Optimization of Paclitaxel Containing pH-Sensitive Liposomes By 3 Factor, 3 Level Box-Behnken Design

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Rane and Prabhakar: Optimization of pH-Sensitive Liposomes of Paclitaxel

The aim of this study was to investigate the combined influence of 3 independent variables in the preparation of paclitaxel containing pH-sensitive liposomes. A 3 factor, 3 levels Box-Behnken design was used to derive a second order polynomial equation and construct contour plots to predict responses. The independent variables selected were molar ratio phosphatidylcholine:diolylphosphatidylethanolamine (X_1) , molar concentration of cholesterylhemisuccinate (X_2) , and amount of drug (X_3) . Fifteen batches were prepared by thin film hydration method and evaluated for percent drug entrapment, vesicle size, and pH sensitivity. The transformed values of the independent variables and the percent drug entrapment were subjected to multiple regression to establish full model second order polynomial equation. F was calculated to confirm the omission of insignificant terms from the full model equation to derive a reduced model polynomial equation to predict the dependent variables. Contour plots were constructed to show the effects of X_1 , X_2 , and X_3 on the percent drug entrapment. A model was validated for accurate prediction of the percent drug entrapment by performing checkpoint analysis. The computer optimization process and contour plots predicted the levels of independent variables X_1 , X_2 , and X_3 (0.99, -0.06, 0, respectively), for maximized response of percent drug entrapment with constraints on vesicle size and pH sensitivity.

Key words: Box-Behnken design, pH-sensitive, liposomes, optimization, paclitaxel

Paclitaxel, the first of a new class of microtubule stabilizing agents, has been hailed by National Cancer Institute as the most significant advance in chemotherapy during the past 20-25 years[1]. Paclitaxel is poorly soluble in aqueous medium, and is currently formulated in a vehicle composed of 1:1 blend of cremophor EL and ethanol which is diluted with 5-20 fold in normal saline or dextrose solution (5%) for administration^[2,3]. One of the substantial problems associated with this formulation is that the ethanol:cremophor vehicle is toxic^[4-7]. The primary goal of formulation development for paclitaxel is to eliminate the cremophor vehicle and also to provide the possibility of improving the efficacy of paclitaxel based anticancer therapy. One of the major obstacles in designing the formulation of novel drugs is their limited aqueous solubility. This problem can be overcome by entrapping the drug in vesicular structures like liposomes. Liposomes that can be triggered to release their contents or fuse in

response to pH stimuli are of particular interest in cancer therapy as they can potentially respond to acidic environments *in vivo*^[8].

However, successful development of these systems requires careful consideration of a number of factors influencing the performance of the formulation, including the physicochemical properties of the raw materials (both drug and excipients), the composition and the component's relative amounts in the formulations, as well as the manufacturing process parameters. Many experiments fail in their purpose because they are not properly thought out and designed, and even the best data analysis cannot compensate for lack of planning. Experimental design is thus the preferred strategy, especially when complex formulations, such as liposomes, are to be developed^[9]. In particular, the multi-varied strategy of experimental design allows simultaneous investigation of the effects of several variables, as well as their actual significance on the considered response and possible interrelationship among them, giving the maximum information

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with minimum number of experiments[10,11]. Traditional experiments require more effort, time, and materials when a complex formulation needs to be developed. Various experimental designs[12-15] are useful in developing a formulation requiring less experimentation and providing estimates of the relative significance of different variables. In the work reported here, Box-Behnken design was used to optimize pH-sensitive liposomes containing paclitaxel. Independent variables selected were molar ratio of phosphatidylcholine: diolylphosphatidylethanolamine (PC:DOPE) (X₁), moles of cholesterylhemisuccinate (CHEMS) (X,), and the amount of drug (X₂) to evaluate their separate and combined effects on percent drug entrapment (PDE), pH-sensitivity, and vesicle size expressed as the average vesicle perimeter (AVP). Liposomes are of interest from a technical viewpoint and allow a wider scope to study the influence of various formulation variables, hence liposomes need to be optimized for desired response. In the present study thin film hydration method was used for the preparation and optimization of paclitaxel liposomes as this method is simple and capable of producing small vesicles.

