

Review Article

Novel Regulatory Mechanisms of Pathogenicity and Virulence to Combat MDR in *Candida albicans*

Saif Hameed and Zeeshan Fatima

Amity Institute of Biotechnology, Amity University Haryana, Manesar, Gurgaon 122413, India

Correspondence should be addressed to Saif Hameed; saifhameed@yahoo.co.in

Received 30 June 2013; Revised 15 August 2013; Accepted 15 August 2013

Academic Editor: Isabel Sá-Correia

Copyright © 2013 S. Hameed and Z. Fatima. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Continuous deployment of antifungals in treating infections caused by dimorphic opportunistic pathogen *Candida albicans* has led to the emergence of drug resistance resulting in cross-resistance to many unrelated drugs, a phenomenon termed multidrug resistance (MDR). Despite the current understanding of major factors which contribute to MDR mechanisms, there are many lines of evidence suggesting that it is a complex interplay of multiple factors which may be contributed by still unknown mechanisms. Coincidentally with the increased usage of antifungal drugs, the number of reports for antifungal drug resistance has also increased which further highlights the need for understanding novel molecular mechanisms which can be explored to combat MDR, namely, ROS, iron, hypoxia, lipids, morphogenesis, and transcriptional and signaling networks. Considering the worrying evolution of MDR and significance of *C. albicans* being the most prevalent human fungal pathogen, this review summarizes these new regulatory mechanisms which could be exploited to prevent MDR development in *C. albicans* as established from recent studies.

1. Introduction

In the last decades, the incidence of fungal infections has increased dramatically due to the rise in the number of immunocompromised patients. The most prevalent fungal pathogen of humans is *Candida albicans* which ranks as the fourth most common cause of hospital acquired infectious disease and is the primary cause of systemic candidiasis, with mortality rates approaching 50% [1]. The dimorphic opportunistic pathogen, *C. albicans*, is normally a commensal organism in humans, but when the host is unable to mount an adequate immune response, as in AIDS, organ transplant, diabetes, or in cancer patients, it results in mucosal, cutaneous, or invasive mycoses [2, 3]. Prolonged usage of antifungals in treating infections caused by *C. albicans* has led to the emergence of azole resistance. This acquired azole resistance in clinical isolates of *C. albicans* mostly results in cross-resistance to many unrelated drugs, a phenomenon termed multidrug resistance (MDR) [4–6]. MDR is a serious complication during treatment of opportunistic fungal infections which poses grave concern given the limited number

of clinically useful antifungal drugs available [7, 8]. Fungal species have evolved a multitude of mechanisms to survive exposure to antifungal drugs and some of them include an overexpression or mutations in *ERG11*, encoding the target enzyme of azoles lanosterol 14 α -demethylase [4, 5, 9, 10], an over expression of the drug efflux pumps encoding genes such as *CaCDRI* and *CaCDR2* belonging to the ABC (ATP-binding cassette) [11–13] and *CaMDRI* belonging to the MFS (major facilitator super family) transporters [14–16].

Although MDR is a complex manifestation of factors which are reasonably documented, there are reports to suggest that it may involve many unknown mechanisms which are yet to be elucidated. In the recent years, emerging evidence has demonstrated that there do exist such novel mechanisms which can be helpful in controlling MDR efficiently. Improved knowledge of such molecular mechanisms controlling MDR in pathogenic fungi should facilitate the development of novel therapies to combat these intransigent infections. This review further defines the focus on the exacerbated need of understanding such mechanisms (Figure 1)

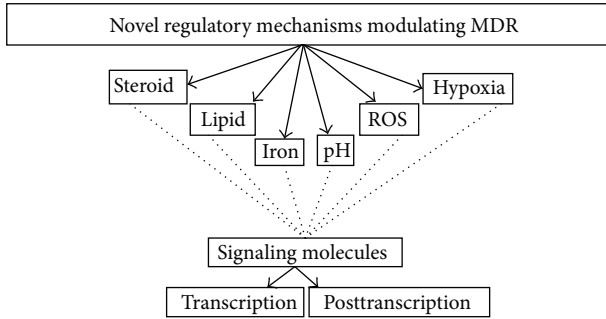


FIGURE 1: Novel regulatory mechanisms modulating MDR in *Candida albicans*.

and attempts to highlight research areas that need to be investigated in greater detail.

2. ROS

In eukaryotic cells, mitochondria are common organelles that represent an important source of reactive oxygen species (ROS). ROS are not just a byproduct of mitochondrial disintegration but also a key regulator for yeast apoptosis [17]. As mediators in signal transduction pathways, ROS participate in early and late steps of the regulation of apoptosis. ROS produced by granulocytes or monocytes are known to exert activity against fungi [18]. Furthermore, *C. albicans* possesses a ROS scavenger, superoxide dismutase suggesting that fungi may require a cytoprotective mechanism against not only exogenous ROS but also endogenous ROS [19]. Kobayashi [20] demonstrated the role of ROS in azoles mediated drug sensitivity in *C. albicans*, thereby establishing a strong correlation between ROS and MDR. They showed that ROS production is directly involved in the cytostatic action of miconazole. In this study complete inhibition of miconazole-induced ROS production resulted in the restoration of 50 to 70% of cell viability, suggesting that ROS production is an important event, in addition to drug-induced inhibition of ergosterol synthesis. Similarly Wu [21] deciphered the antifungal action of plagiocin E (PLE) through mitochondrial-dysfunction-induced ROS accumulation in *C. albicans*. Xu [22] showed that endogenous ROS augmentation contributes to the synergistic action of fluconazole (FLC) and berberine against FLC-resistant *C. albicans*. Rosa [23] explored a mechanistic link between the drug sensitivity, gene expression, and pathogenesis phenotypes of *C. albicans*. They conclude that histone acetyltransferase, Rtt109, is particularly important for fungal pathogenicity, suggesting a unique target for therapeutic antifungal compounds. Recently, the fungicidal activity of Gemini-pyridinium salts and shikonin was found to be mediated through ROS generation only [24, 25]. Thus the fact that ROS production could either form the basis of antifungal action of the compound mentioned above or act in synergism to enhance the cytotoxicity of drugs has become a potential therapeutic strategy nowadays. Therefore, ROS signaling pathways need to be further elucidated in fungal pathogens to enhance the potency of such target.

3. Iron

Pathogens including *C. albicans* colonize various niches which are iron-limited, and iron, being an indispensable micronutrient, is required both by the host and by the microbial community residing within the host [26–31]. Availability of iron in host cells is tightly regulated, since iron is a transition metal and its ability to donate and accept electrons can indulge in the formation of toxic free radicals, and hence iron plays a key role in providing natural resistance to infections in humans [32]. Interestingly, studies suggest that there could be a correlation between intracellular iron and MDR phenomenon. For instance, role of iron in recurrent vulvovaginal candidiasis (RVVC) showed that this element is not only important for pathogenic yeast, but also for normal function of host immunity [30]. Kuipers [33–35] showed that lactoferrin, an iron binding glycoprotein, is synergistic with antifungals against different *Candida* species. However, whether iron affects drug susceptibility of *Candida* cells was not demonstrated experimentally until Prasad [36] reported for the first time that availability of iron could have an impact on defense mechanisms of *Candida* against antifungal drugs. Interestingly, it was observed that iron deprivation enhanced drug susceptibility of *Candida* cells resulting in an increase in membrane fluidity, which in turn leads to enhanced passive diffusion of drugs. A link between changes in membrane fluidity and lowered ergosterol levels was established in iron deprived *Candida* cells probably due to downregulation of *ERG11*. However, the intricate relationship between cellular iron, calcineurin signaling, membrane lipid homeostasis, and drug susceptibility of *Candida* cells was first established by Hameed [37]. Even antifungal action of malachite green is mediated via depletion of labile iron pools as one of its mechanisms [38]. The synergism of lactoferrin with fluconazole has been reported to enhance the antifungal activity of fluconazole against *Candida* spp. [39]. Cap2/Hap43 is essential for *C. albicans* growth under iron-deprivation conditions and for virulence in mouse [40]. Moreover, the usage of fungicidal monoclonal antibodies seems to interfere with iron acquisition in *C. albicans* [41]. Many transcription factors governed by iron homeostasis namely, Sfu1, Hap43, Sef1, Cap2, and Aft2, and regulators of iron uptake genes have already been identified; however, sufficient knowledge of signaling pathways is still lacking, and therefore, understanding the role of these transcription factors which are governed by iron availability will better help in studying the relation between iron and MDR. Kaba [42] demonstrated a link between mitogen activated protein kinase HOG1 and iron availability. Dap1 is a heme binding protein that mediates a functional link between iron homeostasis and azole resistance in *C. glabrata* [43]. Since *dap1* mutants show enhanced azole susceptibility and decreased ergosterol production, hence, its role in *C. albicans* could also be a possible target. Gene expression profiling of BCR1 transcription factor, known to have a role in biofilm formation, has reported that five out of eight genes have been implicated in iron homeostasis [44]. The influence of cellular iron on drug susceptibilities of *Candida* suggests iron to be yet another novel determinant of MDR which merits a closer look.

