# Research Article

# Ischemia Induces Release of Endogenous Amino Acids from the Cerebral Cortex and Cerebellum of Developing and Adult Mice

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Ischemia enhanced release of endogenous neuroactive amino acids from cerebellar and cerebral cortical slices. More glutamate was released in adult than developing mice. Taurine release enhanced by  $K^+$  stimulation and ischemia was more than one magnitude greater than that of GABA or glutamate in the developing cerebral cortex and cerebellum, while in adults the releases were almost comparable. Aspartate release was prominently enhanced by both ischemia and  $K^+$  stimulation in the adult cerebral cortex. In the cerebellum  $K^+$  stimulation and ischemia evoked almost 10-fold greater GABA release in 3-month olds than in 7-day olds. The release of taurine increased severalfold in the cerebellum of 7-day-old mice in high- $K^+$  media, whereas the  $K^+$ -evoked effect was rather small in adults. In 3-month-old mice no effects of  $K^+$  stimulation or ischemia were seen in the release of aspartate, glycine, glutamine, alanine, serine, or threonine. The releases from the cerebral cortex and cerebellum were markedly different and also differed between developing and adult mice. In developing mice only the release of inhibitory taurine may be large enough to counteract the harmful effects of excitatory amino acids in ischemia in both cerebral cortex and cerebellum, in particular since at that age the release of glutamate and aspartate cannot be described as massive.

# 1. Introduction

Glutamate and  $\gamma$ -aminobutyrate (GABA) are the two major amino acid transmitters in the cerebral cortex and cerebellum, glutamate being responsible for excitatory and GABA for inhibitory transmission [1]. In these higher brain regions glycine was earlier assumed to be only an obligatory cotransmitter in the excitatory N-methyl-D-aspartate- (NMDA-) sensitive glutamate receptors, but more recent studies have also demonstrated the existence and function of strychninesensitive inhibitory glycine receptors in these structures [2, 3]. In addition to these established neurotransmitters, taurine also affects neuronal activity as an inhibitory modulator [4]. In the rodent brain the concentrations of taurine are high. In particular, in the developing brain it is the most abundant amino acid, even exceeding the concentration of glutamate [5].

The excessive extracellular accumulation of excitatory amino acids, predominantly that of glutamate but also of aspartate, in ischemia leads to cellular damage in the brain [6, 7]. Their massive release activates glutamate receptors, in particular those of the NMDA class [8], which leads to an excessive influx of  $Ca^{2+}$  and consequent adverse effects [9]. This excitotoxicity may be counteracted by the simultaneous enhanced release of inhibitory GABA and taurine [10, 11]. The functional status in the brain is a delicate balance between these excitatory and inhibitory neurotransmitters under both normal and pathological conditions.

In microdialysis studies *in vivo*, the overflow of endogenous extracellular amino acids can be assessed, (e.g., [6, 12]), but in the vast majority of *in vitro* studies the release of neurotransmitter amino acids has been investigated with the aid of preloaded radioactively labeled compounds. These admix more or less readily with the endogenous homologous pool in the cells and are only thereafter released into the extracellular spaces. Hence, the calculated release rate is affected by the efficacy of this mixing and the sizes of the endogenous amino acid pools and, thus, may not reliably reflect the magnitude of the release. The release of preloaded radioactively labeled amino acids from cerebral cortical preparations has been relatively frequently investigated, whereas relatively few studies have been made with the cerebellum [13–15]. Here we compared the release rates of endogenous GABA, glutamate, aspartate, glycine, and taurine from cerebral cortical and cerebellar slices to estimate the release on a molar basis under normoxic and ischemic conditions. The cerebral cortex and cerebellum represent functionally different brain structures, the cerebellum being predominantly inhibitory and the cerebral cortex with mixed functions, including excitation.

#### 2. Experimental Procedure

2.1. Material. Developing (7-day olds) and young adult (3month olds) NMRI mice of both sexes were used in the experiments. There were no significant differences between male and female mice. The results on male and female mice were therefore combined. The experiments conformed to the European Community Directive (86/609/EEC) for the ethical use of experimental animals, and they were approved by the Tampere University Committee for Animal Experiments. All efforts were made to reduce the number of animals used and their suffering.

2.2. Release Experiments. The mice were killed by fast decapitation and their brains excised without delay. Superficial slices 0.4 mm thick weighing 15–20 mg were manually prepared from the mouse cerebral cortex and cerebellum with a tissue slicer of Stadie-Riggs type. The slices were immediately immersed in 5 mL of preoxygenated medium and preincubated for 30 min under O<sub>2</sub> at 37°C under agitation in standard medium containing (in mmol/L) NaCl 127, KCl 5, CaCl<sub>2</sub> 0.8, MgSO<sub>4</sub> 1.3, Na<sub>2</sub>HPO<sub>4</sub> 1.3, N-2hydroxyethylpiperazine-N<sup>-2</sup>-ethanesulphonic acid (Hepes) 15, NaOH 11, and D-glucose 10 (pH 7.4). The slices were then transferred into 0.25 mL cups and superfused with the above medium (unless otherwise specified) at a rate of 0.25 mL/min for 50 min in a system in which freely floating shaken slices were kept under a continuous flow of oxygen in order to preserve their viability [16]. The superfusion medium was pooled during the first 20 min, whereafter 2 min fractions (0.5 mL) were collected. At 30 min the medium was in many experiments changed to medium containing 50 mM K<sup>+</sup> (potassium stimulation). In our experimental setup this K<sup>+</sup> concentration has yielded the best and most reproducible responses in GABA and taurine release [16]. Under experimental conditions, designated as "ischemia," the glucose-free medium was bubbled with N<sub>2</sub> gas. The effluent fractions were subjected to amino acid assays by highperformance liquid chromatography. The amino acids eluted were visualized by means of o-phthaldialdehyde reagent and the results quantified with both external commercial standards and the internal standard of diamino-N-butyrate, as described in detail by Oja and Kontro [17]. The efflux of amino acids is either shown as a function of superfusion time (GABA, glutamate, and taurine) or calculated as an average

efflux for the period of 32 to 50 min (glycine, glutamine, aspartate, alanine, serine, and threonine).

