SPECIFICITY OF CERTAIN BIOCHEMICAL DERANGEMENTS IN HEPATOCARCINOGENESIS

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SUMMARY.—Earlier work on acid-soluble nucleotides and other liver constituents as affected by azo-dye carcinogenesis has now been extended, with trial of ethionine (weakly carcinogenic) and of α -naphthylisothiocyanate (noncarcinogenic). The effects of ethionine feeding on whole-tissue nucleotide levels were not dramatic, and were generally dissimilar to those produced by azo-dye feeding. However a fall in some or all of the purine nucleotides can still be regarded as a feature of hepatocarcinogenesis.

A fall in mitochondrial nucleotides, as previously found in azo-dye experiments, likewise occurs with ethionine feeding, but also with α -naphthylisothiocyanate. It is suggested that the latter warrants testing as a co-carcinogen. Unlike azo-dyes, ethionine is without adverse effect on the yield of protein in cytoplasmic particles and (in common with α -naphthylisothiocyanate) it raises the yield of RNA in the supernatant fraction.

In liver from ethionine-fed rats and in ethionine-induced hepatomas, the activity of enzymes concerned in UMP synthesis showed a rise more striking than that found with azo-dyes.

PAST work on hepatocarcinogenesis in the author's laboratory has entailed study of primary hepatomas and of "precancerous" liver obtained by azo-dye feeding. Non-carcinogenic azo-dyes were also fed, to check the specificity of the supposed precancerous changes. Amongst the changes which appeared to be specific were the following: a rise in the concentration (per g. of tissue) of certain uridine nucleotides and a fall in that of ADP, GDP, NADP and NADPH₂ (Nodes and Reid, 1963); a rise in the acid-ribonuclease activity of supernatant fractions (Nodes and Reid, 1963); a fall in mitochondrial nucleotides and in microsomal protein and RNA (Reid, 1964a); and a rise in the activity of certain enzymes concerned in UMP synthesis (Reid, 1964b). In the hepatomas the abnormalities were generally similar in character to those in precancerous liver.

It is of interest to know whether these abnormalities as found in azo-dye hepatomas likewise occur in hepatomas which more closely resemble normal liver in histological and biochemical features. Such hepatomas can be induced most readily if the carcinogen is a weak one and is given intermittently rather than continuously (Morris, 1963).

Some of the experiments now reported concern a hepatoma thus induced, with ethionine as the agent. Results are also given for "precancerous" liver from rats fed this agent without interruption. Moreover, work has been done with liver

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from rats fed α -naphthylisothiocyanate. This agent is non-carcinogenic but, more strikingly than the carcinogenic azo-dyes, it causes growth of connective tissue and notably of bile-duct cells (Ungar, Moran, Eisner and Israel, 1962, and other authors as cited by Dessev, Mullock, Reid and Turner, 1969). Evidence can thereby be obtained to help discern metabolic changes which might merely reflect such growth rather than carcinogenesis *per se*.

EXPERIMENTAL

As in the work of Nodes and Reid (1963) and of Dessev *et al.* (1969), the rats were males of about 200 g. body weight initially, given a final overnight fast. A semi-synthetic diet in powder form was used (Griffin, Nye, Noda and Luck, 1948): it contained 20 per cent casein, corresponding to 0.3 per cent methionine in the diet. The α -naphthylisothiocyanate was purchased from Kodak Ltd., and DL-ethionine from the Sigma Chemical Co. ("Grade II") or from Koch-Light Ltd. The concentrations of the two agents in the diet were 0.075 and 0.25 per cent respectively. Only with ethionine was toxicity occasionally encountered, as manifested by anorexia and growth stasis. The data now tabulated refer to rats which took well to the ethionine feeding. Nevertheless they showed a poorer weight gain than the controls and a fall in liver weight relative to body weight.

