

# **Dandelion uses the single-cell adaptive immune receptor repertoire to explore lymphocyte developmental origins**

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## Supplementary Note 1

The 10X Genomics library kit and V(D)J analysis for TCR was tailored for  $\alpha\beta$ TCR contigs and most cellranger versions had difficulties in annotating  $\gamma\delta$ TCR contigs, a problem 10X was aware of and addressed with user-side workaround instructions. When *cellranger vdj* updated its contig annotation pipeline in version 3.1.0

(<https://support.10xgenomics.com/single-cell-gene-expression/software/pipelines/3.1/release-notes>), the priority was  $\alpha\beta$ TCR and the software lost the ability to annotate  $\gamma\delta$ TCR contigs.

However, the same release saw the introduction of custom enrichment primer support, which is integral for proper  $\gamma\delta$ TCR contig reconstruction.  $\gamma\delta$ TCR contig reannotation was reintroduced on an experimental basis in version 7.0.0

(<https://support.10xgenomics.com/single-cell-gene-expression/software/pipelines/7.0/release-notes>). *cellranger vdj* versions between 3.1.0 and 6.1.2 can still reconstruct  $\gamma\delta$ TCR contigs, but cannot natively annotate them.

10X Genomics was aware of the issue, but it was not a priority for them as  $\gamma\delta$ TCR libraries require custom enrichment primers not part of their product line-up. Early support requests would direct users to the last legacy version, 3.0.2, that supported  $\gamma\delta$ TCR annotation (<https://github.com/10XGenomics/cellranger/issues/45>). However, this was not an ideal solution due to the lack of custom enrichment primer support. 10X Genomics subsequently revised their recommended solution to a modification of the reference, wherein all TRG sequences would be renamed to TRA and TRD would be renamed to TRB. This advice used to be available at <https://kb.10xgenomics.com/hc/en-us/articles/360015793931-Can-I-detect-T-cells-with-gamma-delta-chains-in-my-V-D-J-data-> but this has since been overwritten by *cellranger multi* instructions for version 7.0.0.

## Supplementary Figure Legend

**Supplementary Fig. 1 | Non-productive BCR in pDC. a,b,** Boxplot of the proportion of cells with productive (blue) or non-productive (orange) BCR heavy chain (**a**) and light chain (**b**) in different fetal myeloid subsets. Each point represents a sample and data were taken from Suo et al. 2022<sup>1</sup>. Only samples with at least 20 cells are shown. Boxes capture the first to third quartiles and whiskers span a further 1.5X interquartile range on each side of the box. **c,** Expression of genes involved in V(D)J rearrangement in pDCs and cycling pDCs. Data was taken from Suo et al. 2022<sup>1</sup>.

## Supplementary Tables

### **Supplementary Table 1: top\_10\_j\_multimappers.csv (separate file)**

Top 10 J gene combinations with multi-J mapping for each locus in data from Suo et al. 2022<sup>1</sup>, with the number of contigs containing each combination shown next to it.

### **Supplementary Table 2: LR\_results.csv (separate file)**

Logistic regression results exploring factors associated with multi-J mapping presence in data from Suo et al. 2022<sup>1</sup>.

### **Supplementary Table 3: LR\_results\_combined.csv (separate file)**

Logistic regression results exploring factors associated with multi-J mapping presence in control and cycloheximide-treated PBMC data.

### **Supplementary Table 4: j\_sequence\_affect\_j\_multimapper.csv (separate file)**

List of leftmost (5' end) J genes that had significant association with increased or decreased multi-J mapping, together with the sequences of their last 10 nucleotides at 3' ends and the first 11 nucleotides of its 3' end intron.

### **Supplementary Table 5: panimmune\_differential\_VDJ.csv (separate file)**

Differential V(D)J usage across CD4+T, CD8+T, and MAIT cells in data from Conde et al. 2022<sup>2</sup>.

### **Supplementary Table 6: abtentry\_cor\_result.csv (separate file)**

Pearson's correlation coefficients and BH adjusted *P*-values of all genes with branch probabilities to CD8+T lineage within abT(entry) cells.

