Differentiation of Cartilage Repair Techniques Using Texture Analysis from T, Maps

CARTILAGE 2021, Vol. 13(Suppl 1) 718S-728S © The Author(s) 2021

Article reuse guidelines: sagepub.com/journals-permissions DOI: 10.1177/19476035211029698 journals.sagepub.com/home/CAR

Vladimir Juras¹, Pavol Szomolanyi^{1,2}, Veronika Janáčová¹, Alexandra Kirner³, Peter Angele³, and Siegfried Trattnig^{1,4,5,6}

Abstract

Objective. The aim of this study was to investigate texture features from T_2 maps as a marker for distinguishing the maturation of repair tissue after 2 different cartilage repair procedures. *Design.* Seventy-nine patients, after either microfracture (MFX) or matrix-associated chondrocyte transplantation (MACT), were examined on a 3-T magnetic resonance (MR) scanner with morphological and quantitative (T_2 mapping) MR sequences 2 years after surgery. Twenty-one texture features from a gray-level co-occurrence matrix (GLCM) were extracted. The texture feature difference between 2 repair types was assessed individually for the femoral condyle and trochlea/anterior condyle using linear regression models. The stability and reproducibility of texture features for focal cartilage were calculated using intra-observer variability and area under curve from receiver operating characteristics. *Results.* There was no statistical significance found between MFX and MACT for T_2 values (P = 0.96). There was, however, found a statistical significance between MFX and MACT in femoral condyle in GLCM features autocorrelation (P < 0.001), sum of squares (P = 0.023), sum average (P = 0.025), sum variance (P = 0.0048), and sum entropy (P = 0.05); and in anterior condyle/trochlea homogeneity (P = 0.02) and dissimilarity (P < 0.001). *Conclusion.* Texture analysis using GLCM provides a useful extension to T_2 mapping for the characterization of cartilage repair tissue after different cartilage repair procedures, as repair tissue texture (and hence, probably collagen organization) 24 months after MACT more closely resembled healthy cartilage than did MFX repair tissue.

Keywords

cartilage repair, articular cartilage, magnetic resonance imaging, knee

Introduction

To circumvent the invasive biopsy and limitations of clinical evaluation used to monitor patients after cartilage repair surgery, a number of magnetic resonance imaging (MRI) methods have been developed and validated as sensitive approaches with which to assess cartilage repair tissue maturation. MRI methods comprise morphological evaluation, typically using semiquantitative scoring systems, such as Magnetic Resonance Observation of Cartilage Repair Tissue (MOCART)¹⁻³ and glycosaminoglycan- and collagen-specific quantitative MRI techniques.⁴⁻⁷ T₂ mapping, in particular, is the most often used technique for cartilage repair tissue assessment as it reflects the collagen fiber network organization in cartilage, which is represented by the formation of different zones, which is a positive sign of successful tissue maturation after cartilage repair surgery.⁷⁻⁹ Unlike the modern glycosaminoglycan-specific techniques, such as sodium MRI and glycosaminoglycan chemical exchange saturation transfer (gagCEST), it does not require an ultra-high field MR scanner to be clinically feasible.^{6,10} As T_2 is also sensitive to the loading applied to the cartilage, it has been successfully used for functional cartilage repair

 ⁴CD laboratory for Clinical Molecular MR imaging, Vienna, Austria
⁵Austrian Cluster for Tissue Regeneration, Vienna, Austria
⁶Institute for Clinical Molecular MRI in the Musculoskeletal System, Karl Landsteiner Society, Vienna, Austria

Corresponding Author:

Vladimir Juras, High-Field MR Centre, Department of Biomedical Imaging and Image-guided Therapy, Medical University of Vienna, Waehringer Guertel 18-20, Vienna, 1090, Austria. Email: vladimir.juras@meduniwien.ac.at

¹High-Field MR Centre, Department of Biomedical Imaging and Imageguided Therapy, Medical University of Vienna, Vienna, Austria ²Institute of Measurement Science, Slovak Academy of Sciences, Bratislava, Slovakia

³TETEC AG, Reutlingen, Germany

evaluation.^{11,12} In a clinical setup, however, cartilage repair tissue is characterized by a mean T_2 value that can be misleading in some cases as T_2 mapping is limited in ability to detect the subtle details of the repair cartilage architecture and composition.¹³

In such instances, texture analysis of T₂ maps may be of great help, as it quantifies the relationship between individual pixels and some features can be potentially translated to imaging biomarkers. Texture analysis is used for the extracting the quantitative parameters to describe of (not only MRI) image texture. In the past years, it has been applied in MRI as a computer-aided diagnostic tool.¹⁴ As T₂ in cartilage is predominantly related to water content and collagen content and orientation, T₂ map texture provides an important information about collagen matrix status. Despite some technical challenges of the texture analysis of cartilage (including flattening of the cartilage and input parameter selection), the texture analysis using a gray-level co-occurrence matrix (GLCM) was successfully used in studying patients with osteoarthritis.¹⁵⁻¹⁷ There were several attempts to use texture analysis of cartilage on morphological images, but this is very challenging, as the texture of cartilage (i.e., signal differences) are dependent on sequence parameters and MR scanner-related variables (such as receiver gain),¹⁸ which are highly variable and thus suffer from low reproducibility.

There are different strategies for the surgical repair of articular cartilage lesions.¹⁹ The choice of the repair type depends on the location and size of the defect. Larger lesions are typically treated with osteochondral allograft transplantation or autologous chondrocyte transplantation (ACT), while smaller lesions are treated with marrow-stimulating techniques (such as microfracture [MFX]), or osteochondral autograft transfer.²⁰⁻²³ Matrix-associated autologous chondrocyte transplantation (MACT) is very attractive as it uses the patient's own chondrocytes for cartilage regeneration with the cells seeded on a scaffold (matrix) and can be used in larger cartilage defects.²⁴ Various types of scaffold as carriers for chondrocyte transplantation have been designed and tested with different clinical outcome quality.²⁵⁻²⁷

As T_2 values are generated based on the interplay between water molecules and collagen fibers, the absolute values create a texture that reflects the collagen organization. Recently, texture analysis became very popular either as an independent tool or as a part of machine-learning algorithms for advanced image analysis from different imaging modalities.^{14,28,29} Despite the complex shape of articular cartilage, texture analysis using a GLCM was successfully used on cartilage T_2 maps to reveal early stages of cartilage degeneration in osteoarthritis (OA)^{16,17,30} or serve as a marker for risk factors of OA progression.³¹ Along with T_2 mapping, the feasibility of texture analysis of cartilage was tested on T_1 mapping³² and morphological imaging,¹⁸ with inconclusive results. To the best of our knowledge, no study has previously investigated focal cartilage lesions using texture analysis for monitoring of cartilage repair maturation. To this end, we suggested an evaluation pipeline for focal cartilage texture analysis using GLCM, including region of interest (ROI) pre-adjustment and validation of suitable texture features selection. The method was used in 2 patient collectives after 2 different cartilage repair types to investigate the ability to distinguish between the outcomes of repair tissue after the surgical procedures.