The hepatomas now studied were mostly 5th to 15th generation transplants of a primary hepatoma (denoted "U"; usually sub-line "UB") which had been induced by ethionine. Transplantation was by the subcutaneous route, and had to be done every $2-2\frac{1}{2}$ weeks. The original hepatoma arose 8 months after the end of a 7-month period of ethionine feeding in which there were four gaps (amounting to 5 weeks), the first at 5 weeks. The hepatoma was induced in a "hooded" rat, but the incidence of hepatomas was much lower in this strain than in the albino strain used for most of the work. In the albino strain the hepatoma incidence 18 months after the start of feeding had approached 50 per cent. Strain differences in ethionine carcinogenesis have already been noted by Farber (1963). No difference between the strains was evident in respect of the biochemical changes after 2-4 weeks of ethionine feeding.

Estimations of tissue constituents and of enzymic activities were performed as in preceding studies which were concerned respectively with ribonucleotides and ribonucleases (Nodes and Reid, 1963), with cytoplasmic particles (Reid, 1964*a*), and with uridine nucleotide metabolism (Reid, 1964*b*). In the few instances where whole homogenates were inappropriate, suitable tissue fractions were used. For particle-located enzymes which were known to be partly latent, assays were performed with fresh homogenates in an isotonic medium, so that only "free" activity was manifest. It will be noted that tissue weight has been taken as the baseline for expressing results.

The standard abbreviations used for nucleotides (all of which were the 5'isomers) are made up as follows: A = adenosine, G = guanosine, I = inosine, U = uridine; MP = monophosphate, DP = diphosphate, TP = triphosphate. NAD(P) denotes nicotinamide adenine dinucleotide (phosphate); the reduced forms are denoted NADH₂ and NADPH₂.

RESULTS

Histology and biochemical "markers". With α -naphthylisothiocyanate the only histological abnormality of note was the expected proliferation of bile-duct

cells (Fig. 1a). These cells are smaller than the parenchymal cells which comprise the bulk of the mass of the liver. A rise in the concentration of DNA per g. of tissue was therefore expected, and was indeed evident after 3 weeks of feeding. This rise amounted to one-third (Desseev et al., 1969).

Ethionine feeding has been reported to cause diverse histological changes including hyperplasia, notably of bile-duct cells (inter alia; Farber, 1963). In the present work, as in the work of Ito, Marugani, Nakamura, Sasaki, Okajima and Kitamura (1962), there was little evidence of a change in cell population. The only histological change of note was fat deposition (Fig. 1b). The DNA concentration likewise remained close to normal (Dessev et al., 1969).

The primary hepatomas induced by ethionine feeding were heterogeneous in histological appearance. Besides large areas of trabecular carcinoma and adenocarcinoma, there were some areas of necrosis (Fig. 1c), of fibrosis, and of cholangioma-like hyperplasia of bile-duct tissue. Transplantation of the tumour for several generations led to fair homogeneity, with trabecular areas predominating (Fig. 1d). Whilst many mitoses were evident, the DNA concentration was only moderately increased in the hepatomas, whether primary or transplanted (Dessev et al., 1969). In general the hepatomas differed less from normal liver than did azo-dye hepatomas.

Dr. A. A. El-Aaser kindly furnished values for glucose-6-phosphatase, which is a "marker" for healthy parenchymal cells (Reid, 1965). In ethionine-induced hepatomas, the activity (per g.) as per cent of that in controls was typically as high as 60 (primary) and 69 (10th generation transplant), whereas the level is severely depressed in azo-dye hepatomas (Reid, 1964a, 1965). Thus the hepatomas tended to be of "minimal deviation" type in respect of glucose-6-phosphatase, although not in respect of other enzymes. Both deCMP aminohydrolase (G. C. Hartman, unpublished experiments) and glucose-6-phosphate dehydrogenase (McLean and Brown, 1966) were increased, as in azo-dye hepatomas. When ethionine was fed for 2-5 weeks the glucose-6-phosphatase activity was 66 + 6(mean and standard error; 6 experiments). There was no depression in an additional experiment where ethionine was fed for only 7 days. In 7 experiments with α -naphthylisothiocyanate, fed for 1–7 weeks, the activity was 60 ± 4 , the depression being greater than would be expected from mere "dilution" of the parenchymal tissue with bile-duct cells.

