



# *Cryptocotyle lata* (Trematoda: Heterophyidae) Adult from a Korean Raccoon Dog, *Nyctereutes procyonoides koreensis*

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**Abstract:** Total 513 heterophyid flukes were collected from a carcass of wild Korean raccoon dog, *Nyctereutes procyonoides koreensis*, in Korea. With morphological and molecular characteristics, the flukes were identified to *Cryptocotyle lata*. The adult *C. lata* were minute, transparent, pentagonal, 522 µm long by 425 µm wide. Ceca extended into post-testicular region. Ventrogenital sac elliptical, 79 µm by 87 µm with genital pore and ventral sucker. Two testes semielliptical and slightly lobed, located in the posterior region, right testis 173 µm by 155 µm, left testis 130 µm by 134 µm. In a phylogenetic tree, the fluke specimen of this study was grouped with *C. lata* divergent from *Cryptocotyle lingua*. We report here *N. procyonoides koreensis* first as a natural definitive host of *C. lata*.

**Key words:** *Cryptocotyle lata*, *Nyctereutes procyonoides koreensis*, Korean raccoon dog, natural definitive host, Korea

The family Heterophyidae contains a number of intestinal trematodes that can be found in various vertebrates including birds and mammals and they also have potential of zoonosis [1]. In the Republic of Korea (Korea), more than 14 species of heterophyid flukes, i.e., *Metagonimus* spp., *Heterophyes nocens*, *Pygidioopsis summa*, *Heterophyopsis continua*, *Stictodora fuscata*, *Stictodora lari*, *Acanthotrema felis*, *Stellantchasmus falcatus*, *Centrocestus armatus*, *Procerovum varium*, *Cryptocotyle* sp., *Cryptocotyle concava* and *Euryhelminx squamula*, were reported from carnivore (feral or stray cat and raccoon dog) [2-5]. Of these heterophyid flukes, members of genus *Cryptocotyle* are mainly developed in the intestines of fish-eating birds and mammals as adult worms [6-9].

Fluke members in the genus *Cryptocotyle* are mainly divided into 2 types, i.e., linguiform and pentagonal or ovoid form, by the body shape. A total of 4 species, i.e., *C. dominicana*, *C. jeju-na*, *C. lingua* and *C. thapari*, were described as the linguiform species [6,10-15]. As the pentagonal or ovoid form species, more than 9 ones, i.e., *C. concava*, *C. cryptocotyloides*, *C. quin-*

*queangularis*, *C. badamshini*, *C. delamurei*, *C. misgurni*, *C. gyrincola*, *C. yamashitai* and *C. lata*, were recorded in the literatures [5,8,10,15-24].

Recently, Tatonova and Besprozvannykh described a new species of genus *Cryptocotyle*, *C. lata* n. sp. [24]. They recovered adult worms from domestic duck, *Anas platyrhynchos*, which were experimentally infected with metacercariae collected from the second intermediate hosts, *Rhynchocypris percnurus mantschuricus* and *Rana dybowskii*, in Russia [24]. However, the natural definitive hosts of *C. lata* have not been reported yet.

A wild raccoon dog was found in Hyoja-dong (dong = neighborhood) (37°52'02.6"N, 127°44'44.3"E), Chuncheon-si (si = City), Gangwon-do (do = Province), Korea in May 2015. The rescued raccoon dog was moved to the Gangwon Wildlife Medical Rescue Center (WMRC) at Kangwon National University, and found dead at WMRC in the next day (May 15, 2015). A total of 513 trematode worms were recovered from the small intestines of the raccoon dog. For the morphological study, we photographed the flukes before and after fixation with 70% alcohol. A typical image was created using tracing paper. All measurements (n = 10) are presented as arithmetic mean (range) in micrometer (µm).

Genomic DNA was extracted from the flukes using the DNeasy® Blood and Tissue Kit (Qiagen, Alameda, California, USA) according to the manufacturer's instructions. Based on the 18S ribosomal RNA gene sequences of *Euryhelminx costari-*

•Received 22 September 2021, revised 22 November 2021, accepted 22 November 2021.