Materials and Methods

Study Design and Patient Demographics

The study was approved by the appropriate ethics committees and regulatory authorities separately for each individual participating site. The results presented here are derived from the subgroup of patients who participated in the MRI substudy of a prospective, multicenter, randomized, controlled, open-label (blinded MRI reading), phase III study comparing the efficacy and safety of MACT using NOVOCART 3D plus (TETEC AG, Reutlingen, Germany) versus MFX in patients with cartilage defects of the knee. Patients were allocated randomly to the MACT or MFX group in a 2:1 ratio and were to be followed-up for 5 years after cartilage repair surgery. Generally, males and females 18 to 65 years of age (or minors of at least 14 years of age with a closed epiphyseal growth plate) with a localized articular cartilage defect of the femoral condyle or the trochlea of the knee (defect grade of III or IV according to the International Cartilage Repair Society (ICRS) classification; maximum of 2 defects) were eligible for enrollment. The maximum total defect size was limited to 6 cm²; minimum defect size was 2 cm².

The total patient population (n = 262) consisted of 189 males (72.1%) and 73 females (27.9%). The mean age of the patients was (average ± standard deviation) 39.9 ± 10.6 years. Most of the study patients (233 patients, 88.9%) had 1 single lesion, while 29 patients (11.1%) had 2 lesions. All reported lesions had an ICRS grade of 3 or 4. Most of the cartilage lesions in either treatment group were of traumatic origin (77.4% of the lesions in the MFX group and 78.3% of lesions in the MACT group). Both the total lesion size (MFX group: $3.6 \pm 1.3 \text{ cm}^2$; MACT group: $3.8 \pm 1.4 \text{ cm}^2$) and the mean larger lesion size (MFX group: $3.6 \pm 1.2 \text{ cm}^2$) were similar in both treatment groups.

One hundred and ten patients were involved in the MRI substudy (MACT: 75 patients; MFX: 35 patients) and had MRIs performed 3, 12, 24, and 60 months after cartilage repair surgery. For the GLCM analysis presented here (24 months time point), T_2 maps were available for a total of 79 patients (MACT: 55 patients; MFX: 24 patients) after excluding lesions in patella.

Parameter/Sequence	T ₂ Mapping	TSE PD	TSE T ₂ w	SE T _I w	
Orientation plane	Sagittal	Coronal	Sagittal	Sagittal	
Slice thickness (mm)	3	3	2	2	
Slice spacing (mm)	3.3	3.3	2.2	2.2	
Repetition time (ms)	2,000	3,080	3,310	700	
Echo time(s) (ms)	12.5; 25; 37.5; 50; 62.5; 75; 87.5; 100	28	12	12	
Averages	I	2	3	I	
Flip angle (°)	90, 180	180	180	90	
Acquisition matrix	320 imes 256	448 $ imes$ 403	381 $ imes$ 448	448 $ imes$ 381	
Image matrix	320 imes 320	896 $ imes$ 896	448 $ imes$ 448	448 imes448	
Field of view (cm)	16 × 16	16 imes 16	16 imes 16	16 imes 16	
Total acquisition time (minutes:seconds)	10:36	4:46	3:55	3:53	

Table I. Parameters of the MRI Protocol Used in the Multicenter Trial.

 $TSE = turbo spin echo; SE = spin echo; T_2w = T_2 weighting; T_1w = T_1 weighting; PD = proton-density weighted.$

MR Examination

The protocol consisted of a morphological part (TSE PD, TSE T2w, SE T1w) and T₂ mapping using a multi-echo multi-slice sequence. T₂ maps were acquired with the following parameters: orientation, sagittal; slice thickness, 3 mm; echo times, (12.5; 25; 37.5; 50; 62.5; 75; 87.5; 100) ms; acquisition matrix, 320 × 320; field of view, 16 × 16 mm; in-plane resolution 0.5×0.5 mm; and total scan time 10:36 minutes. The sequence parameters of the whole MR examination protocol are listed in detail in **Table 1**. The central reading site collected all images, performed T₂ mapping using a 2-parametric exponential fitting method,³³ and conducted cartilage repair segmentation and scoring.

Image and Texture Analysis

Regions of interest were selected on T₂ weighted images in JiveX (Visus, Bochum, Germany) and subsequently transferred for further processing to MatLab 2020b (Mathworks, Natick, MA). For each patient, 2 to 4 consecutive slices were evaluated to cover the whole cartilage repair site. For each slice, repair cartilage and a reference cartilage were selected by an experienced musculoskeletal radiologist with 28 years of experience (S.T.). As the ROI size was similar between slices, mean T2 was calculated and averaged through the slices ROI-wise resulting in 2 T₂ values per patient (one for the lesion, one for the reference). Zonal T₂ value of cartilage was calculated as a ratio of superficial and deep zone by dividing the cartilage in halves along the axis perpendicular to cartilage surface. Each ROI was transferred to MatLab and processed with in-house-written scripts. This script automatically loaded the ROIs selected by a reader and performed a texture analysis. The calculation itself took approximately 10 seconds per patient. First, the ROI was automatically rotated (using MatLab function "imrotate" with an argument "orientation" from

the function "regionprops") to be as close as possible to a rectangular shape. Then, GLCM was produced using the GLCM features1 function from the MatLab Repository and 21 textural features were extracted (autocorrelation, contrast, correlation, cluster prominence, cluster shade, dissimilarity, energy, entropy, homogeneity, maximum probability, sum of squares, sum average, sum variance, sum entropy, difference variance, difference entropy, information measure of correlation1, information measure of correlation2, inverse difference, inverse difference normalized, and inverse difference moment normalized).³⁴ All features were calculated individually for each slice and then averaged. GLCM was processed with the following setup: offset 0° (parallel to the cartilage surface), a direction parallel to the cartilage surface, 16 gray levels, and a step of one pixel. The optimization of GLCM setup (offset, number of gray levels, and step) for focal cartilage texture was done prior to this study and published elsewhere.³⁵ The complete ROI processing pipeline prior to texture analysis is depicted in Figures 1 and 2.