MATERIALS AND METHODS

Paclitaxel was received as a gift sample from Naprod life sciences, Mumbai. Phosphatidylcholine (PC) was received as a gift sample from Phospholipid GmbH, Nattermannalle, Germany. Cholesteryl hemisuccinate (CHEMS) was received as a generous gift from Merk Eprova AG Switzerland. Dioleoyl phosphatidylethanolamime (DOPE) was received as a gift sample from Lipoid GmbH, Germany. HPLC grade solvents and other chemicals were purchased from local supplier.

Box-Behnken experimental design:

A Box-Behnken optimization design with three variables was applied to find the optimum conditions and to analyze how sensitive the responses were to variations in the settings of the experimental variables^[16]. This design is a factorial design with three levels, using middle points instead of corner points and is useful for estimating the coefficients in a second degree polynomial. A total of 15 experiments were performed including triplicates of the center point. The center points improve the assessment of the response surface curvature and

simplify the estimation of the model error. The traditional approach to develop a formulation is to change one variable at a time. It is difficult to develop an optimized formulation, as the method reveals nothing about the interactions among the variables. Hence, a Box–Behnken statistical design with three factors, three levels was selected for the optimization study. The dependent and independent variables are listed in Table 1. The polynomial equation generated by this experimental design (Instat + software) is as follows:

$$\begin{aligned} \mathbf{Y}_{i} &= \mathbf{b}_{0} + \mathbf{b}_{1} \mathbf{X}_{1} + \mathbf{b}_{2} \mathbf{X}_{2} + \mathbf{b}_{3} \mathbf{X}_{3} + \mathbf{b}_{12} \mathbf{X}_{1} \mathbf{X}_{2} + \mathbf{b}_{13} \mathbf{X}_{1} \mathbf{X}_{3} + \mathbf{b}_{23} \mathbf{X}_{2} \mathbf{X}_{3} \\ &+ \mathbf{b}_{11} \mathbf{X}_{1}^{2} + \mathbf{b}_{22} \mathbf{X}_{2}^{2} + \mathbf{b}_{33} \mathbf{X}_{3}^{2} \end{aligned} \tag{1},$$

Where Y_i is the dependent variable; b_0 is the intercept; b_1 to b_{33} are regression coefficients: and X_1 , X_2 and X_3 are the independent variable selected from the preliminary experiments.

Preparation of liposomes:

Liposomes were prepared by the thin film hydration method. Paclitaxel and the required quantities of CHEMS, Phospholipon 90G, and DOPE were dissolved in chloroform. All the batches were prepared according to the experimental design in Table 2. Chloroform was evaporated using rotary vacuum evaporator and kept overnight under vacuum. Nitrogen gas was passed over the thin film. Then it was hydrated with 5% dextrose solution, above the phase transition temperature of lipids, using glass beads. The suspension of liposomes was sonicated to reduce the size of liposomes. This was transferred to vials and stored at $4^{\circ[17]}$.

Microscopy:

The liposomes were mounted on glass slides and viewed under a microscope (Motic) for morphological observation after suitable dilution. Particle size was measured as average object perimeter.