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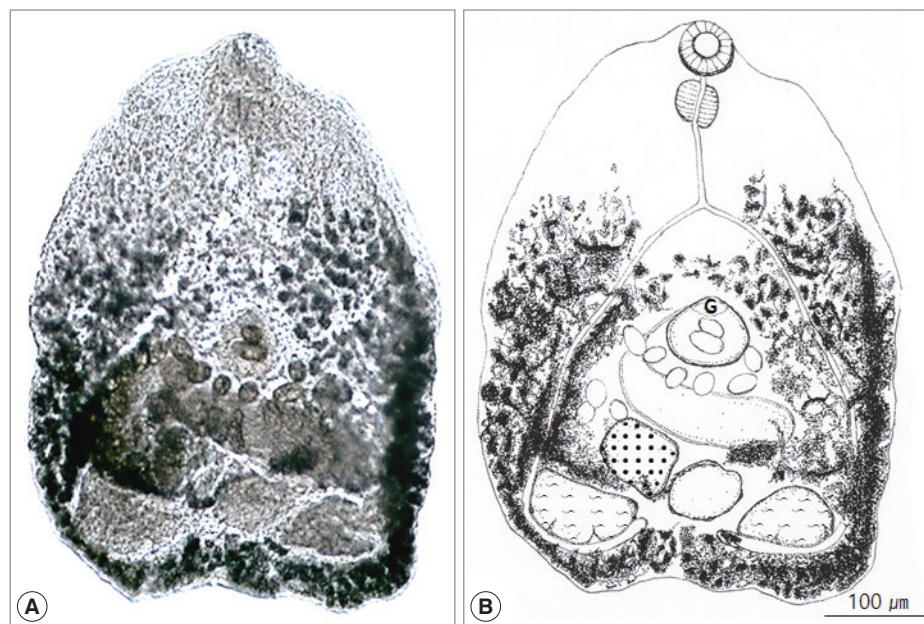
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*censis* (AB5217897), primers were designed using the online tool Primer3Plus (<http://www.bioinformatics.nl/cgi-bin/primer3plus/primer3plus.cgi>). The sequences of primers employed to detect the 18S rRNA gene (DNA) of *Cryptocotyle* specimens were as follows: 5'-TGCCTTATCCCAGGTACGAC-3' (forward primer) and 5'-CAGTTCAACTCCACCCGTTT-3' (reverse primer). The primer set was designed to yield 979 bp amplicon. The reaction mix was prepared with EmeraldAmp GT PCR Master Mix (Takara, Shiga, Japan) and 1  $\mu$ l of the fluke's DNA aliquot. PCR was performed using MyCycler Personal Thermal Cycler (Bio-Rad Laboratories, Hercules, California, USA). PCR was performed under standard condition with 30-35 cycles of 95°C for 30 sec, annealing at 60°C for 30 sec, and extension at 72°C for 1 min. The PCR amplicons were electrophoresed on 1.2% agarose gel, and the target DNA bands were purified using the QIAquick PCR Purification Kit (Qiagen, Alameda, California, USA). The PCR amplicons were directly sequenced using the ABI Prism Big Dye terminator v. 3.0 ready reaction cycle sequencing kits (Applied Biosystems, Foster City, California, USA) and the PCR primers. After trimming, the amplicon revealed 784 bp sequence, and aligned using Clustal Omega program (Clustal O, EMBL-EBI, Hinxton, Cambridgeshire, United Kingdom). A phylogenetic tree was constructed with homologous sequences retrieved from NCBI by using neighbor-joining (NJ) method [25].

The morphological characters of total 513 heterophyid flukes in this study were as follow (Fig. 1). Body small, transparent, pentagonal, 522 (339-646)  $\mu$ m long and 425 (300-493)  $\mu$ m wide. Oral sucker subterminal, 59 (52-65)  $\mu$ m by 66 (53-71)  $\mu$ m. Pharynx large, spherical, 56 (38-61)  $\mu$ m by 47 (43-49)  $\mu$ m. Esophagus slender, straight, long, 87 (79-96)  $\mu$ m in length (Fig. 1B). Ceca extended into post-testicular region. Ventrogenital sac elliptical, 79 (71-88)  $\mu$ m by 87 (83-91)  $\mu$ m with genital pore and ventral sucker. Seminal vesicle saccate, 98 (59-121)  $\mu$ m by 185 (117-205)  $\mu$ m. Ovary spherical, 128 (86-148)  $\mu$ m by 164 (109-180)  $\mu$ m, lying dextral to median line. Seminal receptacle elliptical and large, 89 (68-118)  $\mu$ m by 95 (77-131)  $\mu$ m, just behind of the left margin of ovary. Two testes semielliptical and slightly lobed, located pararely in the posterior region, right testis 173 (168-190)  $\mu$ m by 155 (150-157)  $\mu$ m, left testis 130 (110-145)  $\mu$ m by 134 (120-147)  $\mu$ m. Vitellaria follicular along mainly extra-cecal margin from posterior bifurcation of the intestinal ceca to posterior end of body. Uterus passed transversally from left to right, then from right to left in field surrounded by ventrogenital sac, ceca and gonads, containing 10-45 eggs. Eggs operculated, with slight polar thickening, undeveloped when laid, and 35 (32-37)  $\mu$ m by 18 (17-20)  $\mu$ m. These morphological characteristics implied the minute heterophyid flukes to be *C. lata*.

