treatment: systematic reviews and metaanalysis. Although conclusions favour the intervention arm, a thorough analysis of the composition of cell therapies used in the studies included in these reviews unveiled very different methodologies. Like comparing apples and oranges, recent systematic reviews and metaanalyses have pooled data from studies of MSC treatments performed with different types and concentrations of cell therapy products, which could be deemed a methodological limitation of the analysis.^{20–23,25,28,39,40}

In addition, we have detected a lack of well-categorised MSC products. Albeit MSCs are the focus of current research in multiple medical areas, many of their subtypes' properties and therapeutic potential are still unknown.⁴¹ Although the efficacy of these cells to modulate inflammation has been shown in different animal models, the results obtained in human clinical trials have been more modest.42 Diverse controversial issues on their biology (including their specific phenotype, the requirement of an inflammatory environment to induce immunosuppression or the cell delivery route, among others) persist.¹³ Therefore, a consensus on the definition, composition, action mechanism and production process of MSC is still missing.⁴³ Besides, the minimum

effective dose needed for cell therapies is also unknown. Although Muthu *et al.*¹⁹ already stated superior functional outcomes with non-cultured products of MSC to treat KOA over cultured ones, researchers are still considering different action mechanism hypotheses. The debate is still open on this matter with solid arguments for cultured and non-cultured products.^{13,44–46}

Herein, to tackle the absence of a review focused solely on the efficacy of SVF, only interventional studies testing SVF treatment have been included, but a paucity of high-level evidence studies (e.g. RCTs evaluating clinical and radiological effects) has been observed. This led us to include low-level evidence studies as long as the SVF treatment was well-described and categorised. Although our inclusion criteria have been well-established, only a few articles could finally be included in our analysis. This explains why studies using cultured ADSCs,⁴⁷⁻⁴⁹ processed adipose tissue containing ADSCs,^{4,50} a mixture of extracellular matrix and ADSCs⁵¹ or microfragmented lipoaspirate⁵² have been withdrawn in our screening. Of the included studies, only two were RCTs^{31,32} and just one compared SVF treatment with placebo.32 Despite the potential limitation of pooling and comparing such data – considered with a high risk of bias by the RoB2 and ROBINS-1 tools - and the few articles included, the trials analysed here pointed towards the superiority of SVF treatment over other therapies for KOA on all outcomes considered.

On the other hand, previous clinical trials have indicated the ability of intra-articular ADSC therapy to modify the progression of osteoarthritis.53,54 Progression rates previously reported have been variable,^{55,56} likely related to differences in stage of disease, definition of progression and studied population.⁵⁷ As already suggested by Iolascon et al.,⁵⁸ detecting early-stage KOA (defined as early osteoarthritis) could help to better prescribe these innovative therapeutic approaches. Furthermore, the recommended follow-up period to assess cartilage changes in MRI is at least 12 months.⁵⁹ Among the studies included in our review that assessed MRI changes, Fodor and Paulseth³³ and Garza et al.32 could not find changes in MRI scans, but their follow-up was shorter than 12 months (3 and 6 months, respectively), which could explain these observations.

Clinical and functional outcomes must also be evaluated over time. Except for the study of Yokota *et al.*,³⁰ with a follow-up period of just 6

Study	Randomisation process	Deviation from the intended intervention	Missing outcome data	Measurement of the outcome	Selection of the reported result	Overall
Hong <i>et al.</i> ³¹	+	+	+	+	+	+
Garza <i>et al.</i> ³²	+	!	+	!	+	!
Tran <i>et al.</i> ³⁸	-	!	+	-	!	-
Fodor and Paulseth ³³	-	-	+	-	-	-
Tsubosaka <i>et al.</i> ³⁴	-	+	+	-	-	-
Simunec <i>et al.</i> ³⁶	-	-	+	-	-	-
Gibbs <i>et al.</i> ³⁵	-	-	+	-	+	-
Yokota <i>et al.</i> ³⁰	•	+	+	+	•	-
Nguyen <i>et al.</i> 37	!	+	!	!	-	-

Table 3. Risk of bias for each included study.

months, the follow-up of all other studies has been over 12 months, which provides some consistency to our analysis of their results. Nguyen *et al.*³⁷ presented the only negative pain result, reporting a gradually increasing mean VAS score in the treatment group. Notably, these authors have compared two groups who previously underwent an AM procedure, which can mask results due to persistent post-operative pain or worsening of the knee cartilage surface. On the contrary, the other studies using coadjutant treatments to SVF (Tran *et al.*³⁸ and Simunec *et al.*³⁶) have shown positive results in the intervention group.