Acid-soluble nucleotides (Table I). The few experiments with α -naphthylisothiocyanate gave no distinct decreases in nucleotide levels. Indeed, the levels of AMP and of GDP tended to rise. Further experiments would be needed to confirm a possibility suggested by a single experiment, that UDPglucose, GMP and

EXPLANATION OF PLATE

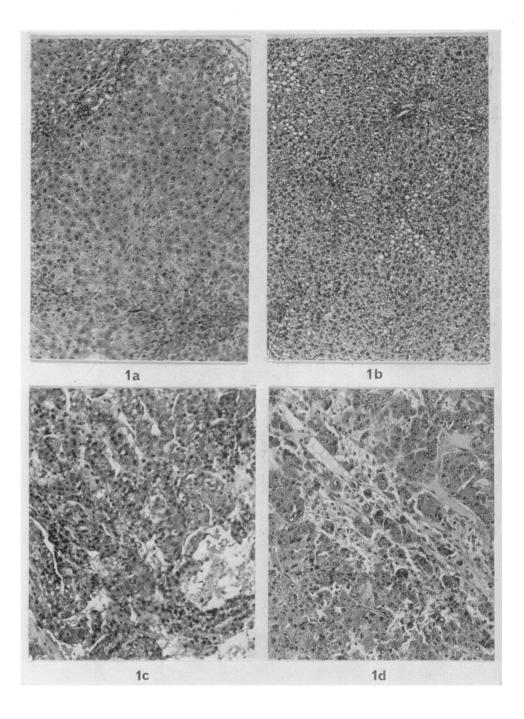
FIG. 1a.—Liver from a rat fed α -naphthylisothiocyanate for 35 days. \times 130.

FIG. 1.—Photomicrographs of tissue sections stained with haematoxylin and eosin. For comments, see text

FIG. 1b.—Liver from a rat fed DL-ethionine for 27 days. imes 75.

FIG. 1c.—Primary hepatoma induced by ethionine feeding (and subsequently transplanted; series "U") $\times 160$. FIG. 1d.—Third-generation transplant of the hepatoma shown in Fig. 1c. $\times 160$. (The trend

towards homogeneity was sharper in a 5th-generation transplant.)



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In Tables I-III, the values for experimental rats are expressed as per cent of those for controls studied simultaneously, and the number of comparisons is given in parentheses (). Values for the probability *P* are given only where the probability that the difference from controls could be due to chance was 0.1 or less.

re given only write the provability may the difference from controls could be due to chance was of or ress. UDP- UDP- activity- glucuro- glucos- nake amine +ATP* AMP ADP GMP GDP +UTP* +UMP NAD NADP NADPH. [†]	$\begin{array}{cccccccccccccccccccccccccccccccccccc$	Liver from rats fed a maphthylisothiocyanate (non-carcinogenic) $\begin{array}{cccccccccccccccccccccccccccccccccccc$	Liver from rats fed ethionine (moderately carcinogenic) 114 (2) . 92 (2) . 103 (2) . 78 (2) . 66 (2) . 66 (2) . 107 (1) . 79 (1) . 158 (2) . 88 (2) (14 (2) . 73 (2) . 88 (2) . 97 (7) . 50 (2) (17 (2) . 73 (2) . 88 (2) . 97 (7) . 50 (17 (1) . 123 (1) . 37 (1) . 37 (1) . 59 (2) (16 (1) . 94 (1) . 33 (1) . 123 (1) . (11) (11) (11) (11) (11) (11) (11	Ethionine-induced hepatomas (transplants)§ (5; $P < 0.005$); (5; $P < 0.001$) (5; $P < 0.005$); (5; $P < 0.001$)	Only in a few experiments (not tabulated) was the peak rechromatographed to give the individual components; these include a small amount of UDP in the case of peak healled UDP gineuronate $+$ ATP. Isolated as its breakdown product, ADP. Iso derive μ mids basic write the form "denixy units" (i.u.; riz, E, if present in 1 ml, of concentrate), it would be necessary to divide by 10-0 (U), 14-2 (A or 1), or 11-8 (G).
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AMP	. 8.9 (=100)	Liver from rats 94 (1) \cdot 137 152 89 (2) \cdot 157 \cdot 152 89 (2) \cdot 158 $\begin{pmatrix} P < \\ P < \\ 0 \cdot 1 \end{pmatrix}$	Liver from . 114 (2) . 104 (2) . 163 (1)	Ethionii $5; \frac{25\pm7}{P<0.005}$	e peak rechron: .; riz. E ₂₆₀ if pi
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UDP- UDP- Mean value in	controls, d.u./g.‡	13 days' feeding . 27-37 days	15 days 27-29 days 37 days	·	 Only in a few experiments (not tabulat the peak labelled UDP glueuronate + ATP † Isolated as its breakdown product, ADI † To derive "mole values from " density."