Texture Features Reproducibility

To assess the reproducibility of individual texture features, 35 patients were evaluated by 2 independent readers performing GLCM feature selection. To define the impact of ROI selection on the texture features, intra-observer variability was calculated and expressed as a coefficient-of-variation (CV, %) for each parameter, separately for reference and repaired cartilage. In order to quantify the interrelationship between the textural features of cartilage, a cross-correlation matrix was constructed using a paired *t*-test. To evaluate the ability of the texture features to distinguish between reference and repaired cartilage, receiver operation curve (ROC) analysis was used on the entire patient cohort. The area under the curve (AUC) was calculated for each parameter, and interpreted as follows: 0.80 to



Figure 1. A diagram of texture analysis of cartilage focal lesion.



Figure 2. An example of a lesion and reference cartilage ROI selection $(\mathbf{A}-T_2$ -weighted turbo-spin echo image; $\mathbf{B}-T_2$ map of segmented cartilage overlaid on the first echo time image); cartilage repair (referred to as the lesion) was selected to include the maximum amount of repaired tissue and a reference cartilage was selected to match the texture of a repair tissue. Extracted ROIs of reference (\mathbf{C}) and lesion (\mathbf{F}) were rotated by automatic angle detection (\mathbf{D} , \mathbf{G}) and, finally, a GLCM was constructed (\mathbf{E} , \mathbf{H}).

Statistical Evaluation

As a T_2 map texture of cartilage in weight-bearing and nonweight-bearing zones may differ due to different zonal cartilage stratification, patients were divided into 2 groups according to the repair site location (weight-bearing cartilage = condyle, non-weight-bearing cartilage = anterior condyle/trochlea).

An exploratory analysis of GLCM parameters was performed using a linear model with the fixed categorical effects of "treatment" and "region." For all parameters, the Shapiro-Wilk test for normality evaluation was used. Treatment differences were estimated using contrast *t*-tests between MACT and MFX for each GLCM parameter. P values lower than 0.05 were considered statistically significant.

Results

All GLCM features as well as T₂ values were normally distributed. The interobserver variability showed a low coefficient of variation for T2 values and for some texture features (autocorrelation, contrast, correlation, dissimilarity, energy, sum average, sum entropy). For some features, the CV was relatively high (cluster shade [$35.79 \pm 46.52\%$] and sum variance $[15.15 \pm 13.64\%]$), and in the case of the cluster prominence, even extremely high (516.50 \pm 1005.50%). In general, CVs were lower by 1.84% (P = 0.344) for the reference cartilage compared to the repair site. ROC analysis revealed moderate to high AUC, and the higher values were recorded in the condyle patient group compared to the anterior condyle/trochlea group. The highest AUC values were found in autocorrelation (0.799), sum of squares (0.810), and sum average (0.795). All CVs and AUC values are listed in Table 2. The cross-correlation matrix revealed the interrelationship between autocorrelation and 3 features (sum of squares, sum average, and sum variance). Dissimilarity and contrast were highly correlated as well. The entire cross-correlation matrix is depicted in **Figure 3**.

From the total number of 57 patients in the condyle group, 16 had MFX treatment, and 41 had MACT treatment. There was no statistical significance between MFX and MACT for T₂ values (46.57 ± 8.14 ms and 46.22 ± 7.46 ms, respectively, P = 0.96) neither for zonal T₂ analysis (1.11 ± 0.3 a.u. and 1.11 ± 0.5 a.u., respectively, P = 0.98). In case of texture features, statistical significance between MFX and MACT was found in autocorrelation (33.71 ± 7.83 and 50.07 ± 5.02, P < 0.001), sum of squares (35.63 ± 7.17 and 48.25 ± 4.59, P < 0.001), sum average (10.47 ± 1.18 and 12.38 ± 0.76, P = 0.01), sum variance

 $(92.00 \pm 23.64 \text{ and } 130.29 \pm 15.16, P < 0.001)$, and sum entropy $(2.84 \pm 0.07 \text{ and } 2.92 \pm 0.04, P = 0.05)$.

From the total number of 22 patients in the trochlea/anterior condyle group, 8 had MFX treatment, and 14 had MACT treatment. There was no statistical significance between MFX and MACT for T₂ values (46.41 ± 9.5 and 53.25 ± 8.7, respectively, P = 0.43) neither for zonal T₂ analysis (1.12 ± 0.12 a.u. and 1.05 ± 0.09 a.u., respectively, P = 0.98). In case of texture features, there was statistical significance between MFX and 1.03 ± 0.22, respectively, P < 0.001), homogeneity (0.52 ± 0.06 and 0.63 ± 0.06, respectively, P = 0.23), and inverse difference normalized INN (0.92 ± 0.02 and 0.94 ± 0.01, respectively P = 0.03).

The individual parameters (T_2 and texture features) are listed for the condyle patient group and the trochlea/anterior condyle patient groups in **Table 3** and **Table 4**, respectively.

Discussion

The findings presented in this study showed that texture features based on a GLCM extracted from cartilage T_2 maps are able to discriminate tissue originating from different cartilage repair methods and potentially can serve as a marker of cartilage repair tissue maturation. Several texture features acquired from GLCM demonstrated high reproducibility, as well as the ability to depict the cartilage texture, especially zonal stratification and overall homogeneity. Unlike T_2 values, which describe a cartilage tissue with a single mean value over the whole region of interest and, to some extent, a cartilage heterogeneity through standard deviation, texture features reflect the differences of the pixels and their distribution in the ROI.

In this work, some texture features demonstrated relatively high reproducibility in combination with a high AUC, namely, autocorrelation, contrast, homogeneity, sum of squares, sum average, and sum entropy, in particular. This suggests the independence of cartilage lesion and reference selection by differently trained readers, as well as the relatively high ability to distinguish cartilage repair tissue from healthy cartilage. On the other hand, other texture features seem to be extremely dependent on cartilage lesion and reference selection and a small difference in cartilage delineation may result in large discrepancies between readers. It could be due to the fact that it is often challenging to correctly define the cartilage-bone interface on T_2 maps, as well as the cartilage surface and synovial fluid boundary.