As previously mentioned, there is yet no proper definition and reproducible manufacturing procedure for SVF treatments. Many different commercially available manufactured kits exist, but although they may look similar, they are not. Kits differ in the time allowed for the collagenase to digest, the neutralisation process, the time and intensity of centrifugation, the SVF dose cell yield, their viability and composition, cost and total processing time.⁶⁰ Given this heterogeneity, although some authors defend the use of kits instead of the reference method (manually manufactured in a laboratory by a technician),⁶¹ the best method to obtain SVF is still debated. In our review, just three studies have used the reference manual method,^{30,31,35} and one of them³⁵ added ultrasonic cavitation to the collagenase step.

Finally, in agreement with previous systematic reviews that included SVF among analysed MSC,^{20–23} only minor adverse effects in the treatment group have been found, most of them caused by the liposuction procedure. They all resolved spontaneously or with mild treatment and in a short time, supporting our conclusion that SVF is a safe treatment for KOA in adult patients.

Limitations

The results of the current study should be interpreted in the context of its limitations. First, only a small number of high-quality studies could be found. Because of the lack of placebo-controlled RCTs, low-quality studies (e.g. case series) were included and a meta-analysis could not be performed. There was also significant variability in the preparation and concentration of SVF products, and, in one study, the number of injected cells was unknown.³⁰ Although the minimum SVF effective dose is yet to be established, these inconsistencies could have limited our analysis. In addition, although pain and functionality were assessed as the main outcomes in most studies, different validated scores were used and some data were missing. Therefore, our forest plot analysis was incomplete, missing data from some studies. Although Satué et al.62 have described the immunomodulatory and cartilage regeneration effects of SVF *in vitro*, the time that SVF cells maintain their properties intra-articularly is still unknown. Consequently, a follow-up period shorter than 1 year might be insufficient to completely evaluate the effect of SVF treatment. Furthermore, not all studies have assessed anatomical changes. Albeit histological samples (which would better describe the anatomical changes) could not be analysed for obvious ethical concerns, MRI evaluation with validated scores has been rarely undertaken. All these methodological pitfalls between studies could confound our comparisons and limit our conclusions.

Performing a clinical trial following the International Standards of Good Clinical Practice⁶³ is a great challenge in cell therapies. Limited access to cell therapy biologists and specialised laboratories by clinicians, strict legal and governmental policies imposed by each country's concerned authorities and the high cost of cell therapy procedures prevent the development of clinical trial protocols and their use in the clinic. From our point of view, placebo-controlled RCTs testing exclusively well-categorised SVF treatment would be mandatory to unequivocally confirm that SVF is an effective treatment for KOA. However, since the technique requires a previous liposuction procedure, a placebo-controlled trial with sham liposuction would be difficult to perform. However, comparing SVF to other approved treatments could also be a breakthrough.

Conclusion

With a low level of evidence, our systematic review suggests that SVF treatment could be a promising therapy for KOA in terms of pain, functionality and anatomical structure improvement. In addition, SVF has proved to be a safe treatment for KOA. However, SVF products need to be standardised to be compared and concomitant treatments should be avoided, as they can mask SVF's therapeutic effect. In addition, the number of cells in the SVF should be similar to allow the comparison of different treatments. Our results have highlighted the lack of prospective RCTs exclusively comparing a well-categorised SVF treatment with placebo or standard approved treatments to better understand the effect on KOA. Therefore, we consider that the most appropriate study to evaluate SVF for KOA would be a controlled RCT testing a well-defined, standardised and categorised SVF product in a large number of patients.

Declarations

Ethics approval and consent to participate Not applicable.

Consent for publication Not applicable.