DERANGEMENTS WITH HEPATOCARCINOGENESIS

131

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		Mito	Mitochondrial fraction	tion		RNA	DWA wot	Gunamotont
	Au	Acid-soluble nucleotides	NAD only	Protein	Protein	extractable at pH 9 with Mg ²⁺ present	run nuu readily extractable	supernatant fraction, RNA
Mean value in controls, d.u./g.† or mg./g.		$19 \ (=100)$	$2 \cdot 6 \; (=100)$	49 (=100)	26 (=100)	$1 \cdot 4 \ (=100)$	$1 \cdot 2 \; (= 100)$	$1 \cdot 8 \; (=100)$
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	13 (1)	•		Transplanted hepatoma 	oma	I	1	I
† Nucleotide values are in density units (viz. E260 if present in 1 ml. of concentrate); the column for acid-soluble nucleotides includes NAD, the amount of which is given in the next column.	are in den given in t T booded	isity units (viz. the next colum	. E260 if presei m.	are in density units (<i>viz.</i> E ₂₈₀ if present in 1 ml. of concentrate); the column for acid-soluble nucleotides includes NAD, given in the next column. IT hooded we have then allying more used for the transmission and for two feading extrements in each set (other then	intrate); the co	lumn for acid-solu	uble nucleotide. riments in each	s includes NAD, set (other then

TABLE II.—Constituents of the Mitochondrial, Microsomal and Supernatant Fractions*

* In Tables II and III, hooded rather than albino rats were used for the transplants and for two feeding experiments in each set (other than the microsomal extractions).

 $NADPH_2$ are transiently increased at 13 days. With hepatocarcinogens there are many precedents for such transient effects (Reid, 1962).

With ethionine feeding, triphosphates tended to fall as judged by values for the mixed peak containing UTP and GTP and for the mixed peak rich in ATP. The two diphosphates examined, ADP and GDP, were significantly although not sharply decreased. The monophosphates AMP and GMP were normal except for an isolated result with 37 days of feeding when AMP was high and GMP was low. The mixed peak containing IMP and UMP showed no consistent changes in level. UDPacetylglucosamine was of the normal order, but UDPglucose was somewhat decreased.

The present experiments with ethionine feeding also indicated decreases in NAD and in NADPH₂; but in two experiments where the feeding lasted only 15 days the values for NADP were high rather than low. In another 15-day experiment (not tabulated) the final overnight fast was omitted, with little effect on the trends for the various nucleotides.

Table I further shows that NADP and NADPH₂ were sharply decreased in ethionine-induced hepatomas. All the purine nucleotides were depressed, as was shown conclusively for AMP and ADP. For the mixed peak containing ATP, the decrease was in part attributable to a fall in UDPglucuronic acid. Yet the levels of UDPacetylglucosamine and of UDPglucose were normal.

Constituents of subcellular fractions (Table II). Both with α -naphthylisothiocyanate and with ethionine; the mitochondrial fraction showed a decrease in the content of acid-soluble nucleotides, particularly after 3 weeks of feeding. Chromatography of the nucleotide mixture showed that NAD was undiminished. Untabulated results for the other nucleotides indicated that AMP, ATP and NADPH₂ (measured as ADPriboseP) were somewhat diminished after 3–7 weeks of ethionine feeding; but there was no fall in ADP or in a peak containing NADP and IMP. With a shorter period of feeding, neither ethionine nor α -naphthylisothiocyanate depressed the ATP level. The total nucleotide content was sharply depressed in an ethionine-induced hepatoma (Table II).