2.3. Statistical Calculations. The presence of statistically significant differences between the sample means was detected by the two-way analysis of variance. Comparison of individual means was made by Hartley's sequential method of testing.

#### 3. Results

3.1. Cerebral Cortex. In 7-day-old mice taurine is the most prominent amino acid, followed by glutamate and aspartate (Table 1). The concentration of taurine decreases dramatically as the mice get older, when glutamate is the most abundant amino acid. The concentrations of glycine, aspartate, alanine, serine, and threonine are also significantly lower in 3-month-old mice, but no marked changes occurred in GABA, glutamate, and glutamine. It should be noted that the concentrations given in Table 1 represent the amino acid levels in the preincubated slices at the onset of superfusion, not the original tissue content.

The basal release of GABA was rather negligible in both 7-day- and 3-month-old mice, but K<sup>+</sup> stimulation evoked marked release at both ages (Figure 2). The magnitude of the evoked release was however about fourfold greater in adult mice. Ischemia induced GABA release at both ages, more so in adults. K<sup>+</sup> stimulation was clearly also preserved in ischemia in both age groups. In developing mice the release was stable during the experiments but in adults it was attenuated with prolonged ischemia (Table 2). In 7-day-old mice there occurred no enhancement of glutamate release upon K<sup>+</sup> stimulation either from oxygenated or ischemic slices (Figure 3). However, the release was increased by ischemia at both ages. The increase was more pronounced during the early phase in developing mice (Table 2). In 3month-old mice K<sup>+</sup> stimulation was marked under normal incubation conditions. K<sup>+</sup> stimulation was also effective in ischemia but the enhancement was less in magnitude. The basal release of taurine was markedly greater than that of GABA or glutamate (Figure 4). The release was increased severalfold when slices from 7-day-old mice were exposed to high-K<sup>+</sup> medium, whereas the stimulation was rather small in 3-month olds. The release of taurine was markedly increased in ischemia in both developing and adult mice, but no K<sup>+</sup>-stimulated release was discernible at either age. In developing mice the release was somewhat diminished with prolonged ischemia (Table 2).

In 7-day-old mice no effects of  $K^+$  stimulation or ischemia were seen in the release of aspartate, glycine, glutamine, alanine, and serine (Figure 5). On the other hand, aspartate release was enhanced by both treatments in 3-month-old mice, the combined effect of ischemia and  $K^+$  stimulation being particularly prominent. The release of glycine was also increased when slices from adult mice were exposed to  $K^+$ stimulation and ischemia, whereas the release of glutamine was diminished. No effects were seen in the release of alanine, serine, and threonine.

Amino acid (mmol/kg)	Cerebral cortex		Cerebellum				
	7-day olds	3-month olds	7-day olds	3-month olds			
Taurine	296.19 ± 23.41	$26.81 \pm 2.75^*$	$194.37 \pm 12.71$	$13.17 \pm 3.06^{*}$			
GABA	$16.38 \pm 1.13$	$18.41 \pm 1.36$	$13.25 \pm 1.80$	$16.03 \pm 1.88$			
Glycine	$24.17 \pm 7.42$	$3.82 \pm 0.27^{*}$	$18.23 \pm 1.49$	$8.52\pm1.08^*$			
Aspartate	$57.82 \pm 4.25$	$20.92 \pm 1.54^{*}$	$50.26 \pm 2.64$	$17.78 \pm 1.90^{*}$			
Glutamate	$75.13 \pm 5.71$	$64.99 \pm 3.38$	$54.82 \pm 3.18$	$53.70\pm6.40$			
Glutamine	$7.54\pm0.68$	$6.53 \pm 1.03$	$9.58 \pm 1.60$	$6.26 \pm 1.27$			
Alanine	$10.34 \pm 3.38$	$3.62 \pm 0.29^{*}$	$8.50\pm0.38$	$4.99\pm0.74^*$			
Serine	$21.33 \pm 9.59$	$5.58\pm0.40^*$	$9.93 \pm 0.38$	$9.55 \pm 1.62$			
Threonine	$5.93 \pm 2.06$	$2.13 \pm 0.16^{*}$	$5.31 \pm 0.20$	$3.11\pm0.84^*$			

TABLE 1: Amino acid contents in slices.

Amino acid concentrations (mean values  $\pm$  SEM) are given in mmol/kg protein in preincubated slices; that is, they show the amino acid contents before the onset of superfusion. Number of independent experiments 8 (cerebral cortex) and 16 (cerebellum). Significance of differences between 7-day-old and 3-month-old mice: \*P < 0.01.

TABLE 2: Early and late ischemia-induced release of GABA, glutamate, and taurine.