Author contributions

Anna Boada-Pladellorens: Formal analysis; Funding acquisition; Investigation; Methodology; Project administration; Supervision; Writing – original draft; Writing – review & editing.

Mercè Avellanet: Formal analysis; Methodology; Validation; Writing – review & editing.

Esther Pages-Bolibar: Formal analysis; Supervision; Writing – review & editing.

Anna Veiga: Formal analysis; Supervision; Writing – review & editing.

Acknowledgements

The authors thank Matías Rey-Carrizo, PhD, at BCN Medical Writing for providing editorial support.

Funding

The authors disclosed receipt of the following financial support for the research, authorship, and/or publication of this article: A.B.-P. gratefully acknowledges the Government of the Principality of Andorra for the grant on Andorran issues (APTA0015-AND/2018). CIMERA S.L.U funded the medical writing services without participating in the review's design or data analysis.

Competing interests

The authors declared the following potential conflicts of interest with respect to the research, authorship, and/or publication of this article: A.B.-P. and M.A. have a service provision contract with Celular Clinic, a clinic owned by CIMERA S.L.U.

Availability of data and materials

Data are available from the corresponding author on reasonable request.

ORCID iD

Anna Boada-Pladellorens D https://orcid.org/ 0000-0003-4048-3310

Supplemental material

Supplemental material for this article is available online.

References

- 1. Murray CJL, Vos T, Lozano R, *et al.* Disabilityadjusted life years (DALYs) for 291 diseases and injuries in 21 regions, 1990-2010: a systematic analysis for the global burden of disease study 2010. *Lancet* 2012; 380: 2197–2223.
- Spitaels D, Mamouris P, Vaes B, et al. Epidemiology of knee osteoarthritis in general practice: a registry-based study. BMJ Open 2020; 10: 1–9, http://bmjopen.bmj.com/
- 3. Charlesworth J, Fitzpatrick J, Perera NKP, *et al.* Osteoarthritis – a systematic review of long-term safety implications for osteoarthritis of the knee. *BMC Musculoskelet Disord* 2019; 20: 1–12.
- Pintat J, Silvestre A, Magalon G, et al. Intraarticular injection of mesenchymal stem cells and platelet-rich plasma to treat patellofemoral osteoarthritis: preliminary results of a longterm pilot study. *J Vasc Interv Radiol* 2017; 28: 1708–1713.
- Quicke JG, Conaghan PG, Corp N, et al. Osteoarthritis year in review 2021: epidemiology & therapy. Osteoarthr Cartil 2022; 30: 196–206.
- 6. Turkiewicz A, Petersson IF, Björk J, *et al.* Current and future impact of osteoarthritis on health care: a population-based study with projections to year 2032. *Osteoarthr Cartil* 2014; 22: 1826–1832.
- Kompel AJ, Roemer FW, Murakami AM, et al. Intra-articular corticosteroid injections in the hip and knee: perhaps not as safe as we thought? *Radiology* 2019; 293: 1–8.
- Lin KY, Yang CC, Hsu CJ, et al. Intra-articular injection of platelet-rich plasma is superior to hyaluronic acid or saline solution in the treatment of mild to moderate knee osteoarthritis: a randomized, double-blind, triple-parallel, placebo-controlled clinical trial. Arthrosc – *J Arthrosc Relat Surg* 2019; 35: 106–117.
- Belk JW, Kraeutler MJ, Houck DA, et al. Platelet-rich plasma versus hyaluronic acid for knee osteoarthritis: a systematic review and metaanalysis of randomized controlled trials. Am J Sports Med 2021; 49: 249–260.
- Migliore A, Paoletta M, Moretti A, et al. The perspectives of intra-articular therapy in the management of osteoarthritis. *Expert Opin Drug Deliv* 2020; 17: 1213–1226.
- Canovas F and Dagneaux L. Quality of life after total knee arthroplasty. *Orthop Traumatol Surg Res* 2018; 104: S41–S416.
- 12. Dominici M, Le Blanc K, Mueller I, *et al.* Minimal criteria for defining multipotent mesenchymal stromal cells. The International

Society for Cellular Therapy position statement. *Cytotherapy* 2006; 8: 315–317.