To determine the phylogenetic relationship with other para-



**Fig. 1.** *Cryptocotyle lata* adult recovered from the small intestine of a Korean raccoon dog. (A) Unstained fluke. (B) Schematic drawing. G, ventrogenital sac.

sites, we analyzed the genomic DNA of the specimens using phylogenetic analysis. The partial sequence of 18S rRNA of *Cryptocotyle* specimens was registered into GenBank under an accession number (MZ436998.1) for *Cryptocotyle* specimens. The sequencing information was compared to those of other parasites, including *E. costaricensis* (AB521800.1), *Euryhelms zelleri* (KM594133.1) and *C. lingua* (MW361240.1). The partial 18S rRNA sequence (784 bp) of the *Cryptocotyle* specimens contained 1 variable point (99% similarity) that belonged to the same clade of *C. lata*. The 18S rRNA sequence of *Cryptocotyle* specimens was inferred using the NJ method. The *Cryptocotyle* specimen showed high similarity (97%) to *C. lingua* (Fig. 2). The sequence of *Cryptocotyle* specimen was closely grouped with *C. lata* divergent from *C. lingua*. Based on these morphological characteristics and phylogenetic features, our *Cryptocotyle* specimen (MZ436998.1) was identified to *C. lata*.

The species of the genus *Cryptocotyle* are world-widely distributed in the marine and freshwater environment [6]. Although the literature on the life cycle of *Cryptocotyle* species is scarce, snails and fish are known as first and second intermediate hosts, respectively. And the definitive hosts for *Cryptocotyle* spp. are also known as fish-eating birds and mammals [8,9,

17].

As far as we know, the species known only from mammals are *C. badamshini* from Caspian seals [20], *C. delamurei* from Commander fur seal [21], *C. quinqueangularis* from cat [15], *C. yamashitai* from red fox [23], and *C. thapari* from River Otter [15]. Only two, *C. cryptocotyloides* has been reported only from birds (the arctic loon, *Gavia arctica*) in Russia [18] and *C. dominicana* from Kelp gull [6]. Four species are known from both birds and mammals; *C. concava* from many different birds and mammals [5,10,15-17,26]; *C. jejuna* from gulls and dogs [11,12,15]; *C. lingua* primarily from marine birds, marine mammals, dogs, cats, and *Rattus* [12,15]. Among them, *C. concava* of carnivorous mammals such as feral cats and raccoons and *C. lingua* of *Larus argentatus*, a herring gull, have been reported by former researchers in Korea [2,3,27]. On the other hand, there are some species whose natural final host is unknown. Adults of *C. gyrenicola* were obtained experimentally by feeding cats and pigeons with metacercariae from tadpoles [22], and adults of *C. misgurni* were obtained experimentally by feeding golden hamsters with metacercariae encysted in loaches [8]. Adults of *C. lata* were obtained experimentally by feeding ducklings with metacercariae encysted in tadpoles and