	7-day-	7-day-old mice mmol/min/kg protein Release period		3-month-old mice mmol/min/kg protein Release period	
Amino acid	mmol/mi				
	Releas				
	20-30 min	32–50 min	20-30 min	32-50 min	
Cerebral cortex					
GABA	$0.108 \pm 0.002$	$0.104\pm0.001$	$0.429 \pm 0.007$	$0.235 \pm 0.024^{*}$	
Glutamate	$0.282\pm0.010$	$0.171 \pm 0.012^{*}$	$0.671 \pm 0.057$	$0.870 \pm 0.015^{*}$	
Taurine	$3.357 \pm 0.164$	$2.078 \pm 0.100^{*}$	$1.112 \pm 0.061$	$1.050\pm0.038$	
Cerebellum					
GABA	$0.055 \pm 0.001$	$0.051 \pm 0.001$	$0.384 \pm 0.020$	$0.265 \pm 0.011^{*}$	
Glutamate	$0.044\pm0.001$	$0.046 \pm 0.001$	$0.229 \pm 0.029$	$0.082 \pm 0.014^{*}$	
Taurine	$0.859 \pm 0.028$	$0.933 \pm 0.022$	$0.826 \pm 0.015$	$0.687 \pm 0.018^{*}$	

The average release rates ( $\pm$ SEM) were calculated for the release periods indicated. Number of independent experiments 8 and release estimates 5 (20–30 min) and 10 (32–50 min). Significance of differences between the early and late release periods: \*P < 0.01.

*3.2. Cerebellum.* Taurine is also the most prominent amino acid in the cerebellum of 7-day-old mice (Table 1). In 3-month olds glutamate is present at the highest concentration, followed by aspartate, glycine, and taurine. In the cerebellum the concentrations of GABA, glutamate, glutamine, and serine are at about the same level in both 7-day-old and 3-month-old mice, whereas the levels of taurine, glycine, aspartate, alanine, and threonine are lower in 3-month olds than in 7-day olds.

There occurred only hardly detectable basal release of GABA from cerebellar slices from 7-day-old mice (Figure 1).  $K^+$  stimulation also evoked almost 10-fold greater GABA release in 3-month-old than in 7-day-old mice. Ischemia enhanced the release, and  $K^+$  stimulation was preserved in ischemic slices at both ages. However, the effects were much greater in adults in which the ischemia-induced enhancement was diminished with prolonged experiments (Table 2). The basal release of glutamate and the  $K^+$  stimulation were likewise severalfold greater in 3-month-old than in 7-day-old mice (Figure 6). The release of glutamate was enhanced in ischemia, but no  $K^+$ -evoked release was discernible in adults. The ischemia-induced enhancement of release was greatly attenuated with time in adults (Table 2). The release of taurine

increased several-fold in 7-day-old mice in high- $K^+$  media, whereas  $K^+$  stimulation was rather small in magnitude in adults (Figure 7). The increase in release in ischemia was fairly stable during the experiments at both ages (Table 2). In ischemia  $K^+$  stimulation was partially preserved in 7-day-old mice, whereas no  $K^+$  effect was seen in 3-month-old mice (Figure 7).

Ischemia increased the release of glycine, glutamine, alanine, serine, and threonine in 7-day-old mice, but  $K^+$  stimulation had no effect on the release of these amino acids in ischemia (Figure 8). Aspartate release was in all cases somewhat marginal at this age. In 3-month-old mice no effects of  $K^+$  stimulation or ischemia were seen in the release of aspartate, glycine, glutamine, alanine, serine, or threonine.

### 4. Discussion

4.1. Mechanisms of Release. Although the amino acid levels in preincubated slices are not identical to the concentrations in intact tissue, the contents measured here reflect those obtaining in vivo. A characteristic finding is the very high taurine concentration in the developing mouse brain. In spite of the much higher content in the developing cerebral



FIGURE 1: Time course of basal (-O-) and (- $\bullet$ -) K<sup>+</sup>-evoked GABA release from cerebellar slices in normoxia and basal (- $\Delta$ -) and K<sup>+</sup>-stimulated (-A-) GABA release in ischemia in 7-day-old (a) and 3-month-old (b) mice. The results are means of 4–8 independent experiments with SEMs indicated. Note the threefold differences in the scale of the *y*-axis in panels (a) and (b). Ischemia significantly (*P* < 0.01) enhanced the release at both ages. K<sup>+</sup> stimulation was likewise significantly effective in both cases.



FIGURE 2: Time course of basal (-O-) and K<sup>+</sup>-evoked (- $\bullet$ -) GABA release from cerebral cortical slices in normoxia and basal (- $\Delta$ -) and K<sup>+</sup>stimulated (- $\bullet$ -) GABA release in ischemia in 7-day-old (a) and 3-month-old (b) mice. The results are means of 4–8 independent experiments with SEMs indicated. Ischemia significantly (*P* < 0.01) enhanced the release at both ages. K<sup>+</sup> stimulation was likewise significantly effective. Note the threefold difference in the scale of the *y*-axis in panels (a) and (b).