4. Hypoxia

Organisms often encounter sites within the host during infection which have inadequate vascularization and irregular blood flow and thus present hypoxic areas. The transcription factor complex hypoxia inducible factor 1 (*HIF-1*) controls the expression of most genes involved in adaptation to hypoxic conditions and therefore represents an important regulator of MDR development. Hypoxia is known to cause resistance to chemotherapy by the induction of human *MDR1* in growing tumors, via activation of hypoxia inducible factor-1 (*HIF1*) [45–48]. Although till date no homolog of *MDR1* has yet been identified in *C. albicans*, as this organism encounters sites during infection which are hypoxic, it is indeed inevitable to understand the hypoxic responses of this pathogen. The relevance to explore hypoxic responses in *C. albicans* is depicted from a wide range of studies. For instance, transcriptome response of *C. albicans* under hypoxia reveals metabolic adaptation to scarce amount of oxygen availability [49]. Following hypoxic growth in vagina-simulative conditions, part of *C. albicans* proteome that is covalently linked to the cell wall has been determined [50]. *Efg1* plays a major role in hypoxic responses of *C. albicans* as suggested by the fact that *efg1* mutants, though inhibit hyphae formation during normoxia, are able to express filaments under microaerobic conditions if grown on or within (embedded) agar at temperatures from 25°C to 35°C [51, 52]. Another study revealed that *efg1* mutants are hyperfilamentous as a response to low oxygen [49]. Role of *Efg1p* in biofilm formation, which is one of the major threat towards antifungal chemotherapy, under hypoxia has been established where *Efg1p* induces all the major classes of genes required for biofilm formation [53]. Moreover, adaptation to hypoxia forms an integral component of biofilm formation in *C. albicans* [54]. Carvalho [55] demonstrated that aspartyl protease activity could be modulated by oxygen availability in *C. albicans*. Another study revealed that regulation of gene expression in response to hypoxia in *C. albicans* could be signaled via lowered sterol levels and induction of filamentation under hypoxic conditions requires the Ras1- and Cdc35-dependent pathway [56]. Recently, it has been demonstrated that kinase Sch9 integrates both hypoxia and CO₂ sensing to inhibit yeast to hyphal transition in *C. albicans* [57]. Responses to hypoxic conditions in pathogenic fungi including *C. albicans* have been already extensively reviewed [58, 59]. Certainly further research on the effect of hypoxia on infection by fungal pathogens should be focused. Antifungal compounds that act specifically on adaptive mechanisms of pathogens required for hypoxic adaptation could be promising alternatives to existing strategies.

5. Steroids

Steroids have been known to affect cell growth, germination, morphogenesis, and virulence in fungi [60–62]. Some lines of evidence for the presence of steroid binding proteins in yeast, such as estradiol binding protein [63], corticosteroid binding protein [64], and progesterone binding protein [65], have already been documented but their exact function in the

steroid response is not known. Many studies have shown that steroids can induce a pleiotropic drug resistance (PDR) state in both pathogenic *C. albicans* and nonpathogenic *Saccharomyces cerevisiae*. Oestrogen regulation and its mechanism have been extensively studied [66, 67] and it is reported that the *CDR1* promoter region does contain steroid responsive elements (SRE) and a drug response element (DRE). Oestrogen mediated binding of the transcription factor *TAC1* to the DRE and induction of *CDR1* expression have been studied [68]. Banerjee [69] for the first time demonstrated that yeast cells, which do not possess steroid receptor cascade, probably perceive steroids as a cellular stress. They examined the genome-wide changes in the gene expression profile following exposure to progesterone. Interestingly, an inverted CCAAT box which in combination with other conserved sequences is attributed towards human *MDR1* responsiveness to cellular stresses is also present in steroid responsive region consensus sequences [70–72]. Banerjee [73] deciphered a more accurate evolutionary significance of the steroid response in yeasts by exposing *S. cerevisiae* and *C. albicans* cells to several doses of progesterone for different time periods. The study revealed the conserved and divergent features of PDR network in yeasts. Recently, antifungal potentials of steroidal quinolones and chalcones, respectively, have been reported and results seems to be quite promising [74, 75]. These studies showed that the steroid response in the absence of any known signaling cascade is a global phenomenon in yeast cells. Yet, in view of the importance of steroids in the physiology of both pathogenic and nonpathogenic yeasts, it would be interesting to examine the steroid-dependent regulatory cascade in these organisms and thereby decipher a new mechanism to combat MDR.

6. Morphogenesis

In response to various environmental stimuli, *C. albicans* is able to switch from the unicellular yeast form into either of the two distinct filamentous forms, that is, cells with pseudo-hyphae or true hyphae. This ability to switch is considered as an important virulence trait which is also coregulated with other virulence factors that are associated with cellular morphology [76, 77]. The morphological form of *C. albicans* is directly related to environmental conditions and these cues trigger separate signal transduction pathways which regulate common targets required to initiate hyphal growth [51, 76, 77]. The transcription factor *Efg1p* is a well known regulator of morphogenesis of *C. albicans* since it induces the yeast-to-hyphal transition and also regulates phenotypic switching and chlamyospore formation of this pathogen [52].

Lo [78] established that *Efg1p* is involved in drug resistance by regulating the expression of *ERG3* gene of ergosterol biosynthetic pathway. Ergosterol is an important target for many antifungal drugs particularly on the plasma membrane. Considering the significance of *Efg1p* regulator in morphogenesis, Prasad [79] have evaluated if disruption in morphogenic signaling cascade would also affect MDR status of *C. albicans* cells. The study showed that null mutant of the morphogenic regulator *EFG1* displayed enhanced drug

sensitivity of *C. albicans* cells by a mechanism that is not dependent on the drug efflux pumps. This study establishes a convergence of *EFG1* and *MDR* pathways and thus proposes an additional new role for this important morphogenic regulator of *C. albicans*. The role of a newly discovered regulator of hyphae formation, *Rca1*, in drug susceptibility of *C. albicans* was established recently [80]. Certainly more extensive analyses are required to elucidate the commonality between *EFG1* and *MDR* signaling cascades to find newer targets for antifungal chemotherapy.

