



Mycoplasma pneumoniae 23S rRNA A2063G mutation does not influence chest radiography features in children with pneumonia

Huan Deng, Jun Rui, Deyu Zhao and Feng Liu

Abstract

Objective: To measure the rate of the A2063G mutation in the *Mycoplasma pneumoniae* (*M. pneumoniae*) 23S rRNA domain V in children with pneumonia and to determine the correlation between radiographic findings and the presence of the A2063G mutation.

Methods: Patients who were hospitalized with a confirmed diagnosis of *M. pneumoniae* pneumonia were enrolled in this study. *M. pneumoniae* strains were collected for genotype analysis. Chest radiography was performed on all children prior to and following macrolide treatment. Clinical and imaging data were obtained.

Results: Of 211 patients, 195 (92.42%) harboured *M. pneumoniae* with the A2063G mutation. No significant differences were identified in inflammation score, chest radiography inflammation absorption grade before and after macrolide treatment, or pulmonary complications (atelectasis, hydrothorax, or pleuritis) prior to macrolide treatment when children were stratified based on the presence or absence of the A2063G mutation.

Conclusions: A high proportion of children with pneumonia harboured strains of *M. pneumoniae* with the A2063G mutation in the 23S rRNA domain V. However, no obvious chest radiographic features of *M. pneumoniae* pneumonia were associated with the A2063G variant.

Keywords

Mycoplasma pneumoniae pneumonia, A2063G mutation, chest radiography, children

Date received: 21 September 2016; accepted: 30 May 2017

Introduction

Mycoplasma pneumoniae (*M. pneumoniae*) is a common agent of community-acquired pneumonia in children and young adults.¹ As *M. pneumoniae* has no cell wall, it is intrinsically resistant to β -lactams and other antibiotics that target the cell wall.

Department of Respiratory Medicine, Children's Hospital of Nanjing Medical University, Nanjing, Jiangsu Province, China

Corresponding authors:

Deyu Zhao and Feng Liu, Department of Respiratory Medicine, Children's Hospital of Nanjing Medical University, 72 Guangzhou Road, Nanjing, Jiangsu 210008, Jiangsu Province, China.

Emails: zhaodeyu98@126.com; axslu@163.com



Therefore, macrolides, especially 14- and 15-membered ring macrolides, are used as first-line agents in children to avoid the potential age-related side-effects of other therapies; tetracyclines have possible adverse effects on enamel hypoplasia and bone, and quinolones may influence the growth of bone and articular cartilage.² However, recently increasing numbers of infections are leading to refractory or life-threatening pneumonia with pulmonary and/or extrapulmonary complications.³⁻⁵ Multiple factors including *M. pneumoniae* load,⁶ macrolide-resistance,⁷ systemic inflammatory response^{2,8} and mixed infection may underlie the development of severe *M. pneumoniae* pneumonia in patients.

Macrolide antibiotics are known to inhibit protein synthesis by binding to domain II and/or domain V of the *M. pneumoniae* 23S rRNA.^{9,10} In turn, the primary mechanism of macrolide-resistance comprises mutations in target genes.⁹⁻¹¹ In particular, point mutations at nucleotide positions A2063, A2064, A2067 and C2617 in domain V have been identified as common mutations.^{10,12} *M. pneumoniae*, with an A-to-G or A-to-C transition at position 2063 of the 23S rRNA gene results in high resistance to 14- and 15-membered ring macrolides, whereas mutations at positions A2067 and C2617 confer lower levels of resistance.^{2,12} The presence of *M. pneumoniae* mutants has been reported to be mainly associated with persistent clinical symptoms such as fever, resulting in prolonged hospital stay.^{3,13} To the best of our knowledge, few studies have investigated the correlation between radiographic findings and gene mutations in *M. pneumoniae* pneumonia in children.

