

Major Article

Monitoring *Rhodnius neglectus* (Lent, 1954) populations' susceptibility to insecticide used in controlling actions in urban areas northwest of São Paulo state

Rubens Antonio da Silva^[1][®], Lis Adriana Maldonado^[1][®], Grasielle Caldas D'Ávila Pessoa^[2][®] and Liléia Diotaiuti^[3][®]

[1]. Secretaria de Estado da Saúde de São Paulo, Superintendência de Controle de Endemias, Laboratório Especializado de Mogi Guaçu: Triatomíneos, São Paulo, SP, Brasil.
[2]. Universidade Federal de Minas Gerais, Departamento de Parasitologia-ICB, Laboratório de Fisiologia de Insetos Hematófagos, Belo Horizonte, MG, Brasil.
[3]. Eurodação Oswaldo Cruz, Instituto René Pachou, Grupo de Pasquica Triatomíneos, Relo Horizonte, MG, Brasil.

[3]. Fundação Oswaldo Cruz, Instituto René Rachou, Grupo de Pesquisa Triatomíneos, Belo Horizonte, MG, Brasil.

ABSTRACT

Background: Chagas disease (CD) is caused by the flagellate protozoan *Trypanosoma cruzi* and can be carried by different species of triatomines, including *Rhodnius neglectus*, which is wild, well distributed in Brazil, and has formed colonies in palm trees located in urban areas of municipalities in the state of São Paulo. Chemical control has been routinely used to reduce population density, but each year, there has been an increase in species dispersion and density. This study aimed to evaluate the susceptibility of insects to insecticides used in control.

Methods: The reference population was collected from Araçatuba municipality, Nilce Maia. Dilutions of deltamethrin were prepared and applied to the back of the first-stage nymphs, which were biologically synchronized. The control group received pure acetone only. Mortality was assessed after 72 h.

Results: The mortality rate with respect to diagnostic dose was 100%. The susceptibility profile observed for this population showed RR₅₀ ranging from 1.76 to 3.632.

Conclusions: The populations were susceptible to the insecticides tested. It is possible that the insecticide residual effect on this ecotope has decreased the lifespan, and controlling failures may be the cause of recolonization in this environment.

Keywords: Insecticide resistance. Rhodnius neglectus. Deltamethrin. Chagas disease. Triatominae control.

INTRODUCTION

Chagas disease (CD) is caused by *Trypanosoma cruzi*, a flagellate protozoan. The infection occurs by contact with contaminated excrement of this triatomine vector, a hematophagous insect, in all of its developmental phases. Infection may also occur by intake of contaminated food, vertical transmission, blood transfusion, and organ transplant¹.

CD is endemic to 21 Latin American countries and affects approximately 6-7 million people worldwide². Recently, efforts have been made to reduce the occurrence of this infection in Latin

America, but high migration rates of individuals from this region to other places have enabled DCs to spread to non-endemic countries, thereby making CD a global health issue³.

São Paulo state was the pioneer in controlling the main vector species, *Triatoma infestans*, and the measures adopted have become a model for other states in Brazil and South American countries^{4,5}. Because of successful control measures in the domiciliary environment of *T. infestans*, entomological surveillance has focused on controlling native species that are usually found in the peridomiciliary environment⁶.

Corresponding author: Dr. Rubens Antonio da Silva. e-mail: rubensantoniosilva@gmail.com

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Conflict of Interest: The authors declare that there is no conflict of interest.

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A total of 18 genera and 154 species of triatomine are known⁷. In Brazil, 64 species have been registered, 13 of these in São Paulo state, including *Rhodnius neglectus* (Lent 1954), which is mainly found in Ribeirão Preto, São José do Rio Preto, and Araçatuba. *R. neglectus* is a wild species, and its natural ecotope consists of palm trees, where it finds shelter and food. However, recent studies have reported that this species is also found in urban areas, domiciles, and peridomiciles^{8,9}.

Palm trees belonging to the genus *Acrocomia* infested by *R. neglectus* feeding on *Pionnus maximiliani*, which make their nests in these palm trees, were detected in urban areas in Araçatuba, Birigui, Guararapes, and Piacatu municipalities, located in the northwest of São Paulo state⁹. Palm trees are used in landscaping design of urban areas in many cities of the state because they present low maintenance costs and are visually pleasant in the environment. The cities analyzed have a significant quantity of palm trees in their public squares and streets, thereby generating a favorable environment for these triatomines.