7. pH

The ability of microorganisms to sense and adapt to changes in the environment is essential for their survival. One environmental factor that microorganisms must respond to is extracellular pH. Environmental pH has dramatic effects on the cell, particularly at the plasma membrane, including effects on protein activity, maintenance of the proton gradient, and nutrient availability. Furthermore, in the opportunistic fungal pathogen *C. albicans*, environmental pH serves as one potent signal for morphological differentiation, although serum N-acetyl glucosamine and agar-embedding represent other several potent signals [2, 3]. Most chemotherapeutic drugs in use today are hydrophobic small, molecules that are also typically either weakly basic, weakly acidic, or charged. Thus, changes in the electrochemical parameters of microorganism's cell membranes have important effects on their transmembranous diffusion and cellular retention [81]. Changes in these parameters can also modulate the function of immunological agents and affect the signal transduction. The *RIM101* pathway that has been identified in *C. albicans* governs pH responses, dimorphism, and pathogenesis [82]. The *RIM101* pathway and pH responses, in general, play an intimate role in pathogenesis beyond simply allowing the organism to grow [83]. Apart from the major *RIM101* pathway, some other pathways have also been identified in *C. albicans*, which act in parallel. For instance a novel *RIM101* independent pH pathway was proposed which was mediated by *PHR2* [83]. *Phr1p* and *Phr2p* are differentially expressed cell wall proteins in response to environmental pH. *Phr1p* is expressed under alkaline conditions and *Phr2p* is expressed under acidic conditions, and importantly both are required for the pathogenesis of the organism for causing systemic candidiasis (pH 7.4) and vaginal candidiasis (pH 4) [84]. Another signaling pathway that was identified for adaptation to neutral/alkaline pH is calcineurin signaling [85]. Calcineurin is a highly conserved calcium-dependent serine/threonine-specific protein phosphatase that mediates various stress responses inside the cell including conferring tolerance to alkaline pH in *C. albicans* [86]. Furthermore, Hameed [37] provided the first evidence to establish relationship between cellular iron, calcineurin signaling, membrane lipid homeostasis, and drug susceptibility of *Candida* cells. They showed that iron deficiency leads to downregulation of calcineurin signaling pathway leading to abrogated sensitivity at alkaline pH, salinity, and membrane stress. In general the ability of pathogenic fungi to adapt to host pH is critical for

survival and disease progression. This highlights the importance of continuing studies of these fundamental pH response pathways in pathogenic fungi in order to understand how these pathogens are adapted to the mammalian host and potentially identify new approaches for preventing or treating infections.

8. Lipid

Recent lines of evidence have provided a comprehensive amount of data regarding the role of lipids and many other lipid derivatives in establishment of various infectious diseases. *MDR* in yeast is closely linked to the status of membrane lipids. It has been already established that the associated changes in membrane lipid composition (phospholipid and ergosterol), its order (fluidity), and asymmetry could be important determinants in the drug susceptibilities of *Candida* cells [87]. Similarly, changes in membrane lipid composition between sensitive and resistant could also influence the action of antifungal drugs like azoles and thereby can form one of the factors responsible for drug resistance [12]. Most strikingly it has been observed that clinical as well as adapted azole-resistant isolates of *C. albicans* exhibit altered membrane phospholipid and sterol compositions [88]. Mukhopadhyay [89] demonstrated that there is an interaction between membrane ergosterol and sphingolipids, and a reduction in the content of either of these two components results in a disruption of this interaction, which has deleterious effects on the drug susceptibilities of *C. albicans* cells. Thus the fact that lipid could also play an important role in drug susceptibilities is becoming apparent from a wide range of recent studies. The importance of lipid signaling molecules in the development and pathogenicity of clinically important fungi has been highlighted [90]. In *C. albicans*, exposure to the oxylipin farnesol causes the regulation of specific genes involved in hyphal development, drug resistance, and iron acquisition. Farnesol increases resistance to oxidative stress in *C. albicans*. Through technologies such as lipidomics, a wider vision has been obtained to know about the diversity of lipid molecule which is not only limited to cellular functions but also to pathogenesis of diseases. Nowadays, lipidome-wide quantification of individual molecular lipid species (molecules with defined chemical structure) by absolute quantification provided a new approach to relate lipidomics and functional genomics studies [91]. Hameed [37] demonstrated the role of lipid homeostasis in iron mediated drug susceptibility of *C. albicans* where iron deprivation leads to enhanced drug susceptibility due to lowered ergosterol levels and increases membrane fluidity. Similarly the antifungal effect of curcumin has also been shown to be influenced by membrane lipid composition depicting marked changes in phosphoglyceride (PGL) species and also ergosterol depletion [92]. Recently, a high throughput lipid profiling has revealed some differences in lipid composition between azole sensitive and azole-resistant isolates highlighting fluctuations in phosphatidyl serine, mannosylinositolphosphorylceramides, and sterol esters levels indicating their compensatory role in

maintaining lipid homeostasis among most of the resistant *Candida* isolates [93]. Cross-talk between mitochondrial lipid homeostasis, cell wall integrity, and azole tolerance has been revealed showing significant changes in several lipid classes, particularly in plasma membrane microdomain-specific lipids such as mannosylinositolphosphorylceramides and ergosterol, and in a mitochondrial-specific phosphoglyceride, phosphatidyl glycerol [94]. The overall cellular lipid homeostasis is a critical factor in the observed FLC resistance [95]. Although the transcriptome, proteome, and interactome of several eukaryotic model organisms have been described in detail, lipidomes still remain relatively uncharacterized and will also improve our understanding of the molecular architecture of membrane domains and cellular organelles. Hence, the lipid associated changes of pathogenic fungi induced in response to infection might help in better understanding of the role of lipids in pathogenesis of *C. albicans* and thereby development of better therapeutic strategies.

9. Signaling

A focal point among the fungal pathogens is that signaling molecules have a key role in mediating cellular stress responses. Signal transduction pathways are crucial mechanisms that allow cells to sense and respond to diverse environmental cues. Exploiting these stress responses through blocking of signaling pathways may provide the foundation for new combination therapies to enhance the efficacy of our limited resources of clinically useful antifungal drugs. Jain [96] have examined the relationship between azole susceptibility and the cyclic AMP (cAMP) protein kinase A (PKA) signaling pathway. Likewise, a key regulator of cell signaling in all eukaryotes, calcineurin, provides a perfect example of the role of signaling molecules in mediating crucial responses to antifungal drugs [97]. Alonso-Monge [98] describe that MAPK cascades control most of the virulence factors characterized in *C. albicans*: cell morphology, superficial antigen (cell wall biogenesis), and response to oxidative and nitrosative stresses. These cascades allow opportunistic pathogens to recognize changes in their environment and take advantage of an impaired immunological system to cause infection. Recent studies reveal that Hsp90, a component of a chaperone complex induced by heat stress, governs drug resistance in fungi [99, 100]. Bastidas [101] showed the significance of signaling cascades in pathogenic fungi. They not only depicted the well known calcineurin pathway and that Hsp90 is an important antifungal target but also described the TOR signaling pathway in context with antifungal drug resistance. Thakur [102] showed mechanistically similar regulation of MDR like in vertebrates by the PXR nuclear receptor, revealing an unexpected functional analogy of fungal and metazoan regulators of MDR. Robbins [103] establish a novel role of nutrient signaling in azole resistance. They revealed that compromising the function of Tor kinase, a global regulator of growth and metabolism, could be an efficient strategy to control drug resistance. Another key cellular stress response pathway having implications in basal tolerance to azoles is the protein kinase C (PKC) mediated cell wall integrity pathway (CWI). Lafayette [104] established

a novel role of CWI and calcineurin signaling pathway in *C. albicans*, while de Dios [105] revealed the relevance of MAPK signaling cascades in fungal virulence. The emerging roles of GlcNAc as an activator and mediator of cellular signaling in *C. albicans* have also been reviewed [106]. Recently, even the significance of metabolic pathways as therapeutic targets and how their disruption can have both physiological and regulatory consequences have also been demonstrated [107]. The diterpene acid, phorbacin H, affects the activity of the cAMP-Efg1 pathway leading to an alteration of *C. albicans* morphology as was demonstrated recently [108]. Likewise antidimorphism activity of catechin was depicted by interfering with Cek1 phosphorylation and cAMP synthesis [109]. Kaba [42], demonstrated the involvement of MAPK HOG1 pathway with iron availability, and as already discussed in the previous sections, iron plays a vital role if one considers drug susceptibility in *C. albicans*. A new study showed for the first time that even lower eukaryote like *C. albicans* possesses a two-component response regulator protein [110]. Thus a deeper knowledge of the mechanism and regulation of various signaling cascades could help in the control of candidiasis as well as in the development of effective therapeutics against these severe infections.