The most common radiographic findings in *M. pneumoniae* pneumonia consist of unilateral or bilateral areas of air-space consolidation and ground-glass opacities. However, these findings are variable and can include reticular or nodular opacities.¹⁴ The current study examined the prevalence

of the A2063G mutation in the *M. pneumoniae* 23S rRNA domain V in children with pneumonia and the correlation between radiographic findings and the presence of this mutation.

Patients and methods

Study population

Patients were enrolled from the Department of Respiratory Medicine, Children's Hospital of Nanjing Medical University Nanjing, Jiangsu Province, China between 1 March 2014 and 31 May 2015. Patients were included in the study if they fulfilled the following three criteria:¹⁴ (i) diagnosis of pneumonia based on symptoms at admission, including fever, cough, productive sputum, chest pain, dyspnoea, abnormal breathing sounds, and a new infiltrate on chest radiography that was at least segmental; (ii) acute *M. pneumoniae* infection was confirmed by polymerase chain reaction (PCR) and/or serology (detecting *M. pneumoniae* immunoglobulin M by enzyme-linked immunosorbent assay in the acute phase or *M. pneumoniae* antibody titres increased ≥ 4 times in the recovery phase); (iii) the absence of other pathogens (bacteria, respiratory syncytial virus, influenza virus, adenovirus, parainfluenza virus, *Chlamydia pneumoniae*, *Coxiella burnetii*, tuberculosis and *Legionella pneumophila*) and the ineffectiveness of penicillin, cephalosporin, and sulfonamide. The following patients were excluded: convalescent patients, patients with immunosuppressive illness, asthma, chronic lung disease, or other systematic disease. Chest radiography was undertaken both before and after macrolide treatment. Demographic data and chest radiography findings were noted.

The study protocol was approved by the Ethics Committee of the Children's Hospital of Nanjing Medical University and written informed consent was obtained from at least one parent/guardian of each patient included.

Study methods

Nasopharyngeal aspirates were collected at admission and assayed for *M. pneumoniae* DNA copy number using the *M. pneumoniae* DNA PCR kit (ACON Biotech Co., Ltd, Hangzhou, China). The 23S rRNA domain V was amplified by nested PCR as described previously.¹⁵ Amplification products were then shipped at 4°C within 2 days for sequencing at Yingweijie, Shanghai, China. Finally, DNA sequences were compared using BLAST® to the *M. pneumoniae* strain M129 (ATCC 29342).¹⁶ Patients were treated with 10 mg/kg per day azithromycin by oral administration or intravenous infusion for the first 3 days, after which patients did not receive treatment for the next 4 days. Finally, patients received a second 3-day treatment with 10 mg/kg per day azithromycin or with 30 mg/kg per day erythromycin by intravenous infusion. The course of macrolide treatment was approximately 1–3 weeks before the clinical symptoms improved and radiographic inflammation was absorbed. Tetracyclines and quinolones were not permitted to avoid possible adverse reactions. According to clinical manifestations, some drugs were selectively used, such as expectorants, cough medicine, inhaled corticosteroid, bronchodilators and immune modulators (corticosteroid and/or intravenous immunoglobulin). Clinical data

were collected from medical records including sex, age, febrile days prior to macrolide treatment, immune modulator use, and chest radiography results prior to and after macrolide treatment.

Chest radiography scoring

Chest radiography findings were scored using the following two questionnaires: (i) the evaluation questionnaire of pneumonia inflammation, which is a recognized evaluation questionnaire for pneumonia chest radiography findings (Tables 1 and 2);¹⁷ and (ii) the absorption questionnaire of pneumonia inflammation. Four grades were obtained by using ‘grade’ evaluation and ‘total score’ evaluation methods. When the two evaluation methods were not consistent, the final absorption decision comprised the optimum result of the two evaluation methods. The two methods utilized the questionnaires as shown in Table 2.¹⁸ Two senior radiologists blinded to the clinical details of each case independently read and assigned scores for each chest radiograph.

