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Association between *APOE*, *SCARB1*, *PPARα* polymorphisms and serum lipids in a population of Lithuanian adults

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Abstract

Background: Dyslipidemia is one of several known risk factors for coronary heart disease, a leading cause of death in Lithuania. Blood lipid levels are influenced by multiple genetic and environmental factors. Epidemiological studies demonstrated the impact of nutrition on lipid levels within the Lithuanian population although the role of genetic factors for dyslipidemias has not yet been studied. The objective of this study was to assess the distribution of the *APOE*, *SCARB1*, *PPARα* genotypes in the Lithuanian adult population and to determine the relationship of these genotypes with dyslipidemia.

Methods: A cross-sectional health survey was carried out in a representative random sample of the Lithuanian population aged 25–64 (n=1030). A variety of single-nucleotide polymorphisms (SNPs) of the *APOE* (rs429358 and rs7412), *SCARB1* (rs5888) and *PPARα* (rs1800206) genes were assessed using real-time polymerase chain reaction. Serum lipids were determined using enzymatic methods.

Results/Principal findings: Men and women with the *APOE2* genotype had the lowest level of total and low-density lipoprotein cholesterol (LDL-C). Men with the *APOE2* genotype had significantly higher levels of triglycerides (TG) than those with the *APOE3* genotype. In men, the carriers of the *APOE4* genotype had higher odds ratios (OR) of reduced (<1.0 mmol/L) high density lipoprotein cholesterol (HDL-C) levels versus *APOE3* carriers (OR=1.98; 95% CI=1.05-3.74). The odds of having elevated (>1.7 mmol/L) TG levels was significantly lower in *SCARB1* genotype CT carriers compared to men with the *SCARB1* genotype CC (OR=0.50; 95% CI=0.31-0.79). In men, carriers of the *PPARα* genotype CG had higher OR of elevated TG levels versus carriers of *PPARα* genotype CC (OR=2.67; 95% CI=1.15-6.16). The odds of having high LDL-C levels were lower in women with the *APOE2* genotype as compared to *APOE3* genotype carriers (OR=0.35; 95% CI=0.22-0.57).

Conclusions/Significance: Our data suggest a gender difference in the associations between *APOE*, *SCARB1*, *PPARα* genotypes and lipid levels. In men, the *APOE4* genotype and *PPARα* genotype CG were correlated with an atherogenic lipid profile while the *SCARB1* genotype CT had an atheroprotective effect. In women, *APOE2* carriers had the lowest odds of high LDL-C.

Keywords: Apolipoprotein E (*APOE*) genotype, Scavenger receptor class B type 1 (*SCARB1*) genotype, Peroxisome proliferator-activated receptor-alpha (*PPARα*) genotype, Dyslipidemia

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Background

A high prevalence of cardiovascular diseases is a major public health problem in Lithuania. In 2010, the age-standardized mortality from coronary heart disease (CHD) for Lithuanian men aged 25–64 years was 198.2 per 100 000 population and for women 44.0 per 100 000 population, while the average rate in the European Union was 102.0 per 100 000 population for men and 24.8 per 100 000 population for women of comparable age [1]. Dyslipidemia is one of several known risk factors for CHD. Blood lipid levels are influenced by multiple genetic and environmental factors [2].

Diet is the most important environmental determinant of lipid levels. Epidemiological studies have demonstrated some positive changes in the diet of the Lithuanian population over the last decades. The use of animal fats has declined, while the use of vegetable fats and the frequency of consumption of fresh vegetables have increased [3]. Consequently, the proportion of saturated fatty acids in the daily energy intake has decreased and the proportion of polyunsaturated fatty acids has increased [4]. Positive changes in diet have contributed to a decline in serum cholesterol levels; however, the mortality rate from CHD has remained high.