- García-Bernal D, García-Arranz M, Yáñez RM, et al. The current status of mesenchymal stromal cells: controversies, unresolved issues and some promising solutions to improve their therapeutic efficacy. Front Cell Dev Biol 2021; 9: 650664.
- Wagner W, Wein F, Seckinger A, et al. Comparative characteristics of mesenchymal stem cells from human bone marrow, adipose tissue, and umbilical cord blood. *Exp Hematol* 2005; 33: 1402–1416.
- Biazzo A, D'Ambrosi R, Masia F, et al. Autologous adipose stem cell therapy for knee osteoarthritis: where are we now? *Phys Sportsmed* 2020; 48: 392–399.
- Andia I, Maffulli N and Burgos-Alonso N. Stromal vascular fraction technologies and clinical applications. *Expert Opin Biol Ther* 2019; 19: 1289–1305.
- Gimble JGF. Adipose-derived adult stem cells: isolation, characterization, and differentiation potential. *Cytotherapy* 2003; 5: 362–369.
- 18. Zuk PA, Zhu M, Mizuno H, *et al.* Multilineage cells from human adipose tissue: implications for cell-based therapies. *Tissue Eng* 2001; 7: 211–228.
- 19. Muthu S, Kartheek RR, Jeyaraman N, *et al.* Is culture expansion necessary in autologous mesenchymal stromal cell therapy to obtain superior results in the management of knee osteoarthritis ? Meta-analysis of randomized controlled trials. *Bioengineering* 2021; 8: 220.
- Agarwal N, Mak C, Bojanic C, et al. Metaanalysis of adipose tissue derived cell-based therapy for the treatment of knee osteoarthritis. *Cells* 2021; 10: 1–31.
- Ha CW, Park YB, Kim SH, et al. Intra-articular mesenchymal stem cells in osteoarthritis of the knee: a systematic review of clinical outcomes and evidence of cartilage repair. *Arthroscopy* 2019; 35: 277–288.e2.
- 22. Shanmugasundaram S, Vaish A, Chavada V, *et al.* Assessment of safety and efficacy of intraarticular injection of stromal vascular fraction for the treatment of knee osteoarthritis – a systematic review. *Int Orthop* 2021; 45: 615–625.
- 23. Maheshwer B, Polce EM, Paul K, *et al.* Regenerative potential of mesenchymal stem cells for the treatment of knee osteoarthritis and chondral defects: a systematic review and metaanalysis. *Arthroscopy* 2021; 37: 362–378.
- 24. Jevotovsky DS, Alfonso AR, Einhorn TA, *et al.* Osteoarthritis and stem cell therapy in humans:

Volume 14

a systematic review. *Osteoarthr Cartil* 2018; 26: 711–729.

- 25. Gentile P, Sterodimas A, Pizzicannella J, *et al.* Systematic review: allogenic use of stromal vascular fraction (SVF) and decellularized extracellular matrices (ECM) as advanced therapy medicinal products (ATMP) in tissue regeneration. *Int J Mol Sci* 2020; 21: 1–14.
- Hurley ET, Yasui Y, Gianakos AL, et al. Limited evidence for adipose-derived stem cell therapy on the treatment of osteoarthritis. *Knee Surg Sport Traumatol Arthrosc* 2018; 26: 3499–3507.
- Ranmuthu CDS, Ranmuthu CKI and Khan WS. Evaluating the current literature on treatments containing adipose-derived stem cells for osteoarthritis: a progress update. *Curr Rheumatol Rep* 2018; 20.
- 28. Kader N, Asopa V, Baryeh K, *et al.* Cell-based therapy in soft tissue sports injuries of the knee: a systematic review. *Expert Opin Biol Ther* 2021; 21: 1035–1047.
- Salaffi F, Stancati A, Silvestri CA, et al. Minimal clinically important changes in chronic musculoskeletal pain intensity measured on a numerical rating scale. Eur J Pain 2004; 8: 283–291.
- Yokota N, Hattori M, Ohtsuru T, et al. Comparative clinical outcomes after intraarticular injection with adipose-derived cultured stem cells or noncultured stromal vascular fraction for the treatment of knee osteoarthritis. *Am J Sports Med* 2019; 47: 2577–2583.
- Hong Z, Chen J, Zhang S, *et al.* Intra-articular injection of autologous adipose-derived stromal vascular fractions for knee osteoarthritis: a double-blind randomized self-controlled trial. *Int Orthop* 2019; 43: 1123–1134.
- Garza JR, Campbell RE, Tjoumakaris FP, et al. Clinical efficacy of intra-articular mesenchymal stromal cells for the treatment of knee osteoarthritis clinical trial. Am J Sports Med 2020; 48: 588–598.
- 33. Fodor PB and Paulseth SG. Adipose derived stromal cell (ADSC) injections for pain management of osteoarthritis in the human knee joint. *Aesthet Surg J* 2016; 36: 229–236.
- Tsubosaka M, Matsumoto T, Sobajima S, et al. The influence of adipose-derived stromal vascular fraction cells on the treatment of knee osteoarthritis. BMC Musculoskelet Disord 2020; 21: 207.
- 35. Gibbs N, Diamond R, Sekyere EO, et al. Management of knee osteoarthritis by combined stromal vascular fraction cell therapy, platelet-rich