Statistical analyses

All statistical analyses were performed using the IBM SPSS® statistical package, version

Table 1. Pneumonia radiographic inflammation evaluation questionnaire.¹⁷

Part	Score		Standard
	Left	Right	
Lung			0: no pneumonia (0)
Upper			I: slightly heavier or increased of lung markings (1)
Middle			II: (mild): lung markings heavier, indistinct of lung markings, or fibrous stripes (2)
Lower			III: (moderate): sheet infiltration, increased density (3)
			IV: (severe): patchy infiltration, high density (4)
			V: (very severe): large infiltration, high density (5)
Hydrothorax			No (0); little (blunting costophrenic angle) (1); lot (2)
Pleuritis			No (0); yes (2)
Interstitial change			No (0); yes (ground-glass or reticular opacity) (2)
Total			

Table 2. Pneumonia radiological inflammatory absorption questionnaire.¹⁸

Final absorption decision	Grade evaluation	Total score evaluation
Complete absorption	Drop from V-I to 0	≥95%
Mostly absorption	Drop from V, IV, or III to II, I	≥70%, <95%
Partial absorption	Drop from V to IV, III; drop from IV to III; drop from II to I	≥30%, <70%
Non-absorption	No downgrade or progression	<30%

Grade evaluation was taken as the highest grade in the six lung fields. Total score evaluation was calculated as the difference between radiographic inflammation scores prior to and following treatment/radiographic inflammation scores prior to treatment $\times 100\%$.

20.0 (IBM, Armonk, NY, USA) for Windows®. Data are presented as mean \pm SD. For data with a skewed distribution, median values and range are reported. Student's *t*-test was used to compare continuous variables, whereas χ^2 -test and Fisher's exact test were used for categorical variables. The linear correlation was determined between two variables. A *P*-value < 0.01 was considered statistically significant.

Results

A total of 211 children were enrolled in the current study, of whom 108 (51.18%) were male and 103 (48.82%) were female. The median age was 4.8 years. Chest radiography was performed on all children prior to and after macrolide treatment.

Regarding the *M. pneumoniae* 23S rRNA genotypes, of the 211 study patients, 195 (92.42%) harboured the A2063G mutation in the *M. pneumoniae* 23S rRNA domain V, whereas the remaining 16 (7.58%) patients had no mutation. The children were stratified into two groups based on the presence or absence of the A2063G mutation. The mean age among the children infected with mutated *M. pneumoniae* was 4.789 years (98 males, 97 females) and was 4.974 years in the remaining patients (10 males, six females) infected with non-mutated strains. No significant differences were identified in age or sex distribution between the two groups.

When considering the clinical characteristics of the children based on their *M. pneumoniae* 23S rRNA genotypes, there was no significant difference in febrile days prior to macrolide treatment (Table 3). Moreover, the use of immune modulators was similar between the two groups. The scores of radiographic inflammation prior to and after macrolide treatment were comparable between the mutated and non-mutated groups, as was the inflammation score difference prior and subsequent to treatment between the two groups.

Radiographic inflammatory absorption was classified into four grades on the basis of the radiological inflammatory absorption questionnaire. No significant differences were observed in the mutated and non-mutated groups between each grade (Table 4). Furthermore, pulmonary complications did not differ between the mutated and non-mutated groups (Table 5).

On the basis of immune modulator use, 159 patients (84 males, 75 females) were not administered immune modulators, while 52 (24 males, 28 females) were administered immune modulators. There was no significant difference in sex and age distribution between the two groups (data not shown). Patients treated with immune modulators had significantly higher radiography inflammation scores before and after macrolide treatment compared with patients who were not treated with immune modulators ($P < 0.001$, $P = 0.002$, respectively)

Table 3. Clinical characteristics of children infected with mutated and non-mutated *Mycoplasma pneumoniae* strains.

Variable	Mutated n = 195	Non-mutated n = 16
Febrile days prior to macrolide treatment	2.559 ± 1.396	2.688 ± 1.537
Immune modulators (with/without)	49/146	3/13
Radiographic inflammation score prior to macrolide treatment	10.446 ± 2.936	10.188 ± 2.713
Radiographic inflammation score after macrolide treatment	6.246 ± 2.895	6.063 ± 2.235
Inflammation score difference prior to and after macrolide treatment	4.200 ± 2.875	4.125 ± 2.605

Data presented as mean ± SD or n of patients.