Several genes were found to be associated with lipid levels and the risk of cardiovascular diseases. Apolipoprotein E (APOE) is a glycoprotein that plays a fundamental role in lipid metabolism. APOE participates in the clearance of chylomicron remnants and very low-density lipoprotein (VLDL) by serving as a ligand for LDL receptors [5]. The APOE isoforms are coded by three alleles ($\epsilon 2$, $\epsilon 3$ and $\epsilon 4$) resulting in six different genotypes ($\epsilon 2/2$, $\epsilon 2/3$, $\epsilon 2/4$, $\epsilon 3/3$, $\epsilon 3/4$, and $\epsilon 4/4$) [6]. Many studies assessing the role of APOE polymorphism on plasma lipids have shown that the presence of the $\epsilon 4$ allele is associated with elevations in LDL-C, while the presence of $\epsilon 2$ is associated with decreased levels [7,8].

The human Class B Type I Scavenger Receptor (SCARB1) protein is a hepatic receptor with large affinity for HDL-C [9]. After HDL-C binds to SCARB1 in the liver, cholesterol is transferred to the cells to be released in the bile. Some prior studies of the effect of SCARB1 genotypes on HDL-C have shown conflicting results. Studies among diabetics [10], CHD patients [11,12] and community-based populations [13] have reported that the SCARB1 exon 8 rs5888 (C>T) polymorphism is associated with HDL-C. However, results of other studies have been inconsistent [14,15].

Peroxisome proliferator-activated receptor- α (PPAR α) is a member of the nuclear hormone receptor superfamily of ligand-inducible transcription factors and regulates the expression of genes involved in peroxisomal and mitochondrial β -oxidation pathways such as fatty acid uptake and the catabolism of circulating triglycerides

(TG) [16]. PPAR α is encoded by the PPAR α gene located on chromosome 22q12-q13.1 [17]. It is expressed mainly in tissues demonstrating high capacity for fatty acid oxidation such as liver, kidney, heart, and skeletal muscle [18,19]. In studies of healthy subjects a relationship between the PPAR α L162V polymorphism and serum concentrations of TG [20,21] and LDL-C [22] has been suggested.

There is a lack of data regarding the role of genetic factors for lipid levels and cardiovascular risk in Lithuania. Our aim was therefore to assess the distribution of the APOE, SCARB1, PPAR α genotypes in the Lithuanian adult population and to determine the relationship of these genotypes with dyslipidemia.

Results

The characteristics of study subjects according to gender are given in Table 1. An association between lipid levels and gender was observed: men had lower levels of HDL-C and higher levels of TG compared to women. The prevalence of dyslipidemias was higher in men, except for a reduction in HDL, which was similar in both men and women. No significant differences in the frequencies of the APOE, SCARB1 and PPAR α genotypes or alleles between men and women were observed (Table 2). Frequencies of genotypes did not differ significantly from those predicted by a Hardy-Weinberg equilibrium.

Table 1 Characteristics of the study population

Characteristics	Men	Women	p value
Age, years	47.6 (0.51)	46.2 (0.43)	0.045
BMI, kg/m ² †	27.5 (0.26)	28.1 (0.22)	n.s.
TC, mmol/L ‡	5.32 (0.05)	5.30 (0.04)	n.s.
LDL-C, mmol/L ‡	3.40 (0.05)	3.21 (0.04)	0.002
HDL-C, mmol/L ‡	1.29 (0.02)	1.43 (0.02)	<0.001
LnTG, mmol/L ‡	0.39 (0.02)	0.31 (0.02)	0.001
High TC [#] , n (%)	253 (59.6)	278 (46.3)	<0.001
High LDL-C [#] , n (%)	265 (62.4)	279 (46.5)	<0.001
Reduced HDL [#] , n (%)	105 (24.7)	138 (22.9)	0.430
Elevated TG [#] , n (%)	140 (33.0)	136 (22.7)	<0.001
Obesity [#] , n (%)	105 (24.8)	151 (25.2)	n.s.
Diabetes ^{§,¶} , n (%)	7 (1.7)	9 (1.5)	n.s.
Arterial hypertension [#] , n (%)	252 (59.4)	206 (34.3)	<0.001
Coronary heart disease [#] , n (%)	36 (8.6)	35 (5.9)	0.027

† Data adjusted by age.