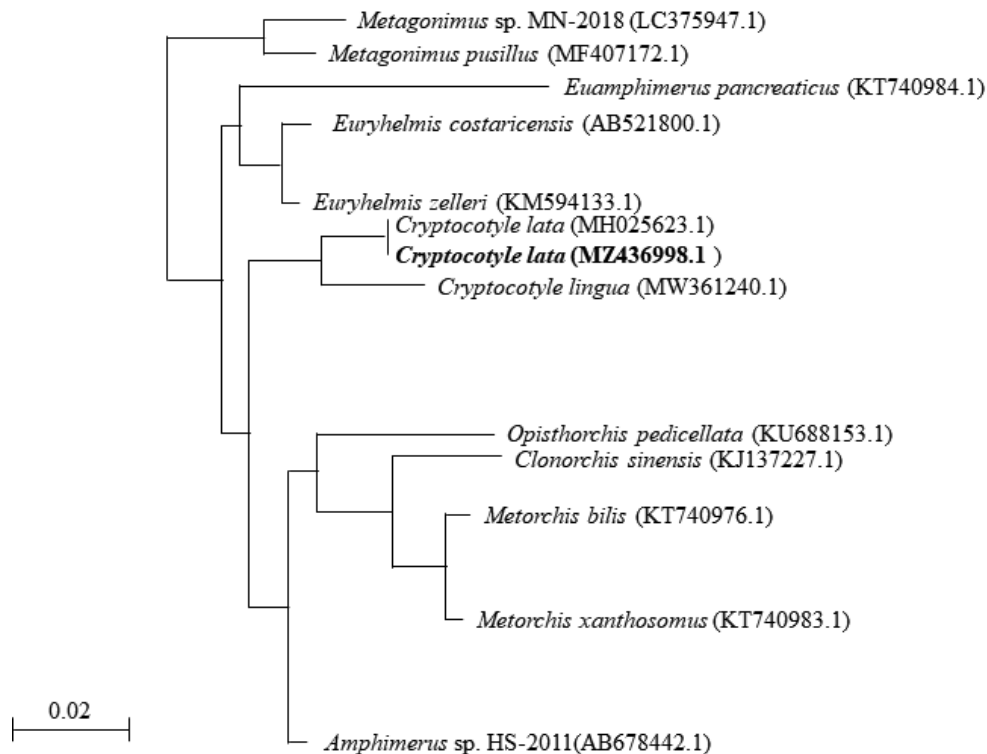


Fig. 2. A phylogenetic tree based on *Cryptocotyle lata* 18S rRNA sequences.

**Table 1.** Comparative measurements of *Cryptocotyle* species of which body is pentagonal or ovoid form

	This study	<i>C. lata</i> [24]	<i>C. concava</i> [2]	<i>C. cryptocotyloides</i> [18]	<i>C. quinqueangularis</i> [24]	<i>C. badamshini</i> [20]	<i>C. delamurei</i> [21]	<i>C. misgurni</i> [8]	<i>C. gyriticola</i> [22]	<i>C. yamashitai</i> [23]
Body length	339-646 (522)	554-739	500-1,170	1,039-1,092	850	564	220-505	301-424	442-700	550-585
Body width	300-493 (425)	570-770	468-732	625-689	740	434	176-384	348-455	411-775	480-505
Oral sucker length	52-65 (59)	50-73	50-84	64-74	60	48	40-60	29.5-45.6	71.2	26-32
Oral sucker width	53-71 (66)	58-85	70-105	74-85	60	52	44-60	38.0-50.1	46	59-78
Pharynx length	38-61 (56)	39-62	28-49	47	46	42	29	27.7-42.2	47.5	35-40
Pharynx width	43-49 (47)	39-62	25-55	47	46	34	21	24.3-34.9	47.5	40-45
Esophagus length	79-96 (87)	35-96	63-230	65-95	-	-	-	16.1-48.1	83.12	37-62
Ventrogenital sac length	71-88 (79)	69-85	35-88	127-150	60	113	46-92	33.7-55.2	120	62-81
Ventrogenital sac width	83-91 (87)	81-96	48-88	127-150	60	114	46-93	45.5-62.2	120	62-81
Ovary length	86-148 (128)	92-154	38-133	53-85	40	39	40-48	78.7-147	-	155
Ovary width	109-180 (164)	112-173	50-120	159	80	109	24-36	43.8-74.6	-	62
Testes length	110-190 (153)	96-154	109-320	105-187	90-100	93-99	40-68	114-171	-	142-167
Testes width	120-157 (144)	154-243	50-179	207-212	130-150	149-165	52-96	54.1-112	-	42-62
Egg length	32-37 (35)	35-39	23-28	40	32	43	28-44	31.2-36.7	30-33	32-37
Egg width	17-20 (18)	19-23	11-17	20	18	15	10-16	15.8-18.9	15-16.6	18-21

Units: micrometer ( $\mu\text{m}$ ).

fish [24]. We also reported a heterophyid fluke, *E. squamula*, similar to *C. lata* from a Korean raccoon dog in 2021 [5]. The morphological characteristic of *E. squamula* is different from other heterophyid flukes in body having rectangular and broader than length. In the present study, the fluke observed in Korean raccoon dog was identified as *C. lata*. This is the first record as the natural definitive host of this parasite.