No significant between-group differences (P -value ≥ 0.01); Student's t -test was used to compare continuous variables, χ^2 -test and Fisher's exact test were used for categorical variables.

Table 4. Radiographic inflammatory absorption grade prior to and after treatment in children infected with mutated and non-mutated *Mycoplasma pneumoniae* strains.

Final absorption decision	Mutated n = 195	Non-mutated n = 16
Complete absorption	1	0
Mostly absorption	75	4
Partial absorption	76	8
Non-absorption	43	4

Data presented as n of patients.

No significant between-group differences (P -value ≥ 0.01); χ^2 -test.

(Table 6). Moreover, there were no linear relationships between radiography inflammation score prior to macrolide treatment and age or between radiography inflammation score prior to macrolide treatment and febrile days prior to macrolide treatment. For the two groups identified by immune modulator use, the chest radiography inflammation score after macrolide treatment had no linear relationship with age and febrile days prior to macrolide treatment.

Discussion

Mycoplasma pneumoniae represents a common agent of acute respiratory

Table 5. Radiographic pulmonary complications in children infected with mutated and non-mutated *Mycoplasma pneumoniae* strains.

Variable	Mutated n = 195	Non-mutated n = 16
Atelectasis (yes/no)	8/187	2/14
Pleuritis (yes/no)	9/186	1/15
Hydrothorax (yes/no)	14/181	1/15

Data presented as n of patients.

No significant between-group differences (P -value ≥ 0.01); χ^2 -test.

infections, exhibiting various clinical manifestations and involving multiple systems. Although *M. pneumoniae* infection is considered self-limiting, an increasing number of recent cases have progressed to severe, life-threatening pneumonia.^{1,3} In addition, the extensive use of macrolides as first-line therapeutics for children has fuelled worldwide macrolide resistance, especially in East Asia.⁶ It is currently believed that genetic mutations in the domains targeted by macrolides result in the resistance and numerous studies have focused on the associations between clinical characteristics and the genetic mutations.^{3,10,14} However, to the best of our knowledge, few reports have explored the chest radiographic findings of

Table 6. Score of radiographic inflammation in children infected with mutated and non-mutated *Mycoplasma pneumoniae* strains stratified according to the use of immune modulators.

Variable	Without immune modulator n = 159	With immune modulator n = 52	Statistical significance ^a
Radiography inflammation score prior to macrolide treatment	9.952 + 2.704	11.846 + 3.121	$P < 0.001$
Radiography inflammation score after macrolide treatment	5.892 + 2.578	7.308 + 3.359	$P = 0.002$
Inflammation score difference prior to and after macrolide treatment	4.069 + 2.726	4.539 + 3.214	NS

Data presented as mean \pm SD.

^aStudent's t-test.

NS, no significant between-group difference (P -value ≥ 0.01).

M. pneumoniae pneumonia upon infection by the A2063G mutated strain.

Mycoplasma pneumoniae pneumonia exhibits diverse chest radiographic manifestations that have been classified into four types: (a) sheet or patchy infiltration similar to that of lobular pneumonia; (b) interstitial changes similar to viral pneumonia; (c) segmental or lobar infiltration similar to bacterial pneumonia; and (d) hilar node enlargement.^{13,18} Therefore, it is difficult to distinguish the pneumonia resulting from infection by *M. pneumoniae* from that arising from other pathogens. In the present study, there were no statistical differences between the two groups stratified by the presence or absence of the A2063G mutation in the chest radiological inflammation scores prior to and after macrolide treatment, the difference in these scores, the inflammatory absorption grade prior to and following treatment, or in chest radiological pulmonary complications prior to treatment. As a consequence, it was difficult to determine from the chest radiographs alone whether the infecting *M. pneumoniae* strain carried the A2063G mutation or to judge whether the pathogen was resistant to macrolide. The rate of immune modulator use was similar between the mutated (49/195 [25.13%]) and non-mutated groups

(3/16 [18.75%]). The children treated with immune modulators had significantly higher chest radiography inflammation scores prior to macrolide treatment than those not treated with immune modulators. It might be that immune modulators reduce effusion and promote the absorption of pulmonary inflammation.

