

### Supporting information

#### **Nanocapillary sampling coupled to liquid chromatography mass spectrometry delivers single cell drug measurement and lipid fingerprints**

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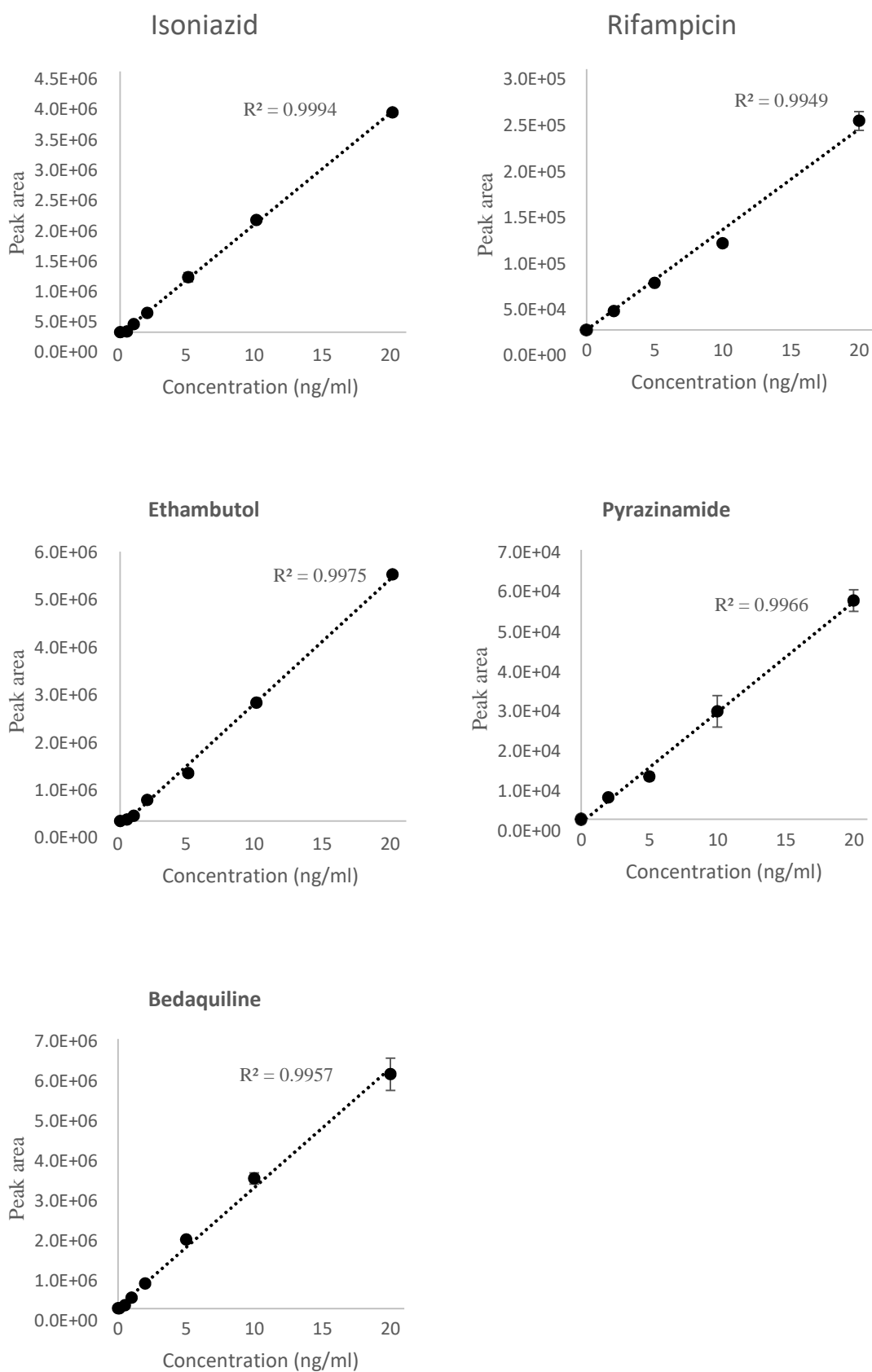
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**Table S1: The liquid chromatography gradient applied in this work, where mobile phase solvent A) 60:40 acetonitrile/water + 0.1% formic acid and mobile phase solvent B) 90:10 isopropanol/acetonitrile + 0.1% formic acid**