‡ Data adjusted by age and BMI.

Data weighted by age.

Data are presented as mean (standard error) unless otherwise indicated.

[§]Physician-diagnosed diabetes.

High TC level ≥ 5 mmol/L; high LDL-C level ≥ 3 mmol/L; reduced HDL-C level - for men < 1.0 mmol/L and for women < 1.2 mmol/L; elevated TG level > 1.7 mmol/L.

Abbreviations: BMI body mass index, HDL-C high density lipoprotein cholesterol, LDL-C low density lipoprotein cholesterol, LnTG logarithmically transformed triglycerides, TC total cholesterol.

Table 2 Distribution of the APOE, SCARB1, PPARα genotypes and frequency of the alleles (%) in the study population

Characteristics	Men n (%)	Women n (%)	p value
<i>APOE</i> ,			
ε 2/2 genotype	2 (0.5)	2 (0.3)	n.s.
ε 3/2 genotype	76 (17.9)	94 (15.7)	
ε 3/3 genotype	270 (63.6)	359 (59.7)	
ε 4/2 genotype	13 (3.0)	21 (3.5)	
ε 4/3 genotype	61 (14.5)	116 (19.3)	
ε 4/4 genotype	2 (0.5)	9 (1.5)	
ε3 allele	79.8	77.2	n.s.
ε2 allele	11.0	9.9	
ε4 allele	9.2	12.9	
<i>SCARB1</i> ,			
CC genotype	149 (35.2)	215 (35.8)	n.s.
CT genotype	212 (50.0)	287 (47.8)	
TT genotype	63 (14.8)	99 (16.4)	
C allele	58.9	58.4	n.s.
T allele	41.1	41.6	
<i>PPARα</i> ,			
CC genotype	399 (94.2)	568 (94.5)	n.s.
CG genotype	25 (5.8)	32 (5.3)	
GG genotype	0	1(0.2)	
C allele	97.1	97.3	n.s.
G allele	2.90	2.70	

Abbreviations: APOE apolipoprotein E, PPARα Peroxisome proliferator-activated receptor-alpha, SCARB1 Scavenger Receptor Class B Type 1.

The fasting serum lipid levels of the study population with respect to the APOE, SCARB1, PPARα genotypes are shown in Table 3. Men and women with the APOE2 genotype had the lowest level of TC and LDL-C. Mean values of TC and LDL-C were the highest in men and women with the APOE4 genotype; however, they did not differ significantly from the mean values of those lipids in the APOE3 genotype carriers. Men with the APOE2 genotype had significantly higher levels of TG than those with the APOE3 genotype. There were no significant differences in the levels of TG in women and in the levels of HDL-C in either gender among different genotype groups.

In men, the SCARB1 TT genotype carriers had a significantly higher levels of LDL-C than men with the SCARB1 genotype CC (Table 3). Moreover, men with the SCARB1 CT genotype had lower levels of TG in comparison to CC genotype carriers. No significant differences in the levels of HDL-C between different SCARB1 genotype groups were found in men. Serum lipid levels were not associated with SCARB1 genotypes in women.

Table 3 The fasting serum lipid levels in mmol/L (mean and standard error) according to the APOE, SCARB1, PPARα genotypes