The pathogenic mechanism of *M. pneumoniae* is considered to include adhesion to respiratory epithelial cells, resistance to clearing and phagocytosis, cytotoxic effects caused by oxidative stress, community-acquired respiratory distress syndrome toxin, and *M. pneumoniae*-derived lipopeptides.^{19–22} Although increasing evidence suggests that the host immune response represents an important cause of inflammatory injury to the lung and other systems,⁸ these pathogenic mechanisms are clearly irrelevant as macrolide targets, which might explain the lack of an observed relationship between chest radiography inflammation scores prior to treatment and A2063G mutation. However, no significant differences were observed in the chest radiographic inflammatory absorption grade prior to and after macrolide treatment between patients carrying mutated or non-mutated strains, or in the difference of inflammation scores after treatment.

A possible explanation might be due to the host immune response after *M. pneumoniae* infection whereby the macrolide might have exerted an immunomodulatory function that provided additional benefits of the treatment.

In conclusion, the infecting *M. pneumoniae* strain in 195 of 211 (92.42%) patients in the current study was found to harbour the A2063G mutation in the 23S rRNA domain V. The A2063G mutation was determined to have no effect on the chest radiographic inflammation scores prior to and after macrolide treatment, their difference, the inflammatory absorption grade before and after macrolide treatment, or in the chest radiographic pulmonary complications occurring prior to treatment. This present study confirms that in the clinic, early chest radiographs will likely contribute to the diagnosis of *M. pneumoniae* pneumonia, but will not help to determine if the infected *M. pneumoniae* strain has the A2063G mutation. Immune modulators should be used early in severe pulmonary inflammation because they effectively alleviate the inflammatory reaction. In addition, these current findings support the continued use of macrolides as the first-line treatment of choice for paediatric patients with *M. pneumoniae* pneumonia owing to their potential immunomodulatory function and the age-related side-effects of other therapies.

Author contributions

Huan Deng and Jun Rui contributed equally to the work as the first authors. Huan Deng and Jun Rui participated in the design of the study, sample preparation, gene amplification and sequence alignment. Huan Deng wrote and submitted the manuscript. Feng Liu conceived of the study, participated in its design and helped to draft the manuscript. Deyu Zhao participated in the design of the study. All authors read and approved the final manuscript.

Acknowledgements

The authors wish to thank everyone who helped with this study.

Declaration of conflicting interests

The authors declare that there are no conflicts of interest.

Funding

This study was supported by grants from the National Natural Science Foundation of China (no. 81370132; Deyu Zhao) and the Nanjing Medical Science and Technique Development Foundation, Nanjing, China (Feng Liu).

References

1. Lee KY. Pediatric respiratory infections by *Mycoplasma pneumoniae*. *Expert Rev Anti Infect Ther* 2008; 6: 509–521.
2. Waites KB and Talkington DF. *Mycoplasma pneumoniae* and its role as a human pathogen. *Clin Microbiol Rev* 2004; 17: 697–728.
3. Zhou Y, Zhang Y, Sheng Y, et al. More complications occur in macrolide-resistant than in macrolide-sensitive *Mycoplasma pneumoniae* pneumonia. *Antimicrob Agents Chemother* 2014; 58: 1034–1038.
4. Azumagawa K, Kambara Y, Murata T, et al. Four cases of arthritis associated with *Mycoplasma pneumoniae* infection. *Pediatr Int* 2008; 50: 511–513.
5. Hawkins S, Rausch CM and McCanta AC. Constrictive pericarditis secondary to infection with *Mycoplasma pneumoniae*. *Curr Opin Pediatr* 2011; 23: 126–129.
6. Nilsson AC, Björkman P, Welinder-Olsson C, et al. Clinical severity of *Mycoplasma pneumoniae* (MP) infection is associated with bacterial load in oropharyngeal secretions but not with MP genotype. *BMC Infect Dis* 2010; 10: 39.
7. Kurata S, Taguchi H, Sasaki T, et al. Antimicrobial and immunomodulatory effect of clarithromycin on macrolide-resistant *Mycoplasma pneumoniae*. *J Med Microbiol* 2010; 59(Pt 6): 693–701.