In previously published studies, different selections of texture features were used. To investigate the spatial variation of T_2 values in cartilage of postmenopausal osteoarthritis patients and age-matched healthy volunteers, Blumenkrantz *et al.* analyzed angular second moment and

		Interobserve	ROC Analysis			
Parameters	Repair		R	eference	Repair vs. Reference	
	Mean CV	95% CI	Mean CV	95% CI	Condyle (AUC)	Anterior Condyle/ Trochlea (AUC)
Τ,	2.31	(2.19, 2.43)	1.78	(1.70, 1.86)	0.540	0.590
Autocorrelation	4.94	(4.74, 5.14)	6.31	(6.09, 6.53)	0.799	0.703
Contrast	0.31	(0.30, 0.32)	0.41	(0.38, 0.44)	0.649	0.660
Correlation	0.02	(0.02, 0.02)	0.02	(0.02, 0.02)	0.529	0.533
Cluster prominence	516.50	(483.91, 549.09)	477.21	(461.12, 493.30)	0.661	0.669
Cluster shade	35.79	(34.72, 36.86)	28.72	(26.90, 30.54)	0.715	0.612
Dissimilarity	0.07	(0.07, 0.07)	0.09	(0.09, 0.09)	0.681	0.676
Energy	0.01	(0.01, 0.01)	0.00	(0.02, 0.02)	0.702	0.689
Entropy	0.09	(0.08, 0.10)	0.09	(0.09, 0.09)	0.633	0.649
Homogeneity	0.02	(0.02, 0.02)	0.02	(0.02, 0.02)	0.703	0.689
Maximum probability	0.01	(0.01, 0.01)	0.01	(0.01, 0.01)	0.695	0.692
Sum of squares	5.11	(4.74, 5.48)	6.29	(5.90, 6.68)	0.810	0.705
Sum average	0.77	(0.75, 0.79)	0.80	(0.74, 0.86)	0.795	0.703
Sum variance	15.15	(14.26, 16.04)	20.18	(19.49, 20.87)	0.799	0.698
Sum entropy	0.04	(0.04, 0.04)	0.05	(0.05, 0.05)	0.714	0.705
Difference variance	0.31	(0.30, 0.32)	0.41	(0.39, 0.43)	0.649	0.660
Information measure	0.02	(0.02, 0.02)	0.02	(0.02, 0.02)	0.630	0.651
Information measure of correlation2	0.01	(0.01, 0.01)	0.01	(0.01, 0.01)	0.533	0.510
Inverse difference normalized INN	0.08	(0.07, 0.09)	0.07	(0.06, 0.08)	0.598	0.608
Inverse difference moment normalized	0.16	(0.17, 0.19)	0.15	(0.14, 0.16)	0.688	0.678

Table 2. Coefficient of Variation (CV [%]) and ROC Calculated for Different Parameters (T₂ and Texture Features) Extracted from Reference Cartilage and Repair Cartilage.

ROC = receiver operation curve; CI = confidence interval; AUC = area under the curve.

entropy. Entropy was found to be higher in OA patients, suggesting that T₂ values in osteoarthritic cartilage are not only elevated but are also more heterogeneous.¹⁵ Joseph et al. investigated patients at risk of OA, and compared to healthy controls, they found higher T₂ values as well as higher contrast and variance (but not entropy).³¹ Carballido-Gamio et al. showed that texture features detected both a different laminar organization and longitudinal laminar changes of cartilage in patients with knee OA. They investigated 2 directions of texture analysis; tendencies showed higher dissimilarity, contrast, energy, and angular second moment perpendicular to the cartilage layers, and higher variance, entropy, homogeneity, and correlation parallel to them.³⁶ Texture analysis of T₂ maps was previously used to predict the need for total knee replacement; contrast was associated with an up to 40% increased risk for total knee replacement in the lateral femur and tibia, and variance in the lateral femur.³⁷ Chanchek et al. used T₂ and 3 texture features, variance, contrast, and entropy, to distinguish between diabetes mellitus patients and healthy controls. For all 4 parameters, the statistically significant higher values

were found in diabetes mellitus patients, suggesting that this disease negatively influences cartilage tissue.³⁸

However, several aspects need to be considered when performing texture analysis of cartilage, such as cartilageflattening methods, parameter selection from a GLCM (number of gray levels, orientation, and step size), and feature selection and interpretation. As knee articular cartilage is of an extremely irregular shape with various curvatures and thicknesses, it has to be flattened before an actual texture analysis to maintain the selected GLCM direction for all pixels. Flattening methods might have a substantial impact on the resulting texture features. Carballido-Gamio et al. compared 3 different flattening approaches: classical reshaping; a parallel method using Bezier spline along the cartilage layers; and a warping method using nonlinear deformation. They found differences in texture features when using different flattening methods; warping was found to be the most appropriate flattening strategy for texture analysis.¹⁶ An alternative to the flattening methods is the variable angle texture analysis of cartilage using the adaptive offset based on the pixel location within the cartilage.¹⁷



Figure 3. Cross-correlation matrix of T2 values, standard deviation of T2, and 20 GLCM texture features.

In our study, the mean T_2 value was not sensitive enough to distinguish between MFX and MACT repair types (in condyle mean difference $T_2 = +0.32$ ms, P = 0.96; in trochlea/anterior condyle mean difference $T_2 = +6.84$ ms, P = 0.43). Zonal T₂ analysis did not show any significant differences between MFX and MACT either. Previous studies demonstrated the ability of T₂ mapping to distinguish between cartilage repair in various knee locations,³⁹ to monitor the patients during repair maturation⁹ and also to differentiate between repair types.^{40,41} Welsch et al. found significantly lower T₂ values in MFX (47.9 \pm 9.8 ms) compared to T₂ values in MACT (53.6 \pm 11.9 ms) and a significantly lower T₂ index in MFX (0.89 \pm 0.12) and MACT (0.99 ± 0.16) , and they attributed this difference to more fibrocartilaginous-like characterization of MFX and hyaline-like nature of MACT.⁴¹ We could not reproduce those findings in our study; one of the reasons could be that they used patients at 36 months after surgery when the tissue differentiation may be more pronounced compared to 24 months follow-up used in our study.