FIGURE 3: Time course of basal (-O-) and K<sup>+</sup>-evoked (- $\bullet$ -) glutamate release from cerebral cortical slices in normoxia and basal (- $\Delta$ -) and K<sup>+</sup>-stimulated (- $\blacktriangle$ -) glutamate release in ischemia in 7-day-old (a) and 3-month-old (b) mice. The results are means of 4–8 independent experiments with SEMs indicated. Ischemia significantly (P < 0.01) enhanced the release in both age groups, but K<sup>+</sup> stimulation was effective only in normoxia in both. Note the fivefold difference in the scale of the *y*-axis in panels (a) and (b).

cortex and cerebellum, the basal release rates of endogenous taurine were not markedly different, but the high content to a great extent obviously underlies the several-fold greater release evoked by  $K^+$  stimulation and ischemia. These effects were clearly more marked in the cerebral cortex than in the cerebellum in both experimental groups. However, the differences in concentration of other amino acids between developing and adult mice are also in any case of such magnitude that they will have affected the present estimated release rates.

Ischemia induced the release of neuroactive amino acids in all brain areas studied. This release could result, for instance, from destabilization and deterioration of plasma membranes owing to phospholipid hydrolysis by phospholipases, resulting in diffusion of amino acids along their concentration gradients [18]. However, in ischemia here there occurred no measurable increase in the extrusion of lactate dehydrogenase (a common marker of plasma membrane damage and nonspecific lysis of neural cells) from the slices upon incubation (data not shown). Nor has it been increased in other experiments of the present type [19, 20]. However, longer periods of ischemia are associated with increasing degrees of plasma membrane disruption, allowing for a greater leakage of intracellular contents [21]. In keeping with the assumption that deterioration of cell plasma membranes may not have been a major factor underlying amino acid release, there now occurred no increase in the release of the nontransmitter amino acids, alanine, serine, and threonine in

the cerebral cortex in ischemia in either experimental group. However, the release of all these amino acids increased in ischemia in slices prepared from the developing cerebellum. It is thus reasonable to surmise that any major damage to neural membranes will not have been involved in the present release rates of amino acids, with a reservation to the preparations from the developing cerebellum.

The opening of swelling-induced  $Cl^-$  channels also allows the passage of amino acids down their concentration gradients. Edema is a frequent consequence of cerebral ischemia [22]. When cells swell, they attempt to restore their normal volume by extruding osmotically active solutes such as amino acids [23]. In particular, release of taurine has often been shown to be evoked by cell swelling in the cerebral cortex, exhibiting typically a delayed time course [24, 25], similar to those obtained in the present study. Moreover, the apparent release of amino acids from slices also depends on the efficacy of reuptake. The amino acids released must traverse the extracellular spaces to reach the medium and be detected. This is the reason why uptake mechanisms also affect the estimated release rates.

In addition to the cell plasma membrane damage discussed previously, and in the case of neurotransmitter amino acids  $Ca^{2+}$ -dependent exocytosis from synaptic vesicles [26], the mechanisms involved in the release of amino acids in ischemia may include swelling-evoked release via anion channels [27], reversal of amino acid transporters in depolarized cells [28], or acidosis-induced failure of reuptake



FIGURE 4: Time course of basal (-O-) and K<sup>+</sup>-evoked (- $\bullet$ -) taurine release from cerebral cortical slices in normoxia and basal (- $\Delta$ -) and K<sup>+</sup>stimulated (- $\bullet$ -) taurine release in ischemia in 7-day-old (a) and 3-month-old (b) mice. The results are means of 4–8 independent experiments with SEMs indicated. Ischemia significantly (P < 0.01) enhanced the release in both age groups, but K<sup>+</sup> stimulation was similarly effective only in normoxia, the effect being less pronounced (P < 0.05) in ischemia. Note the 2.5-fold difference in the scale of the *y*-axis in panels (a) and (b).

into neurons and glia [29]. In particular, the release of glutamate and aspartate has been postulated to involve, in addition to  $Ca^{2+}$ -dependent exocytotic release,  $Ca^{2+}$ -independent release due to a depolarization-induced reversal of the Na<sup>+</sup>-dependent high affinity acidic amino acid transporters [30, 31].  $Ca^{2+}$ -dependent release may account for the initial efflux of neurotransmitter amino acids at the onset of ischemia, whereas  $Ca^{2+}$ -independent nonvesicular release, mediated by Na<sup>+</sup>-dependent amino acid transporters in plasma membranes operating in a reversed mode, could be mostly responsible for the later phase of release [32]. This may signify that the release of glutamate from cerebellar slices could have been due mainly to exocytosis in the present experiments, whereas in cerebral cortical slices the function of transporters may have markedly contributed.

4.2. Ischemia and Excitotoxicity. Excessive release of the excitotoxic amino acids glutamate and aspartate is considered to be a significant contributor to neuronal death in the ischemic brain [33, 34]. There is evidence for a positive feedback loop, in which glutamatergic neurons can be stimulated to release additional glutamate as a consequence of activation of its receptors [35, 36]. In particular, the N-methyl-D-aspartate (NMDA) receptors may occupy a central position here, since they have been shown to regulate the release of both excitatory and inhibitory amino acids from rat

fetal cortical neurons [37]. In human cerebral cortical slices K<sup>+</sup> stimulation and ischemia have also markedly enhanced the release of glutamate, aspartate, GABA, and glycine, but not that of glutamine. The K<sup>+</sup>-evoked release was Ca<sup>2+</sup>dependent, whereas the ischemia-induced release was not [38]. Substantial amounts of glutamate were now released in the adult cerebral cortex under ischemic conditions, and the K<sup>+</sup> stimulation was partially preserved, but the release could not be described as massive in comparison with the release of GABA or taurine. The release of glutamate was markedly smaller in the developing cerebral cortex. In the cerebellum the release was likewise greater in adults, being however significantly smaller than in the cerebral cortex and rapidly attenuated with time. This finding may indicate that the adult cerebral cortex is more susceptible to excitotoxicity. The marked release of aspartate, in particular with the concomitant K<sup>+</sup> stimulation, could contribute to this effect in the cerebral cortex, but not in the cerebellum.