The genotypes	TC Mean (SE)	LDL-C Mean (SE)	HDL-C Mean (SE)	LnTG Mean (SE)
MEN				
<i>APOE</i>				
APOE3	5.38 (0.06)	3.49 (0.06)	1.30 (0.03)	0.35 (0.02)
APOE2	5.03 (0.12)*	3.00 (0.11)*	1.24 (0.05)	0.47 (0.04)*
APOE4	5.56 (0.13)**	3.60 (0.12)**	1.34 (0.05)	0.44 (0.05)
<i>SCARB 1</i>				
CC	5.30 (0.09)	3.29 (0.08)	1.26 (0.04)	0.45 (0.03)
CT	5.29 (0.07)	3.40 (0.07)	1.32 (0.03)	0.33 (0.03) #
TT	5.56 (0.13)	3.66 (0.12) #	1.27 (0.05)	0.39 (0.05)
<i>PPARα</i>				
CC	5.35 (0.05)	3.42 (0.05)	1.30 (0.02)	0.37 (0.02)
CG	5.31 (0.21)	3.33 (0.19)	1.29 (0.09)	0.58 (0.08) # #
WOMEN				
<i>APOE</i>				
APOE3	5.34 (0.05)	3.31 (0.05)	1.42 (0.02)	0.31 (0.02)
APOE2	5.04 (0.10)*	2.74 (0.09)*	1.48 (0.04)	0.30 (0.04)
APOE4	5.39 (0.09)**	3.35 (0.08)**	1.45 (0.03)	0.30 (0.03)
<i>SCARB 1</i>				
CC	5.31 (0.07)	3.27 (0.06)	1.42 (0.03)	0.30 (0.03)
CT	5.29 (0.06)	3.23 (0.05)	1.42 (0.02)	0.32 (0.02)
TT	5.25 (0.10)	3.09 (0.09)	1.49 (0.04)	0.29 (0.04)
<i>PPARα</i>				
CC	5.28 (0.04)	3.21 (0.04)	1.42 (0.03)	0.31 (0.02)
CG	5.54 (0.18)	3.43 (0.16)	1.41 (0.02)	0.36 (0.07)

Data adjusted by age and body mass index.

APOE2 - carriers of the ε 2/2 and ε 2/3 genotype, APOE3 - carriers of the ε 3/3 genotype, and APOE4 - carriers of the ε 3/4 and ε 4/4 genotype.

* p <0.05 compared with APOE3 genotype; ** p <0.05 compared with APOE2 genotype; # p <0.05 compared with SCARB1 genotype CC; ## p <0.05 compared with PPARα genotype CC.

Abbreviations: APOE apolipoprotein E, HDL-C high density lipoprotein cholesterol, LDL-C low density lipoprotein cholesterol, LnTG logarithmically transformed triglycerides, PPARα Peroxisome proliferator-activated receptor-alpha, SE Standard Error, SCARB1 Scavenger Receptor Class B Type 1, TC total cholesterol.

Men with the PPARα genotype CG had higher levels of triglycerides than did CC genotype carriers (Table 3). In women, no significant differences in serum lipid levels were found between different PPARα genotypes.

Multivariate logistic regression analysis was performed to identify the associations of the APOE, SCARB1, PPARα genotypes with the prevalence of dyslipidemias in the study population (Table 4). After adjustment for age, body mass index, physical activity and alcohol consumption, the odds of having high TC and high LDL-C levels were lower in women with the APOE2 genotype compared to APOE3 genotype carriers (respectively OR=0.52; p=0.006 and OR=0.35; p<0.001). In men, the

Table 4 Odds ratios for the likelihood of dyslipidemias according to the *APOE*, *SCARB1*, *PPARα* genotypes in the study population