Table II further shows that ethionine feeding did not diminish the protein of the mitochondrial and microsomal fractions. With α -naphthylisothiocyanate the protein of mitochondrial fractions was diminished. Although neither agent diminishes total microsomal RNA (Dessev *et al.*, 1969), an extraction procedure already used in azo-dye experiments (Reid, 1964a) warranted trial. Evidently there was no change in the extractability of microsomal RNA. However, with both agents there was some increase in supernatant-fraction RNA (Table II)—an effect not encountered with azo-dye feeding (Reid, 1962). *Enzymes of uridine nucleotide metabolism* (Table III). No significant changes

Enzymes of uridine nucleotide metabolism (Table III). No significant changes in the levels of these enzymes were seen with α -naphthylisothiocyanate. In an experiment entailing prolonged feeding the rate of conversion of uridine into UMP (uridine kinase activity) was high, but this represents an isolated observation. This enzymic activity was dramatically increased by ethionine feeding, as was the activity of the multi-enzyme system that catalyses the conversion of carbamoylphosphate into UMP. Conversely, the activity of uracil reductase—the ratelimiting enzyme in uracil catabolism—was depressed by ethionine feeding. UTPase activity was unchanged. The few assays performed on ethionine-induced hepatomas gave high values for aspartate transcarhomylase and uridine kinase activities, both with primary tumours and with transplants.

TABLE III.—Enzymes Concerned in Uridine Nucleotide MetabolismAs in Tables I and II, the values are expressed relative to controls as 100CarbamoylCarbamoylCarbamoylCarbamoylCarbamoylCarbamoylCarbamoylCarbamoylCarbamoylOrotate <td co<="" th=""><th>0.043 (=100) . 0.018 (=100) . 12.0 (=100) . 0.11 (=100)</th><th>$\begin{array}{cccccc} \mbox{Liver from rate fed \$\alpha\$-naphthylisothiocyanate (non-carcinogenic)} \\ 2) & : & 100 (2) & : & 132 (1) & : & 87 (2) & : & 118 (2) & : & 77 (2) \\ 1) & : & 104 (3) & : & 88 (2) & : & 116 (6) & : & 77 (2) & : & 86 (2) \\ 1) & : & 126 (2) & : & - & & : & 290 (1) & : & - & & : & 103 (1) \\ \end{array}$</th><th>Liver from rats fed ethionine (moderately carcinogenic) $\begin{array}{c} 138 \ (3) \\ \vdots \ 143 \ (3) \\ \vdots \ 206 \ (4) \\ (P < 0 \cdot 02) \ 108 \ (1) \\ \end{array} \begin{pmatrix} 189 \pm 34 \\ (P < 0 \cdot 05) \ 174 \ (2) \\ \end{array} \begin{pmatrix} 49 \\ 60 \\ 58 + 63 \\ 06 \\ \end{array} \begin{pmatrix} 98 \\ 60 \\ 268 + 63 \\ 06 \\ \end{array} \begin{pmatrix} 98 \\ 60 \\ 268 + 63 \\ 06 \\ 06 \\ \end{array} \begin{pmatrix} 43 \\ 288 + 63 \\ 268 + 63 \\ 06 \\ 06 \\ 06 \\ 06 \\ 06 \\ 06 \\ 06 \\$</th><th>Ethionine-induced hepatomas </th></td>	<th>0.043 (=100) . 0.018 (=100) . 12.0 (=100) . 0.11 (=100)</th> <th>$\begin{array}{cccccc} \mbox{Liver from rate fed \$\alpha\$-naphthylisothiocyanate (non-carcinogenic)} \\ 2) & : & 100 (2) & : & 132 (1) & : & 87 (2) & : & 118 (2) & : & 77 (2) \\ 1) & : & 104 (3) & : & 88 (2) & : & 116 (6) & : & 77 (2) & : & 86 (2) \\ 1) & : & 126 (2) & : & - & & : & 290 (1) & : & - & & : & 103 (1) \\ \end{array}$</th> <th>Liver from rats fed ethionine (moderately carcinogenic) $\begin{array}{c} 138 \ (3) \\ \vdots \ 143 \ (3) \\ \vdots \ 206 \ (4) \\ (P < 0 \cdot 02) \ 108 \ (1) \\ \end{array} \begin{pmatrix} 189 \pm 34 \\ (P < 0 \cdot 05) \ 174 \ (2) \\ \end{array} \begin{pmatrix} 49 \\ 60 \\ 58 + 63 \\ 06 \\ \end{array} \begin{pmatrix} 98 \\ 60 \\ 268 + 63 \\ 06 \\ \end{array} \begin{pmatrix} 98 \\ 60 \\ 268 + 63 \\ 06 \\ 06 \\ \end{array} \begin{pmatrix} 43 \\ 288 + 63 \\ 268 + 63 \\ 06 \\ 06 \\ 06 \\ 06 \\ 06 \\ 06 \\ 06 \\$</th> <th>Ethionine-induced hepatomas </th>	0.043 (=100) . 0.018 (=100) . 12.0 (=100) . 0.11 (=100)	$ \begin{array}{cccccc} \mbox{Liver from rate fed α-naphthylisothiocyanate (non-carcinogenic)} \\ 2) & : & 100 (2) & : & 132 (1) & : & 87 (2) & : & 118 (2) & : & 77 (2) \\ 1) & : & 104 (3) & : & 88 (2) & : & 116 (6) & : & 77 (2) & : & 86 (2) \\ 1) & : & 126 (2) & : & - & & : & 290 (1) & : & - & & : & 103 (1) \\ \end{array} $	Liver from rats fed ethionine (moderately carcinogenic) $\begin{array}{c} 138 \ (3) \\ \vdots \ 143 \ (3) \\ \vdots \ 206 \ (4) \\ (P < 0 \cdot 02) \ 108 \ (1) \\ \end{array} \begin{pmatrix} 189 \pm 34 \\ (P < 0 \cdot 05) \ 174 \ (2) \\ \end{array} \begin{pmatrix} 49 \\ 60 \\ 58 + 63 \\ 06 \\ \end{array} \begin{pmatrix} 98 \\ 60 \\ 268 + 63 \\ 06 \\ \end{array} \begin{pmatrix} 98 \\ 60 \\ 268 + 63 \\ 06 \\ 06 \\ \end{array} \begin{pmatrix} 43 \\ 288 + 63 \\ 268 + 63 \\ 06 \\ 06 \\ 06 \\ 06 \\ 06 \\ 06 \\ 06 \\$	Ethionine-induced hepatomas
TABLE III.—Enzymes Con As in Tables I and II, the ve Carbamoyl- carbamoyl- phosphate Carbamoyl expartate Carbamoyl- carbamoyl- expondete Software exportate Phosphate orotate, free "starte Software 3.5.2.3 and 2.1.3.2	0.15(=100) . $0.0033(=100)$.	Liver from rats fed α-77 (2) 100 (2) 106 (4) . 104 (3) 133 (1) . 126 (2)	Liver from r72 (2)138 (3)132 (4)143 (3)150 (1)215 (1)	266 (2)	
I.U.B. no. of enzyme	Mean activity in controls, . µmoles/g./min.	8-16 days' feeding 20-41 days 50-51 days	8-22 days 27-41 days 50-51 days	Primary hepatomas Transplanted hepatomas .	