There are 13 species of genus *Cryptocotyle* worldwide and the species have two form of body shape: linguiform and pentagonal or ovoid form. When identifying the specimen of the present study, we only compared it with 9 species having pentagonal or ovoid form, excluding 4 species which have linguiform: *C. lingua*, *C. jejuna*, *C. thapari*, *C. dominicana*. In this study, the body length measured in 522 (339-646)  $\mu\text{m}$ , so we also excluded pentagonal or ovoid form species having difference in body length with the specimen; *C. concava* 500-1,170  $\mu\text{m}$ , *C. quinqueangularis* 850  $\mu\text{m}$ , *C. cryptocotyloides* 1,039-1,092  $\mu\text{m}$  (Table 1). *C. badamshini* and *C. delamureis* had much smaller ovaries, and *C. misgurni* and *C. gyriticola* had significantly different body length and width ratios from those of this study. There are few morphological differences between *C. yamashitai* and *C. lata*, except for very small differences. However, the genetic information of *C. yamashitai* is not available. It is impossible to compare phylogenetic similarity of them now.

In this study, the genetic divergence of the *Cryptocotyle* specimens revealed their conspecific relationship with *C. lingua* and exact correspondence with *C. lata*.

Conclusively, it was confirmed for the first time that based on the morphological and molecular characteristics, *C. lata* indigenously infected the Korean raccoon dog, *N. procyonoides koreensis* in Korea. Additional life cycle studies on the intermediate hosts and other natural definitive hosts of this fluke should be performed in near future in Korea.

## ACKNOWLEDGMENT

This research was supported by Basic Science Research Program through the National Research Foundation of Korea (NRF) funded by the Ministry of Education (Grant No. 2017 RIDIAIB06031728).

## CONFLICT OF INTEREST

The authors declare no conflict of interest related to this study.