The genotypes	High TC OR (95% CI)	High LDL-C OR (95% CI)	Reduced HDL-C OR (95% CI)	Elevated TG OR (95% CI)
MEN				
<i>APOE</i>				
<i>APOE3</i>	1	1	1	1
<i>APOE2</i>	0.60 (0.33-1.02)	0.60 (0.35-1.02)	1.56 (0.85-2.85)	1.36 (0.79-2.34)
<i>APOE4</i>	1.04 (0.58-1.88)	1.37 (0.74-2.54)	1.98 (1.05-3.74)	1.50 (0.84-2.67)
<i>SCARB 1</i>				
CC	1	1	1	1
CT	1.20 (0.77-1.84)	1.09 (0.69-1.72)	0.76 (0.45-1.26)	0.50 (0.31-0.79)
TT	1.67 (0.89-3.23)	1.91 (0.98-3.73)	0.75 (0.36-1.58)	0.66 (0.35-1.26)
<i>PPARα</i>				
CC	1	1	1	1
CG	1.12 (0.47-2.66)	0.79 (0.34-1.84)	0.52 (0.15-1.84)	2.67 (1.15-6.16)
WOMEN				
<i>APOE</i>				
<i>APOE3</i>	1	1	1	1
<i>APOE2</i>	0.52 (0.32-0.83)	0.35 (0.22-0.57)	0.62 (0.34-1.11)	0.97 (0.57-1.66)
<i>APOE4</i>	0.84 (0.54-1.30)	0.96 (0.62-1.49)	0.78 (0.48-1.29)	1.10 (0.69-1.76)
<i>SCARB 1</i>				
CC	1	1	1	1
CT	1.18 (0.80-1.74)	1.14 (0.78-1.68)	1.43 (0.92-2.23)	1.16 (0.76-1.77)
TT	1.39 (0.83-2.33)	0.83 (0.50-1.37)	0.82 (0.43-1.53)	1.03 (0.58-1.80)
<i>PPARα</i>				
CC	1	1	1	1
CG	1.71 (0.77-3.80)	1.95 (0.86-4.43)	1.23 (0.54-2.84)	0.82 (0.34-1.96)

Data adjusted by age, body mass index, physical activity, and alcohol consumption.

APOE2 - carriers of the ϵ 2/2 and ϵ 2/3 genotype, *APOE3* - carriers of the ϵ 3/3 genotype, and *APOE4* - carriers of the ϵ 3/4 and ϵ 4/4 genotype.

Abbreviations: *APOE* apolipoprotein E, *Elevated TG* triglycerides level >1.7 mmol/L, *High TC* total cholesterol \geq 5.0 mmol/L, *High LDL-C* low density lipoprotein cholesterol \geq 3.0 mmol/L, *PPARα* Peroxisome proliferator-activated receptor-alpha, *Reduced HDL-C* high density lipoprotein cholesterol level for men <1.0 mmol/L and for women <1.20 mmol/L, *SCARB 1* Scavenger Receptor Class B Type 1.

odds of having reduced HDL-C levels was statistically higher in *APOE4* genotype carriers than in those with the *APOE3* genotype (OR=1.98; p=0.035). The associations of the *SCARB1* and *PPARα* genotypes with dyslipidemia were found only in men (Table 4). The odds of having elevated TG levels was statistically lower in the *SCARB1* genotype CT carriers as compared to men with the *SCARB1* genotype CC (OR=0.50; p=0.003). In men, carriers of the *PPARα* genotype CG had higher OR of elevated TG levels versus carriers of the *PPARα* genotype CC (OR=2.67; p=0.022).

Discussion

The overall pattern of the allele frequency distribution analyzed in the Lithuanian population was similar to other European populations. Nevertheless, some differences in *APOE* allele frequencies were observed between Northern

European countries and Lithuania. Namely, a lower prevalence of the *APOE* ϵ 2 allele and a higher prevalence of the *APOE* ϵ 4 allele was found in Finland (4% and 20% respectively) [23], Denmark (8% and 17% respectively) [24] and Sweden (7% and 17% respectively) [25], as compared to the Lithuanian population. Findings from 45 populations around the world have shown that the frequency of the *APOE* ϵ 4 allele appears to be higher in northern regions of Europe (the Nordic countries, Scotland, Germany, and the Netherlands) than in southern regions (Switzerland, Tyrol, France, Italy, and Spain) [26]. The distribution of the *APOE* allele in Lithuania was more comparable to Southern European populations. The studies demonstrated a lower risk of CHD in ϵ 2 carriers and a higher risk in ϵ 4 carriers as compared to ϵ 3 homozygotes [8,27,28]. Bearing in mind the facts above, only a modest risk of CHD should be expected in Lithuania, although the mortality from CHD

in the population of Lithuania is one of the highest in Europe [1].