* Assay conditions were such that latent activity was not liberated (see text).

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			Correlation with concer from	
Biochemical change	Correlation with carcinogenicity of the agent, from work with azo-dyes*	Correlation as affected by present results with other agents†	results for azo-dys and Morris 123 rat hepatomas and for a mouse hepatoma*	is Correlation as affected by present work
Fall in NADP Fall in NADPH2 Rise in uridine nucleotides, in precancerous liver	t bad	No clearcut evidence Slightly strengthened Weakened (no rises observed)	Fair Good	Strengthened Strengthened
Fall (moderate) in UDPglucose and UDP- glucuronate in hepatomas		I	. Fair (UDPglucose) or good	Weakened (UDPglucose)
Fall in purine nucleotides	Good for ADP and GDP	Slightly strengthened for ADP and GDP	. Good	Strengthened
Fall in nucleotides of mitochondrial fraction	Good (except for NAD)	Equivocal (fall with αN as well as with athionine)	. Good	Strengthened
Fall in protein of cytoplasmic particles and in microsomal RNA	Good	Weakened	. Uncertain or good	I
Rise in supernatant fraction Good acid ribonuclease		Equivocal (rise with aN as well as with ethionine)?	. Fair	Strengthened [‡]
**	ocal	Strengthened	. Equivocal	Strengthened
Rise in uridine kinase Fall in uracil reductase	Fair Poor	Strengthened Strengthened	. Equivocal . (No fall)	Strengthened
* See Nodes and Reid (19	* See Nodes and Reid (1963) and Reid (1964a, b).			