## REFERENCES

- Santos CP, Borges JN. Current knowledge of small flukes (Digenea: Heterophyidae) from South America. *Korean J Parasitol* 2020; 58: 373-386. <https://doi.org/10.3347/kjp.2020.58.4.373>
- Chai JY, Bahk YY, Sohn WM. Trematodes recovered in the small intestine of stray cats in the Republic of Korea. *Korean J Parasitol* 2013; 51: 99-106. <https://doi.org/10.3347/kjp.2013.51.1.99>
- Sohn WM, Chai JY. Infection status with helminthes in feral cats purchased from a market in Busan, Republic of Korea. *Korean J Parasitol* 2005; 43: 93-100. <https://doi.org/10.3347/kjp.2005.43.3.93>
- Shin EH, Park JH, Guk SM, Kim JL, Chai JY. Intestinal helminth infections in feral cats and a raccoon dog on Aphaedo Island, Shinan-gun, with a special note on *Gymnophalloides seoi* infection in cats. *Korean J Parasitol* 2009; 47: 189-191. <https://doi.org/10.3347/kjp.2009.47.2.189>
- Kim HC, Hong EJ, Ryu SY, PJ, Cho JG, Yu DH, Chae JS, Choi KS, Park BK. *Euryhelms squamula* (Digenea: Heterophyidae) recovered from Korean raccoon dog, *Nyctereutes procyonoides koreensis*, in Korea. *Korean J Parasitol* 2021; 59: 303-309. <https://doi.org/10.3347/kjp.2021.59.3.303>
- Yamaguti S. A synopsis of digenetic trematodes of vertebrates. Vol. I. Keigaku Publishing Co. Tokyo, Japan. 1971, pp 1074.
- Casalins LM, Arbetman MP, Viozzi GP, Flores VR. A new species of *Cryptocotyle* (Eigenea: Heterophyidae) infecting kelp gull and a galaxiid fish in Patagonian freshwater environments: morphological and molecular analyses. *J Parasitol* 2020; 106: 203-210. <https://doi.org/10.1645/18-44>
- Ohyama F, Okino T, Ushirogawa H. *Massaliatrema misgurni* n. sp. (Trematoda: Heterophyidae) whose metacercariae encyst in loaches (*Misgurnus anguillicaudatus*). *Parasitol Int* 2001; 50: 267-271. [https://doi.org/10.1016/S1383-5769\(01\)00084-8](https://doi.org/10.1016/S1383-5769(01)00084-8)
- Pearson J. Family heterophyidae leiper, 1909. In Bray R, Gibson D, Jones A eds, In Keys to the Trematoda, Vol. 3. CAB International and Natural History Museum. London, UK. 2008, pp 113-141.
- Ransom BH. Synopsis of the trematode family Heterophyidae with descriptions of a new genus and five new species. *Pro U S Natl Mus* 1920; 57: 527-573. <https://doi.org/10.5479/si.00963801.57-2322.527>
- Rothschild M. Preliminary note on the life-history of *Cryptocotyle jejuna* Nicoll, 1907 (Trematoda). *Ann Mag Nat Hist* 1938; 1: 238-239. <https://doi.org/10.1080/00222933808526760>
- Nicoll W. Observations on the trematode parasites of British birds. *Ann Mag Nat Hist* 1907; 20: 245-271. <https://doi.org/10.1080/00222930709487332>
- Stunkard HW. Life history of *Cryptocotyle lingua* (Creplin) from the gull and tern. *J Morph Physiol* 1930; 50: 143-191. <https://doi.org/10.1002/jmor.1050500106>
- Kitamura Y. *Cryptocotyle lingua* (Creplin) from the red fox *Vulpes vulpes schrencki* Kishida. *Res bull Meguro Parasit Mus* 1973; 7: 15-16. [http://kiseichu.la.coocan.jp/publ/Res\\_Bull\\_MPM7\\_p15-16.pdf](http://kiseichu.la.coocan.jp/publ/Res_Bull_MPM7_p15-16.pdf)
- Gardner SL, Thew PT. Redescription of *Cryptocotyle thapari* McIntosh, 1953 (Trematoda: Heterophyidae), in the River Otter *Lutra longicaudis* from Bolivia. *Comp Parasitol* 2006; 73: 20-23. <https://doi.org/10.1654/0001.1>
- Zander CD, Kollra HG, Antholz B, Meyer W, Westphal D. Small-sized euryhaline fish as intermediate hosts of the digenetic trematode *Cryptocotyle concavum*. *Helgol Meeresunter* 1984; 37: 433-443. <https://doi.org/10.1007/BF01989322>
- Wootton DM. The life history of *Cryptocotyle concavum* (Creplin, 1825) Fiscoeder, 1903 (Trematoda: Heterophyidae). *J Parasitol* 1957; 43: 271-279. <https://doi.org/10.2307/3274345>
- Issaitschikoff IM. *Ciureana cryptocotyloides* nov. sp. (a new bird trematode and its systematic position within the family Heterophyidae), *Trudy Gosudarstv. Inst Eksper Vet* 1923; 1: 155-158 (in Russian).
- Skryabin KI. Studies on the parasitic worms of carnivores II-IV. *Trudy Gos Inst Exsp Vet* 1923; 1: 67-71.
- Kurochkin YV. Helminth fauna of the Caspian seal during its breeding season, Raboti Po Gelminthoogii K 80-etiyyu Akademiika K.I. Skryabina. Academy of Science of USSR. Moscow, Russia. 1958, pp 188-194 (in Russian).
- Jurachno MV. *Ciureana delamurei* sp. n. (Trematoda, Heterophyidae), a parasite of the northern fur seal. *Parazitologiya* 1987; 21: 752-754 (in Russian).
- Dollfus RP, Timon-David J. Sur une larve de Distome, parasite têtards de *Rana esculenta* L., devenant adulte chez le chat domestique *Felis catus* L. *domest.* et le pigeon domestique *Columba livia* Gmel. *Domest.* *CR Acad Sci Paris* 1960; 250: 1909-1911 (in French).
- Kamiya H, Obayashi M. Some helminths of the red fox, *Vulpes vulpes schlencki* Kishida, in Hokkaido, Japan, with a description of a new trematode, *Massaliatrema yamashitai* n. sp. *Jpn J Vet Res* 1975; 23: 60-68. <http://hdl.handle.net/2115/2066>
- Tatonova YV, Besprozvannykh VV. Description of a new species, *Cryptocotyle lata* sp. nov., and discussion of the phylogenetic relationships in Opisthorchioidea. *Parasitol Inter* 2019; 72: 1-11 <https://doi.org/10.1016/j.parint.2019.101939>
- Hong EJ, Sim C, Chae JS, Kim HC, Park J, Choi KS, Yu DH, Yoo JG, Park BK. A horsehair worm, *Gordius* sp. (Nematomorpha: Gordiida), passed in a canine feces. *Korean J Parasitol* 2015; 53: 719-724. <https://doi.org/10.3347/kjp.2015.53.6.719>
- Hoffman GL. Studies on the life cycle of *Cryptocotyle concavum* from the common sucker and experimentally in the chick. *Proc North Dakota Acad Sci* 1957; 11: 55-56.
- Lee YI, Seo M, Chai JY. Intestinal flukes recovered from a herring gull, *Larus argentatus*, in the Republic of Korea. *Korean J Parasitol* 2020; 58: 81-86. <https://doi.org/10.3347/kjp.2020.58.1.81>