TABLE 1: VARIABLES AND THEIR LEVELS IN BOXBEHNKEN DESIGN

| | Levels | | |
|---|--------|--------|--------|
| | Low | Medium | High |
| Independent variables | | | |
| X,=Molar ratio of PC: DOPE | 8:2 | 6:4 | 4:6 |
| X,=Molar concentration of CHEMS | 2 | 4 | 6 |
| X,=Amount of drug | 2.8 mg | 5.6 mg | 8.4 mg |
| Transformed values | -1 | 0 | 1 |
| Dependent variables | | | |
| Y ₁ =Percent drug entrapment | | | |
| Y ₂ =Vesicle size | | | |
| Y¸=pH-sensitivity | | | |

 ${\tt DOPE=Dioleoyl\ phosphatidylethanolamime}$

Percent drug entrapment:

The amount of paclitaxel incorporated in liposomes was determined using HPLC (Perkin Elmer). Five hundred microliter of liposomal suspension was diluted with water and acetonitrile to 1 ml. Extraction of paclitaxel was accomplished by adding 4 ml of tert-butylmethyl ether, vortex mixing for 1 min, and centrifuging the mixture for 15 min. Three milliliter of the organic layer was separated and evaporated to dryness. Residue was reconstituted with 1 ml methanol. Twenty microliter of the above solution was injected into a C₁₈ column, 5 μm. The column was eluted with acetonitrile/water (60/40). The drug was estimated by UV absorption measurement at 227 nm (flow rate 1 ml/min)^[18,19].

In vitro release from pH-sensitive liposomes:

Liposomal formulations were diluted with phosphate buffer pH 5 in 1:2 ratios, and incubated at 37° for 15 min. The drug released was separated from liposomal paclitaxel, extracted, and quantified using the same procedure described above for the determination of incorporated drug.

Checkpoint analysis:

A checkpoint analysis was performed to confirm the role of the derived polynomial equation and contour plots in predicting the responses. Values of independent variables were taken at three points, one from each contour plot, and the theoretical values of PDE were calculated by substituting the values in the polynomial equation. Liposomes were prepared experimentally at three checkpoints, and evaluated for the responses.

Optimum formula:

After developing the polynomial equations for the responses PDE, AVP, and pH sensitivity with the independent variables, the formulation was optimized for the response PDE. Optimization was performed to find out the level of independent variables $(X_1, X_2, \text{ and } X_3)$ that would yield a maximum value of PDE with constraints on AVP and pH sensitivity.

RESULTS AND DISCUSSION

Liposomes were observed under a microscope to examine their morphology and were observed to be mostly spherical, with a few being slightly elongated. A Box-Behnken experimental design with three independent variables at three different levels was

used to study the effects on dependent variables. All the batches of liposomes within the experimental design were evaluated for PDE, pH sensitivity, and vesicle size. A Box–Behnken experimental design has the advantage of requiring fewer experiments (15 batches) than would a full factorial design (27 batches). Transformed values of all the batches along with their results are shown in Table 2. Formulations 7, 12, 13, 14, and 15 had the highest PDE (>90%). Table 3 shows the observed and predicted values with residuals and percent error of responses for all the batches.

The PDE (dependent variable) obtained at various levels of the three independent variables $(X_1, X_2, \text{ and } X_3)$ was

TABLE 2: BOX-BEHNKEN EXPERIMENTAL DESIGN WITH MEASURED RESPONSES

| Batch no. | X ₁ | X ₂ | X ₃ | Y ₁ (PDE±SD)* | Y ₂ (µm) | Y ₃ (%) |
|-----------|----------------|----------------|----------------|--------------------------|---------------------|--------------------|
| 1 | 0 | -1 | -1 | 84.1±0.68 | 6.4 | 70.69 |
| 2 | 0 | -1 | 1 | 80.31±1.17 | 6.9 | 73.27 |
| 3 | 0 | 1 | -1 | 82.26±1.54 | 2.6 | 81.00 |
| 4 | 0 | 1 | 1 | 79.7±1.23 | 2.7 | 79.56 |
| 5 | -1 | 0 | -1 | 85.52±1.92 | 6.2 | 60.43 |
| 6 | -1 | 0 | 1 | 78.22±0.86 | 3.8 | 65.30 |
| 7 | 1 | 0 | -1 | 92±1.41 | 4.4 | 83.12 |
| 8 | 1 | 0 | 1 | 89.64±1.52 | 4 | 85.11 |
| 9 | -1 | -1 | 0 | 84.04±1.87 | 6.3 | 66.91 |
| 10 | -1 | 1 | 0 | 80.62±2.12 | 3.3 | 75.14 |
| 11 | 1 | -1 | 0 | 94.68±0.98 | 6.5 | 79.87 |
| 12 | 1 | 1 | 0 | 88.73±1.28 | 2.4 | 94.77 |
| 13 | 0 | 0 | 0 | 90.37±1.04 | 4 | 86.07 |
| 14 | 0 | 0 | 0 | 91.14±1.59 | 3.8 | 88.90 |
| 15 | 0 | 0 | 0 | 92.53±1.33 | 4.1 | 85.18 |