Controlling actions employed for palm trees located in urban areas consist of removing leaves and dry bunches, as well as chemical control by applying pyrethroid insecticide⁹. Despite this, every year, there is an increase in the dispersion and infestation of palm trees¹⁰. In cases of re-infestation after pyrethroid insecticide application, recolonization may occur due to insects that survived the spraying process, operational failures when applying the insecticide, or insect resistance to the product¹¹.

Susceptibility to insecticide resistance has already been studied for different species, but not for *R. neglectus*. Due to infestation persistence observed in these municipalities, where chemical control has been systemically employed for over 15 years, it is necessary to assess insect susceptibility to the insecticide used in controlling measures aiming to verify its efficacy, therefore guiding actions carried out by the teams involved in such actions.

METHODS

Triatomines for Assessment

Samples were obtained from Araçatuba, Birigui, Guararapes, and Piacatu municipalities, all of which belong to the Araçatuba administrative region, located northwest of São Paulo state (**Table 1**). The insects were collected from the urban areas of these municipalities from palm trees that were regularly treated with pyrethroid insecticides provided by the Brazilian Ministry of Health under systematic applications¹².

The insects were manually collected during the investigation procedure in the palm trees by municipal teams, kept in pots for transportation, and sent to the laboratory, where they were identified using a dichotomous key for triatomine species and classified according to their evolutionary phase¹³.

At the laboratory, they were kept in a breeding room with a constant temperature of 25 °C \pm 3, an air relative humidity of approximately 75% \pm 3, and a photoperiod of 12 h. The insects remained in crystallizers measuring 10 cm (height) x 23.5 cm (diameter), lined with filtering paper to absorb excrement and humidity, with a hive-shaped 2 mm thick pressed cardboard sheet support and covered with thick dark cotton to protect them from light. The insects were fed through an artificial method with the rustic chicken, *Rhodia sp.*, on a weekly basis.

Reference Population

A reference population of *R. neglectus* was selected, in which laboratory breeding started on 12/01/2004 and the insects were from Araçatuba municipality, Nilce Maia location. Specifically, it was necessary to identify the relevant diagnostic dose under which mortality of 50% of the population was observed (LD_{50}). The criteria for selecting the reference population followed the World Health Organization guidelines¹⁴.

Tests were performed to identify the LD₅₀ and LD₉₉ in this population. After reading the nymph death for the eight dilution rates used, the numbers obtained were organized in a reading database for the Polo Plus[®] software. RR₅₀ and RR₉₉ were obtained by dividing the LD₅₀ of the field population by the LD₅₀ of the susceptible population, with correspondence to calculate the RR₉₉. A confidence interval of 95% (CI 95%) of each population was also calculated for each population.

Insecticide Susceptibility Test

The methodology used to monitor insect resistance in the laboratory was standardized by Pessoa (2016)¹⁵ and defines insect generation, nymph age, insecticide application *locus*, and the ideal diagnostic dose for each species. The insects collected were kept

TABLE 1: Triatomine populations assessed in insecticide susceptibility test, by municipality and location. Araçatuba region, 2020.

Municipalities	Name of Location	Georeference	Insects
	João Arruda Brasil	-21.19291 -50.44186	300
Araçatuba	Alcides Chagas	-21.19555 -50.42841	210
	Escola Nilce Maia	-21.20652 -50.45417	270
Disiani	Pedro de Toledo	-21.28772 -50.34005	240
Birigui	James Mellor	-21.29140 -50.34022	240
	Princesa Isabel	-21.24231 -50.64913	270
	Eurídes Amaral	-21.24397 -50.65012	270
Current	Gaudêncio José Pereira	-21.24214 -50.65025	240
Guararapes	Rachel Caldas	-21.24389 -50.64647	240
	Praça Central	-21.25428 -50.64396	270
	Praça Mohamed	-21.25802 -50.64618	270
Diacotu	José Benetti	-21.58903 -50.59716	240
PidCalu	Câmara Municipal	-21.58701 -50.59553	240