plasma, and musculoskeletal exercises: a case series. \Im *Pain Res* 2015; 8: 799–806.

- Simunec D, Salari H and Meyer J. Treatment of grade 3 and 4 osteoarthritis with intraoperatively separated adipose tissue-derived stromal vascular fraction: a comparative case series. *Cells* 2020; 9: 2096.
- Nguyen PD, Tran TD, Nguyen HT, et al. Comparative clinical observation of arthroscopicmicrofracture in the presence and absence of astromal vascular fraction injection for osteoarthritis. *Stem Cells Transl Med* 2017; 6: 187–195.
- Tran TDX, Wu C-M, Dubey NK, et al. Timeand Kellgren–Lawrence grade-dependent changes in intra-articularly transplanted stromal vascular fraction in osteoarthritic patients. *Cells* 2019; 8: 308.
- 39. Di Matteo B, Vandenbulcke F, Vitale ND, *et al.* Minimally manipulated mesenchymal stem cells for the treatment of knee osteoarthritis: a systematic review of clinical evidence. *Stem Cells Int* 2019; 2019: 1735242.
- Biazzo A, D'Ambrosi R, Masia F, et al. Autologous adipose stem cell therapy for knee osteoarthritis: where are we now? *Phys Sportsmed* 2020; 48: 392–399.
- Zhou W, Lin J, Zhao K, *et al.* Single-cell profiles and clinically useful properties of human mesenchymal stem cells of adipose and bone marrow origin. *Am J Sports Med* 2019; 47: 1722–1733.
- Hervás-Salcedo R, Fernández-García M, Hernando-Rodríguez M, et al. Enhanced antiinflammatory effects of mesenchymal stromal cells mediated by the transient ectopic expression of CXCR4 and IL10. Stem Cell Res Ther 2021; 12: 1–20.
- 43. Caplan AI. Mesenchymal stem cells: time to change the name! *Stem Cells Transl Med* 2017; 6: 1445–1451.
- 44. Wu X, Jiang J, Gu Z, *et al.* Mesenchymal stromal cell therapies: immunomodulatory properties and clinical progress. *Stem Cell Res Ther* 2020; 11: 1–16.
- Caplan AI. Cell-based therapies: the nonresponder. *Stem Cells Transl Med* 2018; 7: 762–766.
- Murphy MB, Moncivais K and Caplan AI. Mesenchymal stem cells: environmentally responsive therapeutics for regenerative medicine. *Exp Mol Med* 2013; 45: e54–e56.
- 47. Koh YG, Kwon OR, Kim YS, *et al.* Adiposederived mesenchymal stem cells with microfracture versus microfracture alone: 2-year

follow-up of a prospective randomized trial. *Arthroscopy* 2016; 32: 97–109.

