





8 | Bacteriology | Research Article

Longitudinal surveillance of the molecular evolution of methicillin-resistant *Staphylococcus aureus* isolates from pediatric patients in Shanghai, China, from 2013 to 2022

Xiaozhou Pan,^{1,2} Jiao Zhang,³ Fen Pan,^{1,2} Chun Wang,¹ Huihong Qin,¹ Fangyuan Yu,¹ Tiandong Zhang,¹ Wenxin Liu,^{1,2} Qianyue Wu,^{1,2} Zhan Ma,^{1,2} Wenhao Weng,^{1,2} Liang Chen,⁴ Fangyou Yu,³ Hong Zhang,^{1,2} Bingjie Wang³

AUTHOR AFFILIATIONS See affiliation list on p. 13.

ABSTRACT The objective of this research was to investigate the genomic epidemiology of methicillin-resistant Staphylococcus aureus (MRSA) in a pediatric population in Shanghai, China. Whole-genome sequencing was conducted for 492 randomly selected MRSA isolates obtained from a pediatric hospital between 2013 and 2022. ST59 (37.4%), ST398 (22.4%), ST88 (5.7%), and ST22 (5.5%) were the predominant lineages among these children. While ST59 maintained a dominant annual proportion before 2017, the proportion of ST398 gradually increased from 2013 to 2016, with ST398 ultimately emerging as a prevalent clone with a proportion comparable to that of ST59 after 2017. Among the prevalent STs, the spa-SCCmec structure also experienced dynamic changes. Within ST59, the t437-IV subtype experienced a decline and has even been replaced by t172-IV in recent years. In ST398, the t011-V subtype appeared in 2014 and rapidly became the leading subtype. The antibiotic resistance profiles and virulence factors exhibited clone-related features. Compared with other prevalent lineages, ST59 presented high resistance to erythromycin and clindamycin, whereas ST398 presented relatively low resistance to common antimicrobial agents and fewer virulence determinants. Panton-Valentine leucocidin was more common in ST338 and ST1232, whereas toxic shock syndrome toxin was closely associated with ST1 and ST5. The MRSA cases could also be classified into community- and hospital-associated cases, with highly significant differences between the two in terms of demographic characteristics, clindamycin susceptibility, and virulence genes. In conclusion, this study revealed high genetic diversity and dynamic changes in the molecular epidemiology of pediatric MRSA isolates from Shanghai collected over a decade.

IMPORTANCE Methicillin-resistant *Staphylococcus aureus* (MRSA) has emerged as a significant global health concern. Previous research on MRSA epidemiology has predominantly focused on adult populations or targeted specific infection sites, while there was limited research on the long-term evolution of MRSA from the pediatric population. This study addresses this knowledge gap by conducting a comprehensive, 10-year surveillance of pediatric MRSA isolates using whole-genome sequencing. We characterized the molecular typing, as well as the phenotypic and genotypic antimicrobial resistance profiles, and virulence factors present in MRSA isolates obtained from children. Our results highlight the imperative for continuous, vigilant monitoring of MRSA within the pediatric demographic to track its evolving genetic landscape.

KEYWORDS methicillin-resistant *Staphylococcus aureus*, genome sequencing, molecular epidemiology, children, antibiotics

Editor Zoe A. Dyson, London School of Hygiene & Tropical Medicine, London, United Kingdom

Address correspondence to Fangyou Yu, wzjxyfy@163.com, Hong Zhang, schjyk2015@126.com, or Bingjie Wang, wangbingjie2017@163.com.

Xiaozhou Pan, Jiao Zhang, and Fen Pan contributed equally to this article. The author order was determined by mutual agreement.

The authors declare no conflict of interest.

See the funding table on p. 13.

Received 13 March 2025 Accepted 1 April 2025 Published 30 April 2025

Copyright © 2025 Pan et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International license.

S taphylococcus aureus is recognized as one of the primary opportunistic bacterial pathogens affecting humans and is frequently implicated in conditions such as endocarditis, bacteremia, osteomyelitis, and skin and soft tissue infections (1). Multidrug resistance has emerged as a major health concern worldwide (2), with methicil-lin-resistant S. S aureus (MRSA) posing a growing challenge since its initial identification in 1961 (3). In addition to being resistant to oxacillin (OXA) and S-lactam antibiotics, recent studies have indicated that MRSA can also exhibit resistance to vancomycin (VAN), linezolid (LNZ), and daptomycin (DAP), which are considered last-line treatment options (4). Moreover, MRSA is associated with considerable morbidity and mortality, largely attributable to its abundant repertoire of virulence determinants, such as biofilm formation, immune evasion, hemolysis, and enterotoxins (5).

MRSA was once confined largely to the healthcare environment. However, since the 1990s, there has been an explosion in the incidence of MRSA infections in healthy community dwellers without prior healthcare contact, which has been associated with the recognition of novel MRSA clones known as community-associated MRSA (CA-MRSA) (6). CA-MRSA strains were often susceptible to most non-β-lactam antimicrobial agents and displayed enhanced virulence (7), associated with distinct lineages of MRSA, such as ST59, ST8 (USA300), and ST22 (8, 9). Over time, these clones began invading healthcare settings, and the distinctions between CA-MRSA and hospital-associated MRSA (HA-MRSA) have become increasingly blurred. For example, there were increasing reports that USA300 had been a common nosocomial pathogen in America (10). A comprehensive understanding of HA-MRSA and CA-MRSA isolates is essential for the development of effective treatment and prevention of MRSA infections across diverse healthcare and community settings.

MRSA epidemics are characterized by the predominance of varied clonal types in different geographic regions (2). In Asia, sequence type (ST) 239, ST5, and ST22 are the most frequently reported, while ST8 and ST45 are prevalent in North America (11). However, the number of prevalent clones in a given area may change over time. Several studies have suggested that the CA-MRSA clone ST59 is gradually replacing the HA-MRSA clone ST239 in China (12, 13). More importantly, the potential for interspecies transmission underscores the significant capacity of MRSA to adapt to different hosts and environments (14). In particular, livestock-associated MRSA (LA-MRSA) has attracted increasing attention because it has been frequently reported in humans. Since 2005, ST398 has been well described as a predominant lineage of LA-MRSA in Europe (15), with subsequent reports of its presence in the human populations of China and the United States (16, 17).

To date, epidemiological research in China has focused mainly on the adult population and has targeted specific infection sites or patient wards (9, 18). However, there remains a lack of comprehensive understanding of the long-term evolution of MRSA derived from pediatric populations. In this study, we randomly collected a total of 492 MRSA isolates from a tertiary pediatric hospital in Shanghai between 2013 and 2022. We aimed to characterize the genetic background and identify predominant lineages of MRSA in pediatric patients.

MATERIALS AND METHODS

Clinical isolates and information collection

First, we collected data on methicillin-susceptible *S. aureus* and MRSA strains isolated from 2013 to 2022 in a pediatric hospital in Shanghai, China, to calculate the percentage of MRSA strains among *S. aureus* for each year. An MRSA isolate was defined as those carrying the *mecA* gene or exhibiting resistance to cefoxitin (FOX; MIC ≥8 μg/mL). Using the RANDBETWEEN randomization module in Microsoft Excel, we initially selected 50 isolates annually from 2013 to 2022. Eight isolates were excluded due to incomplete clinical information or the low quality of whole-genome sequencing outputs. Finally, a total of 492 non-duplicate MRSA clinical isolates were included in this study. The relevant

clinical information, including sex, age, source, and clinical diagnosis, was extracted from the anonymized laboratory information system. The included patients were divided into six age groups: newborn (0–28 days), infant (1–12 months), toddler (13 months–3 years), preschool (4–7 years), school-aged (8–12 years), and adolescent (13–16 years). CA-MRSA and HA-MRSA were both defined based on the clinical criteria. CA-MRSA strains were isolated from outpatients or inpatients within 48 hours of hospital admission, without a history of hospitalization, long-term care facility, surgery, permanent indwelling catheter use, percutaneous medical device use, or positivity for MRSA culture in the previous 12 months. HA-MRSA strains were defined as those that did not meet the above CA-MRSA standards (19).

