



Draft Genome Sequence of NDM-Encoding *Klebsiella pneumoniae* Isolated from Feral Swine

Huijia Liu,^{a,b} Yuting Zhai,^{a,b} Ting Liu,^{a,b} Peixin Fan,^{a,b} Raoul Boughton,^c  Kwangcheol C. Jeong^{a,b}

^aEmerging Pathogens Institute, University of Florida, Gainesville, Florida, USA

^bDepartment of Animal Sciences, College of Agricultural and Life Sciences, University of Florida, Gainesville, Florida, USA

^cThe Mosaic Company, Lithia, Florida, USA

ABSTRACT New Delhi metallo- β -lactamase (NDM)-producing *Enterobacteriaceae* pose a great threat to public health globally. Most known NDM-producing *Enterobacteriaceae* are associated with human hospital or community infections. Here, we report the draft genome sequence of an NDM-1-encoding *Klebsiella pneumoniae* strain isolated from feral swine (*Sus scrofa*) captured in Florida, USA.

New Delhi metallo- β -lactamase-1 (NDM-1) is a transferable molecular class B (zinc metallo-) β -lactamase which hydrolyzes beta-lactam antibiotics, including all penicillins, cephalosporins, and carbapenems (1). NDM-1 was first identified in *K. pneumoniae* and *E. coli* strains, from a Swedish patient with a hospitalization history in India in 2008 (2), and has been isolated from various hosts, including human, livestock, and companion animals (3, 4). As part of a large collaborative project on feral swine ecology, we collected 393 fecal samples from feral swine captured on Archbold's Buck Island Ranch (27°09'N, 81°11'W). All activities associated with the trapping and sampling of feral swine occurred under Institutional Care and Use Committee-approved protocols (201408495, 201808495). Fecal samples (0.1 g) were incubated in tryptic soy broth containing meropenem (16 μ g/ml) overnight at 37°C; then, resistant bacteria were selected on MacConkey agar containing meropenem (16 μ g/ml). Meropenem-resistant isolates were subjected to PCR after boiling lysis to detect the *bla*_{NDM} gene using primer pairs (forward: 5'-GGTTTGGCGATCTGGTTTC-3'; reverse: 5'-CGGAATGGCTCATCAGATC-3') as described previously (5). One strain (KCJ2K2161) was identified as NDM positive and applied for whole-genome sequencing.

KCJ2K2161 was cultured overnight at 37°C in tryptic soy broth containing meropenem (16 μ g/ml). Genomic DNA was extracted using the DNeasy blood and tissue kit (Qiagen, Valencia, CA), and the DNA concentration was measured using a Qubit 3 fluorometer (Invitrogen, Waltham, MA). To construct a DNA library, the DNA sample was diluted and quantified to 0.2 to 0.4 ng/ μ l. The DNA library was prepared using the Nextera XT sample preparation kit following the manufacturer's instructions (Illumina, San Diego, CA). Genome sequencing was performed using the Illumina MiSeq platform with a 2 \times 250-bp, 500-cycle cartridge (Illumina). The total number of reads and coverage of KCJ2K2161 were 1,581,846 and 74 \times , respectively. Sickle v1.33.2 (6) was used for adaptive trimming of the FASTQ raw sequencing data, with the quality and length thresholds set to 30 and 50 bp, respectively. Genome assembly was performed using SPAdes v3.12.0 (7). In the SPAdes genome assembler, the K-mer used was set as 21, 33, 55, 77, 99, 127, and the coverage cutoff was set as auto. Contigs that are less than 200 bp were removed. QUAST v5.0.2 (8) was used to evaluate the genome assembly quality. The assembled genome size of KCJ2K2161 was 5,340,476 bp, containing a total of 278 contigs, with a GC content of 57.09% and an *N*₅₀ value of 107,917 bp.

KCJ2K2161 was identified as *Klebsiella pneumoniae* sequencing type 1967 using SpeciesFinder v2.0 and MLST v2.0 (9, 10). The genome sequence of *K. pneumoniae* KCJ2K2161

Citation Liu H, Zhai Y, Liu T, Fan P, Boughton R, Jeong KC. 2021. Draft genome sequence of NDM-encoding *Klebsiella pneumoniae* isolated from feral swine. Microbiol Resour Announc 10: e00808-21. <https://doi.org/10.1128/MRA.00808-21>.

Editor David Rasko, University of Maryland School of Medicine

Copyright © 2021 Liu et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to Kwangcheol C. Jeong, kcjeong@ufl.edu.

Received 11 August 2021

Accepted 26 September 2021

Published 14 October 2021

was annotated using the NCBI Prokaryotic Genome Annotation Pipeline v5.2. A total of 5,324 coding sequences, 84 tRNAs, 6 complete rRNAs, 11 noncoding RNAs, and 141 pseudogenes were identified. Using Resistance Gene Identifier v5.2.0 within the Comprehensive Antibiotic Resistance Database v3.1.3 (11), a total of 38 antibiotic-resistant genes were identified, including four beta-lactamase genes (*bla*_{TEM-1}, *bla*_{NDM-1}, *bla*_{SHV-26}, and *bla*_{AmpH}). A total of 14 virulence factors related to adherence, invasion, and iron uptake were detected using the PathoSystems Resource Integration Center v3.6.9 (12). Default parameters were used for all software unless otherwise specified.