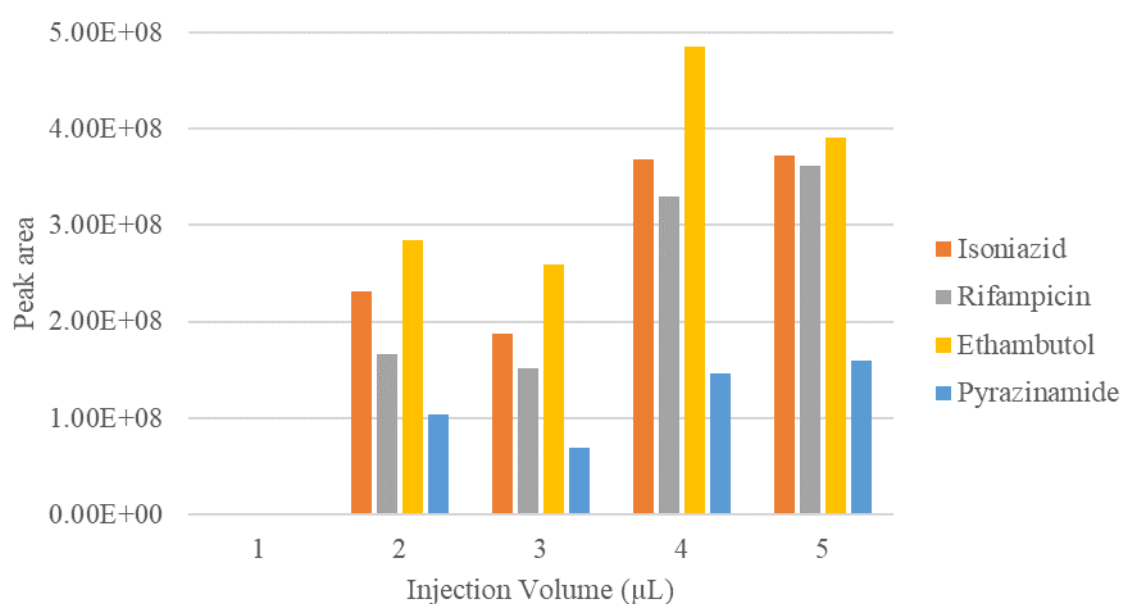
Time (min)	% Solvent <b>A</b>	% Solvent <b>B</b>
0.0	60	40
1.0	50	50
3.6	31	69
12.0	12	88
14.0	60	40
16.0	60	40



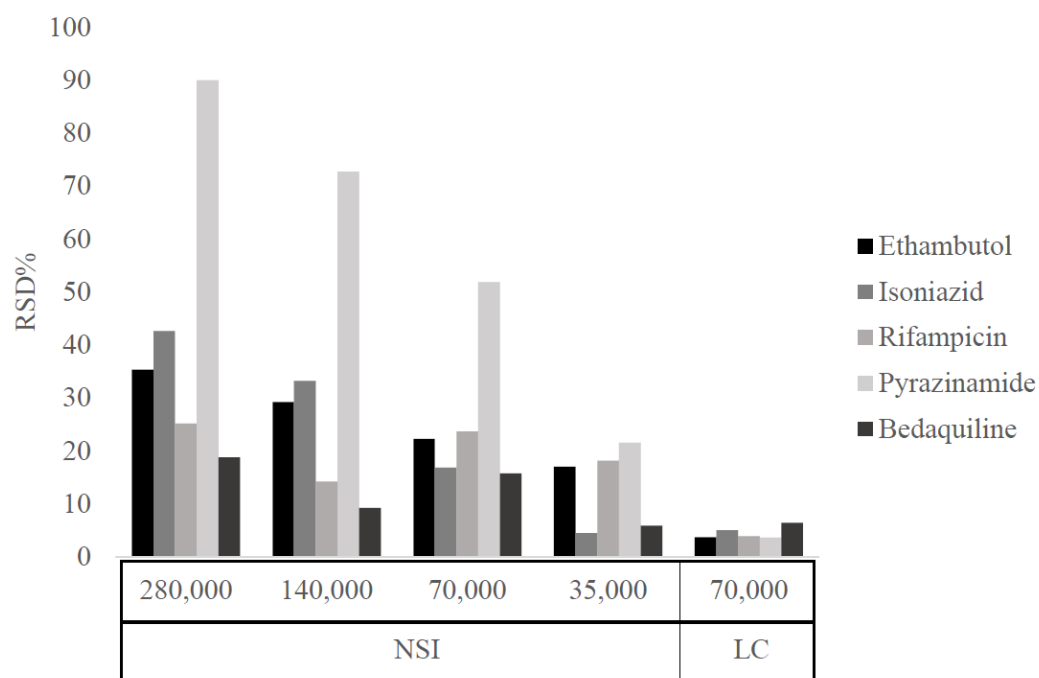
**Figure S1: Calibration curves for isoniazid, rifampicin, ethambutol, pyrazinamide and bedaquiline and concentrations 0, 0.1, 0.5, 1, 2, 5, 10, and 20 ng/ml diluted in MeOH:EtOH using LC-MS method; (n=5) injections per sample where the error bars show the standard deviation between repeats.**