PDE=Percent drug entrapment, n=3, Y2=average vesicular size in μ m, Y3=percent drug release

TABLE 3: OBSERVED AND PREDICTED VALUES WITH RESIDUALS OF THE RESPONSE $\mathbf{Y}_1(\text{PDE})^*$

| Batch no. | Observed PDE | Predicted PDE | Residuals | %Error |
|-----------|--------------|---------------|-----------|--------|
| 1 | 84.1 | 85.169 | -1.069 | 1.27 |
| 2 | 80.31 | 81.167 | -0.857 | 1.07 |
| 3 | 82.26 | 82.213 | 0.047 | 0.06 |
| 4 | 79.7 | 78.211 | 1.489 | 1.87 |
| 5 | 85.52 | 83.667 | 1.853 | 2.17 |
| 6 | 78.22 | 79.665 | -1.445 | 1.85 |
| 7 | 92 | 92.829 | -0.829 | 0.90 |
| 8 | 89.64 | 88.827 | 0.813 | 0.91 |
| 9 | 84.04 | 83.817 | 0.223 | 0.27 |
| 10 | 80.62 | 80.861 | -0.241 | 0.30 |
| 11 | 94.68 | 92.979 | 1.701 | 1.80 |
| 12 | 88.73 | 90.023 | -1.293 | 1.46 |
| 13 | 90.37 | 91.477 | -1.107 | 1.22 |
| 14 | 91.14 | 91.477 | -0.337 | 0.37 |
| 15 | 92.53 | 91.477 | 1.053 | 1.14 |
| | | | | |

PDE=Percent drug entrapment

subjected to multiple regression to yield a second-order polynomial equation (full model):

PDE =
$$91.347+4.581X_1-1.478X_2-2.001X_3-0.6325X_1$$

 $X_2+1.235X_1X_3+0.03075X_2X_3+0.2117X_1^2-$
 $4.541X_2^2-5.213X_3^2$ (2)

The value of the correlation coefficient (r2) of Eqn. 2 was found to be 0.9755, indicating good fit. The PDE values measured for the different batches showed wide variation (i.e., values ranged from a minimum of 79.7 to a maximum of 94.68). The results clearly indicate that the PDE value is strongly affected by the variables selected for the study. This is also reflected by the wide range of values for coefficients of the terms of Eqn 2. The main effects of X_1 , X_2 , and X_3 represent the average result of changing one variable at a time from its low level to its high level. The interaction terms $(X_1X_2, X_1X_3, X_2X_3, X_1^2, X_2^2, \text{ and } X_3^2)$ show how the PDE changes when two variables are simultaneously changed. Positive coefficient of X₁ (molar ratio of PL 90 and DOPE) indicate favorable effect on PDE, while the negative coefficient of X₂ and X₃ indicate unfavorable effect on PDE. Among the three independent variables the lowest coefficient value is for X_2 ($b_2 = -1.478$) indicating that this variable is not significant in predicting PDE.