Data availability. This whole-genome shotgun project has been deposited in DDBJ/ENA/GenBank under the accession number [JAHWDE000000000](https://www.ncbi.nlm.nih.gov/nuclseq/JAHWDE000000000). The version described in this paper is version [JAHWDE010000000](https://www.ncbi.nlm.nih.gov/nuclseq/JAHWDE010000000). The reads are available through the NCBI Sequence Read Archive under accession number [SRR15327870](https://www.ncbi.nlm.nih.gov/sra/SRR15327870).

ACKNOWLEDGMENTS

Financial support for the research was in part supplied by the USDA-APHIS National Feral Swine Damage Management Program through a cooperative agreement between the U.S. Department of Agriculture Animal and Plant Health Inspection Service (APHIS) and R.B.

We thank Bethany Wight, Wesley Anderson, and Tyler Buckley for their dedicated efforts in trapping and sampling feral swine at ABIR.

REFERENCES

1. Sun Z, Hu L, Sankaran B, Prasad BVV, Palzkill T. 2018. Differential active site requirements for NDM-1 beta-lactamase hydrolysis of carbapenem versus penicillin and cephalosporin antibiotics. *Nat Commun* 9:4524. <https://doi.org/10.1038/s41467-018-06839-1>.
2. Yong D, Toleman MA, Giske CG, Cho HS, Sundman K, Lee K, Walsh TR. 2009. Characterization of a new metallo-beta-lactamase gene, *bla*_{NDM-1}, and a novel erythromycin esterase gene carried on a unique genetic structure in *Klebsiella pneumoniae* sequence type 14 from India. *Antimicrob Agents Chemother* 53:5046–5054. <https://doi.org/10.1128/AAC.00774-09>.
3. Moellering RC, Jr. 2010. NDM-1—a cause for worldwide concern. *N Engl J Med* 363:2377–2379. <https://doi.org/10.1056/NEJMp1011715>.
4. Fischer J, Schmoger S, Jahn S, Helmuth R, Guerra B. 2013. NDM-1 carbapenemase-producing *Salmonella enterica* subsp. *enterica* serovar Corvallis isolated from a wild bird in Germany. *J Antimicrob Chemother* 68:2954–2956. <https://doi.org/10.1093/jac/dkt260>.
5. Poirel L, Revathi G, Bernabeu S, Nordmann P. 2011. Detection of NDM-1-producing *Klebsiella pneumoniae* in Kenya. *Antimicrob Agents Chemother* 55:934–936. <https://doi.org/10.1128/AAC.01247-10>.
6. Joshi NA, Fass JN. 2011. Sickle: a sliding-window, adaptive, quality-based trimming tool for FastQ files (version 1.33). <https://github.com/najoshi/sickle>.
7. Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. *J Comput Biol* 19:455–477. <https://doi.org/10.1089/cmb.2012.0021>.
8. Gurevich A, Saveliev V, Vyahhi N, Tesler G. 2013. QUAST: quality assessment tool for genome assemblies. *Bioinformatics* 29:1072–1075. <https://doi.org/10.1093/bioinformatics/btt086>.
9. Larsen MV, Cosentino S, Lukjancenko O, Saputra D, Rasmussen S, Hasman H, Sicheritz-Ponten T, Aarestrup FM, Ussery DW, Lund O. 2014. Benchmarking of methods for genomic taxonomy. *J Clin Microbiol* 52:1529–1539. <https://doi.org/10.1128/JCM.02981-13>.
10. Maiden MCJ. 2006. Multilocus sequence typing of bacteria. *Annu Rev Microbiol* 60:561–588. <https://doi.org/10.1146/annurev.micro.59.030804.121325>.
11. Alcock BP, Raphenya AR, Lau TTY, Tsang KK, Bouchard M, Edalatmand A, Huynh W, Nguyen A-LV, Cheng AA, Liu S, Min SY, Miroschnichenko A, Tran H-K, Werfalli RE, Nasir JA, Oloni M, Speicher DJ, Florescu A, Singh B, Faltyn M, Hernandez-Koutoucheva A, Sharma AN, Bordeleau E, Pawlowski AC, Zubyk HL, Dooley D, Griffiths E, Maguire F, Winsor GL, Beiko RG, Brinkman FSL, Hsiao WWL, Domselaar GV, McArthur AG. 2020. CARD 2020: antibiotic resistance surveillance with the Comprehensive Antibiotic Resistance Database. *Nucleic Acids Res* 48:D517–D525. <https://doi.org/10.1093/nar/gkz935>.
12. Davis JJ, Wattam AR, Aziz RK, Brettin T, Butler R, Butler RM, Chlenski P, Conrad N, Dickerman A, Dietrich EM, Gabbard JL, Gerdes S, Guard A, Kenyon RW, Machi D, Mao C, Murphy-Olson D, Nguyen M, Nordberg EK, Olsen GJ, Olson RD, Overbeek JC, Overbeek R, Parrello B, Pusch GD, Shukla M, Thomas C, VanOeffelen M, Vonstein V, Warren AS, Xia F, Xie D, Yoo H, Stevens R. 2020. The PATRIC Bioinformatics Resource Center: expanding data and analysis capabilities. *Nucleic Acids Res* 48:D606–D612. <https://doi.org/10.1093/nar/gkz943>.