Municipalities	Locations	LD ₅₀ (IC 95%)	LD ₉₉ (IC 95%)	RR ₅₀ **	Slope
Araçatuba	Nilce Maia – LRS*	0.0250 (0.0020 - 0.0320)	0.1960 (0.0108 - 0.3310)	-	2.520+-0.428
Araçatuba	Alcides Chagas	0.0823 (0.0691 - 0.1014)	0.6096 (0.3453 - 1.7103)	3.292	2.675+-0.436
	Arruda Brasil	0.0633 (0.0502 - 0.0819)	0.6209 (0.3293 - 2.1666)	2.532	3.346+-0.294
	Escola Nilce Maia	0.0741 (0.0556 - 0.0977)	0.3904 (0.2178 -2.0769)	2.964	3.223+-0.437
Birigui	James Mellor	0.0484 (0.0349 - 0.0733)	0.6723 (0.2757 - 5.8711)	1.936	2.035+-0.294
	Pedro de Toledo	0.0440 (0.0361 - 0.0528)	0.4049 (0.2543 - 0.8754)	1.76	2.413+-0.320
Guararapes	Eurides Amaral	0.0775 (0.0680 - 0.0903)	0.4439 (0.2863 - 0.9319)	3.1	3.088+-0.446
	Gaudêncio José	0.0871 (0.0738 - 0.1047)	0.6786 (0.4047 - 1.7420)	3.484	2.609+-0.400
	Praça Central	0.0739 (0.0590 - 0.0921)	0.4040 (0.2422 - 1.2799)	2.956	3.154+-0.409
	Praça Mohamed	0.0674 (0.0557 - 0.0817)	0.8291 (0.4623 - 2.3296)	2.696	2.134+-0.311
	Princesa Isabel	0.0845 (0.0707 - 0.1049)	0.9821 (0.5095 - 3.4704)	3.38	2.184+-0.359
	Raquel Caldas	0.0681 (0.0580 - 0.0790)	0.4120 (0.2811 - 0.7801)	2.724	2.976+-0.399
Piacatu	Câmara Municipal	0.0908 (0.0767 – 0.1149)	0.3839(0.2406 - 1.0276)	3.632	3.716+-0.522
	José Benetti	0.0754 (0.0661 - 0.0853)	0.3151 (0.2348 - 0.5057)	3.016	3.744+-0.472

TABLE 2: Result of mortality of first stage triatomine nymphs of *Rhodnius neglectus* after topical application of deltamethrin in a reference population and in samples from different locations. Araçatuba region, 2020.

*Reference population susceptibility, **Resistance ratio 50% ng a.i./treated nymph (nanogram active ingredient / treated nymph.

in a quarantine after arriving at the laboratory and, at the end of this period, eggs were removed and the first stage nymphs' birth was synchronized in order to perform the experiment.

The pyrethroid deltamethrin (99.1% purity) was supplied by Bayer. A total volume of 0.1 *u*L was applied on the back of the nymph on the fifth day after birth. The control groups received only pure acetone on their backs. Readings were performed 72 h after application, considering the normal, intoxicated, and dead nymphs. For each dilution, 30 insects were used at the first stage, with at least eight doses necessary to determine the mortality curve. Three groups were formed, with 10 insects in each group, and the tests were carried out on different days.

Mortality data were analyzed using the Basic Probit Analysis program to estimate the slope and lethal dose (LD) in nanograms of active ingredients per treated nymph. The susceptibility classification was in accordance with the Pan American Health Organization¹⁴. After defining the *R. neglectus* susceptibility baseline for the reference population, 30 nymphs from each field population were subjected to a diagnostic dose of 1xLD₉₉. The survival of at least two insects in three repetitions was interpreted as a resistance indicator.

RESULTS

The susceptibility to deltamethrin in the reference population was 0.025 ng/treated nymphs at LD_{50} . At this diagnostic dose, 100% mortality was observed in all populations. Regarding *R. neglectus* samples from field populations, variations between 0.0440 and 0.0908 ng/treated nymphs were observed (**Table 2**). Populations from Birigui (James Mellor and Pedro de Toledo) and Guararapes (Praça Mohamed and Princesa Isabel) presented lower slopes than the reference population, indicating less homogeneous populations with a higher resistance selection likelihood.

Values obtained for RR_{50} in the field populations were significantly different from those in the reference population, and there was no superposition of confidence interval limits at 95%.

DISCUSSION

Insecticide resistance can be understood as a decrease in mortality observed in a population that has undergone constant exposure to chemical products used for extermination¹¹. In the case of triatomines, such resistance is regarded as rare and unlikely, mainly because of the life cycle of these insects, which hinders the selection of resistant individuals¹¹. However, previous studies have demonstrated cases of triatomine populations resistant to numerous active substances in different regions of America¹⁶⁻¹⁸.