The standardized effect of the independent variables and their interaction on the dependent variable was investigated by preparing a pareto chart (fig. 1) which depicts the main effect of the independent variables

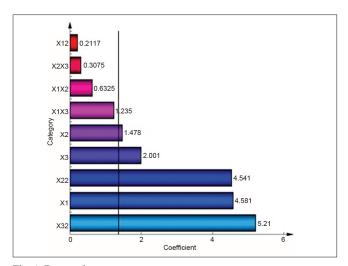


Fig. 1: Pareto chart.

Pareto chart showing the standardized effect of independent variables and their interaction on the percent drug entrapment of liposomes.

and interactions with their relative significance on the PDE. The length of each bar in the chart indicates the standardized effect of that factor on the response. The fact that the bar for X_1X_2 , X_1X_3 , X_2X_3 , and X_1^2 remains inside the reference line in fig. 1 and the small coefficients for these terms in Eqn. 2 indicate that these terms contribute the least in prediction of PDE. Hence, these terms are omitted from the full model to obtain a reduced second-order polynomial equation (equation 3) by multiple regressions of the PDE and the significant terms (P<0.05) of Eqn. 2:

PDE =
$$91.477+4.581X_1-1.478X_2-2.001X_3-4.557X_2^2-5.230X_3^2$$
 (3)

To confirm the omission of nonsignificant terms, F statistic was calculated after applying analysis of variance for the full model and the reduced model (Table 4). The F calculated value (1.04) is less than the tabled value of F (4.95) at a 0.05 confidence interval. Hence, it is concluded that the omitted terms do not significantly contribute to predicting

TABLE 4: RESULTS OF ANOVA OF FULL AND REDUCED MODELS FOR PDE OF PACLITAXEL IN LIPOSOMES

| ANOVA | Df | SS | MS | F value | P value |
|------------|----|--------------------------|--------------------------|---------|---------|
| Regression | | | | | |
| Α | 9 | 392.575 | 43.619 | 22.16 | 0.0016 |
| В | 5 | 384.33 | 76.866 | 38.25 | 0.0000 |
| Residuals | | | | | |
| Α | 5 | 9.842 (C ₁) | 1.9685 (D ₁) | | |
| В | 9 | 18.087 (C ₂) | 2.0097 | | |

ANOVA indicates analysis of variance; PDE, percent drug entrapment; A, full model; B, reduced model; Df, degrees of freedom; SS, sum of squares; MS, mean of squares; F, Fischers ratio. FCAL = [(C2-C1)/NTO]/D1 = 1.040, where NTO is the number of terms omitted (having a *P* value more than 0.05)

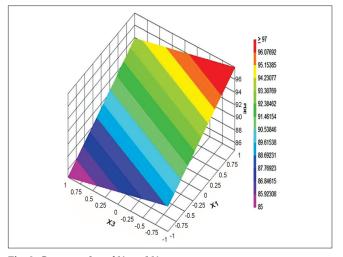


Fig. 2: Contour plot of X_1 and X_3 . Contour plot showing the effect of molar ratio of PC: DOPE (X_1) , and amount of drug added (X_3) on the percent drug entrapment of liposomes.

the PDE. This implies that the main effect of the amount of drug and the molar ratio of phospholipids is significant, as is evident from their high coefficients and the fact that the bars corresponding to variables X_1 , X_2 , X_3 , X_2^2 , X_3^2 extend beyond the reference line in fig. 1.

Vesicle size of liposome batches, measured by using microscopy (Motic), was found to be in the range of 2.4 to 6.9 μ m. A polynomial equation was also developed for AVP which is given as

$$\begin{array}{l} {\rm AVP} = 3.967 - 0.3875\,{\rm X_1} - 1.888\,{\rm X_2} - 0.2750\,{\rm X_3} - \\ 0.2750\,{\rm X_1}\,{\rm X_2} + 0.5\,{\rm X_1}\,{\rm X_3} - 0.1\,{\rm X_2}\,{\rm X_3} + 0.3040\,{\rm X_1}^2 + \\ 0.3542\,{\rm X_2}^2 + 0.3292\,{\rm X_3}^2 \end{array} \tag{4} \end{array}$$