The release of GABA from cerebral cortical slices was greater that that from cerebellar slices in both experimental groups. In keeping with this finding, glutamate decarboxylase-positive cell bodies, that is, GABAergic neuronal density, in the rat neocortex are high already at birth [39]. On the other hand, in adult mice GABA release was of the same order of magnitude in both brain areas. In adults, the release of glutamate was significantly greater than that of GABA. In the adult cortex both these neurotransmitters



FIGURE 5: Average release of aspartate, glycine and glutamine (a) and alanine, serine and threonine (c) from cerebral cortical slices from 7-day-old mice and from cerebral cortical slices from 3-month-old mice ((b) and (d)) during the superfusion period of 32 to 50 min. The first open bars show the basal, and the second right-hatched bars the K<sup>+</sup>-stimulated release in normoxia, the third left-hatched bars the basal, and the fourth cross-hatched bars the K<sup>+</sup>-stimulated release in ischemia. The results are mean values  $\pm$  SEM of 4–8 independent experiments. Note the twofold differences in the scale of the *y*-axes in panels (a), (b), (c), and (d). The statistically significant difference from the corresponding unstimulated release in normoxia: \*\* *P* < 0.01.

function in the vast majority of all synapses, but the glutamatergic synapses outnumber the GABA synapses about fivefold [40]. The greater release of glutamate thus reflects this relation. In cultured rat cerebellar neurons the release of GABA and taurine has been shown to originate from the GABAergic interneurons, the basket and stellate cells, and the release of glutamate and aspartate mainly from the granule neurons [41]. In cultured neurons from the rat cerebellum the glutamate receptor agonists kainate and quisqualate have evoked the release of endogenous glutamate, taurine, and GABA in a dose- and Ca2+-dependent manner [42, 43]. Endogenous aspartate, glutamate, and GABA are also released from such cultures by K<sup>+</sup> depolarization in a doseand Ca<sup>2+</sup>-dependent manner whereas the release of taurine is dose-dependent but not Ca<sup>2+</sup>-dependent [44]. In contrast, the omission of Ca<sup>2+</sup> has enhanced basal taurine release, a finding also recorded, for example, in cerebral cortical slices and in hippocampal slices prepared from young, adult, and aged mice [45, 46].

The greater glycine release from cortical slices from developing than from adult mice may be associated, in addition to the high glycine content, with the presence of glycine receptors in the cerebral cortex during early development [47, 48]. In particular, the  $\alpha$ 2 subunit of glycine receptors declines sharply following the first postnatal week and remains at a low level in the adult neocortex [47, 49, 50]. Moreover, the glycine sensitivity of neurons in the rat prefrontal cortex has been shown to decrease with age [51], and functional glycine receptors in older animals have not been demonstrated. On the other hand, the expression of glycine receptors in the cerebellum is rather weak and does not show a similar decline with maturation [52].

Taurine acts at both GABA and glycine receptors, being however less effective than either of these [3, 53]. It therefore acts as an inhibitory agent in the central nervous system [4]. The release of taurine in the developing cerebral cortex and cerebellum in ischemia is of such magnitude that it could clearly counteract the effects of excitatory amino acid



FIGURE 6: Time course of basal (-O-) and K<sup>+</sup>-evoked (- $\bullet$ -) glutamate release from cerebellar slices in normoxia and basal (- $\Delta$ -) and K<sup>+</sup>stimulated (- $\bullet$ -) glycine release in ischemia in 7-day-old (a) and 3-month-old (b) mice. Note the fourfold differences in the scale of the *y*-axis in panels (a) and (b). The results are means of 4–8 independent experiments with SEMs indicated. Ischemia significantly (*P* < 0.01) enhanced the release in both age groups, but K<sup>+</sup> stimulation was effective only in ischemia in 7-day-old mice and in normoxia in 3-month-old mice.



FIGURE 7: Time course of basal (-O-) and K<sup>+</sup>-evoked (- $\bullet$ -) taurine release from cerebellar slices in normoxia and basal (- $\Delta$ -) and K<sup>+</sup>-stimulated (- $\bullet$ -) taurine release in ischemia in 7-day-old (a) and 3-month-old (b) mice. The results are means of 4–8 independent experiments with SEMs indicated. Ischemia significantly (*P* < 0.01) enhanced the release in both age groups, but K<sup>+</sup> stimulation had no significant effect in ischemia in 3-month-old mice.



FIGURE 8: Average release of aspartate, glycine and glutamine (a) and alanine, serine and threonine (c) from cerebellar slices from 7-day-old mice and from cerebellar slices from 3-month-old mice ((b) and (d)) during the superfusion period of 32 to 50 min. The first open bars show the basal and the second right-hatched bars the K<sup>+</sup>-stimulated release in normoxia, the third left-hatched bars the basal, and the fourth cross-hatched bars the K<sup>+</sup>-stimulated release in normoxia, the third left-hatched bars the basal, and the fourth cross-hatched bars the K<sup>+</sup>-stimulated release in schemia. The results are mean values  $\pm$  SEM of 4–8 independent experiments. The statistically significant difference from the corresponding unstimulated release in normoxia: \**P* < 0.05; \*\**P* < 0.01. Note the differences in the scale of the *y*-axes.

transmitters, in particular since at that age the release of glutamate and aspartate cannot be described to be massive. GABA and glycine cannot protect against excitotoxicity in the developing cerebral cortex and cerebellum, since GABA is during early development rather excitatory than inhibitory [54, 55]. Glycine receptors also tend to mediate excitation in the neonatal rat cerebral cortex [56, 57].