Many studies suggested that variation of the *APOE* gene was associated with variations of lipid levels [7,8,24,25,27]. In line with the findings of other researchers, we found that *APOE2* genotype carriers had the lowest levels of TC and LDL-C. The mechanism of this effect probably involves decreased conversion of VLDL into LDL observed in $\epsilon 2$ carriers [5,6]. More efficient catabolism of chylomicrons and VLDL-remnants, increased intestinal cholesterol absorption, and reduced LDL-receptor activity was shown in $\epsilon 4$ carriers resulting in higher levels of TC and LDL-C when compared to $\epsilon 2$ and $\epsilon 3$ allele carriers. Thus, a high frequency of the $\epsilon 2$ allele would appear to predict a favorable lipid profile in the Lithuanian population. Moreover, positive changes in nutritional habits of Lithuanians occurred over the last decades of the post-communist transition period and has contributed to the decrease in mean TC and LDL-C levels [3,4]. However, the prevalence of other cardiovascular disease risk factors, such as hypertension, smoking and obesity, remains very high in Lithuania [29]. It is obvious, that the risk of cardiovascular disease is determined by multiple genetic and environmental factors, and the impact of one gene is very limited. On the other hand, men with the *APOE2* genotype had higher levels of TG than those with the *APOE3* genotype. It was revealed that *APOE2* binds poorly to lipoprotein receptors leading to accumulation of chylomicron and VLDL-remnants [6]. The higher TG levels in *APOE2* genotype carriers may attenuate the cardioprotective effects of lower LDL-C [24]. The *APOE* genotype also affected HDL-C level. Our data showed the higher odds of reduced HDL-C levels in men with the *APOE4* genotype as compared to those with the *APOE3* genotype. Other studies also found lower HDL-C in $\epsilon 4$ allele carriers [7,24,25].

Much less is known regarding the role of *SCARB1* in humans, although several studies have reported that the SNP of *SCARB1* (rs5888) is associated with the lipid profile and the development of CHD [9,12,13]. The frequencies of the *SCARB1* alleles in our population were similar to frequencies that have been reported for French and North American populations [30]. Our data indicated that in men the carriers of the *SCARB1* genotype CT had two-fold lower odds of high TG levels than the carriers of the genotype CC. The tendency of lower odds of high TG levels was shown also for the *SCARB1* genotype TT carriers. Insufficient numbers of individuals with the *SCARB1* genotype TT in the study population may probably explain why the odds did not reach statistical significance. Men with the *SCARB1* genotype TT had a higher level of LDL-C as compared to *SCARB1* genotype CC carriers. In contrast to our findings,

Morabia A et al. found higher LDL-C in women with the T allele [9]. We did not observe any association between the *SCARB1* genotype and dyslipidemia in women. Several studies have shown that a rare T variant was associated with an atheroprotective lipid profile in men but not in women [9,13]. A previous study carried out in Lithuania revealed that men aged 65–74 who were carriers of the TT genotype had a higher level of HDL-C and a lower risk of myocardial infarction as compared to the CC genotype [31]. Gender and age-specific effects on blood lipid profiles were also demonstrated by other investigators [12,13,15]. Those studies highlighted a protective T allele effect on HDL-C. *SCARB1* was identified as a physiologically important HDL receptor; however, it is also a multiligand receptor participating in the metabolism of other plasma lipoproteins [14]. The mechanism of the *SCARB1* SNP rs5888 effect on the lipid profile remains not finally determined because polymorphisms in this SNP does not lead to a change in the amino acid sequence of the *SCARB1* protein [9,14].