The value of correlation coefficient (r2) of Eqn. 4 was found to be 0.9499 indicating good fit. Among the independent variables selected and their interactions only X, was found to be significant (P<0.05), indicating a major contributing effect of X₂ on AVP. A negative value of the coefficient for X₂ (molar concentration of CHEMS) indicates a favorable effect on AVP. Vesicles obtained at high molar concentration of CHEMS are smaller than are those obtained at low molar concentration of CHEMS. CHEMS improves the fluidity of bilayer membrane. The smaller size of vesicles may result from this property of CHEMS. The effect of CHEMS on the liposome bilayer structure can be mainly ascribed to its charge inducing properties and presumably to a minor extent to its molecular geometry, or to a combination of both.

pH sensitivity (Y_3) of liposomes measured as % drug release at pH 5 was found to be in the range of 60.43 to 94.77%. A polynomial equation was developed for % drug release:

$$Y_{3} = 86.717 + 9.386X_{1} + 4.966X_{2} + 1.00X_{3} + 1.668 X_{1}X_{2} - 0.720X_{1}X_{3} - 1.005 X_{2}X_{3} - 5.092X_{1}^{2} - 2.452X_{2}^{2} - 8.135X_{3}^{2}$$
 (5)

The value of correlation coefficient (r^2) of Eqn. 5 was found to be 0.9799 indicating good fit. Among the independent variables selected X_1 , and X_2 , X_1^2 , and X_3^2 were found to be significant (P<0.05), indicating a major contributing effect of molar ratio of phospholipids and CHEMS on pH sensitivity of liposomes. The positive values of coefficients for X_1 and X_2 indicate a favorable effect on pH sensitivity. Liposomes with high level of DOPE and CHEMS exhibit good pH sensitivity.

The three replicated center points in the Box–Behnken experimental design made it possible to assess the pure error of the experiments and enabled the model's lack of fit to be checked. In this study, the model was checked for lack of fit for all the three responses. For lack of fit (F test) P values obtained were 0.969, 0.949, 0.909 for Y_1 , Y_2 and Y_3 , respectively, and hence the current model provided a good fit to the data (P>0.05) and had no lack of fit.

The relationship between the dependent and independent variables was further elucidated by constructing contour plots. The effects of X₁ and X₃ with their interaction on PDE at a fixed level of X₂ (medium level) are shown in fig. 2. The plots were found to be linear up to 97% PDE, indicating the linear relationship between X_1 and X_3 . It was determined from the contour plot that a higher value of PDE (97%) could be obtained with an X₁ level range from 0.8 to 1, and an X_3 level range from -0.5to -1. It is evident from the contour that the low level of X_3 , and the high level of X_1 favors PDE of liposomes. PC is present in a lower proportion at the high level of X₁, and therefore it is evident that PC decreases the entrapment efficiency. When the coefficient values of two key variables, X₁ and X₃, were compared, the value for variable X_1 (b₂=4.581) was found to be higher, indicating that it contributes the most to predict the PDE. Fig. 3 shows the contour plot drawn at 0 level of X_3 . The contours of all PDE values were found to be curvilinear and indicated that a high value of PDE (96%) can be obtained for a combination of two independent variables, X₁ level 1 and X₂ level in the range of -0.06 to -0.245, indicating that CHEMS decreases PDE. Similarly, fig. 4 shows the contour plot at 0 level of X₁. The plot is found to be curvilinear up to 91% PDE, but above this value, the plots were found to be nonlinear indicating a nonlinear relationship between X_2 and X_3 . This may be due to the interaction between X, and X₃. High value of PDE could be obtained with an X₂ level range of -0.25 to 0.99 and an X₃ level range of 1 to -0.14. All the contour plots for the high value of PDE were found to be nonlinear. This signifies that there is no direct linear relationship among the selected independent variables. A high value of PDE can be obtained up to a certain level of all three independent variables, but above this an increase in the level of independent variables leads to a decrease in the PDE of liposomes.