10. Transcription Factors

Transcriptional regulation can be of significant importance in the development of antifungal resistance. It is important to understand the regulatory network controlling drug resistance in fungal pathogens. Different strategies have been adopted for isolation of regulators of multidrug transporters in *C. albicans*. One such approach is through the analysis of cis-acting elements in genes encoding multidrug efflux transporters. For instance the basal expression element (BEE) responsible for basal expression, the drug-responsive element (DRE) required for the response to drugs such as fluphenazine and estradiol, two steroid-responsive element (SRE) involved in the response to steroid hormones and the negative regulatory element (NRE), have been already deciphered [66, 67, 111]. One of the major breakthroughs was the discovery of *TAC1* regulator whose deletion abrogates the *CDR1/CDR2* expression in *C. albicans* clinical isolates resistant to azoles, thus demonstrating that *TAC1* was a major mediator of azole resistance due to the upregulation of the ABC transporter [68]. Apart from it other potential regulators of *CDR1* have also been reported which were identified through functional complementation in *S. cerevisiae*. For instance functional homologue of *PDR1/PDR3* in *S. cerevisiae*, namely, fluconazole resistance 1 (*FCR1*), was reported in *C. albicans*. Deletion of *FCR1* in *C. albicans* resulted in reduced susceptibility to FLC [112]. Similarly, *NDT80* inactivation in *C. albicans*, a gene similar to the *S. cerevisiae* *NDT80* gene, resulted in a decreased basal *CDR1* expression and a decreased *CDR1* inducibility in the presence of drugs [113]. Genome-wide transcription profiling was employed for the identification of an *MDR1* regulator or of factors binding to the *MDR1* promoter. By comparing the transcriptional profiles of three different *C. albicans* clinical

isolates over expressing *MDR1* with azole-susceptible parents, one of the commonly upregulated genes in the three isolates was orf19.7372 which was subsequently known as *MRR1* [114]. Another study reported that regulator of efflux pump 1 (*REPI*) acts as a negative regulator of *MDR1* which belongs to the transcription factor family including *NDT80* [115]. Similarly, *C. albicans* gene (*UPC2*) with homology to both *S. cerevisiae* genes has been identified and characterized [116]. Deletion of *UPC2* in *C. albicans* abrogates *ERG11* upregulation in response to azole drugs. A recent study has demonstrated the role of the mediator complex in the transcriptional response of multidrug transporter genes in *S. cerevisiae* and *C. glabrata* [102]; however, in *C. albicans*, the binding to the transcriptional activator of drug resistance genes (*TAC1*, *MRR1*, and *UPC2*) still remains hypothetical.

Gain-of-function (GOF) mutations in alleles of the transcription factors (*TAC1*, *MRR1*, *CgPDR1*, and *UPC2*) from azole-resistant isolates cause constitutive high expression of their drug resistance gene targets and thus azole resistance when expressed in an azole-susceptible background [68, 114, 117–122]. Nowadays, genome-wide analysis is an important tool to yield regulons of selected transcription factors for the elucidation of transcriptional regulatory network of drug resistance. Moreover, systematic deletion of transcription factor genes could also be an alternative approach to reveal transcriptional circuits responsible for drug resistance. Genome-wide genetic screens for the identification of additional targets involved in drug resistance or participating in the response of fungal pathogens to drug exposure will likely result in elucidating regulatory mechanisms. Recently, it has been uncovered that a network comprises 800 target genes and a tightly knit transcriptional regulatory circuit indicating that many aspects of commensalism and pathogenicity are crosslinked [123]. Dhamgaye [124] deciphered through RNA sequencing some novel genes including the transcription factor *CZF1* which are responsible for drug resistance.

Although the transcriptional regulation is considered to be the major step in MDR regulation, reports do exist nowadays revealing the significance of posttranscriptional regulation as well. Manoharlal [125] demonstrated that *CDR1* mRNA half-life was increased in azole-resistant as compared to azole sensitive isolates. Moreover, Manoharlal [126] further dissected the molecular basis of the above observed increased mRNA stability where it was demonstrated that loss of heterozygosity at the *PAP1* locus is linked to hyperadenylation and subsequent mRNA stability of *CDR1* transcripts in azole resistant isolates. Recently, posttranscriptional regulation of transcription factor Sef1 has been demonstrated in controlling virulence of *C. albicans* [127]. Thus despite poor knowledge about the posttranscriptional regulation of MDR in *C. albicans*, certainly, this aspect needs to be further elucidated to gain deeper insights.

11. Concluding Remarks

Today, the rapidly evolving issue of MDR demands the obvious need for dissecting completely new regulatory mechanisms that could be targeted to control MDR. Novel

mechanisms described earlier for pathogenic fungi clearly hold promise that can facilitate the development of better antifungal strategies to efficiently control the human fungal diseases. Elucidating these mechanisms may provide new foundations for antifungal chemotherapy and can present an exciting challenge for the future investigation.

Acknowledgment

The financial assistance in the form of Young Scientist Award (SR/FT/LS-12/2012) to Saif Hameed from Science and Engineering Research Board (SERB), New Delhi, is deeply acknowledged. The authors thank Professor S. M. Paul Khurana, Director, Amity Institute of Biotechnology for encouragement.

References

- [1] M. A. Pfaller and D. J. Diekema, "Epidemiology of invasive candidiasis: a persistent 873 public health problem," *Clinical Microbiology Reviews*, vol. 20, no. 1, pp. 133–163, 2007.
- [2] R. A. Calderone, *Candida and Candidiasis*, American Society for Microbiology Press, Washington, DC, USA, 2002.
- [3] F. C. Odds, *Candida and Candidosis: A Review and Bibliography*, London, UK, 1988.
- [4] T. C. White, K. A. Marr, and R. A. Bowden, "Clinical, cellular, and molecular factors that contribute to antifungal drug resistance," *Clinical Microbiology Reviews*, vol. 11, no. 2, pp. 382–402, 1998.
- [5] T. C. White, S. Holleman, F. Dy, L. F. Mirels, and D. A. Stevens, "Resistance mechanisms in clinical isolates of *Candida albicans*," *Antimicrobial Agents and Chemotherapy*, vol. 46, no. 6, pp. 1704–1713, 2002.
- [6] R. Franz, S. L. Kelly, D. C. Lamb, D. E. Kelly, M. Ruhnke, and J. Morschhäuser, "Multiple molecular mechanisms contribute to a stepwise development of fluconazole resistance in clinical *Candida albicans* strains," *Antimicrobial Agents and Chemotherapy*, vol. 42, no. 12, pp. 3065–3072, 1998.
- [7] J. B. Anderson, "Evolution of antifungal-drug resistance: mechanisms and pathogen fitness," *Nature Reviews Microbiology*, vol. 3, no. 7, pp. 547–556, 2005.
- [8] L. E. Cowen and W. J. Steinbach, "Stress, drugs, and evolution: the role of cellular signaling in fungal drug resistance," *Eukaryotic Cell*, vol. 7, no. 5, pp. 747–764, 2008.
- [9] D. C. Lamb, D. E. Kelly, W.-H. Schunck et al., "The mutation T315A in *Candida albicans* sterol 14 α -demethylase causes reduced enzyme activity and fluconazole resistance through reduced affinity," *Journal of Biological Chemistry*, vol. 272, no. 9, pp. 5682–5688, 1997.
- [10] R. Prasad, N. Gupta, and M. Gaur, "Molecular basis of antifungal resistance in pathogenic fungi," in *Pathogenic Fungi—Host Interactions and Emerging Strategies For Control*, G. San-Blas and R. A. Calderone, Eds., pp. 357–414, Caister Academic Press, Norfolk, UK, 2004.
- [11] G. D. Albertson, M. Niimi, R. D. Cannon, and H. F. Jenkinson, "Multiple efflux mechanisms are involved in *Candida albicans* fluconazole resistance," *Antimicrobial Agents and Chemotherapy*, vol. 40, no. 12, pp. 2835–2841, 1996.
- [12] A. Kohli, S. Smriti, K. Mukhopadhyay, A. Rattan, and R. Prasad, "In vitro low-level resistance to azoles in *Candida albicans* is associated with changes in membrane lipid fluidity and