The prevalence of the *PPAR α* G allele in our population was lower as compared to the prevalence that has been reported in other studies [20,22,32]. The results from a Quebec study indicated that carriers of the G allele may be at increased risk of abdominal obesity, hypertriglyceridemia, and low HDL-C levels [20]. The Framingham Offspring Study found that the G allele was associated with increased plasma concentrations of TC, LDL-C, and apoB [22]. In our study, the *PPAR α* CG genotypes were associated with elevated TG levels only in men. It is known that *PPAR α* stimulates proteins of the transport and binding of fatty acids. Moreover, it regulates genes involved in fatty acid oxidation [16,19]. Taking this in mind it is reasonable to believe that the functional L162V polymorphism located in the DNA-binding domain of the *PPAR α* gene may have an effect on lipid metabolism, particularly on TG levels [33].

Study strength and limitations

A major strength of our study is an investigation of a nationally representative random sample of the Lithuanian population, allowing assessment of the frequencies of alleles related to lipid levels. However, this study had some limitations. The cross-sectional study design did not allow us to consider the effects of all possible genetic and environmental factors and their interactions on lipid levels. Another issue involves the unhealthy diet of the majority of the Lithuanian population. A high intake of fats, especially saturated fats, might influence lipid levels and have an important interaction with the genotypes under study. Furthermore, we did not have data about hormone replacement therapy in postmenopausal

women which might also affect the association between genotypes and lipid levels.

Conclusions

Our data suggested a gender difference of the associations between *APOE*, *SCARB1*, *PPAR α* genotypes and lipid levels. In men, the *APOE4* genotype and the *PPAR α* genotype CG were correlated with an atherogenic lipid profile while the *SCARB1* genotype CT had an atheroprotective effect on lipid levels. In women, *APOE2* carriers had the lowest odds of high LDL-C. Further studies, particularly evaluating genetic-environmental interactions on lipid levels, are necessary to confirm our established associations.

Materials and methods

Study design and sample

The cross-sectional health survey was carried out in five randomly selected municipalities of Lithuania. The random sample was obtained from lists of inhabitants aged 25–64. Health examinations were conducted for 1739 participants (58% of the eligible sample). From these, 1030 individuals (425 men and 605 women) had the *APOE*, *SCARB1* and *PPAR α* genotypes determined. We excluded 5 subjects taking lipid lowering medications from our analysis (1 man and 4 women). Also we excluded 34 subjects (13 men and 21 women) with the rare *APOE* ϵ 2/4 genotype from the analysis of associations between the *APOE* genotypes and lipid levels.

APOE, *SCARB1* and *PPAR α* genotyping

For DNA extraction, blood samples were collected from each individual in ethylenediaminetetraacetic (EDTA) tubes during their health examination. DNA was extracted from peripheral blood leukocytes using a reagent kit (NucleoSpin Blood L Kit; Macherey & Nagel, Düren, Germany).

Two single-nucleotide polymorphisms (SNPs) of *APOE* gene (rs429358 and rs7412) were assessed using commercially available genotyping kit C__3084793_20 and C__904973_10. SNP of *SCARB1* gene (rs5888) was assessed using commercially available genotyping kit C_7497008_1_ and SNP of *PPAR α* gene (rs1800206) was assessed using commercially available genotyping kit C__8817670_20 (Applied Biosystems, Foster City, CA, USA). We used the Applied Biosystems 7900HT Real-Time Polymerase Chain Reaction System for detecting the SNPs. The cycling program started with heating at 95°C for 10 min, followed by 40 cycles (at 95°C for 15 s and at 60°C for 1 min). Allelic discrimination was carried out using the software of Applied Biosystems.

Three *APOE* genotype groups were analyzed in this study: *APOE2* (carriers of the ϵ 2/2 and ϵ 2/3 genotype),

APOE3 (carriers of the ϵ 3/3 genotype), and *APOE4* (carriers of the ϵ 3/4 and ϵ 4/4 genotypes).