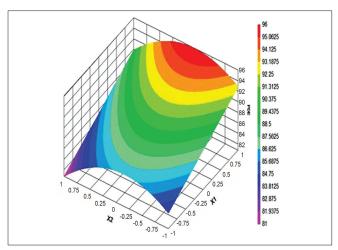


Fig. 3: Contour plot of X_1 and X_2 . Contour plot showing the effect of molar ratio of PC: DOPE (X_1) , and molar concentration of CHEMS (X_2) on the percent drug entrapment of liposomes.

TABLE 5: CHECKPOINT BATCHES WITH THEIR PREDICTED AND MEASURED VALUE OF PDE

| Batch no. | X ₁ | X ₂ | X ₃ | PI | PDE | |
|-----------|----------------|----------------|----------------|----------|-----------|--|
| | | | | Measured | Predicted | |
| 16 | 0 | -0.5 | 0.5 | 87.92 | 88.76 | |
| 17 | 0.5 | 0 | -0.5 | 91.7 | 93.46 | |
| 18 | -0.5 | 0.5 | 0 | 89.15 | 87.31 | |

PDE = Percent drug entrapment

Three checkpoint batches were prepared and evaluated for PDE, as shown in Table 5. Results indicate that the measured PDE values were as expected. When measured PDE values were compared with predicted PDE values using student *t*-test, the differences were found to be insignificant (*P*>0.05) indicating that the obtained mathematical equation is valid for predicting the PDE.

After studying the effect of the independent variables on the responses, the levels of these variables that give the optimum response were determined. It is evident from the polynomial equation and contour plots that CHEMS decreases the PDE within liposomes. Also CHEMS is known to abolish the gel-to-liquid phase transition of liposomes and the resulting liposomes are less leaky. Hence, medium level was selected as optimum for the molar concentration of CHEMS (X₂), as up to this level high value of PDE can be obtained. The optimum formulation is one that gives a high value of PDE ($\geq 91\%$), high pH sensitivity and a low AVP (≤4 µm) along with a high total amount of drug entrapped and low amount of carrier in the resultant liposomes. Using a computer optimization process and the contour plot shown in fig. 3, for X_1 the

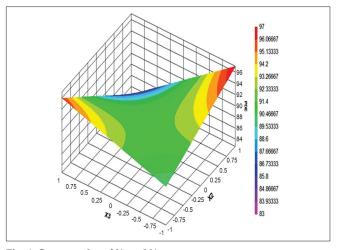


Fig. 4: Contour plot of X_2 and X_3 . Contour plot showing the effect of molar concentration of CHEMS (X_2) and amount of drug added (X_3) on the percent drug entrapment of liposomes.

level of 0.99, for X_2 level of -0.06 which gives the theoretical values of 96.08% PDE, 90.61% release, and 3.58 μ m AVP were selected. A decrease in the level of amount of drug (X_3) below the selected level leads to a decrease in the total amount of entrapped drug and increase in the level above the selected level leads to low entrapment efficiency. Hence, a 0 level of the amount of drug was selected as optimum. For confirmation, a fresh formulation was prepared at the optimum levels of the independent variables, and evaluated for responses. The observed values of PDE, % release and AVP were found to be 94.37%, 89.6% and 3.24 μ m, respectively, which were in close agreement with the theoretical values.

Multicomponent liposomal formulations may include more factors during their preparation, making the interpretation of the system extremely complicated. In order for all the factors to be used at their optimal level and the best responses to be achieved, a lot of experiments must be performed, including all the possible combinations between the different factors. The use of fractional factorial design as described in the present study, can decrease the number of experiments, give useful conclusions for the main effects and interactions between the examined factors, and clarify complicated interactions through graphical presentations.

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