Triatomine resistance to pyrethroids associated with ineffective field treatment has been reported in *R. prolixus* in Venezuela and in *Triatoma infestans* in Brazil, Argentina, and Bolivia¹⁹. After these findings, studies aimed at verifying susceptibility in Brazilian populations of *Triatoma infestans, T. sordida*, and *T. brasiliensis* were carried out and have found, in some situations, resistance to the deltamethrin pyrethroid insecticide^{18,20-24}.

The first report of pyrethroid insecticide resistance in triatomines of the genus *Rhodnius* was observed in populations of *R. prolixus* species in Venezuela using dieldrin²⁵. There are no reports of studies conducted in Brazil to verify insecticide resistance susceptibility in populations of *Rhodnius* genus, which may be due to the fact that most species from these genus do not inhabit homes, therefore posing secondary risk to human beings.

In São Paulo state, *R. neglectus* is the second most collected species and, currently, most samples are from urban areas where there is a superposition of controlling actions aimed at arboviruses and visceral leishmaniasis²⁶. To chemically control vectors, the Brazilian Ministry of Health provides states and municipalities with alphacypermethrin, a pyrethroid insecticide, as this insecticide has a low residual effect, does not remain on treated surfaces for long periods, and is subjected to weather changes, which may impair its useful life¹⁷. This insecticide is also widely used because of its low toxicity in mammals and because it does not persist in the environment²⁷.

Chemical control employed frequently to decrease infestation of this species in urban areas has not presented satisfactory results, which may lead to metabolic changes in the insects, resulting in resistance to the insecticides applied to control them. However, results from bioassays for this triatomine species did not indicate resistance; yet, there is evidence concerning the need for management strategies to maintain the insecticide lifespan, but it is necessary to continuously test insect resistance in these insect populations.

Values obtained for RR₅₀ in this assessment, and considering that values higher or equal to 5 are parameters to characterize insect resistance to the insecticide, indicated that controlling actions may proceed with the same insecticide; however, new management methodologies must be considered for the palm trees. Mechanical control of this vector species in this ecotope must also be prioritized, since it is a fact that this insect currently inhabits an urban area in São Paulo state and its widening distribution will bring new challenges.

Pessoa et al. $(2016)^{15}$ defined a reference population for *R. neglectus* in which the LD₅₀ value was 0.001 ng i. a/ninfa. If this value is considered, all populations tested in this assessment would be classified as resistant, indicating that it may be necessary to locally standardize the species under study. Obara et al. $(2011)^{20}$ and Pessoa et al. $(2014)^{21}$ also found such a need when working with *T. sordida*.

It is important to note that the reference population in this study was collected from the same area where samples used in this assessment were collected, that is, Araçatuba municipality, Nilce Maria location. However, spaying insecticide on the palm trees to control insect populations was not a technical standard at the time. As other vectors are also found in this area, pyrethroids are frequently used in controlling actions, which must be considered for the reference population, as they may show signs of resistance.

The fact that the slope was the same or higher than that of the reference population shows that, in most populations tested, there is little heterogeneity among them. For populations with lower slopes, resistance can be observed, which justifies the follow-up. Molecular studies have indicated less genetic diversity in areas treated with chemical treatment²⁸. It is important to highlight that the mortality at the diagnostic dose was 100% in all populations. Genetic variation in populations must be considered as a factor that may directly interfere with test results^{21,29}.

Finally, it is essential to analyze triatomine resistance to insecticides through studies aiming to better understand which factors may affect the control of these vectors, in order to evaluate and enhance intervention measures, if necessary. Previous studies have reported *R. neglectus* populations in palm trees located in urban areas in the Federal District of Goiás, Minas Gerais, and Mato do Grosso do Sul states^{8,30,31}. It is not uncommon to find notifications to public authorities concerning these insects by inhabitants of these areas, indicating that new epidemiological settings may occur.

Recolonization of this species in the environment may be related to behavioral factors and to the fact that they are also associated with birds, which facilitate its wide distribution in municipalities within the São Paulo state^{9,32}. Therefore, studies using field and laboratory insecticides simultaneously must be carried out to obtain results that are closer to local reality.