**Table S2. Limits of detection for the drug analytes in solution, determined by a method of infinite dilutions.**





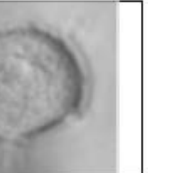


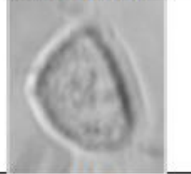
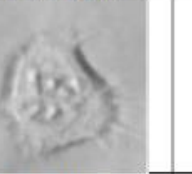
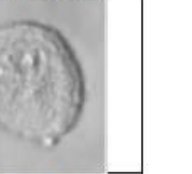




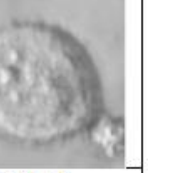



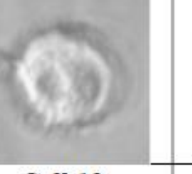


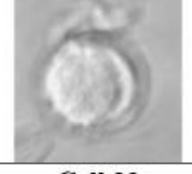







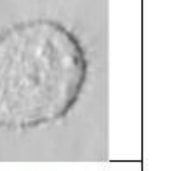
	Limit of detection (ng/ml)	Limit of detection (pg)
Isoniazid	1	5
Rifampicin	2	10
Ethambutol	0.5	2.5
Pyrazinamide	2	10
Bedaquiline	0.5	2.5