Antimicrobial susceptibility testing

We conducted antimicrobial susceptibility testing via the broth microdilution method. The antibiotics tested included FOX, OXA, erythromycin (ERY), clindamycin (CLI), VAN, sulfamethoxazole/trimethoprim (SXT), LNZ, rifampicin (RIF), gentamicin (GEN), ciprofloxacin (CIP), tetracycline (TCY), fusidic acid (FA), mupirocin (MOP), teicoplanin (TCL), DAP, and dalbavancin (DAL). The interpretative criteria followed the CLSI guidelines (20), except for FA, for which EUCAST breakpoints were applied (21).

Whole-genome sequencing and bioinformatic analyses

The total DNA of the MRSA isolates was extracted via a TIANamp Bacteria DNA Kit (TIANGEN) with additional lysostaphin. The isolated genomic DNA was sequenced on the Illumina NovaSeq platform in 2 \times 150 bp paired-end mode. The raw data were filtered with fastp v0.20.1 (22) and then *de novo* assembled into contigs using CLC Genomics Workbench software (version 12.0; CLCbio). The N50 contig number was 55.28 \pm 48.15, with 137.69 \pm 44.89 kbp. Multilocus sequence typing (MLST) was performed via the PubMLST database (https://pubmlst.org/organisms/staphylococcusaureus). The *spa* and SCC*mec* types were predicted using SpaFinder and SCCmecFinder, respectively (23, 24).

Moreover, the virulence factor genes and antimicrobial resistance genes were identified by ABRicate v1.01 (https://github.com/tseemann/abricate) using the VFDB and CARD databases (25, 26). The candidate virulence genes were associated with adhesion, enterotoxins, immune evasion, secretion, toxins, hemolysis, serine protease, and iron uptake. The genes were annotated via Prokka v1.14.5 (27). For phylogenetic analyses, a core-genome alignment based on the concatenation of 1,702 core genes was obtained via Panaroo v1.5.0 (28) with default parameters. A maximum-likelihood phylogenetic tree was constructed via FastTree v2.1.11 (model GTR + GAMMA) with 1,000 bootstrap replicates (29). The tree was visualized using iTOL v6 (30).

Statistical analyses

Statistical analyses were performed using GraphPad Prism 9.0. The data were analyzed using Pearson's χ^2 test, Fisher's exact test, and Student's t test, as appropriate. A P value of <0.05 was considered to indicate statistical significance.

RESULTS

Molecular typing characteristics

In the present study, a total of 492 non-duplicated MRSA clinical isolates were collected over 10 years. Thirty-one distinct STs were identified, with ST59 (37.4%, 184/492), ST398 (22.4%, 110/492), ST88 (5.7%, 28/492), and ST22 (5.5%, 27/492) being the most prevalent (Table S1). Additionally, three isolates belonging to novel STs were identified, and their genome sequences have been submitted to PubMLST (https://pubmlst.org/organisms/staphylococcus-aureus/, id: 47326–47328), where they have been assigned to new STs (ST9203, ST9204, and ST9205). The identified STs can be classified into 11 clonal complexes (CCs), with CC59 (42.9%, 211/492), CC398 (23.6%, 116/492), and CC5 (7.7%,

38/492) being the most common CCs, collectively accounting for more than 70% of the isolates.

All the MRSA isolates were assigned to 77 types by *spa* typing, with 15 new types identified. ST88 dominated among the novel *spa* types, accounting for 40% (6/15) of cases. The *spa* types of the MRSA isolates exhibited a strong correlation with their CCs. The predominant *spa* type was t437 (24.2%, 119/492), which was the major type within CC59 (56.4%, 119/211). The second most common type was t011 (12.6%, 62/492), classified under CC398. Within CC59, t172 was also frequently observed (20.4%, 43/211). Notably, the ST59 lineage presented a greater diversity of *spa* types than the other CC59 clones did (Table S2).

Four SCC*mec* types were identified, namely, II, III, IV, and V. SCC*mec* IV (57.5%, 283/492) was the most prevalent, followed by the SCC*mec* V cassette (36.4%, 179/492). Twenty-four isolates could not be classified into any SCC*mec* type, with nearly one-third of them belonging to ST88 (28.6%, 8/28). CC59 and CC398 were closely correlated with SCC*mec* IV (85.3%, 180/211) and SCC*mec* V (96.6%, 112/116), respectively. Notably, ST338, a variant clone of ST59, was classified under SCC*mec* V. Additionally, SCC*mec* IV was subdivided into IVa, IVc, IVg, and IVi. CC59 exhibited a strong association with SCC*mec* IVa (84.4%, 178/211), whereas CC88 was more likely to be associated with SCC*mec* IVc (67.9%, 19/28; Table S2).

In general, the population structure of the MRSA isolates in our study was dominated by ST59-t437-IV (17.3%, 85/492), ST398-t011-V (11.4%, 56/492), and ST59-t172-IV (7.1%, 35/492).

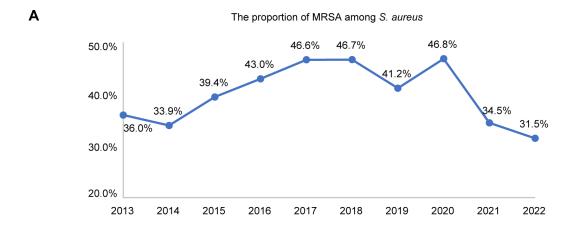
Demographic characteristics of the MRSA isolates

Among the 492 MRSA isolates, a majority were collected from children under 1 year of age (61.0%, 300/492). The sample sources of isolates varied widely and included sputum, pharyngeal swabs, pus, secretions, blood, cerebrospinal fluid, and central venous catheters. Sputum was the predominant sample source (60.6%, 298/492), especially among neonates and children under 3 years of age. The most frequently identified clones from the sputum samples were ST59 and ST398. For the second most common source, pus (22.0%, 108/492), isolates were more frequently obtained from school-aged individuals and adolescents, with ST59 and ST22 being the most common clones in this group. Moreover, in our study, several isolates were derived from vaginal secretions, predominantly within the preschool subgroup (65.0%, 13/20; Table S2). The percentage of MRSA strains among *S. aureus* was slightly higher in male patients, constituting 58.5% (288/492) of the total. The annual proportions of MRSA among *S. aureus* tended to increase from 2014 to 2018, corresponding with an increase in the prevalence of ST398. Nevertheless, a significant decline in this rate was observed in 2021 (34.5% vs 46.8%, χ^2 test, P < 0.001; Fig. 1A).

On the basis of the clinical data, the MRSA isolates were categorized into CA-MRSA (23.4%, 115/492) and HA-MRSA (76.6%, 377/492). Statistical analysis revealed no significant differences in the annual proportions of CA-MRSA and HA-MRSA (Table 1, χ test, P=0.33). HA-MRSA was more commonly detected in children under 1 year of age, whereas CA-MRSA was more prevalent among preadolescent children in this study. HA-MRSA was predominantly isolated from the neonatology ward, whereas CA-MRSA was significantly more common in the pulmonology ward (Table 2).