- asymmetry," *Antimicrobial Agents and Chemotherapy*, vol. 46, no. 4, pp. 1046–1052, 2002.
- [13] D. Sanglard, F. Ischer, M. Monod, and J. Bille, "Cloning of *Candida albicans* genes conferring resistance to azole antifungal agents: characterization of CDR2, a new multidrug ABC transporter gene," *Microbiology*, vol. 143, no. 2, pp. 405–416, 1997.
- [14] R. Ben-Yaacov, S. Knoller, G. A. Caldwell, J. M. Becker, and Y. Koltin, "*Candida albicans* gene encoding resistance to benomyl and methotrexate is a multidrug resistance gene," *Antimicrobial Agents and Chemotherapy*, vol. 38, no. 4, pp. 648–652, 1994.
- [15] V. Gupta, A. Kohli, S. Krishnamurthy et al., "Identification of polymorphic mutant alleles of CaMDR1, a major facilitator of *Candida albicans* which confers multidrug resistance, and its in vitro transcriptional activation," *Current Genetics*, vol. 34, no. 3, pp. 192–199, 1998.
- [16] S. S. Pao, I. T. Paulsen, and M.H. Saier Jr., "Major facilitator superfamily," *Microbiology and Molecular Biology Reviews*, vol. 62, pp. 1–34, 1998.
- [17] G. G. Perrone, S.-X. Tan, and I. W. Dawes, "Reactive oxygen species and yeast apoptosis," *Biochimica et Biophysica Acta*, vol. 1783, no. 7, pp. 1354–1368, 2008.
- [18] N. Vázquez, T. J. Walsh, D. Friedman, S. J. Chanock, and C. A. Lyman, "Interleukin-15 augments superoxide production and microbicidal activity of human monocytes against *Candida albicans*," *Infection and Immunity*, vol. 66, no. 1, pp. 145–150, 1998.
- [19] C.-S. Hwang, G.-E. Rhie, S.-T. Kim et al., "Copper- and zinc-containing superoxide dismutase and its gene from *Candida albicans*," *Biochimica et Biophysica Acta*, vol. 1427, no. 2, pp. 245–255, 1999.
- [20] D. Kobayashi, K. Kondo, N. Uehara et al., "Endogenous reactive oxygen species is an important mediator of miconazole antifungal effect," *Antimicrobial Agents and Chemotherapy*, vol. 46, no. 10, pp. 3113–3117, 2002.
- [21] X.-Z. Wu, A.-X. Cheng, L.-M. Sun, S.-J. Sun, and H.-X. Lou, "Plagiochin E, an antifungal bis(benzyl), exerts its antifungal activity through mitochondrial dysfunction-induced reactive oxygen species accumulation in *Candida albicans*," *Biochimica et Biophysica Acta*, vol. 1790, no. 8, pp. 770–777, 2009.
- [22] Y. Xu, Y. Wang, L. Yan et al., "Proteomic analysis reveals a synergistic mechanism of fluconazole and berberine against fluconazole-resistant *Candida albicans*: endogenous ROS augmentation," *Journal of Proteome Research*, vol. 8, no. 11, pp. 5296–5304, 2009.
- [23] J. L. Da Rosa, V. L. Boyartchuk, L. J. Zhu, and P. D. Kaufman, "Histone acetyltransferase Rtt109 is required for *Candida albicans* pathogenesis," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 107, no. 4, pp. 1594–1599, 2010.
- [24] A. Shirai, S. Ueta, H. Maseda, H. Kourai, and T. Omasa, "Action of reactive oxygen species in the antifungal mechanism of gemini-pyridinium salts against yeast," *Biocontrol Science and Technology*, vol. 17, no. 2, pp. 77–82, 2012.
- [25] H. Miao, L. Zhao, C. Li, Q. Shang, H. Lu, and Z. Fu, "Inhibitory effect of shikonin on *Candida albicans* growth," *Biological & Pharmaceutical Bulletin*, vol. 35, no. 11, pp. 1956–1963, 2012.
- [26] R. S. Almeida, D. Wilson, and B. Hube, "*Candida albicans* iron acquisition within the host," *FEMS Yeast Research*, vol. 9, no. 7, pp. 1000–1012, 2009.
- [27] J. J. Bullen, H. J. Rogers, P. B. Spalding, and C. G. Ward, "Natural resistance, iron and infection: a challenge for clinical medicine," *Journal of Medical Microbiology*, vol. 55, no. 3, pp. 251–258, 2006.
- [28] M. A. Fischbach, H. Lin, D. R. Liu, and C. T. Walsh, "How pathogenic bacteria evade mammalian sabotage in the battle for iron," *Nature Chemical Biology*, vol. 2, no. 3, pp. 132–138, 2006.
- [29] I. Nyilasi, T. Papp, M. Takó, E. Nagy, and C. Vágvölgyi, "Iron gathering of opportunistic pathogenic fungi: a mini review," *Acta Microbiologica et Immunologica Hungarica*, vol. 52, no. 2, pp. 185–197, 2005.
- [30] J. Spacek, P. Jilek, V. Buchta, M. Forstl, M. Hronek, and M. Holeckova, "The serum levels of calcium, magnesium, iron and zinc in patients with recurrent vulvovaginal candidosis during attack, remission and in healthy controls," *Mycoses*, vol. 48, no. 6, pp. 391–395, 2005.
- [31] E. D. Weinberg, "The role of iron in protozoan and fungal infectious diseases," *Journal of Eukaryotic Microbiology*, vol. 46, no. 3, pp. 231–238, 1999.
- [32] D. Radisky and J. Kaplan, "Regulation of transition metal transport across the yeast plasma membrane," *Journal of Biological Chemistry*, vol. 274, no. 8, pp. 4481–4484, 1999.
- [33] M. E. Kuipers, H. G. De Vries, M. C. Eikelboom, D. K. F. Meijer, and P. J. Swart, "Synergistic fungistatic effects of lactoferrin in combination with antifungal drugs against clinical *Candida* isolates," *Antimicrobial Agents and Chemotherapy*, vol. 43, no. 11, pp. 2635–2641, 1999.
- [34] M. E. Kuipers, L. Beljaars, N. Van Beek et al., "Conditions influencing the in vitro antifungal activity of lactoferrin combined with antimycotics against clinical isolates of *Candida*: impact on the development of buccal preparations of lactoferrin," *APMIS*, vol. 110, no. 4, pp. 290–298, 2002.
- [35] M. E. Kuipers, J. Heegsma, H. I. Bakker et al., "Design and fungicidal activity of mucoadhesive lactoferrin tablets for the treatment of oropharyngeal candidosis," *Drug Delivery*, vol. 9, no. 1, pp. 31–38, 2002.
- [36] T. Prasad, A. Chandra, C. K. Mukhopadhyay, and R. Prasad, "Unexpected link between iron and drug resistance of *Candida* spp.: iron depletion enhances membrane fluidity and drug diffusion, leading to drug-susceptible cells," *Antimicrobial Agents and Chemotherapy*, vol. 50, no. 11, pp. 3597–3606, 2006.
- [37] S. Hameed, S. Dhamgaye, A. Singh, S. K. Goswami, and R. Prasad, "Calcineurin signaling and membrane lipid homeostasis regulates iron mediated multidrug resistance mechanisms in *Candida albicans*," *PLoS ONE*, vol. 6, no. 4, Article ID e18684, 2011.
- [38] S. Dhamgaye, F. Devaux, R. Manoharlal et al., "In vitro effect of malachite green on *Candida albicans* involves multiple pathways and transcriptional regulators UPC2 and STP2," *Antimicrobial Agents and Chemotherapy*, vol. 56, no. 1, pp. 495–506, 2012.
- [39] T. Kobayashi, H. Kakeya, T. Miyazaki et al., "Synergistic antifungal effect of lactoferrin with azole antifungals against *Candida albicans* and a proposal for a new treatment method for invasive candidiasis," *Japanese Journal of Infectious Diseases*, vol. 64, no. 4, pp. 292–296, 2011.
- [40] R. P. Singh, H. K. Prasad, I. Sinha, N. Agarwal, and K. Natarajan, "Cap2-HAP complex is a critical transcriptional regulator that has dual but contrasting roles in regulation of iron homeostasis in *Candida albicans*," *Journal of Biological Chemistry*, vol. 286, no. 28, pp. 25154–25170, 2011.
- [41] S. Brena, J. Cabezas-Olcoz, M. D. Moragues et al., "Fungicidal monoclonal antibody C7 interferes with iron acquisition in *Candida albicans*," *Antimicrobial Agents and Chemotherapy*, vol. 55, no. 7, pp. 3156–3163, 2011.