[cor\_tcr] Pearson's correlation coefficients for pseudotime inferred from neighborhood V(D)J space

[pval\_tcr] Pearson's correlation *P*-values for pseudotime inferred from neighborhood V(D)J space

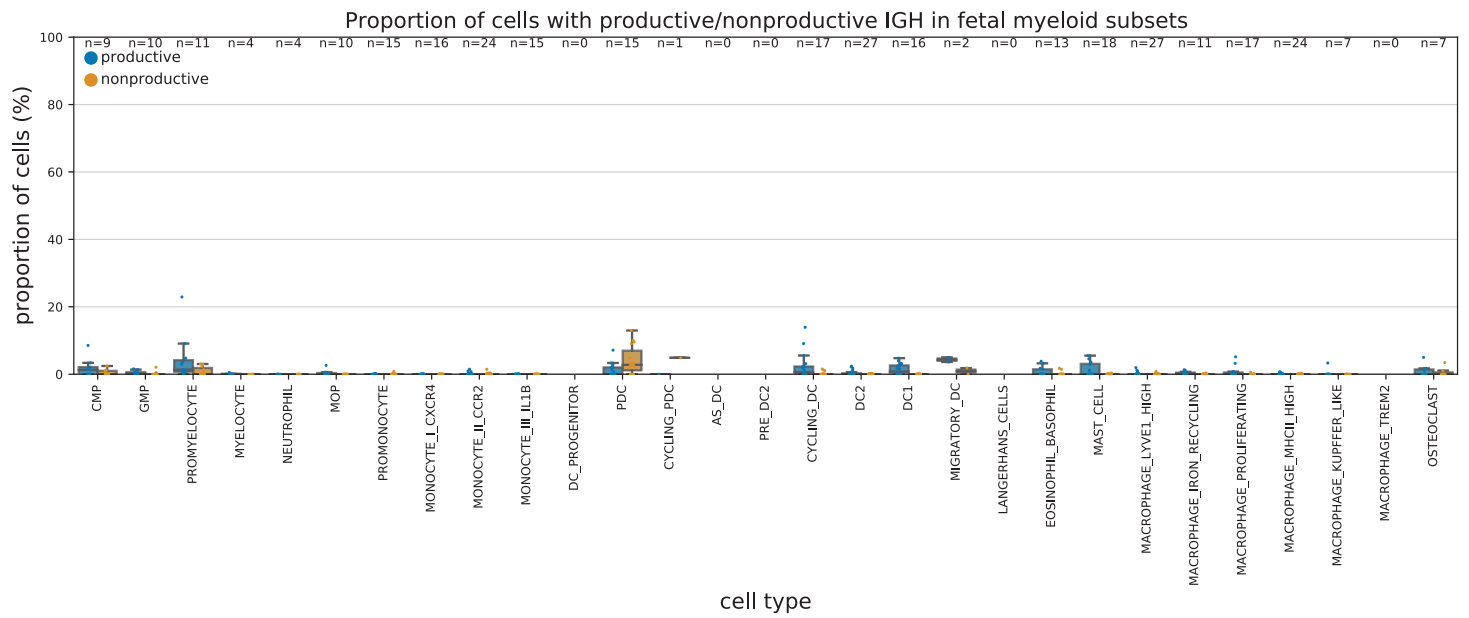
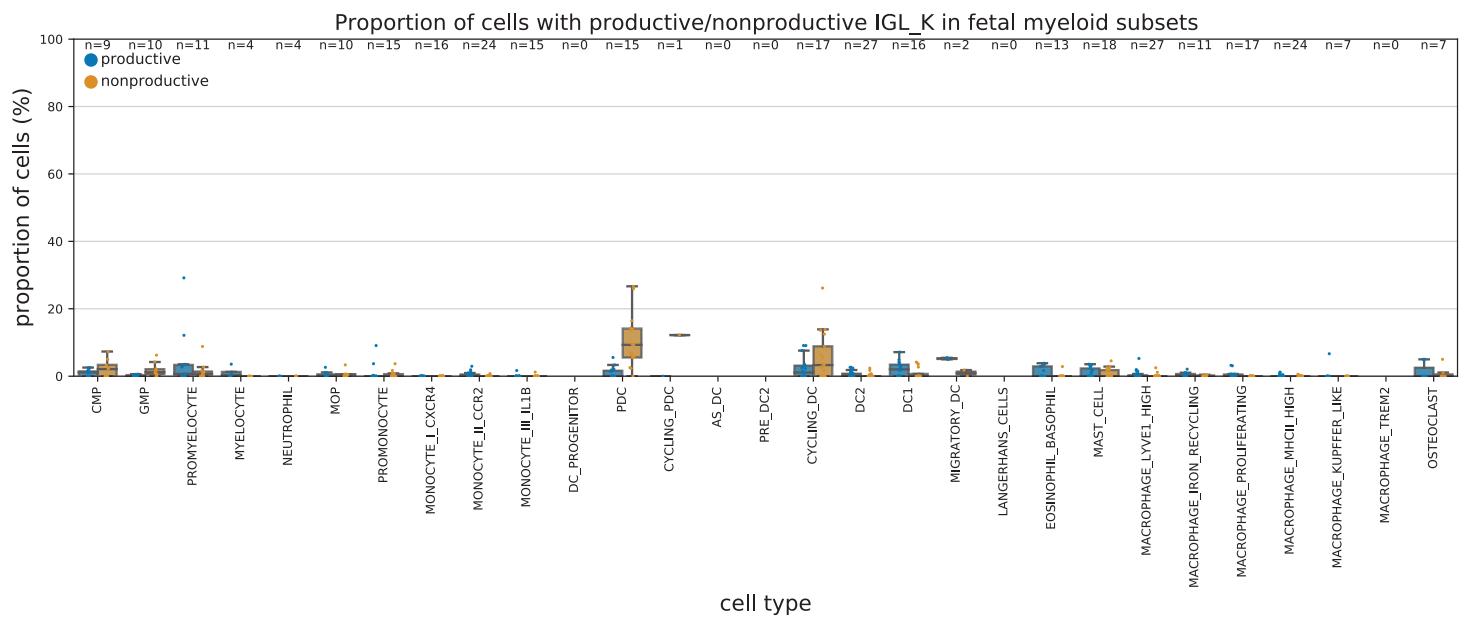
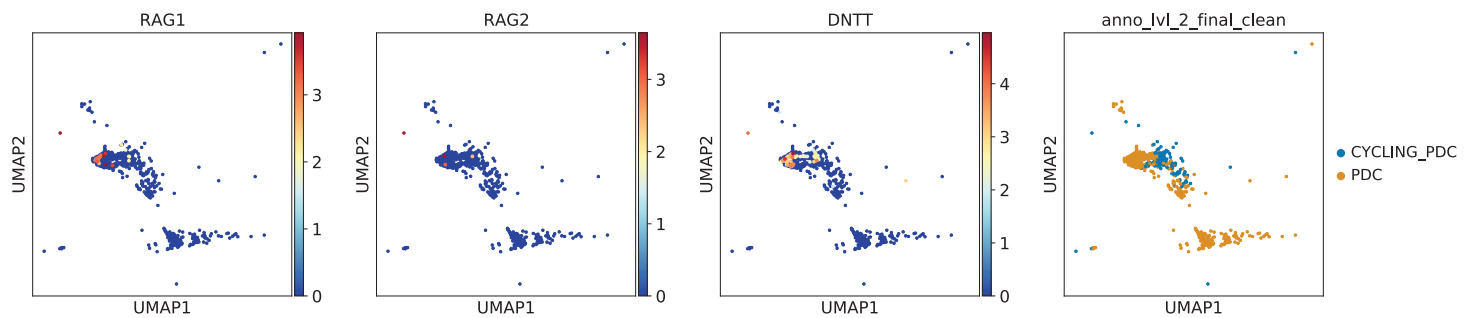
[adjp\_tcr] *P*-values from pval\_tcr adjusted by BH procedure

[cor\_gex] Pearson's correlation coefficients for pseudotime inferred from neighborhood GEX space

## Supplementary References

1. Suo, C. *et al.* Mapping the developing human immune system across organs. *Science* **376**, eabo0510 (2022).
2. Domínguez Conde, C. *et al.* Cross-tissue immune cell analysis reveals tissue-specific features in humans. *Science* **376**, eabl5197 (2022).

## Supplementary Figure

**a****b****c**

Suo et al. Supplementary Fig. 1