Dynamic evolution of MRSA from 2013 to 2022

The prevalence of different molecular types over the 10 years showed dynamic fluctuations (Fig. 1B). From 2013 to 2016, ST59 maintained a dominant position in terms of its annual proportion. However, its proportion started to decline in 2017 (from a maximum of 55.1% to a minimum of 24.0%). In contrast, ST398 exhibited a consistent increase from 2013 to 2017, with the proportion rising from 6.0% to 34.0%. Subsequently, the prevalence of ST398 either matched or exceeded that of ST59, except in the year 2020. ST88 exhibited a relatively stable distribution throughout the study



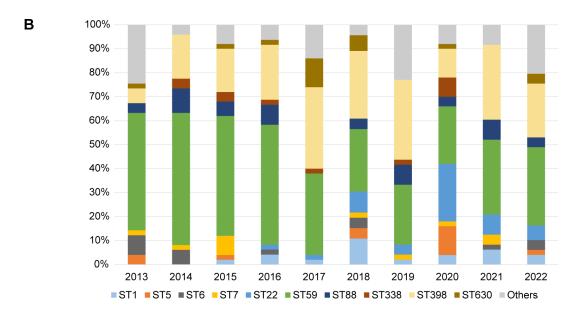


FIG 1 (A) The proportion of MRSA among S. aureus in each year. (B) The annual proportion of each ST. STs with <10 isolates were included in the "others" group.

period. Notably, ST22 emerged in 2016, and its proportion sharply increased in 2020, reverting to the previous levels in 2021 (Fig. 2A). The spa types were conserved within their respective CCs, yet they also underwent notable changes over the decade (Fig. 2B). The proportion of the predominant type t437 slightly increased in the initial years, followed by a gradual decline until 2022. Conversely, the prevalence of t011 showed a persistent increase, starting from a low value and ultimately reaching parity with t437, which aligns with the trends observed for their associated lineages ST59 and ST398, respectively. The prevalence of another spa type of CC59, t172, initially displayed a declining trend in the early years but has increased again in recent years (Table S3; Fig. 2B). Further analysis of the two predominant clones, ST59 and ST398, revealed temporal dynamics in their population structures over the 10-year period. For ST59, the t437-IV subtype was the most dominant subtype from 2013 to 2019, but in recent years, its proportion within ST59 has decreased, with t172-IV emerging as the dominant subtype in 2020 and 2022 (Fig. 2C). For ST398, the leading type, t011-V, was first identified in 2014, and it rapidly became dominant beginning in 2015. The isolation frequency of t1250-V fluctuated from 2014 to 2022 (Fig. 2D).

We constructed phylogenetic trees for all 492 MRSA isolates to determine their evolutionary relationships. The analysis revealed the genetic diversity of each clone over

TABLE 1 The yearly demographic characteristics of the samples involved in this research between 2013 and 2022 (%)

	2013	2014	2015	2016	2017	2018	2019	2020	2021	2022
	(n = 50)	(n = 49)	(n = 50)	(n = 49)	(n = 50)	(n = 47)	(n = 48)	(n = 50)	(n = 49)	(n = 50)
Specimen source							'			'
Sputum	46.0	69.4	90.0	73.5	58.0	55.3	66.7	40.0	57.1	50.0
Pus	22.0	16.3	8.0	16.3	24.0	19.2	14.6	48.0	26.5	24.0
Swab	4.0	4.1	0.0	4.1	4.0	10.6	14.6	4.0	6.1	6.0
Secretion	20.0	4.1	0.0	2.0	4.0	2.1	2.1	0.0	4.1	2.0
Blood/cerebrospinal fluid	2.0	4.1	0.0	0.0	0.0	4.3	0.0	2.0	4.1	12.0
Others	6.0	2.0	2.0	4.1	10.0	8.5	2.1	6.0	2.0	6.0
Epidemiological classification										
CA-MRSA	22.0	16.3	14.0	24.5	20.0	19.1	41.7	26.0	24.5	26.0
HA-MRSA	78.0	83.7	86.0	75.5	80.0	80.9	58.3	74.0	75.5	74.0

the 10-year period and their evolutionary relationships. The CC22 strains (ST22) isolated in 2020 clustered together, which was consistent with their rapid dissemination in 2020. Notably, several ST398 strains isolated in 2017 were also closely related, indicating their evolution from a common ancestor (Fig. 3).

It is important to highlight the atypical epidemic trends observed in 2020. First, the annual percentage of MRSA strains among *S. aureus* peaked in 2020 and then plunged immediately in 2021 (Fig. 1). Furthermore, the decline in ST398 abundance interrupted a previously established upward trend, although this ST experienced a resurgence in 2021. In contrast, the abundance of ST22 increased clearly in 2020 and then reverted (Fig.

TABLE 2 Demographic characteristics of CA-MRSA and HA-MRSA cases, n (%)

	CA-MRSA (n = 115)	HA-MRSA (n = 377)	<i>P</i> -value
Sex			
Male	66 (57.4)	222 (58.9)	>0.05
Female	49 (42.6)	155 (41.1)	>0.05
Age			
0–28 days	1 (0.9)	105 (27.9)	< 0.001
1–12 months	36 (31.3)	159 (42.2)	< 0.05
13 months to 3 years	31 (27.0)	59 (15.9)	< 0.05
4–7 years	27 (23.5)	22 (5.8)	< 0.001
8–12 years	18 (15.7)	25 (6.6)	< 0.05
13–16 years	2 (1.7)	6 (1.6)	>0.05
Specimen source			
Sputum	37 (32.2)	261 (69.2)	< 0.001
Pus	44 (38.3)	64 (17.0)	< 0.001
Swab	11 (9.6)	17 (4.5)	< 0.05
Secretion	12 (10.4)	8 (2.1)	< 0.05
Blood/CSF ^a	5 (4.3)	9 (2.4)	>0.05
Others	6 (5.2)	18 (4.8)	>0.05
Outpatient department			
Otorhinolaryngology	10 (8.7)	3 (0.8)	< 0.05
Dermatology	7 (6.1)	0	< 0.05
Others	11 (9.6)	8 (2.1)	< 0.05
Inpatient department			
Neonatology	5 (4.3)	156 (41.4)	< 0.001
$PICU^a$	5 (4.3)	29 (7.7)	>0.05
Pulmonology	25 (21.7)	27 (7.2)	< 0.05
General surgery	19 (16.5)	14 (3.7)	< 0.05
Orthopedic	13 (11.3)	1 (0.3)	< 0.05
Others	20 (17.4)	139 (36.9)	< 0.05

 ${}^a\!$ CSF, cerebrospinal fluid; PICU, pediatric intensive care unit.

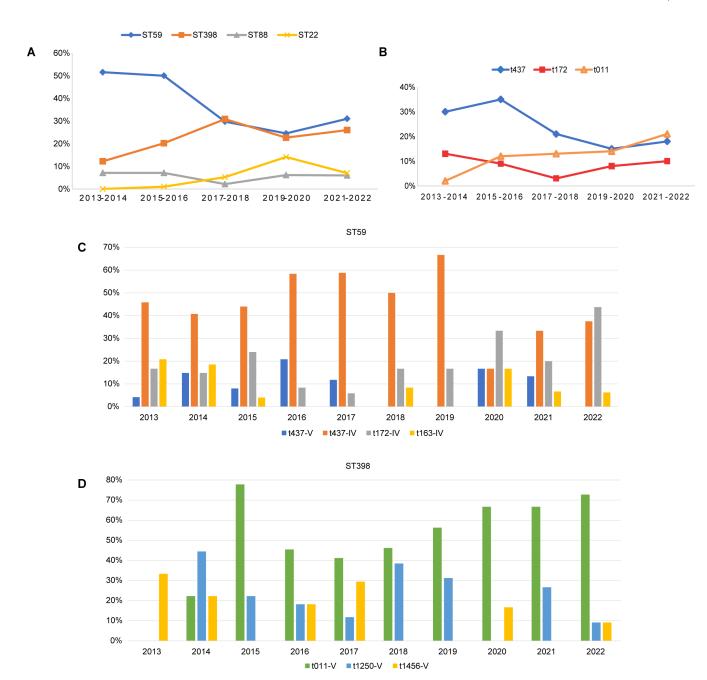


FIG 2 The changing molecular characteristics by year. (A) The 2-year temporal changes of four predominant STs. (B) The 2-year temporal changes of three main spa types. (C) The proportion of major molecular structures of ST398 during the 10 years. (D) The proportion of major molecular structures of ST398 during the 10 years.