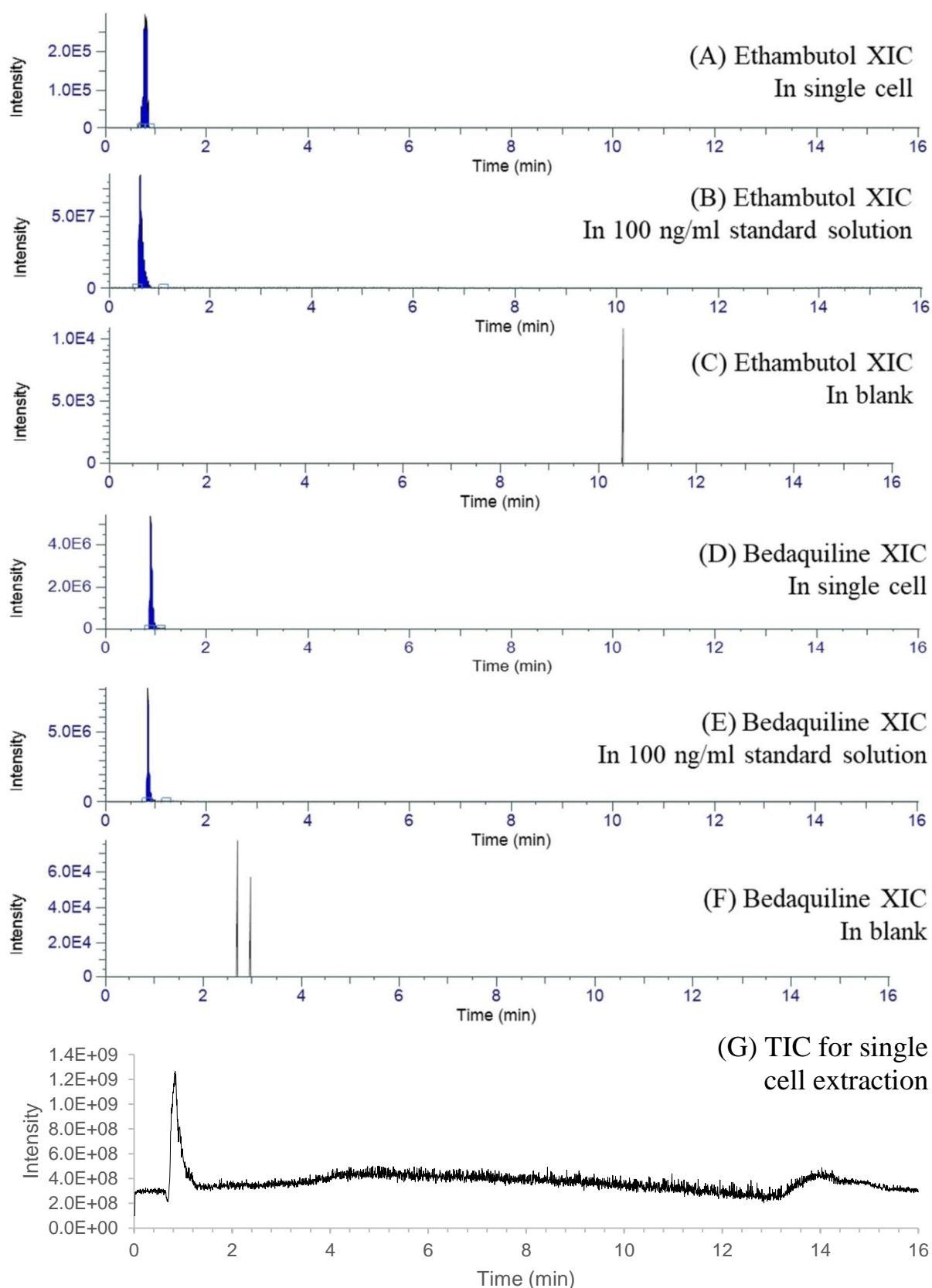
**Figure S2: Signal intensity for isoniazid, rifampicin, ethambutol and pyrazinamide where the same mass (10 ng) of analyte is diluted in different injection volumes (n=1)**



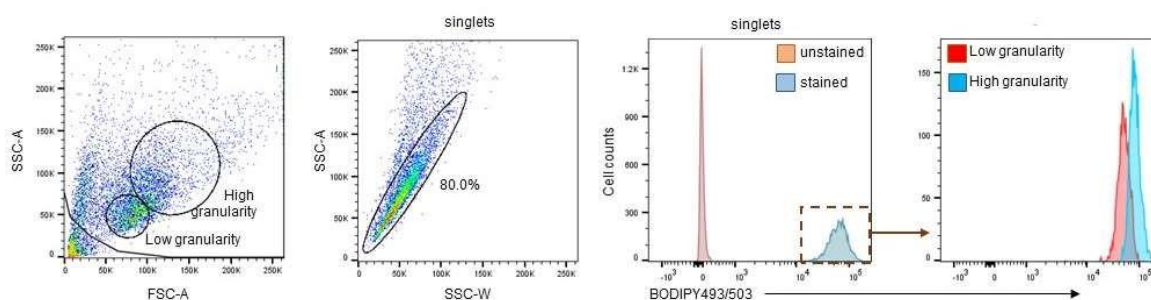
**Figure S3: A graph to show the RSD% (n=5) of the [M+H]<sup>+</sup> intensity of the five anti-TB drugs: ethambutol, isoniazid, rifampicin, pyrazinamide and bedaquiline (100 ng/ml), using NSI (3.5 minute acquisition) at four different mass resolutions: 280,000; 140,000; 70,000 and 35,000 and also LC at 70,000 (n=5) for comparison**

				
<b>Cell 1</b>	<b>Cell 2</b>	<b>Cell 3</b>	<b>Cell 4</b>	<b>Cell 5</b>
38 $\mu\text{m}$	27 $\mu\text{m}$	34 $\mu\text{m}$	35 $\mu\text{m}$	30 $\mu\text{m}$
28700 $\mu\text{m}^2$	10300 $\mu\text{m}^2$	20600 $\mu\text{m}^2$	22400 $\mu\text{m}^2$	14100 $\mu\text{m}^2$
				