- [42] H. E. Kaba, M. Nimtz, P. P. Müller, and U. Bilitewski, "Involvement of the mitogen activated protein kinase Hog1p in the response of *Candida albicans* to iron availability," *BMC Microbiology*, vol. 13, article 16, 2013.
- [43] N. Hosogaya, T. Miyazaki, M. Nagi, K. Tanabe, A. Minematsu, and Y. Nagayoshi, "The heme-binding protein Dap1 links iron homeostasis to azole resistance via the P450 protein Erg11 in *Candida glabrata*," *FEMS Yeast Research*, vol. 13, no. 4, pp. 411–421, 2013.
- [44] T. Srikantha, K. J. Daniels, C. Pujol, E. Kim, and D. R. Soll, "Identification of genes upregulated by the transcription factor Bcr1 that are involved in impermeability, impenetrability and drug-resistance of *Candida albicans* $\alpha\alpha$ biofilms," *Eukaryot Cell*, 2013.
- [45] K. M. Comerford, T. J. Wallace, J. Karhausen, N. A. Louis, M. C. Montalto, and S. P. Colgan, "Hypoxia-inducible factor-1-dependent regulation of the multidrug resistance (MDR1) gene," *Cancer Research*, vol. 62, no. 12, pp. 3387–3394, 2002.
- [46] K. M. Comerford and S. P. Colgan, "Assessing oxygen sensitivity of the multidrug resistance (MDR) gene," *Methods in Enzymology*, vol. 381, pp. 376–387, 2004.
- [47] K. M. Comerford, E. P. Cummins, and C. T. Taylor, "c-Jun NH2-terminal kinase activation contributes to hypoxia-inducible factor 1 α -dependent P-glycoprotein expression in hypoxia," *Cancer Research*, vol. 64, no. 24, pp. 9057–9061, 2004.
- [48] C. K. Mukhopadhyay, B. Mazumder, and P. L. Fox, "Role of hypoxia-inducible factor-1 in transcriptional activation of ceruloplasmin by iron deficiency," *Journal of Biological Chemistry*, vol. 275, no. 28, pp. 21048–21054, 2000.
- [49] E. R. Setiadi, T. Doedt, F. Cottier, C. Noffz, and J. F. Ernst, "Transcriptional response of *Candida albicans* to hypoxia: linkage of oxygen sensing and Efg1p-regulatory networks," *Journal of Molecular Biology*, vol. 361, no. 3, pp. 399–411, 2006.
- [50] G. J. Sosinska, P. W. J. de Groot, M. J. T. de Mattos et al., "Hypoxic conditions and iron restriction affect the cell-wall proteome of *Candida albicans* grown under vagina-simulative conditions," *Microbiology*, vol. 154, no. 2, pp. 510–520, 2008.
- [51] A. J. P. Brown and N. A. R. Gow, "Regulatory networks controlling *Candida albicans* morphogenesis," *Trends in Microbiology*, vol. 7, no. 8, pp. 333–338, 1999.
- [52] A. Sonneborn, D. P. Bockmühl, and J. F. Ernst, "Chlamyospore formation in *Candida albicans* requires the Efg1p morphogenetic regulator," *Infection and Immunity*, vol. 67, no. 10, pp. 5514–5517, 1999.
- [53] C. Stichernoth and J. F. Ernst, "Hypoxic adaptation by Efg1 regulates biofilm formation by *Candida albicans*," *Applied and Environmental Microbiology*, vol. 75, no. 11, pp. 3663–3672, 2009.
- [54] J. Bonhomme, M. Chauvel, S. Goyard, P. Roux, T. Rossignol, and C. D'Enfert, "Contribution of the glycolytic flux and hypoxia adaptation to efficient biofilm formation by *Candida albicans*," *Molecular Microbiology*, vol. 80, no. 4, pp. 995–1013, 2011.
- [55] A. P. Carvalho, L. C. Gursky, R. T. Rosa et al., "Non-steroidal anti-inflammatory drugs may modulate the protease activity of *Candida albicans*," *Microbial Pathogenesis*, vol. 49, no. 6, pp. 315–322, 2010.
- [56] J. M. Synnott, A. Guida, S. Mulhern-Haughey, D. G. Higgins, and G. Butler, "Regulation of the hypoxic response in *Candida albicans*," *Eukaryotic Cell*, vol. 9, no. 11, pp. 1734–1746, 2010.
- [57] C. Stichernoth, A. Fraund, E. Setiadi, L. Giasson, A. Vecchiarelli, and J. F. Ernst, "Sch9 kinase integrates hypoxia and CO₂ sensing to suppress hyphal morphogenesis in *Candida albicans*," *Eukaryotic Cell*, vol. 10, no. 4, pp. 502–511, 2011.
- [58] J. F. Ernst and D. Tielker, "Responses to hypoxia in fungal pathogens," *Cellular Microbiology*, vol. 11, no. 2, pp. 183–190, 2009.
- [59] N. Grahl, K. M. Shepardson, D. Chung, and R. A. Cramer, "Hypoxia and fungal pathogenesis: to air or not to air?" *Eukaryotic Cell*, vol. 11, no. 5, pp. 560–570, 2012.
- [60] L. Romani, F. Bistoni, and P. Puccetti, "Adaptation of *Candida albicans* to the host environment: the role of morphogenesis in virulence and survival in mammalian hosts," *Current Opinion in Microbiology*, vol. 6, no. 4, pp. 338–343, 2003.
- [61] P. R. Gujjar, M. Finucane, and B. Larsen, "The effect of estradiol on *Candida albicans* growth," *Annals of Clinical and Laboratory Science*, vol. 27, no. 2, pp. 151–156, 1997.
- [62] X. Zhang, M. Essmann, E. T. Burt, and B. Larsen, "Estrogen effects on *Candida albicans*: a potential virulence-regulating mechanism," *Journal of Infectious Diseases*, vol. 181, no. 4, pp. 1441–1446, 2000.
- [63] N. D. Madani, P. J. Malloy, P. Rodriguez-Pombo, A. V. Krishnan, and D. Feldman, "*Candida albicans* estrogen-binding protein gene encodes an oxidoreductase that is inhibited by estradiol," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 91, no. 3, pp. 922–926, 1994.
- [64] P. J. Malloy, X. Zhao, N. D. Madani, and D. Feldman, "Cloning and expression of the gene from *Candida albicans* that encodes a high-affinity corticosteroid-binding protein," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 90, no. 5, pp. 1902–1906, 1993.
- [65] M. Das and A. Datta, "Steroid binding protein(s) in yeasts," *Biochemistry International*, vol. 11, no. 2, pp. 171–176, 1985.
- [66] M. De Micheli, J. Bille, C. Schueller, and D. Sanglard, "A common drug-responsive element mediates the upregulation of the *Candida albicans* ABC transporters CDR1 and CDR2, two genes involved in antifungal drug resistance," *Molecular Microbiology*, vol. 43, no. 5, pp. 1197–1214, 2002.
- [67] N. Kamani, N. Akhtar Gaur, S. Jha et al., "SRE1 and SRE2 are two specific steroid-responsive modules of candida drug resistance gene I (CDRI) promoter," *Yeast*, vol. 21, no. 3, pp. 219–239, 2004.
- [68] A. T. Coste, M. Karababa, F. Ischer, J. Bille, and D. Sanglard, "TAC1, transcriptional activator of CDR genes, is a new transcription factor involved in the regulation of *Candida albicans* ABC transporters CDR1 and CDR2," *Eukaryotic Cell*, vol. 3, no. 6, pp. 1639–1652, 2004.
- [69] D. Banerjee, B. Pillai, N. Karnani, G. Mukhopadhyay, and R. Prasad, "Genome-wide expression profile of steroid response in *Saccharomyces cerevisiae*," *Biochemical and Biophysical Research Communications*, vol. 317, no. 2, pp. 406–413, 2004.
- [70] K. M. Barnes, B. Dickstein, G. B. Cutler Jr., T. Fojo, and S. E. Bates, "Steroid transport, accumulation, and antagonism of P-glycoprotein in multidrug-resistant cells," *Biochemistry*, vol. 35, no. 15, pp. 4820–4827, 1996.
- [71] M. Sukhai and M. Piquette-Miller, "Regulation of the multidrug resistance genes by stress signals," *Journal of Pharmacy & Pharmaceutical Sciences*, vol. 3, no. 2, pp. 268–280, 2000.
- [72] S. Labialle, L. Gayet, E. Marthinet, D. Rigal, and L. G. Baggetto, "Transcriptional regulators of the human multidrug resistance 1 gene: recent views," *Biochemical Pharmacology*, vol. 64, no. 5-6, pp. 943–948, 2002.
- [73] D. Banerjee, G. Lelandais, S. Shukla et al., "Responses of pathogenic and nonpathogenic yeast species to steroids reveal the functioning and evolution of multidrug resistance transcriptional networks," *Eukaryotic Cell*, vol. 7, no. 1, pp. 68–77, 2008.