In conclusion, we may state that the releases from the cerebral cortex and cerebellum were markedly different and also differed between developing and adult mice. In developing mice with 7 days of age, only the release of inhibitory taurine may be large enough to counteract the harmful effects of excitatory amino acids in ischemia in both cerebral cortex and cerebellum, in particular since at that age the release of glutamate and aspartate cannot be described as massive. The present experiments in which rather long exposure times were studied cannot reveal any fast or transient effects on amino acid release. Moreover, it is open to what extent the results obtained are applicable to the human brain, though the basic mechanisms are likely similar in the mouse and the man.

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## References

- G. E. Fagg and A. C. Foster, "Amino acid neurotransmitters and their pathways in the mammalian central nervous system," *Neuroscience*, vol. 9, no. 4, pp. 701–719, 1983.
- [2] M. Darstein, G. B. Landwehrmeyer, C. Kling, C. M. Becker, and T. J. Feuerstein, "Strychnine-sensitive glycine receptors in rat caudatoputamen are expressed by cholinergic interneurons," *Neuroscience*, vol. 96, no. 1, pp. 33–39, 2000.
- [3] O. A. Sergeeva and H. L. Haas, "Expression and function of glycine receptors in striatal cholinergic interneurons from rat and mouse," *Neuroscience*, vol. 104, no. 4, pp. 1043–1055, 2001.
- [4] P. Saransaari and S. S. Oja, "Taurine in neurotransmission," in *Handbook of Neurochemistry and Molecular Neurobiology: Neurotransmitter Systems*, A. Lajtha and S. E. Vizi, Eds., vol. 3rd, pp. 326–341, Springer, New York, NY, USA, 2008.

- [5] P. Klivenyi, K. A. Kekesi, Z. Hartai, G. Juhasz, and L. Vecsei, "Effects of mitochondrial toxins on the brain amino acid concentrations," *Neurochemical Research*, vol. 30, no. 11, pp. 1421–1427, 2005.
- [6] S. P. Butcher, R. Bullock, D. I. Graham, and J. McCulloch, "Correlation between amino acid release and neuropathologic outcome in rat brain following middle cerebral artery occlusion," *Stroke*, vol. 21, no. 12, pp. 1727–1733, 1990.
- [7] D. L. Small, P. Morley, and A. M. Buchan, "Biology of ischemic cerebral cell death," *Progress in Cardiovascular Diseases*, vol. 42, no. 3, pp. 185–207, 1999.
- [8] M. Szatkowski and D. Attwell, "Triggering and execution of neuronal death in brain ischaemia: two phases of glutamate release by different mechanisms," *Trends in Neurosciences*, vol. 17, no. 9, pp. 359–365, 1994.
- [9] P. Morley, M. J. Hogan, and A. M. Hakim, "Calcium-mediated mechanisms of ischemic injury and protection," *Brain Pathol*ogy, vol. 4, no. 1, pp. 37–47, 1994.
- [10] P. Saransaari and S. S. Oja, "Enhanced GABA release in cell-damaging conditions in the adult and developing mouse hippocampus," *International Journal of Developmental Neuroscience*, vol. 15, no. 2, pp. 163–174, 1997.
- [11] P. Saransaari and S. S. Oja, "Release of endogenous glutamate, aspartate, GABA, and taurine from hippocampal slices from adult and developing mice under cell-damaging conditions," *Neurochemical Research*, vol. 23, no. 4, pp. 563–570, 1998.
- [12] P. Andine, M. Sandberg, R. Bagenholm, A. Lehmann, and H. Hagberg, "Intra- and extracellular changes of amino acids in the cerebral cortex of the neonatal rat during hypoxic-ischemia," *Developmental Brain Research*, vol. 64, no. 1-2, pp. 115–120, 1991.
- [13] T. Kubo, A. Takano, N. Tokushige, N. Miyata, M. Sato, and S. Hatakeyama, "Electrical stimulation-evoked release of endogenous taurine from slices of the hippocampus, cerebral cortex and cerebellum of the rat," *Journal of Pharmacobio-Dynamics*, vol. 15, no. 9, pp. 519–525, 1992.
- [14] H. Huang, L. Barakat, D. Wang, and A. Bordey, "Bergmann glial GlyT1 mediates glycine uptake and release in mouse cerebellar slices," *Journal of Physiology*, vol. 560, no. 3, pp. 721–736, 2004.
- [15] L. A. C. Reno, W. Zago, and R. P. Markus, "Release of [<sup>3</sup>H]-Lglutamate by stimulation of nicotinic acetylcholine receptors in rat cerebellar slices," *Neuroscience*, vol. 124, no. 3, pp. 647–653, 2004.
- [16] P. Kontro and S. S. Oja, "Taurine and GABA release from mouse cerebral cortex slices: potassium stimulation releases more taurine than GABA from developing brian," *Developmental Brain Research*, vol. 37, no. 1-2, pp. 277–291, 1987.
- [17] S. S. Oja and P. Kontro, "Release of endogenous taurine and γ-aminobutyric acid from brain slices from the adult and developing mouse," *Journal of Neurochemistry*, vol. 52, no. 4, pp. 1018–1024, 1989.
- [18] J. W. Phillis and M. H. O'Regan, "Mechanisms of glutamate and aspartate release in the ischemic rat cerebral cortex," *Brain Research*, vol. 730, no. 1-2, pp. 150–164, 1996.
- [19] D. E. Pellegrini-Giampietro, G. Cherici, M. Alesiani, V. Carla, and F. Moroni, "Excitatory amino acid release and free radical formation may cooperate in the genesis of ischemia-induced neuronal damage," *Journal of Neuroscience*, vol. 10, no. 3, pp. 1035–1041, 1990.
- [20] G. G. Haddad and C. Jiang, "O<sub>2</sub> deprivation in the central nervous system: on mechanisms of neuronal response, differential