1B). Interestingly, sputum samples consistently represented a majority of the samples collected each year, but the number of isolates derived from pus exceeded that derived from sputum in 2020. We subsequently collected annual data on the total number of pus and upper respiratory tract samples (except for 2013). The analysis revealed an increase in the ratio of pus/upper respiratory tract samples in 2020, mainly due to a decrease in upper respiratory tract samples, while the number of pus samples stayed stable (9.1% vs $5.7\% \pm 0.71\%$, Student's t test, P < 0.05). The reduction in upper respiratory tract samples may be attributed to non-pharmaceutical interventions during the COVID-19 outbreak (31, 32). This may account for the shift in the percentage of MRSA isolates originating from pus (Table S4).

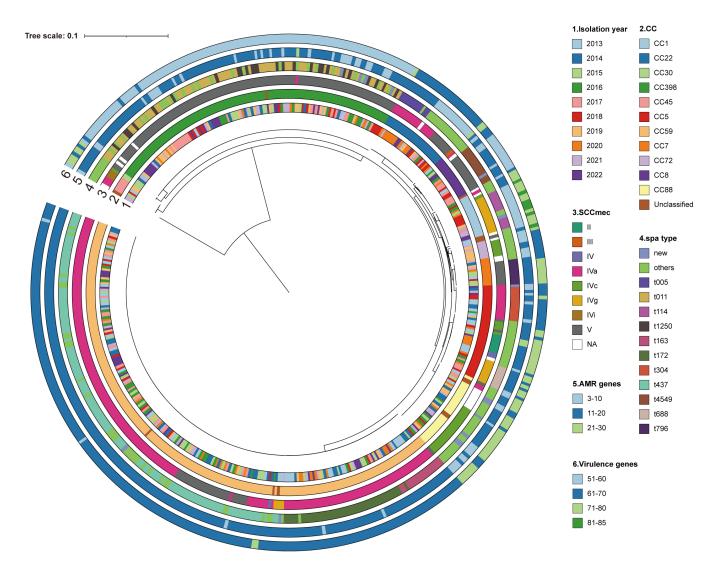


FIG 3 Phylogenetic tree of all the 492 MRSA. Isolation year, CC types, SCC*mec* types, *spa* types, the carrying number of antimicrobial resistance genes (AMR), and virulence genes are color coded in the outside rings.

Phenotypic and genotypic antimicrobial susceptibility

The phenotypic antimicrobial susceptibility profile of the MRSA isolates was determined using the broth microdilution method (Fig. 4; Fig. S1). All of the MRSA isolates were resistant to cefoxitin, and as expected, a majority of the strains harbored multiple β-lactam resistance genes, including blal, blaR1, blaZ, mecA, and mecR1 (Table S5). A total of 97.8% (480/492) of the MRSA strains carried mecA. Notably, three mecA-positive isolates exhibited susceptibility to oxacillin. A high rate of erythromycin resistance (71.8%) was observed, particularly within CC5 (86.8%) and CC59 (83.4%), which corresponds with the relatively high carriage rates of erm(C) in CC5 and erm(B) in CC59. Clindamycin resistance was more common in CC59 (80.1% vs 11.1%–25.0%, χ^2 test, P < 0.001). However, the direct lincosamide resistance genes Inu(A), Inu(G), and Isa(A) were infrequently identified across all the isolates, while the macrolide-associated gene ermB affecting susceptibility to clindamycin had a relatively high prevalence within the CC59 lineage (65.4%, 138/211; Table S5). In contrast, CC398 presented a relatively low level of resistance to erythromycin and clindamycin (Fig. 4B). Therefore, the rates of resistance to both antibiotics have declined since 2017 (Fig. 4C). Although the overall resistance to ciprofloxacin in this study was low (10.3%), it was significantly greater in

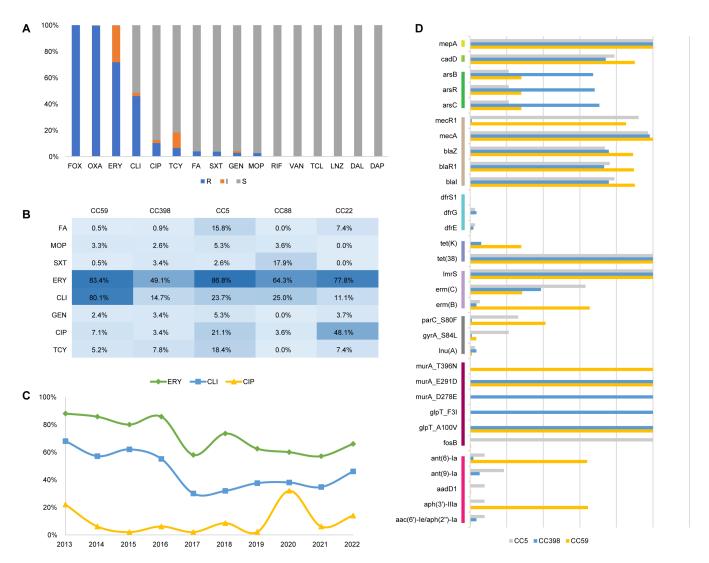


FIG 4 Phenotypic and genetic antimicrobial susceptible characteristics. (A) The overall rates of antibiotic resistance (R), intermediate susceptibility (I), and susceptibility (S). (B) The resistance rate of eight antibiotics in dominant lineages. (C) Annual trends in resistance rates for ERY, CLI, and CIP. (D) The antimicrobial resistance genes carried by CC59, CC398, and CC5.

CC22 than in other prevalent lineages (48.1% vs 3.6%–21.1%, χ^2 test, P < 0.001), which was attributed to more frequent mutations in $gyrA_S84L$ and $parC_S80F$ in CC22. This trend was further reflected by an increased rate of resistance to ciprofloxacin in 2020 (Fig. 4C). The MRSA isolates in this study overall demonstrated low resistance rates to fusidic acid (3.9%), mupirocin (2.6%), sulfamethoxazole/trimethoprim (3.7%), and gentamicin (2.8%). However, ST630 may tend to be resistant to fusidic acid, as 86.7% of the strains carried fusB. Importantly, none of the isolates were resistant to vancomycin, linezolid, rifampicin, teicoplanin, daptomycin, or dalbavancin. The tetracycline-related gene tet (38) and the efflux-related gene tet (38) and the efflux-related gene tet (39) and the efflux-related gene tet (30) as statistically significant difference in resistance rate to clindamycin was observed between CA-MRSA and HA-MRSA (Table 3).

Virulence factors vary among clones

Diverse virulence genes enable MRSA to invade and adapt to humans. A total of 134 virulence factors were examined (Table S6). Among them, 49 virulence genes associated with various virulence characteristics presented carriage rates exceeding 90%, indicating a high level of conservation in MRSA. Compared with other prevalent clones, CC398

TABLE 3 Comparative antibiotic resistance of CA-MRSA and HA-MRSA isolates, n (%)

	CA-MRSA (n = 115)	HA-MRSA (n = 377)	<i>P</i> -value
			r-value
FA	6 (5.2)	13 (3.4)	>0.05
MOP	4 (3.5)	9 (2.4)	>0.05
SXT	4 (3.5)	14 (3.7)	>0.05
ERY	88 (76.5)	265 (70.3)	>0.05
CLI	62 (53.9)	165 (43.8)	< 0.05
GEN	3 (2.6)	11 (2.9)	>0.05
CIP	9 (7.8)	41 (10.9)	>0.05
TCY	9 (7.8)	23 (6.1)	>0.05

carried fewer virulence genes, whereas CC1 harbored more virulence factors (Fig. 3; Table S6). In our cohort, 16.1% of the MRSA isolates carried *lukF-PV* and *lukS-PV*, which encode Panton-Valentine leucocidin (PVL), a well-studied toxin in *S. aureus* and a molecular marker in CA-MRSA (33). Indeed, its carriage rate was significantly higher in CA-MRSA in the present study, at 33.9% (Table 4). Notably, all the ST338 and ST1232 isolates were positive for PVL, and all the CC5 strains were PVL negative. The carriage rate of PVL in ST59-t437-V was significantly higher than that in other subtypes of ST59 (85.7%, 12/14; χ^2 test, P < 0.001).