<b>Cell 6</b>	<b>Cell 7</b>	<b>Cell 8</b>	<b>Cell 9</b>	<b>Cell 10</b>
20 $\mu\text{m}$	34 $\mu\text{m}$	22 $\mu\text{m}$	35 $\mu\text{m}$	42 $\mu\text{m}$
4200 $\mu\text{m}^2$	20600 $\mu\text{m}^2$	5600 $\mu\text{m}^2$	22400 $\mu\text{m}^2$	38800 $\mu\text{m}^2$
				
<b>Cell 11</b>	<b>Cell 12</b>	<b>Cell 13</b>	<b>Cell 14</b>	<b>Cell 15</b>
27 $\mu\text{m}$	31 $\mu\text{m}$	35 $\mu\text{m}$	27 $\mu\text{m}$	43 $\mu\text{m}$
10300 $\mu\text{m}^2$	15600 $\mu\text{m}^2$	22400 $\mu\text{m}^2$	10300 $\mu\text{m}^2$	41600 $\mu\text{m}^2$
				
<b>Cell 16</b>	<b>Cell 17</b>	<b>Cell 18</b>	<b>Cell 19</b>	<b>Cell 20</b>
58 $\mu\text{m}$	30 $\mu\text{m}$	33 $\mu\text{m}$	27 $\mu\text{m}$	31 $\mu\text{m}$
102100 $\mu\text{m}^2$	14100 $\mu\text{m}^2$	18800 $\mu\text{m}^2$	10300 $\mu\text{m}^2$	15600 $\mu\text{m}^2$
				
<b>Cell 21</b>	<b>Cell 22</b>	<b>Cell 23</b>	<b>Cell 24</b>	<b>Cell 25</b>
37 $\mu\text{m}$	32 $\mu\text{m}$	40 $\mu\text{m}$	24 $\mu\text{m}$	30 $\mu\text{m}$
26500 $\mu\text{m}^2$	17200 $\mu\text{m}^2$	33500 $\mu\text{m}^2$	7200 $\mu\text{m}^2$	14100 $\mu\text{m}^2$
				
<b>Cell 26</b>	<b>Cell 27</b>	<b>Cell 28</b>	<b>Cell 29</b>	<b>Cell 30</b>
41 $\mu\text{m}$	46 $\mu\text{m}$	32 $\mu\text{m}$	51 $\mu\text{m}$	32 $\mu\text{m}$
36100 $\mu\text{m}^2$	50900 $\mu\text{m}^2$	17200 $\mu\text{m}^2$	69500 $\mu\text{m}^2$	17200 $\mu\text{m}^2$

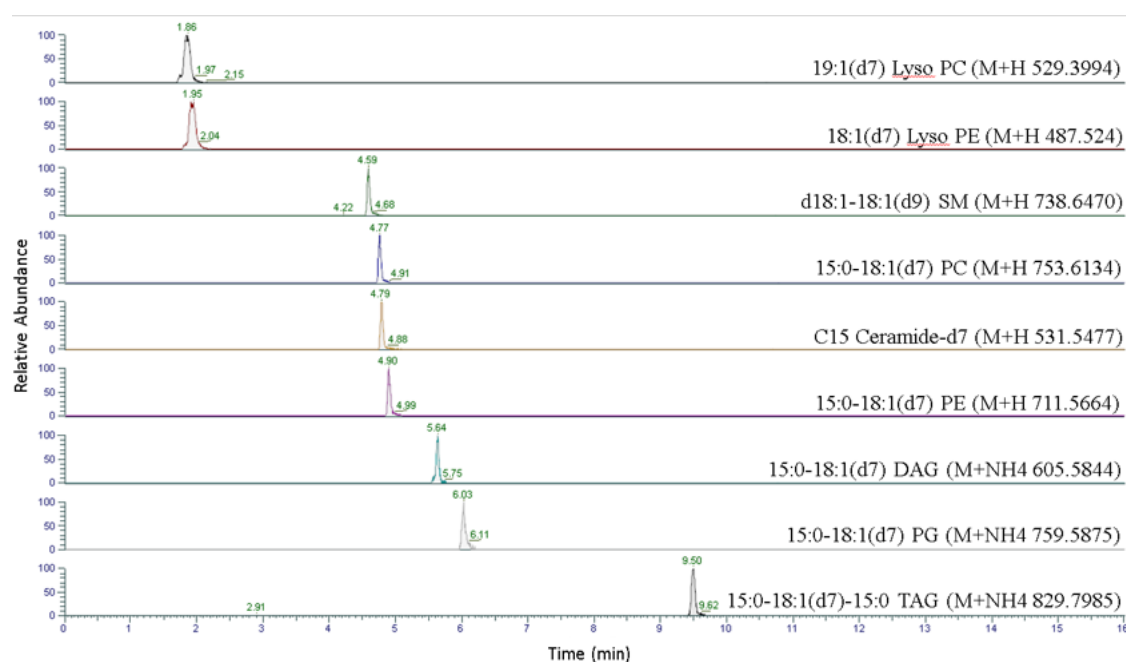
**Figure S4. Microscope images of the 30 extracted cells before nanocapillary extraction alongside the corresponding cell diameters and volumes.**