sensitivity and injury," *Progress in Neurobiology*, vol. 40, no. 3, pp. 277–318, 1993.

- [21] J. W. Phillis, M. Smith-Barbour, and M. H. O'Regan, "Changes in extracellular amino acid neurotransmitters and purines during and following ischemias of different durations in the rat cerebral cortex," *Neurochemistry International*, vol. 29, no. 2, pp. 115–120, 1996.
- [22] Y. Katayama, T. Tamura, D. P. Becker, and T. Tsubokawa, "Early cellular swelling during cerebral ischemia in vivo is mediated by excitatory amino acids released from nerve terminals," *Brain Research*, vol. 577, no. 1, pp. 121–126, 1992.
- [23] R. O. Law, "Amino acids as volume-regulatory osmolytes in mammalian cells," *Comparative Biochemistry and Physiology A*, vol. 99, no. 3, pp. 263–277, 1991.
- [24] E. R. Korpi, P. Kontro, and K. Nieminen, "Spontaneous and depolarization-induced efflux of hypotaurine from mouse cerebral cortex slices: comparison with taurine and GABA," *Life Sciences*, vol. 29, no. 8, pp. 811–816, 1981.
- [25] A. Y. Estevez, D. Song, J. W. Phillis, and M. H. O'Regan, "Effects of the anion channel blocker DIDS on ouabain- and high K<sup>+</sup>induced release of amino acids from the rat cerebral cortex," *Brain Research Bulletin*, vol. 52, no. 1, pp. 45–50, 2000.
- [26] D. Nicholls and D. Attwell, "The release and uptake of excitatory amino acids," *Trends in Pharmacological Sciences*, vol. 11, no. 11, pp. 462–468, 1990.
- [27] J. W. Phillis, D. Song, and M. H. O'Regan, "Inhibition by anion channel blockers of ischemia-evoked release of excitotoxic and other amino acids from rat cerebral cortex," *Brain Research*, vol. 758, no. 1-2, pp. 9–16, 1997.
- [28] R. A. Swanson, J. Chen, and S. H. Graham, "Glucose can fuel glutamate uptake in ischemic brain," *Journal of Cerebral Blood Flow and Metabolism*, vol. 14, no. 1, pp. 1–6, 1994.
- [29] M. C. Longuemare and R. A. Swanson, "Excitatory amino acid release from astrocytes during energy failure by reversal of sodium-dependent uptake," *Journal of Neuroscience Research*, vol. 40, no. 3, pp. 379–386, 1995.
- [30] D. G. Nicholls, T. S. Sihra, and J. Sanchez-Prieto, "Calciumdependent and -independent release of glutamate from synaptosomes monitored by continuous fluorometry," *Journal of Neurochemistry*, vol. 49, no. 1, pp. 50–57, 1987.
- [31] M. Ikeda, T. Nakazawa, K. Abe, T. Kaneko, and K. Yamatsu, "Extracullular accumulation of glutamate in the hippocampus induced by ischemia is not calcium dependent - In vitro an in vivo evidence," *Neuroscience Letters*, vol. 96, no. 2, pp. 202–206, 1989.
- [32] J. W. Phillis and M. H. O'Regan, "Characterization of modes of release of amino acids in the ischemic/reperfused rat cerebral cortex," *Neurochemistry International*, vol. 43, no. 4-5, pp. 461–467, 2003.
- [33] H. Benveniste, "The excitotoxin hypothesis in relation to cerebral ischemia," *Cerebrovascular and Brain Metabolism Reviews*, vol. 3, no. 3, pp. 213–245, 1991.
- [34] D. W. Choi, "Excitotoxic cell death," *Journal of Neurobiology*, vol. 23, no. 9, pp. 1261–1276, 1992.
- [35] S. Carboni, F. Melis, L. Pani, M. Hadjiconstantinou, and Z. L. Rossetti, "The non-competitive NMDA-receptor antagonist MK-801 prevents the massive release of glutamate and aspartate from rat striatum induced by 1-methyl-4-phenylpyridinium (MPP<sup>+</sup>)," *Neuroscience Letters*, vol. 117, no. 1-2, pp. 129–133, 1990.

