

Genome Sequence of *Clostridium tyrobutyricum* ATCC 25755, a Butyric Acid-Overproducing Strain

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Clostridium tyrobutyricum ATCC 25755 is an efficient producer of butyric acid. Here we report a 3.01-Mb assembly of its genome sequence and other useful information, including the coding sequences (CDSs) responsible for an alternative pathway leading to acetate synthesis as well as a series of membrane transport systems.

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utyric acid, one of the four-carbon short-chain organic acids, is a specialty chemical with many applications in different industries, and currently there is an increasing interest in using it from microbial production as bio-based natural ingredients in foods, cosmetics, and pharmaceuticals (1). Clostridium tyrobutyricum ATCC 25755 is the most promising acidogenic bacterium used for the production of butyric acid (2-6). A highest butyric acid concentration of 86.9 g/liter has been achieved by our team with an adapted strain of C. tyrobutyricum ATCC 25755 (7). Moreover, genetic manipulation with this type strain, such as the inactivation of ack and pta genes, which encode enzymes associated with the acetate formation pathway, has been proved to significantly improve butyric acid production (8, 9). However, the formation of acetic acid was not completely eliminated in these mutants, suggesting the existence of other enzymes also leading to acetate synthesis. Therefore, investigation of the genetic information and characteristics of C. tyrobutyricum ATCC 25755 is desired. Genome-scale analysis has been proven useful for the metabolic engineering application (10).

Here we present the draft genome sequence of strain *C. tyrobutyricum* ATCC 25755, obtained using the Illumina Hiseq 2000 system, which was performed by Shanghai Majorbio Bio-Pharm Technology Co., Ltd. The reads were assembled with Velvet (1), and the sequence was annotated using the RAST annotation server (11). A library containing 300-bp inserts was constructed. Sequencing was performed based on the paired-end strategy of 101-bp reads to produce 1,667 Mb of filtered sequences, representing 553.82-fold coverage of the genome. The sequence of *C. tyrobutyricum* ATCC 25755 is 3,011,226 bases with a G+C content of 30.84%, which was assembled into 80 contigs and 67 scaffolds. They contain 3,040 open reading frames (ORFs), 46 tRNA genes, and 3 rRNA genes identified by Glimmer 3.02 (12), Genemark (13), tRNAscan-SE (14), and RNAmmer (15).

Inspiringly, acyl-coenzyme A (CoA):acetoacetyl-CoA transferase was detected among the gene products, which provided the first definitive evidence that there was an alternative pathway leading to acetate synthesis in acidogenic clostridia (8, 9). According to the genomic analysis, strain C. tyrobutyricum ATCC 25755 may have a powerful membrane transport capacity, including 20 protein-coding sequences (CDSs) involved in the phosphotransferase systems and the corresponding regulatory system analogs (e.g., Hpr), 106 CDSs involved in ATP-binding cassette (ABC)type transporters, and 1 involved in P-type ATPases, as well as many uncharacterized transporters. They may play important roles in the production of butyric acid with high productivity (6, 7). Additionally, several putative enzymes were also annotated in the C. tyrobutyricum ATCC 25755 genome, such as glycosyltransferase, aspartate transaminase, sugar phosphate isomerase, amino acid permease, glucokinase, intracellular protease/amidase, amidohydrolase, helicase, and some regulatory proteins. Further studies will be performed to confirm their functions, and a complete genome sequence will be included in the future to reveal the unique molecular characteristics of strain C. tyrobutyricum ATCC 25755.

Nucleotide sequence accession numbers. This whole genome shotgun project has been deposited at DDBJ/EMBL/GenBank under accession no. APMH00000000. The version described in this paper is the first version, with accession no. APMH01000000.

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REFERENCES

- 1. Zerbino DR, Birney E. 2008. Velvet: algorithms for de novo short read assembly using de Bruijn graphs. Genome Res. 18:821–829.
- Jiang L, Wang JF, Liang SZ, Cai J, Xu ZN, Cen PL, Yang ST, Li S. 2011. Enhanced butyric acid tolerance and bioproduction by *Clostridium tyro-butyricum* immobilized in a fibrous bed bioreactor. Biotechnol. Bioeng. 108:31–40.

- 3. Jiang L, Wang JF, Liang SZ, Cai J, Xu ZN. 2011. Control and optimization of *Clostridium tyrobutyricum* ATCC 25755 adhesion into fibrous matrix in a fibrous bed bioreactor. Appl. Biochem. Biotechnol. 165: 98–108.
- Jiang L, Wang JF, Liang SZ, Wang XN, Cen PL, Xu ZN. 2010. Production of Butyric Acid from glucose and xylose with immobilized cells of *Clostridium tyrobutyricum* in a fibrous-bed bioreactor. Appl. Biochem. Biotechnol. 160:350–359.
- Jiang L, Wang JF, Liang SZ, Xu ZN, Yang ST. 2010. Phosphoenolpyruvate-dependent phosphorylation of sucrose by *Clostrid-ium tyrobutyricum* ZJU 8235: evidence for the phosphotransferase transport system. Bioresour. Technol. 101:304–309.
- Jiang L, Wang JF, Liang SZ, Wang XN, Cen PL, Xu ZN. 2009. Butyric acid fermentation in a fibrous bed bioreactor with immobilized *Clostridium tyrobutyricum* from cane molasses. Bioresour. Technol. 100: 3403–3409.
- Jiang L, Li S, Hu Y, Xu Q, Huang H. 2012. Adaptive evolution for fast growth on glucose and the effects on the regulation of glucose transport system in *Clostridium tyrobutyricum*. Biotechnol. Bioeng. 109:708–718.
- Liu XG, Zhu Y, Yang ST. 2006. Construction and characterization of ack deleted mutant of *Clostridium tyrobutyricum* for enhanced butyric acid and hydrogen production. Biotechnol. Prog. 22:1265–1275.
- 9. Zhu Y, Liu XG, Yang ST. 2005. Construction and characterization of pta Gene-deleted mutant of *Clostridium tyrobutyricum* for enhanced Butyric Acid fermentation. Biotechnol. Bioeng. 90:154–166.

- Nölling J, Breton G, Omelchenko MV, Makarova KS, Zeng Q, Gibson R, Lee HM, Dubois J, Qiu D, Hitti J, Wolf YI, Tatusov RL, Sabathe F, Doucette-Stamm L, Soucaille P, Daly MJ, Bennett GN, Koonin EV, Smith DR. 2001. Genome sequence and comparative analysis of the solvent-producing bacterium *Clostridium acetobutylicum*. J. Bacteriol. 183:4823–4838.
- 11. Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O. 2008. The RAST server: rapid annotations using subsystems technology. BMC Genomics 9:75.
- 12. Delcher AL, Bratke KA, Powers EC, Salzberg SL. 2007. Identifying bacterial genes and endosymbiont DNA with glimmer. Bioinformatics 23:673–679.
- Besemer J, Lomsadze A, Borodovsky M. 2001. GeneMarkS: a selftraining method for prediction of gene starts in microbial genomes. Implications for finding sequence motifs in regulatory regions. Nucleic Acids Res. 29: 2607–2618.
- Lowe TM, Eddy SR. 1997. tRNAscan-SE: a program for improved detection of transferRNAgenes in genomic sequence. Nucleic Acids Res. 25: 955–964.
- Lagesen K, Hallin P, Rødland EA, Staerfeldt HH, Rognes T, Ussery DW. 2007. RNAmmer: consistent and rapid annotation of ribosomal RNA genes. Nucleic Acids Res. 35:3100–3108.