In our study, some of the texture features were significantly different between the 2 repair tissue types (mean differences in condyle, autocorrelation = +16.36 m, P < 0.001; sum of squares = +12.62, P < 0.001; sum average = +1.91, P = 0.01; sum variance = +38.29, P < 0.001; and sum entropy = +0.08, P = 0.05; in trochlea/anterior condyle, dissimilarity = -0.49, P < 0.001; homogeneity = +0.11, P = 0.02; and inversion difference normalized INN = +0.02, P = 0.03). Comparing these differences to the reference cartilage values, it seems that repair tissue texture (and, hence, probably collagen organization) 24 months after MACT more closely resembles healthy cartilage than does tissue after MFX. However, the interpretation of some texture features with regard to cartilage quality is a difficult task, as some degree of inhomogeneity (zonal stratification) is a characteristic of "normal"

	Reference ($N = 57$)		MFX (N = 16)		MACT ($N = 41$)		D)/alua
Femoral Condyle	Mean	95% CI	Mean	95% CI	Mean	95% CI	(MFX vs. MACT)
Global T ₂	44.06	(36.56, 51.56)	46.57	(38.43, 54.71)	46.22	(38.76, 53.68)	0.96
Superficial/deep T ₂	1.13	(1.09, 1.18)	1.11	(0.98, 1.24)	1.11	(0.96, 1.26)	0.98
Autocorrelation	69.94	(64.76, 75.12)	33.71	(25.88, 41.54)	50.07	(45.05, 55.09)	<0.001
Contrast	2.84	(2.63, 3.05)	3.15	(2.23, 4.07)	3.44	(2.85, 4.03)	0.57
Correlation	0.88	(0.81, 0.94)	0.80	(0.75, 0.85)	0.86	(0.82, 0.89)	0.05
Cluster prominence	3,138	(2,899, 3,376)	2,702	(2,120, 3,283)	2,927	(2,554, 3,300)	0.49
Cluster shade	14.25	(12.75, 14.94)	107.78	(70.92, 142.81)	72.11	(49.40, 95.49)	0.09
Dissimilarity	1.49	(1.38, 1.60)	1.31	(1.10, 1.52)	1.24	(1.10, 1.38)	0.54
Energy	0.04	(0.03, 0.04)	0.04	(0.04, 0.05)	0.04	(0.03, 0.04)	0.18
Entropy	3.56	(3.30, 3.81)	3.58	(3.46, 3.71)	3.70	(3.62, 3.79)	0.09
Homogeneity	0.66	(0.61, 0.71)	0.55	(0.50, 0.60)	0.59	(0.56, 0.62)	0.12
Maximum probability	0.08	(0.07, 0.08)	0.10	(0.08, 0.12)	0.09	(0.08, 0.10)	0.17
Sum of squares	55.69	(51.64, 59.75)	35.63	(28.46, 42.80)	48.25	(43.66, 52.85)	<0.001
Sum average	14.73	(13.59, 15.88)	10.47	(9.29, 11.65)	12.38	(11.62, 13.13)	0.01
Sum variance	155.51	(144.18, 166.84)	92.00	(68.36, 115.64)	130.29	(115.13, 145.44)	<0.001
Sum entropy	3.03	(2.81, 3.25)	2.84	(2.77, 2.92)	2.92	(2.88, 2.97)	0.05
Difference variance	3.89	(3.58, 4.20)	3.15	(2.23, 4.07)	3.44	(2.85, 4.03)	0.57
Difference entropy	1.45	(1.34, 1.56)	1.36	(1.23, 1.49)	1.38	(1.30, 1.46)	0.82
Information measure	-0.39	(-0.36, -0.41)	-0.38	(-0.43, -0.33)	-0.38	(-0.41, -0.35)	0.91
Information measure of correlation	0.90	(0.84, 0.97)	0.90	(0.88, 0.92)	0.90	(0.89, 0.92)	0.68
Inverse difference normalized INN	0.93	(0.92, 0.94)	0.93	(0.92, 0.94)	0.93	(0.92, 0.93)	0.62
Inverse difference moment normalized	0.99	(0.98, 0.99)	0.99	(0.98, 0.99)	0.99	(0.98, 0.99)	0.56

Table 3. T_2 and Texture Features in the Condyle Group for Reference and Repair Tissue after MACT and MFX and Statistical Differences between MACT and MFX^a.

MFX = microfracture; MACT = matrix-associated chondrocyte transplantation; CI = confidence interval.

^aStatistically significant differences (P < 0.05) are marked in bold.

cartilage. Autocorrelation seems to be the most reliable feature, as it reflects the repetitive patterns of the texture that can be translated as cartilage zonal stratification. Homogeneity and its inverse counterpart, dissimilarity, are harder to interpret, as it is impossible to determine whether an inhomogeneity results from disorganized collagen fibers or from zonal stratification. Our results suggest, however, that the former inhomogeneity influence is somehow larger, which resulted in the ability of texture-homogeneity to distinguish between repair tissue types.

Our results support earlier findings on inferior repair tissue quality after MFX compared to MACT. In a meta-analysis, MFX was found to produce primarily fibrocartilage.⁴² Hyaline repair tissue was more common with ACT than with MFX.⁴³ The inferior quality of repair tissue after MFX, and especially, in larger lesions, as well as a worse defect filling compared to MACT, is considered the reason for long-term inferior clinical results of MFX.^{44,45} In this context, it has been shown that filling of smaller, well-shouldered cartilage defects, even with non-hyaline repair tissue, still improves function and clinical complaints within the first few years, while the histological repair tissue quality becomes more important for larger defects and in the longer term.^{43,45-47} This meta-analysis also found a difference with regard to tissue maturation. While repair tissue from MACT becomes more hyaline-like (a process that takes up to 5 years),⁴⁸ demonstrating tissue maturation with increased stiffness of the repair tissue, the amount of fibrocartilage formed after MFX only enlarges over time, but usually without maturation into cartilage with hyaline properties.