8. Shimizu T, Kida Y and Kuwano K. Cytoadherence-dependent induction of inflammatory responses by Mycoplasma pneumoniae. *Immunology* 2011; 133: 51–61.
9. Douthwaite S, Hansen LH and Mauvais P. Macrolide-ketolide inhibition of MLS-resistant ribosomes is improved by alternative drug interaction with domain II of 23S rRNA. *Mol Microbiol* 2000; 36: 183–193.
10. Bébéar CM and Pereyre S. Mechanisms of drug resistance in Mycoplasma pneumoniae. *Curr Drug Targets Infect Disord* 2005; 5: 263–271.
11. Kawai Y, Miyashita N, Kubo M, et al. Therapeutic efficacy of macrolides, minocycline, and tosufloxacin against macrolide-resistant Mycoplasma pneumoniae pneumonia in pediatric patients. *Antimicrob Agents Chemother* 2013; 57: 2252–2258.
12. Morozumi M, Hasegawa K, Kobayashi R, et al. Emergence of macrolide-resistant Mycoplasma pneumoniae with a 23S rRNA gene mutation. *Antimicrob Agents Chemother* 2005; 49: 2302–2306.
13. Ma Z, Zheng Y, Deng J, et al. Characterization of macrolide resistance of Mycoplasma pneumoniae in children in Shenzhen, China. *Pediatr Pulmonol* 2014; 49: 695–700.
14. Guo Q, Li HY, Zhou YP, et al. Associations of radiological features in Mycoplasma pneumoniae pneumonia. *Arch Med Sci* 2014; 10: 725–732.
15. Matsuoka M, Narita M, Okazaki N, et al. Characterization and molecular analysis of macrolide-resistant Mycoplasma pneumoniae clinical isolates obtained in Japan. *Antimicrob Agents Chemother* 2004; 48: 4624–4630.
16. National Center for Biotechnology Information, U.S. National Library of Medicine. <https://blast.ncbi.nlm.nih.gov/Blast.cgi>.
17. Yin R, Li M, Xu H, et al. Preliminary development and application of evaluation questionnaire of pneumonia inflammation (China). *Chinese Journal of Respiratory and Critical Care Medicine* 2012; 11: 185–187.
18. Tanaka H. Correlation between radiological and pathological findings in patients with Mycoplasma pneumoniae pneumonia. *Front Microbiol* 2016; 7: 695.
19. Kannan TR, Provenzano D, Wright JR, et al. Identification and characterization of human surfactant protein A binding protein of Mycoplasma pneumoniae. *Infect Immun* 2005; 73: 2828–2834.
20. Kannan TR, Musatovova O, Balasubramanian S, et al. Mycoplasma pneumoniae community acquired respiratory distress syndrome toxin expression reveals growth phase and infection-dependent regulation. *Mol Microbiol* 2010; 76: 1127–1141.
21. Browning GF, Marendra MS, Noormohammadi AH, et al. The central role of lipoproteins in the pathogenesis of mycoplasmoses. *Vet Microbiol* 2011; 153: 44–50.
22. Bose S, Segovia JA, Somarajan SR, et al. ADP-ribosylation of NLRP3 by Mycoplasma pneumoniae CARDS toxin regulates inflammasome activity. *MBio* 2014; 5: e02186–e02214.