Overall, most of the detected virulence genes presented clone-related features. The toxic shock syndrome toxin (TSST) encoded by *tsst-1* was frequently detected in ST1 (76.5%, 13/17) and ST5 (66.7%, 8/12). Moreover, more than 80% of ST22 isolates harbored either *pvl* or *tsst-1*, with all TSST-positive ST22 isolates belonging to SCC*mec* IV. The adhesion-related virulence gene *cna* was absent in most clones, except ST22. In contrast, the *ebp* gene was absent in ST22, whereas it was detected in nearly all the other clones. The frequency of biofilm-associated genes was generally high in all the MRSA strains, except for ST398 isolates, which largely lacked *fnbA*, *fnbB*, *sdrD*, and *sdrE* (Fig. 5). The secretion system-associated genes *esaC*, *essC*, and *esxB* were all absent in CC59, CC398, and CC22. Additionally, genes of the immune evasion cluster (IEC), including *scn*, *chp*, *sea*, *sep*, and *sak*, exhibited an ST-related distribution. The gene *sak* was prevalent in CC59, except in ST338. A majority of the MRSA isolates exhibited an *scn-chp-sak* pattern, except ST630, which did not carry the IEC. Enterotoxins also exhibited variability among strains. A majority of CC59 isolates harbored *seb-sek-selk-selq-seq-sey*; however, most isolates of subclone CC59-t437 lacked these enterotoxins. Similarly, the *sea* was absent in CC59-t437

TABLE 4 Virulence genes carried by CA-MRSA and HA-MRSA isolates, n (%)

	CA-MRSA (n = 115)	HA-MRSA (n = 377)	<i>P</i> -value
pvl	39 (33.9)	40 (10.6)	<0.001
tsst-1	8 (7.0)	32 (8.5)	>0.05
chp	105 (91.3)	320 (84.9)	< 0.05
sak	92 (80.0)	344 (91.2)	>0.05
scn	109 (94.8)	348 (92.3)	>0.05
sea	20 (17.4)	100 (26.5)	< 0.05
sep	6 (5.2)	40 (10.6)	< 0.05
coa	107 (93.0)	304 (80.6)	< 0.05
vWbp	10 (8.7)	46 (12.2)	>0.05
clfA	48 (41.7)	142 (37.7)	>0.05
clfB	30 (26.1)	135 (35.8)	< 0.05
cna	2 (1.7)	10 (2.7)	>0.05
fnbA	75 (65.2)	277 (73.5)	>0.05
fnbB	68 (59.1)	231 (61.3)	>0.05
sdrD	55 (47.8)	201 (53.3)	>0.05
sdrE	82 (71.3)	262 (69.5)	>0.05

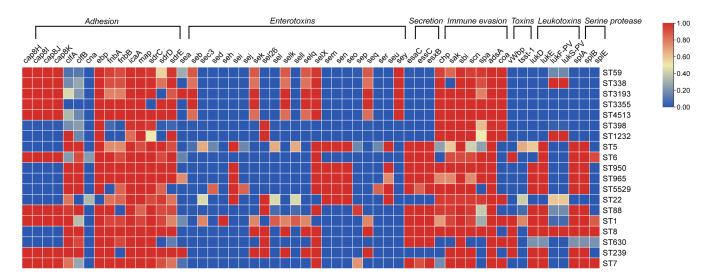


FIG 5 The distribution of virulence determinants. Virulence genes with >99% or 0% carriage rate were not displayed. More details can be found in Table S6.

but was universally present in CC59-t163 and CC59-t172, suggesting a difference in virulence associated with *spa* type (Fig. 5).

DISCUSSION

In the present study, 492 MRSA isolates were randomly collected from a single pediatric cohort over the past decade to investigate the epidemiology and evolution of MRSA in children. The included samples covered a wide range of sources and were collected from individuals aged 0–16 years.

The phylogenetic analysis revealed high genetic diversity of the MRSA isolates across 10 years. During the research period, CC59 emerged as the predominant lineage, with the Asian-Pacific clone ST59-t437-IV being the most prevalent sub-lineage. This finding aligns with previous studies conducted on children in Beijing and Sichuan, as well as on adults in other regions of China (34–37). Notably, the temporal patterns of prevalence varied between children and adults. In an all-age and multicenter study on Chinese MRSA, the prevalence of ST59 increased from 27.8% to 35.6% between 2014 and 2020 (9). In contrast, we found a decline in ST59 prevalence from 55% to 24% over the same 6-year period.

Accordingly, the prevalence of ST398 steadily increased from 2013 to 2017, after which both ST398 and ST59 became the predominant lineages. This dissemination capacity and the prevalence trends suggest that ST398 may have strong host or environmental adaptations (1). In recent years, an obvious increase in the prevalence of ST398 has been observed in adults (35, 38). However, as early as 2013, the prevalence of ST398 in children increased gradually. Notably, unlike ST398-t034, which is disseminated among adults (34), our study revealed that the prevalent subtype in the pediatric population was ST398-t011. Another study on epidemic clones in China revealed that ST59 and ST398 had greater virulence potential than their counterparts ST5 and ST239 (39). However, our findings revealed that ST398 presented relatively low carriage of adhesion virulence genes, minimal levels of enterotoxins, and almost no *pvl* or *tsst-1*, indicating a complex correlation between the virulence phenotype and gene expression and regulation. ST59 and ST398 are more strongly associated with SCC*mec* IV and V, which are smaller cassettes that can reduce the potential fitness burden (40). This may have contributed to their pandemic characteristics.

ST22 was the second most common type among the pus-derived isolates in the present study. Similarly, another study from Shanghai reported that MRSA isolates from bone and joint infections in children were also characterized, with ST22 being the second major genotype (41). MRSA ST22 reportedly exhibits high virulence and has strong

potential to replace other previously epidemic MRSA clones, which can enable ST22 clones to cause more severe infections.

ST88 has been identified as a predominant circulating clone within both hospital and community settings in African countries (42) and has been sporadically reported in the food industry (43). One-third of the non-typeable SCC*mec* elements belonged to ST88. Evidence suggested that Chinese ST88 isolates were identified with lineage-specific pseudo-SCC*mec* genes and lacked *ccr* genes (44), which may explain the challenges encountered in SCC*mec* typing. Intriguingly, ST88 was the most prevalent among the newly identified *spa* types, with the principal types (t1764 and t3622) showing differences from previously reported types (t690) in adults (45).

ST5 MRSA strains have been described as representative emerging epidemic clones in East China, including Shanghai (35, 38). Nonetheless, the findings of our study revealed a different epidemiological pattern, as ST5 accounted for only 2.4%, indicating a divergence in the prevalence of ST5 between children and adults. ST239 was previously demonstrated to be the predominant HA-MRSA type in adults in China (12). Interestingly, despite the observed replacement of ST239 by ST5 and ST59 in adults since 2013 (46, 47), only one ST239 MRSA isolate was detected in our study in 2013, indicating an earlier or more pronounced decrease in the incidence of ST239 in children. In addition to the epidemic clones, some single STs were detected. ST4513, belonging to CC59, was previously reported in an endocarditis patient (48). However, ST5529 (CC5), ST6309, and ST4083 have not been previously reported.