This study had several limitations. First, the sample size was relatively low, particularly the trochlea/anterior condyle group. In theory, these 2 groups could be merged together to increase the statistical sample size, but it would very likely result in lower sensitivity of the calculated texture features, as zonal stratification as well as absolute T_2 values vary in different cartilage locations. Second, the texture analysis was performed slice-wise rather than in 3D fashion. Three-dimensional texture analysis is definitely more robust compared to 2D,^{49,50} but because of the

Trochlea/Anterior Condyle	Reference ($N = 22$)		$MFX\;(N=8)$		MACT ($N = 14$)		P Value
	Mean	95% CI	Mean	95% CI	Mean	95% CI	(MFX vs. MACT)
Τ,	45.21	(41.01, 49.21)	46.41	(36.91, 55.91)	53.25	(44.55, 61.95)	0.43
Superficial/deep T ₂	1.04	(0.94, 1.15)	1.12	(1.00, 1.23)	1.05	(0.96, 1.15)	0.92
Autocorrelation	65.37	(60.62, 70.11)	52.26	(38.96, 65.56)	43.89	(31.66, 56.12)	0.34
Contrast	2.82	(2.61, 3.04)	3.68	(2.62, 4.75)	2.50	(1.53, 3.48)	0.10
Correlation	0.90	(0.83, 0.96)	0.79	(0.72, 0.85)	0.86	(0.80, 0.92)	0.08
Cluster prominence	3,219	(2,969, 3,469)	3,231	(2,376, 4,086)	3,020	(2,233, 3,806)	0.71
Cluster shade	90.60	(72.89, 96.70)	36.34	(-23.11, 94.75)	77.14	(23.06, 131.48)	0.29
Dissimilarity	1.54	(1.43, 1.66)	1.52	(1.29, 1.76)	1.03	(0.81, 1.24)	<0.001
Energy	0.06	(0.06, 0.07)	0.04	(0.02, 0.05)	0.05	(0.04, 0.07)	0.10
Entropy	3.29	(3.03, 3.55)	3.71	(3.49, 3.93)	3.47	(3.27, 3.68)	0.12
Homogeneity	0.66	(0.62, 0.71)	0.52	(0.46, 0.59)	0.63	(0.57, 0.69)	0.02
Maximum probability	0.10	(0.09, 0.11)	0.08	(0.05, 0.11)	0.11	(0.09, 0.14)	0.14
Sum of squares	42.82	(39.59, 46.06)	50.28	(36.62, 63.94)	44.43	(31.86, 57.00)	0.52
Sum average	15.04	(13.97, 16.11)	12.64	(10.60, 14.68)	11.53	(9.66, 13.41)	0.41
Sum variance	147.12	(135.61, 158.64)	138.86	(97.84, 179.87)	120.61	(82.88, 158.34)	0.50
Sum entropy	2.83	(2.62, 3.03)	2.92	(2.78, 3.06)	2.81	(2.68, 2.94)	0.26
Difference variance	3.91	(3.61, 4.21)	3.68	(2.62, 4.75)	2.50	(1.53, 3.48)	0.10
Difference entropy	1.47	(1.37, 1.58)	1.45	(1.30, 1.60)	1.26	(1.12, 1.40)	0.07
Information measure	-0.50	(-0.46, -0.54)	-0.38	(-0.44, -0.32)	-0.41	(-0.46, -0.36)	0.48
Information measure of correlation	0.90	(0.84, 0.96)	0.90	(0.87, 0.93)	0.90	(0.87, 0.93)	0.93
Inverse difference normalized INN	0.97	(0.95, 0.99)	0.92	(0.90, 0.93)	0.94	(0.93, 0.95)	0.03
Inverse difference moment normalized	0.98	(0.98, 0.99)	0.98	(0.98, 0.99)	0.99	(0.99, 0.99)	0.10

Table 4. T₂ and Texture Features in the Trochlea/Anterior Condyle Group for Reference and Repair Tissue after MACT and MFX and Statistical Differences between MACT and MFX^a.

 $\mathsf{MFX}=\mathsf{microfracture};\,\mathsf{MACT}=\mathsf{matrix}\text{-}\mathsf{associated}\;\mathsf{chondrocyte}\;\mathsf{transplantation};\,\mathsf{CI}=\mathsf{confidence}\;\mathsf{interval}.$

^aStatistically significant differences (P < 0.05) are marked in bold.

nonzero slice distance typically used in multi-echo spinecho T_2 mapping, it is impossible to implement 3D texture analysis on T_2 maps.

the status of repaired cartilage may provide additional information about the cartilage structure and composition.

Conclusion

In conclusion, texture analysis using GLCM provides a useful add-on to T₂ mapping for the characterization of cartilage repair tissue by increasing its sensitivity to overall structure. Some texture features, such as autocorrelation, homogeneity, dissimilarity, sum of squares, sum of averages, and sum entropy, were able to distinguish between repair tissue that resulted from MACT and MFX, whereby repair tissue texture (and hence, probably collagen organization) 24 months after MACT more closely resembled healthy cartilage compared with MFX repair tissue. This is in accordance with other publications that have reported better repair tissue quality with ACT compared to MFX. In terms of methodology, it is crucial to individually evaluate weight-bearing and non-weight-bearing cartilage, as their texture substantially differs. The results of this study suggest that using texture analysis in clinical trials monitoring

Acknowledgments and Funding

The author(s) disclosed receipt of the following financial support for the research, authorship, and/or publication of this article: The financial support by the Austrian Federal Ministry for Digital and Economic Affairs and the National Foundation for Research, Technology and Development is gratefully acknowledged. Funding information: Austrian Science Fund, KLIF-541 B30, Recipient: Vladimir Juras. Slovak Scientific Grant Agency VEGA 2/0030/20, Recipient: Pavol Szomolanyi.

Declaration of Conflicting Interests

The author(s) declared no potential conflicts of interest with respect to the research, authorship, and/or publication of this article.

Ethical Approval

Ethical approvals for this study were obtained separately for each individual participating site (study registered on EU Clinical Trials Register under number EUDRACT NO.: 2016-002817-22).

Informed Consent

Written informed consent was obtained from all subjects before the study.