TABLE IV.—Possible Significance for Hepatocarcinogenesis of Certain Biochemical Changes As in the foregoing Tables, results are considered on a tissue-weight hasis

* See Nodes and Keid (1963) and Keid (1964*a*, b). † Agents: ethionine, a moderately active carcinogen; α -naphthylisothiocyanate (" α N "), non-carcinogenic agent. ‡ A. A. El-Aaser and E. Reid, unpublished experiments.

135

DISCUSSION

The findings for nucleotide levels in ethionine-fed rats may be compared with those reported by Caldarera, Budini, Barbiroli and Rabbi (1962). Contrary to the present findings these authors observed a marked rise in GMP and sharp decreases in UMP, IMP, UDP and GDP. Ethionine clearly does not mimic the effect of azo-dyes in raising uridine nucleotide levels. For ATP and for NAD, account has also to be taken of other reports (Bartels and Hohorst, 1963; Stekol, Bedrak, Mody, Burnette and Somerville, 1963; Shull, Villa-Trevino and Oler, 1964; Smith and Salmon, 1965). There is general agreement that ATP and NAD fall in ethionine-treated rats, but disagreement about the magnitude of the decreases even if regard is paid only to results for chronic as distinct from acute treatment. Only small decreases have now been found. The magnitude of the fall in UTP and GTP also varies from one laboratory to another.

Whilst a fall in triphosphate levels represents the most striking change in the nucleotide pattern of whole tissue after ethionine feeding, such a fall is not a consistent feature of azo-dye carcinogenesis (Nodes and Reid, 1963). There is, however, a striking effect common to azo-dyes (Reid, 1964*a*) and to ethionine, namely a depletion of nucleotides (other than NAD) in isolated mitochondrial fractions. There is some evidence that mitochondrial function is impaired after ethionine administration (Stekol *et al.*, 1963; see also Reid, 1965). Possibly the mitochondrial membranes become more fragile, so that nucleotides escape *in vivo* or at least *in vitro*.

It may seem disappointing that the mitochondrial depletion is found even with the non-carcinogenic agent α -naphthylisothiocyanate. One possibility, now being tested, is that this agent may be a co-carcinogen, as might be revealed by potentiation of the carcinogenic action of a weak carcinogen such as ethionine. In agreement with this suggestion of a "pseudo-carcinogenic" role, α -naphthylisothiocyanate simulated carcinogenie azo-dyes in raising AMP aminohydrolase (deaminase) activity and thereby vitiated an otherwise good correlation with carcinogenicity (Kizer, Lovig, Howell and Cox, 1964). Moreover, Sneider and Potter (1969) found that α -naphthylisothiocyanate, in common with a carcinogenic azo-dye, raised the level of deoxycytidine monophosphate aminohydrolase (deaminase), despite earlier negative results (Hartman and Reid, 1964) with a less sensitive method of assay. Another change common to α -naphthylisothiocyanate and carcinogens is a rise in acid ribonuclease in the supernatant fraction (A. A. El-Aaser and E. Reid, unpublished experiments).

Table IV collates evidence concerning the nature and importance of changes found early during hepatocarcinogenesis or in hepatomas. Attention is particularly drawn to the fall in NADPH₂ and, in hepatomas, the fall in purine nucleotides. The capacity of the enzyme pathways for UMP synthesis tends to rise, but hardly so dramatically as to point to a key role in hepatocarcinogenesis. Indeed, the Morris 5123 hepatoma shows no rise in uridine kinase activity (Reid, 1964b). This reinforces the view that trial of different hepatomas and of different dietary agents is helpful for screening the diverse changes observed and eliminating those which are unrelated to hepatocarcinogenesis.

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Note added in proof.—A trial with rats fed α -naphthylisothiocyanate together with ethionine in low concentration has given no evidence of strong co-carcinogenicity (B. M. Bullock and E. Reid, unpublished work).

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