- [36] G. Bustos, J. Abarca, M. I. Forray, K. Gysling, C. W. Bradberry, and R. H. Roth, "Regulation of excitatory amino acid release by *N*-methyl-*D*-aspartate receptors in rat striatum: in vivo microdialysis studies," *Brain Research*, vol. 585, no. 1-2, pp. 105–115, 1992.
- [37] E. López, J. Hernandez, C. Arce, S. Cañadas, M. J. Oset-Gasque, and M. P. González, "Involvement of NMDA receptor in the modulation of excitatory and inhibitory amino acid neurotransmitters release in cortical neurons," *Neurochemical Research*, vol. 35, no. 9, pp. 1478–1486, 2010.
- [38] E. Hegstad, J. Berg-Johnsen, T. S. Haugstad, E. Hauglie-Hanssen, and I. A. Langmoen, "Amino-acid release from human cerebral cortex during simulated ischaemia in vitro," *Acta Neurochirurgica*, vol. 138, no. 2, pp. 234–241, 1996.
- [39] F. Morin and C. Beaulieu, "Equivalent cell density in three areas of neonatal rat cerebral cortex," *Neuroscience Letters*, vol. 176, no. 1, pp. 85–88, 1994.
- [40] C. Beaulieu and M. Colonnier, "A laminar analysis of the number of round-asymmetrical and flat- symmetrical synapses on spines, dendritic trunks, and cell bodies in area 17 of the cat," *Journal of Comparative Neurology*, vol. 231, no. 2, pp. 180–189, 1985.
- [41] M. L. Simmons and G. R. Dutton, "Neuronal origins of K<sup>+</sup>evoked amino acid release from cerebellar cultures," *Journal of Neuroscience Research*, vol. 31, no. 4, pp. 646–653, 1992.
- [42] K. L. Rogers, R. A. Philibert, and G. R. Dutton, "Glutamate receptor agonists cause efflux of endogenous neuroactive amino acids from cerebellar neurons in culture," *European Journal of Pharmacology*, vol. 177, no. 3, pp. 195–199, 1990.
- [43] G. Levi, M. Patrizio, and V. Gallo, "Release of endogenous and newly synthesized glutamate and of other amino acids induced by non-N-methyl-D-aspartate receptor activation in cerebellar granule cell cultures," *Journal of Neurochemistry*, vol. 56, no. 1, pp. 199–206, 1991.
- [44] K. L. Rogers, R. A. Philibert, and G. R. Dutton, "K<sup>+</sup>-stimulated amino acid release from cultured cerebellar neurons: comparison of static and dynamic stimulation paradigms," *Neurochemical Research*, vol. 16, no. 8, pp. 899–904, 1991.
- [45] S. S. Oja and P. Saransaari, "Taurine release and swelling of cerebral cortex slices from adult and developing mice in media of different ionic compositions," *Journal of Neuroscience Research*, vol. 32, no. 4, pp. 551–561, 1992.
- [46] P. Saransaari and S. S. Oja, "Enhanced taurine release in celldamaging conditions in the developing and ageing mouse hippocampus," *Neuroscience*, vol. 79, no. 3, pp. 847–854, 1997.
- [47] M. L. Malosio, B. Marqueze-Pouey, J. Kuhse, and H. Betz, "Widespread expression of glycine receptor subunit mRNAs in the adult and developing rat brain," *EMBO Journal*, vol. 10, no. 9, pp. 2401–2409, 1991.
- [48] A. C. Flint, X. Liu, and A. R. Kriegstein, "Nonsynaptic glycine receptor activation during early neocortical development," *Neuron*, vol. 20, no. 1, pp. 43–53, 1998.
- [49] K. Sato, H. Kiyama, and M. Tohyama, "Regional distribution of cells expressing glycine receptor α2 subunit mRNA in the rat brain," *Brain Research*, vol. 590, no. 1-2, pp. 95–108, 1992.
- [50] S. Jonsson, N. Kerekes, P. Hyytiä, M. Ericson, and B. Söderpalm, "Glycine receptor expression in the forebrain of male AA/ANA rats," *Brain Research*, vol. 1305, supplement, pp. S27–S36, 2009.
- [51] Y. Lu and J. H. Ye, "Glycine-activated chloride currents of neurons freshly isolated from the prefrontal cortex of young rats," *Brain Research*, vol. 1393, pp. 17–22, 2011.

- [52] G. García-Alcocer, C. Mejía, L. C. Berumen, R. Miledi, and A. Martínez-Torres, "Developmental expression of glycine receptor subunits in rat cerebellum," *International Journal of Developmental Neuroscience*, vol. 26, pp. 3119–3322, 2008.
- [53] S. Oja, E. R. Korpi, and P. Saransaari, "Modification of chloride flux across brain membranes by inhibitory amino acids in developing and adult mice," *Neurochemical Research*, vol. 15, no. 8, pp. 797–804, 1990.
- [54] Y. Ben-Ari, "Excitatory actions of GABA during development: the nature of the nurture," *Nature Reviews Neuroscience*, vol. 3, no. 9, pp. 728–739, 2002.
- [55] A. T. Gulledge and G. J. Stuart, "Excitatory actions of GABA in the cortex," *Neuron*, vol. 37, no. 2, pp. 299–309, 2003.
- [56] W. Kilb, I. L. Hanganu, A. Okabe et al., "Glycine receptors mediate excitation of subplate neurons in neonatal rat cerebral cortex," *Journal of Neurophysiology*, vol. 100, no. 2, pp. 698–707, 2008.
- [57] J. H. Ye, "Regulation of excitation by glycine receptors," *Results and Problems in Cell Differentiation*, vol. 44, pp. 123–143, 2008.