- Koh YG, Choi YJ, Kwon SK, *et al.* Clinical results and second-look arthroscopic findings after treatment with adipose-derived stem cells for knee osteoarthritis. *Knee Surg Sport Tr A* 2015; 23: 1308–1316.
- Kim SH, Djaja YP, Park YB, *et al.* Intra-articular injection of culture-expanded mesenchymal stem cells without adjuvant surgery in knee osteoarthritis: a systematic review and metaanalysis. *Am J Sports Med* 2020; 48: 2839–2849.
- Roato I, Belisario DC, Compagno M, et al. Concentrated adipose tissue infusion for the treatment of knee osteoarthritis: clinical and histological observations. Int Orthop 2019; 43: 15–23.
- 51. Pak J, Lee JH, Park KS, *et al.* Regeneration of cartilage in human knee osteoarthritis with autologous adipose tissue-derived stem cells and autologous extracellular matrix. *Biores Open Access* 2016; 5: 192–200.
- Hudetz D, Bori I, Rod E, *et al.* Early results of intra-articular micro-fragmented lipoaspirate treatment in patients with late stages knee osteoarthritis: a prospective study. *Croat Med J* 2019; 60: 227–236.
- Jo CH, Lee YG, Shin WH, *et al.* Intra-articular injection of mesenchymal stem cells for the treatment of osteoarthritis of the knee: a proof-of-concept clinical trial. *Stem Cells* 2014; 32: 1254–1266.
- 54. Freitag J, Bates D, Wickham J, *et al.* Adiposederived mesenchymal stem cell therapy in the treatment of knee osteoarthritis: a randomized controlled trial. *Regen Med* 2019; 14: 213–230.
- 55. Amin S, LaValley MP, Guermazi A, et al. The relationship between cartilage loss on magnetic resonance imaging and radiographic progression in men and women with knee osteoarthritis. *Arthritis Rheum* 2005; 52: 3152–3159.
- 56. Ding C, Cicuttini F, Scott F, *et al.* Natural history of knee cartilage defects and factors affecting change, https://jamanetwork.com/
- 57. Cibere J, Sayre EC, Guermazi A, *et al.* Natural history of cartilage damage and osteoarthritis progression on magnetic resonance imaging in a population-based cohort with knee pain. *Osteoarthr Cartil* 2011; 19: 683–688.
- Iolascon G, Gimigliano F, Moretti A, et al. Early osteoarthritis: how to define, diagnose, and manage. A systematic review. Eur Geriatr Med 2017; 8: 383–396.

- 59. Biswal S, Hastie T, Andriacchi TP, *et al.* Risk factors for progressive cartilage loss in the knee: a longitudinal magnetic resonance imaging study in forty-three patients. *Arthritis Rheum* 2002; 46: 2884–2892.
- 60. Aronowitz JA, Lockhart RA and Hakakian CS. Mechanical versus enzymatic isolation of stromal vascular fraction cells from adipose tissue. *Springerplus* 2015; 4: 713–719.
- 61. Rodriguez J, Pratta AS, Abbassi N, *et al.* Evaluation of three devices for the isolation of the stromal vascular fraction from adipose tissue and for ASC culture: a comparative study. *Stem Cells Int* 2017; 2017: 9289213.
- 62. Satué M, Schüler C, Ginner N, *et al.* Intraarticularly injected mesenchymal stem cells promote cartilage regeneration, but do not permanently engraft in distant organs. *Sci Rep* 2019; 9: 1–10.
- Grimes DA, Hubacher D, Nanda K, *et al.* The good clinical practice guideline: a bronze standard for clinical research. *Lancet* 2005; 366: 172–174.

Appendix 1

Search strategy

Database searched:

MEDLINE (PUBMED): 1960 TO MAY 2021 Date of search: 3 May 2021

Date range searched: January 2011 to April 2021

Search strategy:

- 1. (exp. knee osteoarthritis/) OR (exp. osteoarthritis/)
- 2. (exp. cartilage/)
- (exp. adipose-derived stem cells/) OR (exp. adipose-derived stromal cells/)
- 4. (exp. stromal vascular fraction/)
- 5. (exp. mesenchymal stem cells/) OR (exp. mesenchymal stromal cells/)
- 6. ((#1) OR (#2)) AND ((#3) OR (#4) OR (#5))
- 7. 'last 10 years' [dp]
- 8. (#6) AND (#7)
- 9. humans [mh]
- 10.(#9) AND (#10)