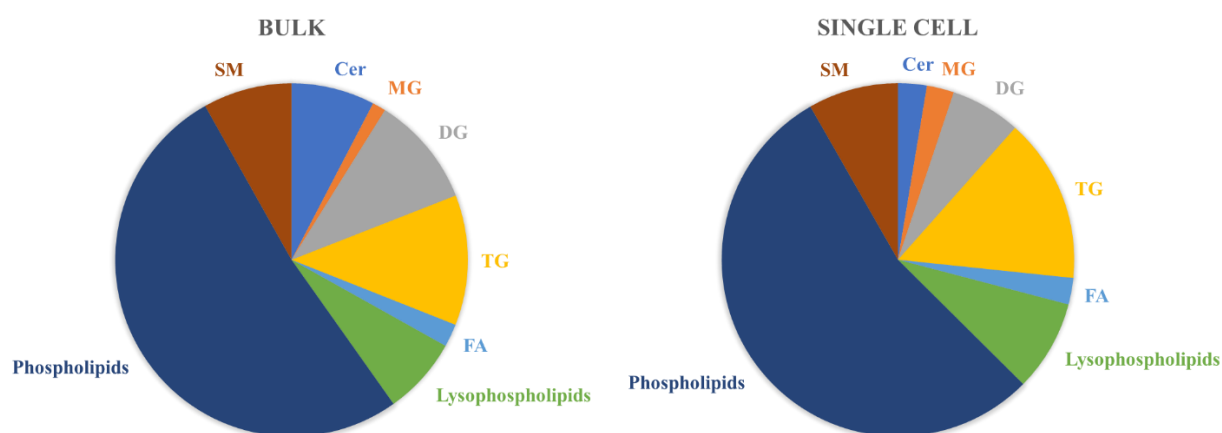
**Figure S5: Extracted ion chromatograms for ethambutol ( $m/z$  205.1911) in a single cell, in a 100 ng/ml standard solution and in a blank (PBS extracted using pressure injector diluted with 5 $\mu$ L 50:50 MeOH/EtOH solution) (A, B and C) and extracted ion chromatograms for bedaquiline ( $m/z$  555.1642) in a single cell, in a 100 ng/ml standard solution and in a blank (PBS extracted using pressure injector diluted with 5 $\mu$ L 50:50 MeOH/EtOH solution) (D, E and F). The single cell Total Ion Chromatogram is also shown (G).**