- [74] S. Gogoi, K. Shekarrao, A. Duarah, T. C. Bora, S. Gogoi, and R. C. Boruah, "A microwave promoted solvent-free approach to steroidal quinolines and their in vitro evaluation for antimicrobial activities," *Steroids*, vol. 77, no. 13, pp. 1438–1445, 2012.
- [75] D. Kakati, R. K. Sarma, R. Saikia, N. C. Barua, and J. C. Sarma, "Rapid microwave assisted synthesis and antimicrobial bioevaluation of novel steroidal chalcones," *Steroids*, vol. 78, no. 3, pp. 321–326, 2013.
- [76] J. F. Ernst and A. Schmidt, *Dimorphism in Human Pathogenic and Apathogenic Yeasts*, Karger, Bern, Switzerland, 2000.
- [77] J. F. Ernst, "Transcription factors in *Candida albicans*-environmental control of morphogenesis," *Microbiology*, vol. 146, no. 8, pp. 1763–1774, 2000.
- [78] H.-J. Lo, J.-S. Wang, C.-Y. Lin et al., "Efg1 involved in drug resistance by regulating the expression of ERG3 in *Candida albicans*," *Antimicrobial Agents and Chemotherapy*, vol. 49, no. 3, pp. 1213–1215, 2005.
- [79] T. Prasad, S. Hameed, R. Manoharlal et al., "Morphogenic regulator EFG1 affects the drug susceptibilities of pathogenic *Candida albicans*," *FEMS Yeast Research*, vol. 10, no. 5, pp. 587–596, 2010.
- [80] P. Vandeputte, S. Pradervand, F. Ischer, A. T. Coste, S. Ferrari, and K. Harshman, "Identification and functional characterization of Rca1, a transcription factor involved in both antifungal susceptibility and host response in *Candida albicans*," *Eukaryot Cell*, vol. 11, no. 7, pp. 916–931, 2012.
- [81] P. D. Roepe, "pH and multidrug resistance," *Novartis Foundation Symposium*, vol. 240, pp. 232–247, 2001.
- [82] D. Davis, "Adaptation to environmental pH in *Candida albicans* and its relation to pathogenesis," *Current Genetics*, vol. 44, no. 1, pp. 1–7, 2003.
- [83] D. Davis, R. B. Wilson, and A. P. Mitchell, "RIM101-dependent and-independent pathways govern pH responses in *Candida albicans*," *Molecular and Cellular Biology*, vol. 20, no. 3, pp. 971–978, 2000.
- [84] M. A. Ghannoum, B. Spellberg, S. M. Saporito-Irwin, and W. A. Fonzi, "Reduced virulence of *Candida albicans* PHR1 mutants," *Infection and Immunity*, vol. 63, no. 11, pp. 4528–4530, 1995.
- [85] R. Serrano, A. Ruiz, D. Bernal, J. R. Chambers, and J. Ariño, "The transcriptional response to alkaline pH in *Saccharomyces cerevisiae*: Evidence for calcium-mediated signalling," *Molecular Microbiology*, vol. 46, no. 5, pp. 1319–1333, 2002.
- [86] T. Bader, B. Bodendorfer, K. Schröppel, and J. Morschhäuser, "Calcineurin is essential for virulence in *Candida albicans*," *Infection and Immunity*, vol. 71, no. 9, pp. 5344–5354, 2003.
- [87] K. Mukhopadhyay, A. Kohli, and R. Prasad, "Drug susceptibilities of yeast cells are affected by membrane lipid composition," *Antimicrobial Agents and Chemotherapy*, vol. 46, no. 12, pp. 3695–3705, 2002.
- [88] J. Löffler, H. Einsele, H. Hebart, U. Schumacher, C. Hrastnik, and G. Daum, "Phospholipid and sterol analysis of plasma membranes of azole-resistant *Candida albicans* strains," *FEMS Microbiology Letters*, vol. 185, no. 1, pp. 59–63, 2000.
- [89] K. Mukhopadhyay, T. Prasad, P. Saini, T. J. Pucadyil, A. Chattopadhyay, and R. Prasad, "Membrane sphingolipid-ergosterol interactions are important determinants of multidrug resistance in *Candida albicans*," *Antimicrobial Agents and Chemotherapy*, vol. 48, no. 5, pp. 1778–1787, 2004.
- [90] J. M. Shea and M. Del Poeta, "Lipid signaling in pathogenic fungi," *Current Opinion in Microbiology*, vol. 9, no. 4, pp. 352–358, 2006.
- [91] C. S. Ejsinga, J. L. Sampaio, V. Surendranatha, E. Duchoslavb, K. Ekroosc, and R. W. Klemma, "Global analysis of the yeast lipidome by quantitative shotgun mass spectrometry," *Proceedings of the National Academy of Sciences*, vol. 106, no. 7, pp. 2136–2141, 2009.
- [92] M. Sharma, S. Dhamgaye, A. Singh, and R. Prasad, "Lipidome analysis reveals antifungal polyphenol curcumin affects membrane lipid homeostasis," *Frontiers in Bioscience*, no. 4, pp. 1195–1209, 2012.
- [93] A. Singh and R. Prasad, "Comparative lipidomics of azole sensitive and resistant clinical isolates of *Candida albicans* reveals unexpected diversity in molecular lipid imprints," *PLoS ONE*, vol. 6, no. 4, Article ID e19266, 2011.
- [94] A. Singh, V. Yadav, and R. Prasad, "Comparative lipidomics in clinical isolates of *Candida albicans* reveal crosstalk between mitochondria, cell wall integrity and azole resistance," *PLoS One*, vol. 7, no. 6, Article ID e39812, 2012.
- [95] A. Singh, K. K. Mahto, and R. Prasad, "Lipidomics and in vitro azole resistance in *Candida albicans*," *OMICS*, vol. 17, no. 2, pp. 84–93, 2013.
- [96] P. Jain, I. Akula, and T. Edlind, "Cyclic AMP signaling pathway modulates susceptibility of *Candida* species and *Saccharomyces cerevisiae* to antifungal azoles and other sterol biosynthesis inhibitors," *Antimicrobial Agents and Chemotherapy*, vol. 47, no. 10, pp. 3195–3201, 2003.
- [97] W. J. Steinbach, J. L. Reedy, R. A. Cramer Jr., J. R. Perfect, and J. Heitman, "Harnessing calcineurin as a novel anti-infective agent against invasive fungal infections," *Nature Reviews Microbiology*, vol. 5, no. 6, pp. 418–430, 2007.
- [98] R. Alonso-Monge, E. Román, D. M. Arana, J. Pla, and C. Nombela, "Fungi sensing environmental stress," *Clinical Microbiology and Infection*, vol. 15, no. 1, pp. 17–19, 2009.
- [99] L. E. Cowen and S. Lindquist, "Cell biology: Hsp90 potentiates the rapid evolution of new traits: drug resistance in diverse fungi," *Science*, vol. 309, no. 5744, pp. 2185–2189, 2005.
- [100] L. E. Cowen, S. D. Singha, J. R. Köhlerb, C. Collinsa, A. K. Zaasc, and W. A. Schellc, "Harnessing Hsp90 function as a powerful, broadly effective therapeutic strategy for fungal infectious disease," *Proceedings of the National Academy of Sciences*, vol. 106, no. 8, pp. 2818–2823, 2009.
- [101] R. J. Bastidas, J. L. Reedy, H. Morales-Johansson, J. Heitman, and M. E. Cardenas, "Signaling cascades as drug targets in model and pathogenic fungi," *Current Opinion in Investigational Drugs*, vol. 9, no. 8, pp. 856–864, 2008.
- [102] J. K. Thakur, H. Arthanari, F. Yang et al., "A nuclear receptor-like pathway regulating multidrug resistance in fungi," *Nature*, vol. 452, no. 7187, pp. 604–609, 2008.
- [103] N. Robbins, C. Collins, J. Morhayim, and L. E. Cowen, "Metabolic control of antifungal drug resistance," *Fungal Genetics and Biology*, vol. 47, no. 2, pp. 81–93, 2010.
- [104] S. L. Lafayette, C. Collins, A. K. Zaas et al., "PKC signaling regulates drug resistance of the fungal pathogen *Candida albicans* via circuitry comprised of mkc1, calcineurin, and hsp90," *PLoS Pathogens*, vol. 6, no. 8, Article ID e1001069, pp. 79–80, 2010.
- [105] C. H. de Dios, E. Román, R. A. Monge, and J. Pla, "The role of MAPK signal transduction pathways in the response to oxidative stress in the fungal pathogen *Candida albicans*: implications in virulence," *Current Protein and Peptide Science*, vol. 11, no. 8, pp. 693–703, 2010.
- [106] J. B. Konopka, "N-acetylglucosamine (GlcNAc) functions in cell signaling," *Scientifica*, vol. 2012, Article ID 489208, 15 pages, 2012.