ORCID iD

Vladimir Juras 🕩 https://orcid.org/0000-0002-4026-5922

References

- Marlovits S, Singer P, Zeller P, Mandl I, Haller J, Trattnig S. Magnetic Resonance Observation of Cartilage Repair Tissue (MOCART) for the evaluation of autologous chondrocyte transplantation: determination of interobserver variability and correlation to clinical outcome after 2 years. Eur J Radiol. 2006;57:16-23.
- Schreiner MM, Raudner M, Marlovits S, Bohndorf K, Weber M, Zalaudek M, *et al.* The MOCART (Magnetic Resonance Observation of Cartilage Repair Tissue) 2.0 Knee Score and Atlas. Cartilage. Epub Aug 17, 2019. doi:10 .1177/1947603519865308
- Roemer FW, Guermazi A, Trattnig S, Apprich S, Marlovits S, Niu J, *et al.* Whole joint MRI assessment of surgical cartilage repair of the knee: Cartilage Repair Osteoarthritis Knee Score (CROAKS). Osteoarthritis Cartilage. 2014;22(6):779-99.
- Trattnig S, Domayer S, Welsch GW, Mosher T, Eckstein F. MR imaging of cartilage and its repair in the knee—a review. Eur Radiol. 2009;19(7):1582-94.
- Welsch GH, Trattnig S, Scheffler K, Szomonanyi P, Quirbach S, Marlovits S, *et al.* Magnetization transfer contrast and T2 mapping in the evaluation of cartilage repair tissue with 3T MRI. J Magn Reson Imaging. 2008;28(4):979-86.
- Zbyn S, Mlynarik V, Juras V, Szomolanyi P, Trattnig S. Evaluation of cartilage repair and osteoarthritis with sodium MRI. NMR Biomed. 2016;29(2):206-15.
- 7. Watanabe A, Boesch C, Anderson SE, Brehm W, Mainil Varlet P. Ability of dGEMRIC and T2 mapping to evaluate cartilage repair after microfracture: a goat study. Osteoarthritis Cartilage. 2009;17(10):1341-9.
- Welsch GH, Mamisch TC, Domayer SE, Dorotka R, Kutscha-Lissberg F, Marlovits S, *et al.* Cartilage T2 assessment at 3-T MR imaging: in vivo differentiation of normal hyaline cartilage from reparative tissue after two cartilage repair procedures initial experience. Radiology. 2008;247(1):154-61.
- 9. Welsch GH, Mamisch TC, Marlovits S, Glaser C, Friedrich K, Hennig FF, *et al.* Quantitative T2 mapping during followup after matrix-associated autologous chondrocyte transplantation (MACT): full-thickness and zonal evaluation to visualize the maturation of cartilage repair tissue. J Orthop Res. 2009;27(7):957-63.
- Brinkhof S, Nizak R, Khlebnikov V, Prompers JJ, Klomp DWJ, Saris DBF. Detection of early cartilage damage: feasibility and potential of gagCEST imaging at 7T. Eur Radiol. 2018;28(7):2874-81.
- Juras V, Welsch GH, Millington S, Szomolanyi P, Mamisch TC, Pinker K, *et al.* Kinematic biomechanical assessment of human articular cartilage transplants in the knee using 3-T MRI: an in vivo reproducibility study. Eur Radiol. 2009; 19(5):1246-52.

- Mamisch TC, Trattnig S, Quirbach S, Marlovits S, White LM, Welsch GH. Quantitative T2 mapping of knee cartilage: differentiation of healthy control cartilage and cartilage repair tissue in the knee with unloading—initial results. Radiology. 2010;254(3):818-26.
- Shimomura K, Hamada H, Hart DA, Ando W, Nishii T, Trattnig S, *et al.* Histological analysis of cartilage defects repaired with an autologous human stem cell construct 48 weeks postimplantation reveals structural details not detected by T2-mapping MRI. Cartilage. Jan 21, 2021. doi: 10.1177/1947603521989423
- Larroza ABV, Moratal D. Texture analysis in magnetic resonance imaging: review and considerations for future applications. In: Constandinides C, ed. Assessment of cellular and organ function and dysfunction using direct and derived MRI methodologies. InTechOpen; 2016.
- Blumenkrantz G, Stahl R, Carballido-Gamio J, Zhao S, Lu Y, Munoz T, *et al*. The feasibility of characterizing the spatial distribution of cartilage T(2) using texture analysis. Osteoarthritis Cartilage. 2008;16(5):584-90.
- Carballido-Gamio J, Link TM, Majumdar S. New techniques for cartilage magnetic resonance imaging relaxation time analysis: texture analysis of flattened cartilage and localized intra- and inter-subject comparisons. Magn Reson Med. 2008;59(6):1472-7.
- 17. Peuna A, Hekkala J, Haapea M, Podlipska J, Guermazi A, Saarakkala S, *et al.* Variable angle gray level co-occurrence matrix analysis of T2 relaxation time maps reveals degenerative changes of cartilage in knee osteoarthritis: Oulu Knee Osteoarthritis Study. J Magn Reson Imaging. 2018; 47(5):1316-27.
- Boutsikou K, Kostopoulos S, Glotsos D, Cavouras D, Lavdas E, Oikonomou G, *et al.* Texture analysis of articular cartilage traumatic changes in the knee calculated from morphological 3.0T MR imaging. Eur J Radiol. 2013;82(8):1266-72.
- Smith GD, Knutsen G, Richardson JB. A clinical review of cartilage repair techniques. J Bone Joint Surg Br. 2005; 87(4):445-9.
- Bugbee WD, Convery FR. Osteochondral allograft transplantation. Clin Sports Med. 1999;18(1):67-75.
- Brittberg M, Lindahl A, Nilsson A, Ohlsson C, Isaksson O, Peterson L. Treatment of deep cartilage defects in the knee with autologous chondrocyte transplantation. N Engl J Med. 1994;331(14):889-95.
- Steadman JR, Rodkey WG, Rodrigo JJ. Microfracture: surgical technique and rehabilitation to treat chondral defects. Clin Orthop Relat Res. 2001;(391 Suppl):S362-S369.
- Hangody L, Rathonyi GK, Duska Z, Vasarhelyi G, Fules P, Modis L. Autologous osteochondral mosaicplasty. Surgical technique. J Bone Joint Surg Am. 2004;86-A(Suppl 1): 65-72.
- Behrens P, Bitter T, Kurz B, Russlies M. Matrix-associated autologous chondrocyte transplantation/implantation (MACT/ MACI)—5-year follow-up. Knee. 2006;13(3):194-202.
- Marcacci M, Berruto M, Brocchetta D, Delcogliano A, Ghinelli D, Gobbi A, *et al.* Articular cartilage engineering with hyalograft C: 3-year clinical results. Clin Orthop Relat Res. 2005;(435):96-105.