The unusual epidemic trend in 2020, which varied from that in other years, can be linked to the emergence of the COVID-19 pandemic during that year. Another pediatric study in Henan, China, revealed that the *S. aureus* positivity in the respiratory system significantly decreased during the COVID-19 pandemic, whereas there were no obvious changes among isolates from non-respiratory system samples (49). This finding parallels the reduction in the number of upper respiratory tract samples noted in our study, which influenced the proportion of MRSA isolated from pus samples.

With respect to virulence determinants, the prevalence of PVL was 16.1%, similar to the reported rate of 15.1% among children in Southwest China (37). These data indicate that the rate of PVL in MRSA isolates from children is still low. Furthermore, PVL was more common in ST59-V than in ST59-IV, which is consistent with the previously reported ST59/SCCmec V/PVL-positive Taiwan clone (50).

In antimicrobial susceptibility testing, we identified three oxacillin-susceptible MRSA (OS-MRSA) isolates that carried the *mecA* gene. Previous studies have indicated that the prevalence of OS-MRSA in Shanghai is 1.8%, with most isolates derived from pediatric inpatients (51). Nucleotide substitutions within the *mecA* promoter (52), frameshift mutations in *mecA* (53), a loss of penicillin-binding protein 4 (54), and a deletion in the *blaR1* gene (55) are all possible mechanisms underlying the low-level oxacillin resistance of the OS-MRSA strains identified. There is evidence that CA-MRSA clone ST59 is more susceptible to antibiotics than the prevalent HA-MRSA clones ST239 or ST5 are (35). Notably, in our study, ST398 exhibited greater antibiotic susceptibility than ST59. Moreover, although it has been reported that CA-MRSA is more likely than HA-MRSA to be susceptible to CIP, CLI, GEN, and SXT (56), our data revealed no significant difference in resistance between CA-MRSA and HA-MRSA.

In conclusion, the molecular epidemiology of pediatric MRSA isolates in Shanghai experienced great dynamic changes during the 10-year period. Compared with the findings of previous studies on adults, there were many differences in the MRSA isolates between adults and children. Our study provides updated insights into pediatric MRSA, including clinical demographic features, molecular types, antibiotic profiles, resistance and virulence genes, and temporal evolution, highlighting the need for long-term surveillance of pediatric MRSA.

ACKNOWLEDGMENTS

This work was supported by the 3-Year Initiative Plan for Strengthening Public Health System Construction in Shanghai (2023–2025; grant number GWVI-3), the Development of High-Sensitivity Multi-pathogen Specific Antibody Detection Kits grant (grant number 21531901900), the Natural Science Fund of China (grant number 82272394) and Shanghai Pulmonary Hospital Young Talents Funding Initiative (grant number Fkcy2306).

AUTHOR AFFILIATIONS

¹Department of Clinical Laboratory, Shanghai Children's Hospital, School of Medicine, Shanghai Jiao Tong University, Shanghai, China

²Institute of Pediatric Infection, Immunity, and Critical Care Medicine, School of Medicine, Shanghai, China

³Department of Clinical Laboratory, Shanghai Pulmonary Hospital, School of Medicine, Tongji University, Shanghai, China

⁴School of Pharmacy and Pharmaceutical Sciences, University at Buffalo, Buffalo, New York, USA

AUTHOR ORCIDs

Xiaozhou Pan (1) http://orcid.org/0000-0003-1034-6008

Liang Chen http://orcid.org/0000-0001-5845-2235

Fangyou Yu http://orcid.org/0000-0003-4924-2484

Hong Zhang http://orcid.org/0000-0002-0886-9531

Bingjie Wang http://orcid.org/0009-0003-8361-5843

FUNDING

Funder	Grant(s)	Author(s)
The Three-year initiative plan for strengthening public health system construction in Shanghai	GWVI-3	Hong Zhang
Development of High sentivity multi-pathogen specific antibody detection kit	21S31901900	Hong Zhang
Shanghai Pulmonary Hospital Young Talents Funding Initiative	grant number Fkcy2306	Bingjie Wang

AUTHOR CONTRIBUTIONS

Fangyou Yu, Supervision | Bingjie Wang, Formal analysis, Investigation, Methodology, Project administration, Supervision, Writing – review and editing.

DATA AVAILABILITY

The raw sequence data for 492 MRSA isolates have been deposited in the Sequence Read Archive under BioProject accession no. PRJNA1211827.

ETHICS APPROVAL

This study was approved by the Shanghai Children's Hospital Ethics Committee (No. 2021R126-E01) and was exempt from the informed consent requirements, as the bacterial strains in this study were collected via routine clinical diagnostics and their collection did not affect the patients' health.

ADDITIONAL FILES

The following material is available online.

Supplemental Material

Fig. S1 (mSystems00371-s0001.docx). The MIC distribution of 16 tested antibiotics in this study.

Supplemental Tables (mSystems00371-s0002.xlsx). Tables S1 to S6.

REFERENCES

- Howden BP, Giulieri SG, Wong Fok Lung T, Baines SL, Sharkey LK, Lee JYH, Hachani A, Monk IR, Stinear TP. 2023. Staphylococcus aureus host interactions and adaptation. Nat Rev Microbiol 21:380–395. https://doi.org/10.1038/s41579-023-00852-y
- Turner NA, Sharma-Kuinkel BK, Maskarinec SA, Eichenberger EM, Shah PP, Carugati M, Holland TL, Fowler VG Jr. 2019. Methicillin-resistant Staphylococcus aureus: an overview of basic and clinical research. Nat Rev Microbiol 17:203–218. https://doi.org/10.1038/s41579-018-0147-4
- BARBER M. 1961. Methicillin-resistant staphylococci. J Clin Pathol 14:385–393. https://doi.org/10.1136/jcp.14.4.385
- Algammal AM, Hetta HF, Elkelish A, Alkhalifah DHH, Hozzein WN, Batiha GE-S, El Nahhas N, Mabrok MA. 2020. Methicillin-resistant *Staphylococcus* aureus (MRSA): one health perspective approach to the bacterium epidemiology, virulence factors, antibiotic-resistance, and zoonotic impact. Infect Drug Resist 13:3255–3265. https://doi.org/10.2147/IDR.S2 72733
- Tong SYC, Davis JS, Eichenberger E, Holland TL, Fowler VG Jr. 2015. Staphylococcus aureus infections: epidemiology, pathophysiology, clinical manifestations, and management. Clin Microbiol Rev 28:603– 661. https://doi.org/10.1128/CMR.00134-14
- Uhlemann AC, Otto M, Lowy FD, DeLeo FR. 2014. Evolution of community- and healthcare-associated methicillin-resistant *Staphylo*coccus aureus. Infect Genet Evol 21:563–574. https://doi.org/10.1016/j.m eegid.2013.04.030
- Herold BC, Immergluck LC, Maranan MC, Lauderdale DS, Gaskin RE, Boyle-Vavra S, Leitch CD, Daum RS. 1998. Community-acquired methicillin-resistant *Staphylococcus aureus* in children with no identified predisposing risk. JAMA 279:593–598. https://doi.org/10.1001/jama.279. 8.593
- King MD, Humphrey BJ, Wang YF, Kourbatova EV, Ray SM, Blumberg HM. 2006. Emergence of community-acquired methicillin-resistant Staphylococcus aureus USA 300 clone as the predominant cause of skin and soft-tissue infections. Ann Intern Med 144:309–317. https://doi.org/ 10.7326/0003-4819-144-5-200603070-00005
- Wang B, Xu Y, Zhao H, Wang X, Rao L, Guo Y, Yi X, Hu L, Chen S, Han L, Zhou J, Xiang G, Hu L, Chen L, Yu F. 2022. Methicillin-resistant Staphylococcus aureus in China: a multicentre longitudinal study and whole-genome sequencing. Emerg Microbes Infect 11:532–542. https://doi.org/10.1080/22221751.2022.2032373
- Patel M, Kumar RA, Stamm AM, Hoesley CJ, Moser SA, Waites KB. 2007. USA300 genotype community-associated methicillin-resistant Staphylococcus aureus as a cause of surgical site infections. J Clin Microbiol 45:3431–3433. https://doi.org/10.1128/JCM.00902-07
- Lakhundi S, Zhang K. 2018. Methicillin-resistant Staphylococcus aureus: molecular characterization, evolution, and epidemiology. Clin Microbiol Rev 31:e00020-18. https://doi.org/10.1128/CMR.00020-18
- Chen H, Yin Y, van Dorp L, Shaw LP, Gao H, Acman M, Yuan J, Chen F, Sun S, Wang X, Li S, Zhang Y, Farrer RA, Wang H, Balloux F. 2021. Drivers of methicillin-resistant *Staphylococcus aureus* (MRSA) lineage replacement in China. Genome Med 13:171. https://doi.org/10.1186/s13073-021-0099
- Li S, Sun S, Yang C, Chen H, Yin Y, Li H, Zhao C, Wang H. 2018. The changing pattern of population structure of *Staphylococcus aureus* from Bacteremia in China from 2013 to 2016: ST239-030-MRSA Replaced by ST59-t437. Front Microbiol 9:332. https://doi.org/10.3389/fmicb.2018.00 332
- Álvarez A, Fernández L, Gutiérrez D, Iglesias B, Rodríguez A, García P. 2019. Methicillin-resistant *Staphylococcus aureus* in Hospitals: latest trends and treatments based on bacteriophages. J Clin Microbiol 57:e01006-19. https://doi.org/10.1128/JCM.01006-19
- Witte W, Strommenger B, Stanek C, Cuny C. 2007. Methicillin-resistant Staphylococcus aureus ST398 in humans and animals, central Europe. Emerg Infect Dis 13:255–258. https://doi.org/10.3201/eid1302.060924
- Zhao C, Liu Y, Zhao M, Liu Y, Yu Y, Chen H, Sun Q, Chen H, Jiang W, Liu Y, Han S, Xu Y, Chen M, Cao B, Wang H. 2012. Characterization of