- [107] J. V. Desai, V. M. Bruno, S. Ganguly, R. J. Stamper, K. F. Mitchell, and N. Solis, "Regulatory role of glycerol in *Candida albicans* biofilm formation," *MBio*, vol. 4, no. 2, 2013.
- [108] S. H. Lee, J. E. Jeon, C. H. Ahn, S. C. Chung, J. Shin, and K. B. Oh, "Inhibition of yeast-to-hypha transition in *Candida albicans* by phorbacin H isolated from phorbas sp," vol. 97, no. 7, pp. 3141–3148, 2013.
- [109] H. Saito, M. Tamura, K. Imai, T. Ishigami, and K. Ochiai, "Catechin inhibits *Candida albicans* dimorphism by disrupting Cek1 phosphorylation and cAMP synthesis," *Microbial Pathogenesis*, vol. 56, pp. 16–20, 2013.
- [110] J. Mavrianos, E. L. Berkow, C. Desai, A. Pandey, M. Batish, and M. J. Rabadi, "Mitochondrial two-component signaling systems in *Candida albicans*," *Eukaryot Cell*, 2013.
- [111] N. A. Gaur, N. Puri, N. Karnani, G. Mukhopadhyay, S. K. Goswami, and R. Prasad, "Identification of a negative regulatory element which regulates basal transcription of a multidrug resistance gene CDR1 of *Candida albicans*," *FEMS Yeast Research*, vol. 4, no. 4-5, pp. 389–399, 2004.
- [112] D. Talibi and M. Raymond, "Isolation of a putative *Candida albicans* transcriptional regulator involved in pleiotropic drug resistance by functional complementation of a *pdr1 pdr3* mutation in *saccharomyces cerevisiae*," *Journal of Bacteriology*, vol. 181, no. 1, pp. 231–240, 1999.
- [113] C.-G. Chen, Y.-L. Yang, H.-I. Shih, C.-L. Su, and H.-J. Lo, "CaNdt80 is involved in drug resistance in *Candida albicans* by regulating CDR1," *Antimicrobial Agents and Chemotherapy*, vol. 48, no. 12, pp. 4505–4512, 2004.
- [114] J. Morschhäuser, K. S. Barker, T. T. Liu, J. Bläß-Warmuth, R. Homayouni, and P. D. Rogers, "The transcription factor *Mrr1p* controls expression of the MDR1 efflux pump and mediates multidrug resistance in *Candida albicans*," *PLoS Pathogens*, vol. 3, no. 11, article e164, 2007.
- [115] C.-G. Chen, Y.-L. Yang, K.-Y. Tseng et al., "Replp negatively regulating MDR1 efflux pump involved in drug resistance in *Candida albicans*," *Fungal Genetics and Biology*, vol. 46, no. 9, pp. 714–720, 2009.
- [116] P. M. Silver, B. G. Oliver, and T. C. White, "Role of *Candida albicans* transcription factor *Upc2p* in drug resistance and sterol metabolism," *Eukaryotic Cell*, vol. 3, no. 6, pp. 1391–1397, 2004.
- [117] A. T. Coste, V. Turner, F. Ischer et al., "A mutation in *Tac1p*, a transcription factor regulating CDR1 and CDR2, is coupled with loss of heterozygosity at chromosome 5 to mediate antifungal resistance in *Candida albicans*," *Genetics*, vol. 172, no. 4, pp. 2139–2156, 2006.
- [118] H.-F. Tsai, A. A. Krol, K. E. Sarti, and J. E. Bennett, "*Candida glabrata* PDR1, a transcriptional regulator of a pleiotropic drug resistance network, mediates azole resistance in clinical isolates and petite mutants," *Antimicrobial Agents and Chemotherapy*, vol. 50, no. 4, pp. 1384–1392, 2006.
- [119] J.-P. Vermitsky, K. D. Earhart, W. L. Smith, R. Homayouni, T. D. Edlind, and P. D. Rogers, "Pdr1 regulates multidrug resistance in *Candida glabrata*: gene disruption and genome-wide expression studies," *Molecular Microbiology*, vol. 61, no. 3, pp. 704–722, 2006.
- [120] N. Dunkel, J. Bläß, P. D. Rogers, and J. Morschhäuser, "Mutations in the multi-drug resistance regulator MRR1, followed by loss of heterozygosity, are the main cause of MDR1 overexpression in fluconazole-resistant *Candida albicans* strains," *Molecular Microbiology*, vol. 69, no. 4, pp. 827–840, 2008.
- [121] N. Dunkel, T. T. Liu, K. S. Barker, R. Homayouni, J. Morschhäuser, and P. D. Rogers, "A gain-of-function mutation in the transcription factor *Upc2p* causes upregulation of ergosterol biosynthesis genes and increased fluconazole resistance in a clinical *Candida albicans* isolate," *Eukaryotic Cell*, vol. 7, no. 7, pp. 1180–1190, 2008.
- [122] S. Ferrari, F. Ischer, D. Calabrese et al., "Gain of function mutations in *CgPDR1* of *Candida glabrata* not only mediate antifungal resistance but also enhance virulence," *PLoS Pathogens*, vol. 5, no. 1, Article ID e1000268, 2009.
- [123] J. C. Pérez, C. A. Kumamoto, and A. D. Johnson, "*Candida albicans* commensalism and pathogenicity are intertwined traits directed by a tightly knit transcriptional regulatory circuit," *PLOS Biology*, vol. 11, no. 3, Article ID e1001510, 2013.
- [124] S. Dhamgaye, M. Bernard, G. Lelandais, O. Sismeiro, S. Lemoine, and J. Y. Coppée, "RNA sequencing revealed novel actors of the acquisition of drug resistance in *Candida albicans*," *BMC Genomics*, vol. 13, article 396, 2012.
- [125] R. Manoharlal, N. A. Gaur, S. L. Panwar, J. Morschhäuser, and R. Prasad, "Transcriptional activation and increased mRNA stability contribute to overexpression of CDR1 in azole-resistant *Candida albicans*," *Antimicrobial Agents and Chemotherapy*, vol. 52, no. 4, pp. 1481–1492, 2008.
- [126] R. Manoharlal, J. Gorantala, M. Sharma, D. Sanglard, and R. Prasad, "PAP1 [poly(A) polymerase 1] homozygosity and hyperadenylation are major determinants of increased mRNA stability of CDR1 in azole-resistant clinical isolates of *Candida albicans*," *Microbiology*, vol. 156, no. 2, pp. 313–326, 2010.
- [127] C. Chen and S. M. Noble, "Post-transcriptional regulation of the *Sef1* transcription factor controls the virulence of *Candida albicans* in its mammalian host," *PLoS Pathogens*, vol. 8, no. 11, Article ID e1002956, 2012.