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REFERENCES

- Jurberg J, Rodrigues JMS, Moreira FFF, Dale C, Cordeiro IRS, Lamas-Jr VD, et al. Atlas iconográfico dos triatomíneos do Brasil (vetores da doença de Chagas). Rio de Janeiro: Instituto Osvaldo Cruz; 2014
- World Health Organization [internet]. Chagas disease (American trypanosomiasis). Disponível em: https://www.who.int/health-topics/ chagas-disease#tab=tab_1. 2021 [acesso em 01 fev 2021].
- Basile L, Jansá JM, Carlier Y, Salamanca DD, Angheben A, Bartoloni A, et al. Chagas disease in European countries: the challenge of a surveillance system. Euro Surveill. 2011;16(37):19968.
- Souza AG, Wanderley DMV, Buralli GM, Andrade JCR. Consolidation of the control of Chagas disease in State of São Paulo. Mem Inst Oswaldo Cruz. 1984;79:125-32.
- Rocha e Silva EO, Rodrigues VLCC, Silva RA, Wanderley DMV. Programa de Controle da Doença de Chagas no estado de São Paulo, Brasil: o controle e a vigilância da transmissão vetorial. Rev Soc Bras Med Trop. 2011;44(Suppl 2):74-84.
- Silva RA. Estado atual da vigilância entomológica da doença de Chagas no estado de São Paulo. Brazilian Journal of Health Review. 2019;2(2):742-55.
- Oliveira J, Alevi KCC, Almeida CE, Mendonça VJ, Costa J, Rosa JA. *Triatoma brasiliensis* species complex: characterization of the external female genitalia. J Vector Ecol. 2020;45(1):57-68.
- Diotaiuti L, Dias JCP. Ocorrência e biologia de *Rhodnius neglectus*, Lent, 1954 em macaubeiras da periferia de Belo Horizonte, Minas Gerais. Mem Inst Oswaldo Cruz. 1984;79(3):293-301.
- Rodrigues VLCC, Pauliquévis-Jr C, Silva RA, Wanderley DMV, Guirardo MM, Rodas LAC, et al. Colonization of palm trees by *Rhodnius neglectus* and household and invasion in an urban area, Araçatuba, São Paulo, Brazil. Rev Inst Med Trop São Paulo. 2014;56(3):213-8.
- Silva RA. Relatório de avaliação do *Rhodnius neglectus* em área urbana de municípios da região de Araçatuba. São Paulo: Superintêndencia de Controle de Endemias; 2018.
- Pessoa GCD, Vinãs PA, Rosa ACL, Diotaiuti L. History of insecticide resistance of triatominae vectors. Rev Soc Bras Med Trop 2015; 48(4):380-389.
- Ministério da Saúde Brasil [internet]. NT-36.2012 Orientações sobre vigilância entomológica e a utilização de inseticida de ação residual no controle de triatomíneos – vetores da doença de Chagas. Brasília; 2012 [acesso em 6 set 2020].
- Lent H, Wygodzinsky PW. Revision of the Triatominae (Hemiptera, Reduviidae), and their significance as vectors of Chagas disease. Bull Am Mus Nat Hist. 1979;163(1):123-520.
- 14. Organizacion Panamericana de la Salud (PAHO). Il Reunion técnica latino-americana de monitoreo de resistência a inseticidas em triatominos vectores de Chagas, OPS. Panamá: PAHO; 2005.
- Pessoa GCD, Silva RA, Alves RV, Costa VM, Cavalcante KRLJ, Diotaiuti L. Fortalecimento da vigilância em saúde no Brasil: Rede de Monitoramento da resistência dos triatomíneos aos inseticidas. Rev Patol Trop 2016; 45(4):417-424.
- Yon C, Balta R, García N, Troyes M, Cumpa H, Valdivia A. Susceptibilidad y resistencia de *Triatoma infestans y Panstrongylus herreri* a los insecticidas piretroides, Perú 2001. Revista Peruana de Medicina Experimental y Salud Pública. 2004; 21(3):179-182.