- Dorotka R, Bindreiter U, Macfelda K, Windberger U, Nehrer S. Marrow stimulation and chondrocyte transplantation using a collagen matrix for cartilage repair. Osteoarthritis Cartilage. 2005;13(8):655-64.
- Panagopoulos A, van Niekerk L, Triantafillopoulos I. Autologous chondrocyte implantation for knee cartilage injuries: moderate functional outcome and performance in patients with high-impact activities. Orthopedics. 2012;35(1): e6-e14.
- Depeursinge A, Foncubierta-Rodriguez A, De Ville DV, Muller H. Three-dimensional solid texture analysis in biomedical imaging: review and opportunities. Med Image Anal. 2014;18(1):176-96.
- Larroza A, Lopez-Lereu MP, Monmeneu JV, Gavara J, Chorro FJ, Bodi V, *et al.* Texture analysis of cardiac cine magnetic resonance imaging to detect nonviable segments in patients with chronic myocardial infarction. Med Phys. 2018;45(4):1471-80.
- Stehling C, Baum T, Mueller-Hoecker C, Liebl H, Carballido-Gamio J, Joseph GB, *et al.* A novel fast knee cartilage segmentation technique for T2 measurements at MR imaging—data from the Osteoarthritis Initiative. Osteoarthritis Cartilage. 2011;19(8):984-9.
- 31. Joseph GB, Baum T, Carballido-Gamio J, Nardo L, Virayavanich W, Alizai H, *et al.* Texture analysis of cartilage T2 maps: individuals with risk factors for OA have higher and more heterogeneous knee cartilage MR T2 compared to normal controls—data from the osteoarthritis initiative. Arthritis Res Ther. 2011;13(5):R153.
- 32. Mayerhoefer ME, Welsch GH, Riegler G, Mamisch TC, Materka A, Weber M, *et al.* Feasibility of texture analysis for the assessment of biochemical changes in meniscal tissue on T1 maps calculated from delayed gadolinium-enhanced magnetic resonance imaging of cartilage data: comparison with conventional relaxation time measurements. Invest Radiol. 2010;45(9):543-7.
- Abragam A. Principles of nuclear magnetism. Oxford University Press; 2004.
- Uppuluri A. GLCM_Features4.m: vectorized version of GLCM_Features1.m [with code changes]. Available from: https://www.mathworks.com/matlabcentral/fileexchange/22354 -glcm_features4-m-vectorized-version-of-glcm_features1-m -with-code-changes
- Janacova V, Juras V, Szomolanyi P, Trattnig S. Optimization of knee cartilage texture analysis from quantitative MRI T2 maps. Osteoarthritis Cartilage. 2021;29(Suppl 1):S329-S330.
- 36. Carballido-Gamio J, Joseph GB, Lynch JA, Link TM, Majumdar S. Longitudinal analysis of MRI T2 knee cartilage laminar organization in a subset of patients from the osteoarthritis initiative: a texture approach. Magn Reson Med. 2011;65(4):1184-94.
- 37. Heilmeier U, Wamba JM, Joseph GB, Darakananda K, Callan J, Neumann J, et al. Baseline knee joint effusion and medial femoral bone marrow edema, in addition to MRI-based T2 relaxation time and texture measurements of knee cartilage, can help predict incident total knee arthroplasty 4-7 years later: data from the Osteoarthritis Initiative. Skeletal Radiol. 2019;48(1):89-101.

- Chanchek N, Gersing AS, Schwaiger BJ, Nevitt MC, Neumann J, Joseph GB, *et al.* Association of diabetes mellitus and biochemical knee cartilage composition assessed by T2 relaxation time measurements: data from the osteoarthritis initiative. J Magn Reson Imaging. 2018;47(2):380-90.
- Welsch GH, Mamisch TC, Quirbach S, Zak L, Marlovits S, Trattnig S. Evaluation and comparison of cartilage repair tissue of the patella and medial femoral condyle by using morphological MRI and biochemical zonal T2 mapping. Eur Radiol. 2009;19(5):1253-62.
- 40. Domayer SE, Apprich S, Stelzeneder D, Hirschfeld C, Sokolowski M, Kronnerwetter C, *et al.* Cartilage repair of the ankle: first results of T2 mapping at 7.0 T after microfracture and matrix associated autologous cartilage transplantation. Osteoarthritis Cartilage. 2012;20(8):829-36.
- 41. Welsch GH, Trattnig S, Domayer S, Marlovits S, White LM, Mamisch TC. Multimodal approach in the use of clinical scoring, morphological MRI and biochemical T2-mapping and diffusion-weighted imaging in their ability to assess differences between cartilage repair tissue after microfracture therapy and matrix-associated autologous chondrocyte transplantation: a pilot study. Osteoarthritis Cartilage. 2009;17(9):1219-27.
- DiBartola AC, Everhart JS, Magnussen RA, Carey JL, Brophy RH, Schmitt LC, *et al.* Correlation between histological outcome and surgical cartilage repair technique in the knee: a meta-analysis. Knee. 2016;23(3):344-49.
- Riboh JC, Cvetanovich GL, Cole BJ, Yanke AB. Comparative efficacy of cartilage repair procedures in the knee: a network meta-analysis. Knee Surg Sports Traumatol Arthrosc. 2017;25(12):3786-99.
- 44. Brown WE, Potter HG, Marx RG, Wickiewicz TL, Warren RF. Magnetic resonance imaging appearance of cartilage repair in the knee. Clin Orthop Relat Res. 2004;(422):214-23.
- 45. Niemeyer P, Feucht MJ, Fritz J, Albrecht D, Spahn G, Angele P. Cartilage repair surgery for full-thickness defects of the knee in Germany: indications and epidemiological data from the German Cartilage Registry (KnorpelRegister DGOU). Arch Orthop Trauma Surg. 2016;136(7):891-7.
- Devitt BM, Bell SW, Webster KE, Feller JA, Whitehead TS. Surgical treatments of cartilage defects of the knee: systematic review of randomised controlled trials. Knee. 2017;24(3):508-17.
- Jones KJ, Kelley BV, Arshi A, McAllister DR, Fabricant PD. Comparative effectiveness of cartilage repair with respect to the minimal clinically important difference. Am J Sports Med. 2019;47(13):3284-93.
- Paatela T, Vasara A, Nurmi H, Kautiainen H, Jurvelin JS, Kiviranta I. Biomechanical changes of repair tissue after autologous chondrocyte implantation at long-term follow-up. Cartilage. Epub May 23, 2020. doi:10.1177/1947603520 921433
- Mahmoud-Ghoneim D, Toussaint G, Constans JM, de Certaines JD. Three dimensional texture analysis in MRI: a preliminary evaluation in gliomas. Magn Reson Imaging. 2003;21(9):983-7.
- Zhang J, Yu C, Jiang G, Liu W, Tong L. 3D texture analysis on MRI images of Alzheimer's disease. Brain Imaging Behav. 2012;6(1):61-9.