- community acquired *Staphylococcus aureus* associated with skin and soft tissue infection in Beijing: high prevalence of PVL+ ST398. PLoS One 7:e38577. https://doi.org/10.1371/journal.pone.0038577
- Smith TC, Male MJ, Harper AL, Kroeger JS, Tinkler GP, Moritz ED, Capuano AW, Herwaldt LA, Diekema DJ. 2009. Methicillin-resistant Staphylococcus aureus (MRSA) strain ST398 is present in midwestern U.S. swine and swine workers. PLoS One 4:e4258. https://doi.org/10.1371/journal.pone.0004258
- Lee YH, Chen CJ, Lien RI, Huang YC. 2022. A longitudinal molecular surveillance of clinical methicillin-resistant *Staphylococcus aureus* isolates in neonatal units in A teaching hospital, 2003-2018. J Microbiol Immunol Infect 55:880–887. https://doi.org/10.1016/j.jmii.2021.10.007
- Henderson A, Nimmo GR. 2018. Control of healthcare- and communityassociated MRSA: recent progress and persisting challenges. Br Med Bull 125:25–41. https://doi.org/10.1093/bmb/ldx046
- Institute CaLS. 2020. M100 performance standards for antimicrobial susceptibility testing, M100.
- Testing ECoAS. 2024. Breakpoint tables for interpretation of MICs and zone diameters Version 14.0.
- Chen S, Zhou Y, Chen Y, Gu J. 2018. Fastp: an ultra-fast all-in-one FASTQ preprocessor. Bioinformatics 34:i884–i890. https://doi.org/10.1093/bioin formatics/bty560
- Bartels MD, Petersen A, Worning P, Nielsen JB, Larner-Svensson H, Johansen HK, Andersen LP, Jarløv JO, Boye K, Larsen AR, Westh H. 2014. Comparing whole-genome sequencing with Sanger sequencing for spa typing of methicillin-resistant *Staphylococcus aureus*. J Clin Microbiol 52:4305–4308. https://doi.org/10.1128/JCM.01979-14
- Kaya H, Hasman H, Larsen J, Stegger M, Johannesen TB, Allesøe RL, Lemvigh CK, Aarestrup FM, Lund O, Larsen AR. 2018. SCC mec finder, a web-based tool for typing of staphylococcal cassette chromosome mec in *Staphylococcus aureus* using whole-genome sequence data. mSphere 3. https://doi.org/10.1128/mSphere.00612-17
- Alcock BP, Raphenya AR, Lau TTY, Tsang KK, Bouchard M, Edalatmand A, Huynh W, Nguyen A-LV, Cheng AA, Liu S, et al. 2020. CARD 2020: antibiotic resistome surveillance with the comprehensive antibiotic resistance database. Nucleic Acids Res 48:D517–D525. https://doi.org/10 .1093/nar/gkz935
- Liu B, Zheng D, Jin Q, Chen L, Yang J. 2019. VFDB 2019: a comparative pathogenomic platform with an interactive web interface. Nucleic Acids Res 47:D687–D692. https://doi.org/10.1093/nar/gky1080
- Seemann T. 2014. Prokka: rapid prokaryotic genome annotation. Bioinformatics 30:2068–2069. https://doi.org/10.1093/bioinformatics/bt u153
- Tonkin-Hill G, MacAlasdair N, Ruis C, Weimann A, Horesh G, Lees JA, Gladstone RA, Lo S, Beaudoin C, Floto RA, Frost SDW, Corander J, Bentley SD, Parkhill J. 2020. Producing polished prokaryotic pangenomes with the Panaroo pipeline. Genome Biol 21:180. https://doi.org/10.1186/s130 59-020-02090-4
- Price MN, Dehal PS, Arkin AP. 2009. FastTree: computing large minimum evolution trees with profiles instead of a distance matrix. Mol Biol Evol 26:1641–1650. https://doi.org/10.1093/molbev/msp077
- Letunic I, Bork P. 2024. Interactive Tree of Life (iTOL) v6: recent updates to the phylogenetic tree display and annotation tool. Nucleic Acids Res 52:W78–W82. https://doi.org/10.1093/nar/gkae268
- Randall MM, Raae-Nielsen J, Tu CJ, Barcega BR, Young TP, Brown LA. 2024. Childhood respiratory illnesses before and after COVID-19 pandemic restrictions. Cureus 16:e72957. https://doi.org/10.7759/cureus .72957
- Li Q, Zhao L, Chen J, Qu S, Niu X, Zhu R, Hu W. 2024. Decreased hospitalacquired respiratory infections among older inpatients during the COVID-19 pandemic: a retrospective observational study in a general hospital in China. BMC Infect Dis 24:904. https://doi.org/10.1186/s12879-024-09779-y