- 17. Santos MAT, Areas MA, Reyes FG. Piretróides uma visão geral. Alimentos e Nutrição Araraquara 2007; 18(3):339-349.
- Pessoa GCD, Rosa AC, Bedin C, Wilhelms T, Mello F, Coutinho HS, et al. Susceptibility characterization of residual Brazilian populations of *Triatoma infestans* Klug, 1834 (Hemiptera: Reduviidae) to deltamethrin pyrethroid. Rev Soc Bras Med Trop 2015; 48:157-161.
- Depickère S, Buitrago R, Siñani E, Baune M, Monje M, Lopez R, et al. Susceptibility and resistance to deltamethrin of wild and domestic populations of *Triatoma infestans* (Reduviidae: Triatominae) in Bolivia: new discoveries. Mem Inst Oswaldo Cruz 2012; 107: 1042-47.
- Obara MT, Otrera VCG, Gonçalves RG, Santos JP, Santalucia M, Rosa JA, et al. Monitoramento da suscetibilidade de populações de *Triatoma sordida* Stal, 1859 (Hemiptera: Reduviidae) ao inseticida deltametrina, na região centro-oeste do Brasil. Rev Soc Bras Med Trop 2011; 44(2):206-212.
- Pessoa GCD, Dias LS, Diotaiuti L. Deltamethrin pyrethroid susceptibility characterization of *Triatoma sordida* Stal, 1859 (Hemiptera: Reduviidae) populations in the northern region of Minas Gerais, Brazil. Rev Soc Bras Med Trop 2014; 47(4):426-429.
- Pessoa GCD, Trevizani NAB, Dias LS, Bezerra CM, Melo BV, Diotaiuti L. Toxicological profile of deltamethrin in *Triatoma brasiliensis* (Hemiptera: Reduviidae) in state of Ceará, northeastern Brazil. Rev Soc Bras Med Trop 2015; 48(1):39-43.
- Pessoa GCD, Santos TRM, Salazra GC, Dias LS, Mello BV, Ferraz ML, et al. Variability of susceptibility to deltamethrin in peridomestic *Triatoma sordida* from Triângulo Mineiro, state of Minas Gerais, Brazil. Rev Soc Bras Med Trop 2015; 48(4):417-421.
- Pessoa GCD, Rosa ACL, Cavalari L, Rezende JG, Mello BV, Diotaiuti L. Susceptibility of *Triatoma sordida* Stal, 1859 (Hemiptera: Reduviidae) to alpha-cypermethrin under natural climatic conditions. Rev Soc Bras Med Trop 2015; 48(4):422-426.

- Vassena C, Picollo M, Zerba E. Insecticide resistance in Brazilian Triatoma infestans and Venezuelan Rhodnius prolixus. Med Vet Entomol 2000; 14(1):51-5.
- Camargo-Neves VLFD, Katz G, Rodas LAC, Poletto DW, Lage LC, Spínola RMF, et al. Utilização de ferramentas de análise espacial na vigilância epidemiológica de leishmaniose visceral americana-Araçatuba, São Paulo, Brasil, 1998-1999. Cad Saúde Pública 2001; 17:1263-67.
- 27. Narahashi T. Neuronal ion channel as the target sites of insecticides. Pharmacol Toxicol 1996; 79(1):1-14.
- Perez de Rosas AR, Segura EL, Garcia BA. Microssatellites analysis of genetic structure in natural *Triatoma infestans* (Hemiptera: Reduvidae) populations from Argentina: its implication in assessing the effectiveness of Chagas disease vector control program. Mol Ecol 2007; 16(7):1401-1412.
- Amelotti I, Catalá SS, Gorla DE. Experimental evaluation of insectidal paints against *Triatoma infestans* (Hemiptera: Reduvidae), under natural climatic conditions. Parasit Vectors 2009; 2:30.
- Zapata MTG, Virgens D, Soares VA, Bosworth A, Mardsen PD. House invasion by secondary triatominae species in Mambaí, Goiás-Brazil. Rev Soc Bras Med Trop 1985; 18(3):199-201.
- 31. Gurgel-Gonçalves R, Duarte MA, Ramalho ED, Romaña CA, Cuba CAC. Distribuição espacial de populações de triatomíneos (Hemiptera, Reduviidae) em palmeiras da espécie *Mauritia flexuosa* no Distrito Federal, Brasil. Rev Soc Bras Med Trop 2004; 37(3):241-247.
- Forattini OP, Ferreira OA, Silva EOR, Rabello ES. Aspectos ecológicos da tripanossomíase americana. XV – Desenvolvimento, variação e permanência de *Triatoma sordida*, *Panstrongylus megistus* e *Rhodnius neglectus* em eótopos artificiais. Rev Saúde Pública 1979; 13(3):220-234.

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