Laboratory analyses and anthropometric measurements

Blood samples for lipids measurements were taken in the morning after fasting at least 12 hours. TC, LDL-C, HDL-C, and TG levels were determined by an automatic analyzer using conventional enzymatic methods. All laboratory analyses were made in the same certified laboratory. Quality control measures were followed for estimation of lipid levels. Dyslipidemias were defined using the following criteria: high TC level - serum TC \geq 5 mmol/L; high LDL-C level - serum LDL-C \geq 3 mmol/L; reduced HDL-C level - serum HDL-C for men $<$ 1.0 mmol/L and for women $<$ 1.2 mmol/L; elevated TG level - TG $>$ 1.7 mmol/L [34].

The height of participants wearing no shoes was measured to the nearest centimeter with a stadiometer. The body weight of participants wearing light indoor clothing and no shoes was measured to the nearest 0.1 kg with standardized medical scales. Body mass index (BMI) was calculated as weight divided by height squared (kg/m^2). Obesity was defined as BMI \geq 30 kg/m^2 .

Assessment of physical activity and alcohol consumption

Information on physical activity and alcohol consumption was gathered by a standard questionnaire. The number of hours spent per week for physical activity at work, travelling to and from work and at leisure time was calculated. The questionnaire on alcohol consumption contained questions about a type and frequency of alcohol consumption and amount of alcohol consumed at one occasion. The amount of alcohol consumed at one occasion was recalculated into standard alcohol units (SAUs) using the following formula: SAU = amount (in liters) \times strength of alcoholic drink (beer – 5%, wine – 12%, strong alcohol – 40%). One SAU equals to 10 g of ethanol. Then the amount of SAUs consumed during a month was calculated.

Statistical analysis

The data were analyzed with statistical software package SPSS version 19.0 for Windows. All data were analyzed in each gender separately. Continuous variables were presented as mean values and standard error (SE). The normality of distribution of continuous variables was tested by Kolmogorov - Smirnov test. Only distribution of triglyceride levels was skewed, and this variable was logarithmically transformed to improve normality for statistical testing. Analysis of variance with Bonferroni multiple comparison tests were used to compare the means of continuous variables across groups.

The mean values of TC, LDL-C, HDL-C and TG levels by the *APOE*, *SCARB1*, *PPAR α* genotypes were calculated

using general linear model and controlling for age, BMI, physical activity and alcohol consumption.

The effects of the *APOE*, *SCARB1*, *PPARα* genotypes on the prevalence of dyslipidemias were evaluated using logistic regression analysis. All models were applied separately for men and women. Each genotype was included into a separate model. The odds of dyslipidemias were calculated with adjustment for age, body mass index, physical activity and alcohol consumption.

Ethics statement

The study protocol was approved by the Lithuanian Bioethics Committee. Written informed consent for participation in the study was obtained from all participants.

Abbreviations

TC: Total cholesterol; LDL-C: Low density lipoprotein cholesterol; TG: Triglycerides; HDL-C: High density lipoprotein cholesterol; CHD: Coronary heart disease; APOE: Apolipoprotein E; VLDL: Very low-density lipoprotein; SCARB1: Scavenger receptor class B type 1; PPARα: Peroxisome proliferator-activated receptor-α; SNPs: Single nucleotide polymorphisms; BMI: Body mass index; SAUs: Standard alcohol units; SE: Standard error; OR: Odds ratio; CI: Confidence intervals.

Competing interests

None of the authors has any proprietary interests or conflicts of interest related to this submission. This submission has not been published anywhere previously, and it is not simultaneously being considered for any other publication.

Authors' contributions

AS participated in the design of the study, has made analysis and interpretation of data; drafted the manuscript. JP participated in the design of the study, has made analysis and interpretation of data; revised manuscript critically for important intellectual content, has given final approval of the version to be published. DL has made analysis and interpretation of data; helped to draft the manuscript. KJ has made analysis and interpretation of data. JK has made substantial contributions to conception and design, coordinated study, participated in acquisition of data, revised manuscript critically for important intellectual content. VL has made substantial contributions to conception and design, revised manuscript critically for important intellectual content, has given final approval of the version to be published. All authors read and approved the final manuscript.

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