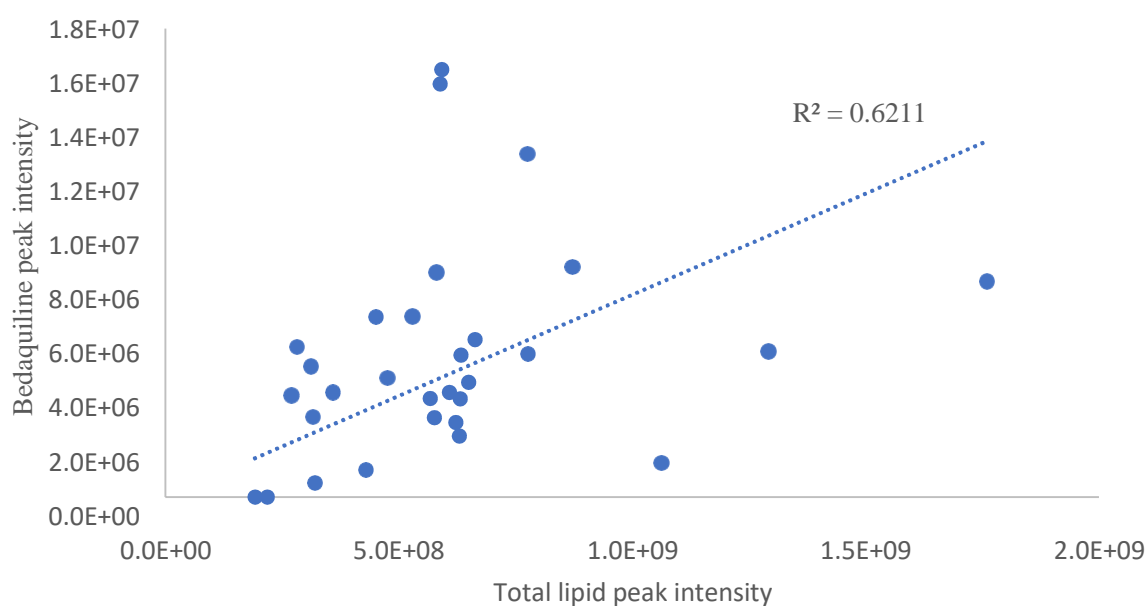
**Figure S6: PANC-1 cells present high levels of lipid droplets as shown by the specific probe BODIPY 493/503.**



**Figure S7: Extracted ion chromatograms of lipids analytes in the Avanti® Polar Lipids EquiSPLASH® diluted to 32 ng/mL dissolved in starting mobile phase, showing separation obtained using the chromatography method**

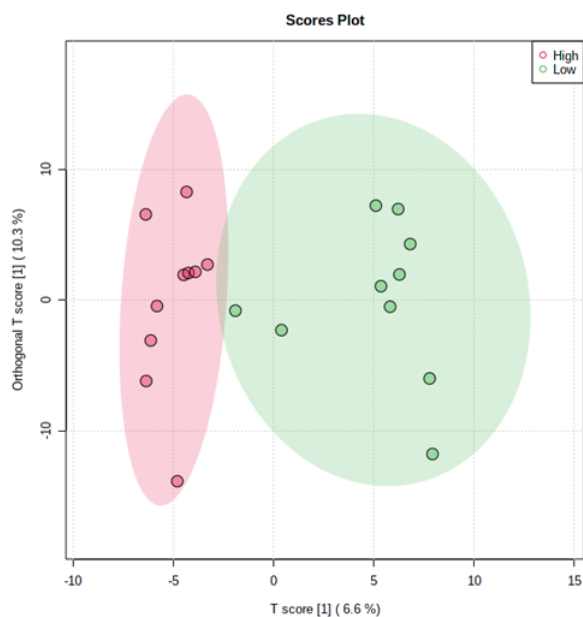


**Figure S8: Charts showing proportion of assigned lipids detected (A) for cell suspension (B) averaged data for 12 single cells. Where SM: sphingolipids, Cer: ceramides, MG: monoglycerides, DG: diglycerides, TG: triglycerides, FA: fatty acids**



**Figure S9: The total lipid peak intensity vs. the bedaquiline peak intensity for each extracted cell**





m/z	Presumptive peak assignment	VIP score
843.7412	TG(14:0e_18:2_20:4)+Li	2.23
732.5150	PC(12:0_20:4)+Li	2.18
734.5119	PC(12:1e_22:6)+H	2.18
694.6133	Cer(d20:1_26:6)+H	2.12
950.7396	PS(24:1_23:0)+Li	2.07
775.7538	OAHA(15:0_36:1)+H	2.06
894.7134	PS(20:0e_24:1)+Li	2.00
415.3030	DG(10:0_11:3)+Li	2.00

**Figure S10: (A) Partial Least Squares Discriminant Analysis (PLS-DA) for the 10 cells with the highest measured ethambutol content and 10 cells with the lowest measured ethambutol content. (B) Top variable importance in projection scores.**