- Vandenesch F, Naimi T, Enright MC, Lina G, Nimmo GR, Heffernan H, Liassine N, Bes M, Greenland T, Reverdy ME, Etienne J. 2003. Communityacquired methicillin-resistant Staphylococcus aureus carrying Panton-Valentine leukocidin genes: worldwide emergence. Emerg Infect Dis 9:978–984. https://doi.org/10.3201/eid0908.030089
- Qiao Y, Ning X, Chen Q, Zhao R, Song W, Zheng Y, Dong F, Li S, Li J, Wang L, Zeng T, Dong Y, Yao K, Yu S, Yang Y, Shen X. 2014. Clinical and molecular characteristics of invasive community-acquired *Staphylococ*cus aureus infections in Chinese children. BMC Infect Dis 14:582. https:// doi.org/10.1186/s12879-014-0582-4
- 35. Jin Y, Zhou W, Zhan Q, Zheng B, Chen Y, Luo Q, Shen P, Xiao Y. 2021. Genomic epidemiology and characterization of methicillin-resistant *Staphylococcus aureus* from bloodstream infections in China. mSystems 6:e0083721. https://doi.org/10.1128/mSystems.00837-21
- Chen C-J, Unger C, Hoffmann W, Lindsay JA, Huang Y-C, Götz F. 2013. Characterization and comparison of 2 distinct epidemic communityassociated methicillin-resistant *Staphylococcus aureus* clones of ST59 lineage. PLoS One 8:e63210. https://doi.org/10.1371/journal.pone.00632
- Sun C, Wang Q, Li WT, Wen DN, Chen CH, Yang X, Shi W, Meng QH, Yao KH, Qian SY. 2020. Molecular characteristics and antimicrobial susceptibility of Staphylococcus aureus among children with respiratory tract infections in Southwest China. World J Pediatr 16:284–292. https://doi.org/10.1007/s12519-019-00317-4
- Chen Y, Sun L, Ba X, Jiang S, Zhuang H, Zhu F, Wang H, Lan P, Shi Q, Wang Z, Chen Y, Shi K, Ji S, Jiang Y, Holmes MA, Yu Y. 2022. Epidemiology, evolution and cryptic susceptibility of methicillin-resistant Staphylococcus aureus in China: a whole-genome-based survey. Clin Microbiol Infect 28:85–92. https://doi.org/10.1016/j.cmi.2021.05.024
- Chen F, Yin Y, Chen H, Jin L, Li S, Wang R, Wang S, Wang Q, Sun S, Wang H. 2022. Phenotypic and genomic comparison of *Staphylococcus aureus* highlight virulence and host adaptation favoring the success of epidemic clones. mSystems 7:e0083122. https://doi.org/10.1128/msystems.00831-22
- Lee SM, Ender M, Adhikari R, Smith JMB, Berger-Bächi B, Cook GM. 2007. Fitness cost of staphylococcal cassette chromosome mec in methicillinresistant Staphylococcus aureus by way of continuous culture. Antimicrob Agents Chemother 51:1497–1499. https://doi.org/10.1128/A AC.01239-06
- Fu P, Nijiati Y, Li T, Wu X, Wang Z, Zhou J, Wang C, Ning B. 2023. Clinical and molecular characteristics of methicillin-resistant *Staphylococcus* aureus in bone and joint infection among children. Ann Clin Microbiol Antimicrob 22:104. https://doi.org/10.1186/s12941-023-00654-3
- Kpeli G, Buultjens AH, Giulieri S, Owusu-Mireku E, Aboagye SY, Baines SL, Seemann T, Bulach D, Gonçalves da Silva A, Monk IR, Howden BP, Pluschke G, Yeboah-Manu D, Stinear T. 2017. Genomic analysis of ST88 community-acquired methicillin resistant *Staphylococcus aureus* in Ghana. PeerJ 5:e3047. https://doi.org/10.7717/peerj.3047
- Hamza M, Sivaraman GK, Mothadaka MP. 2023. Multidrug-resistant phenotypes among biofilm-producing staphylococcal isolates from farm-raised fish: a molecular scrutiny. J Appl Microbiol 134:lxad136. https://doi.org/10.1093/jambio/lxad136
- Zhou W, Jin Y, Shen P, Chen W, Chen Y, Xiao Y. 2023. Novel SCCmec variants in clonal complex 398 and lineage-specific pseudo-SCCmec identified in ST88 MRSA from invasive bloodstream infections in China. J

- Antimicrob Chemother 78:2366–2375. https://doi.org/10.1093/jac/dkad 250
- Wang W, Li H, Li M, Dong Y, Bai Y, Li F, Xiao J. 2022. Molecular evolution and genomic insights into community-acquired methicillin-resistant Staphylococcus aureus sequence type 88. Microbiol Spectr 10:e0034222. https://doi.org/10.1128/spectrum.00342-22
- Dai Y, Liu J, Guo W, Meng H, Huang Q, He L, Gao Q, Lv H, Liu Y, Wang Y, Wang H, Liu Q, Li M. 2019. Decreasing methicillin-resistant *Staphylococcus aureus* (MRSA) infections is attributable to the disappearance of predominant MRSA ST239 clones, Shanghai, 2008-2017. Emerg Microbes Infect 8:471–478. https://doi.org/10.1080/22221751.2019.1595161
- Zhou W, Jin Y, Chen P, Ge Q, Dong X, Chen Y, Jiang M, Xiao Y. 2025. Reshaping the battlefield: a decade of clonal wars among *Staphylococcus aureus* in China. Drug Resist Updat 78:101178. https://doi.org/10.1016/j.drup.2024.101178
- Ji S, Jiang S, Wei X, Sun L, Wang H, Zhao F, Chen Y, Yu Y. 2020. In-host evolution of daptomycin resistance and heteroresistance in methicillinresistant Staphylococcus aureus strains from three endocarditis patients. J Infect Dis 221:S243–S252. https://doi.org/10.1093/infdis/jiz571
- Liang Y, Li J, Hou L, Zhang X, Hou G, Zhang W. 2023. Changes of Staphylococcus aureus infection in children before and after the COVID-19 pandemic, Henan, China. J Infect 86:e70–e71. https://doi.org/ 10.1016/j.jinf.2022.12.024
- Huang YC, Chen CJ. 2011. Community-associated meticillin-resistant Staphylococcus aureus in children in Taiwan, 2000s. Int J Antimicrob Agents 38:2–8. https://doi.org/10.1016/j.ijantimicag.2011.01.011
- Liu JL, Li TM, Zhong N, Wang X, Jiang J, Zhang WX, Tang R, Guo YJ, Liu Y, Hu J, He LH, Tang J, Wu WJ, Li M. 2021. Current status of oxacillinsusceptible mecA-positive *Staphylococcus aureus* infection in Shanghai, China: A multicenter study. J Microbiol Immunol Infect 54:1070–1077. ht tps://doi.org/10.1016/j.jmii.2020.07.021
- 52. Harrison EM, Ba X, Coll F, Blane B, Restif O, Carvell H, Köser CU, Jamrozy D, Reuter S, Lovering A, Gleadall N, Bellis KL, Uhlemann A-C, Lowy FD, Massey RC, Grilo IR, Sobral R, Larsen J, Rhod Larsen A, Vingsbo Lundberg C, Parkhill J, Paterson GK, Holden MTG, Peacock SJ, Holmes MA. 2019. Genomic identification of cryptic susceptibility to penicillins and β-lactamase inhibitors in methicillin-resistant *Staphylococcus aureus*. Nat Microbiol 4:1680–1691. https://doi.org/10.1038/s41564-019-0471-0
- Goering RV, Swartzendruber EA, Obradovich AE, Tickler IA, Tenover FC. 2019. Emergence of oxacillin resistance in stealth methicillin-resistant Staphylococcus aureus due to mecA sequence instability. Antimicrob Agents Chemother 63:e00558-19. https://doi.org/10.1128/AAC.00558-19
- Memmi G, Filipe SR, Pinho MG, Fu Z, Cheung A. 2008. Staphylococcus aureus PBP4 is essential for beta-lactam resistance in communityacquired methicillin-resistant strains. Antimicrob Agents Chemother 52:3955–3966. https://doi.org/10.1128/AAC.00049-08
- Jian Y, Li T, Zhao L, Zhao N, Liu Y, Lv H, Wang Y, Liu Q, Li M. 2022. Regulation of bla system in ST59-related oxacillin-susceptible mecA-positive Staphylococcus aureus. J Antimicrob Chemother 77:604–614. htt ps://doi.org/10.1093/jac/dkab460
- Naimi TS, LeDell KH, Como-Sabetti K, Borchardt SM, Boxrud DJ, Etienne J, Johnson SK, Vandenesch F, Fridkin S, O'Boyle C, Danila RN, Lynfield R. 2003. Comparison of community- and health care-associated methicillin-resistant Staphylococcus aureus infection. JAMA 290:2976–2984. https://doi.org/10.1001/jama.290.22.2976