



Hypoxia and the Tumor Microenvironment

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Abstract

Hypoxia is an important feature of the tumor microenvironment, and is closely associated with cell proliferation, angiogenesis, metabolism and the tumor immune response. All these factors can further promote tumor progression, increase tumor aggressiveness, enhance tumor metastatic potential and lead to poor prognosis. In this review, these effects of hypoxia on tumor biology will be discussed, along with their significance for tumor detection and treatment.

Keywords

hypoxia, tumor microenvironment, angiogenesis, metabolism, hypoxia-targeted therapy

Abbreviations

ATP, adenosine triphosphate; CA9, carbonic anhydrase IX; CAFs, cancer-associated fibroblasts; ECM, extracellular matrix; Ecs, endothelial cells; ¹⁸F-EF5, ¹⁸F-2-(2-nitro-1H-imidazol-1-yl)-N-(2,2,3,3,3-pentafluoropropyl)-acetamide; ¹⁸F-FAZA, ¹⁸F-fluoroazomycin- β -arabinofuranoside; ¹⁸F-FDG, ¹⁸F-fluorodeoxyglucose; ¹⁸F-FMISO, ¹⁸F-misonidazole; GLUT-1, glucose transporter-1; HAPs, hypoxia-activated prodrugs; HIF-1 α , hypoxia inducible factor-1 α ; ³H-FLT, ³H-fluorothymidine; ¹³¹I-IAZGP, ¹³¹I-labeled iodoazomycin galactopyranoside; MDSCs, myeloid-derived suppressor cells; PET, positron emission tomography; TAMs, tumor-associated macrophages; TME, tumor microenvironment; VEGF, vascular endothelial growth factor

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Introduction

Hypoxia is a common and important feature of solid malignancies. Tissue is generally classed as hypoxic if the oxygen tension falls below 10mmHg, compared to 40-60mmHg in most normal tissue.¹ In tumors, hypoxia arises due to a mismatch between oxygen delivery and consumption. Hypoxia is closely associated with tumor progression, increased aggressiveness, enhanced metastatic potential, resistance to radio- or chemo-therapy and lower overall survival of various tumor types.²⁻⁴ Hypoxia also has the ability to facilitate the selection of apoptosis-resistant cell clones and induce tumor metastasis.⁵

Hypoxia may be either acute or chronic. Acute hypoxia is perfusion-related and occurs through transient opening and closing of tumor vessels. Chronic hypoxia is diffusion-related, as the oxygen demand associated with tumor cell proliferation creates a maximum distance beyond which oxygen cannot diffuse.⁶⁻⁸ Chronic hypoxia is long-term or possibly irreversible, and generally occurs when the tumor cells are distant from functional vessels.

Tumor cells response to hypoxia depends in part on the duration of exposure. Hypoxic tumor cells may undergo

necrosis or reoxygenation, and during these hypoxia-reoxygenation cycles, they may acquire a more aggressive phenotype.⁹

Tumor Microenvironment (TME)

Solid malignancies consist of transformed cells embedded in a mass of connective tissue or mesenchyme, which forms the tumor microenvironment, and includes vascular endothelial

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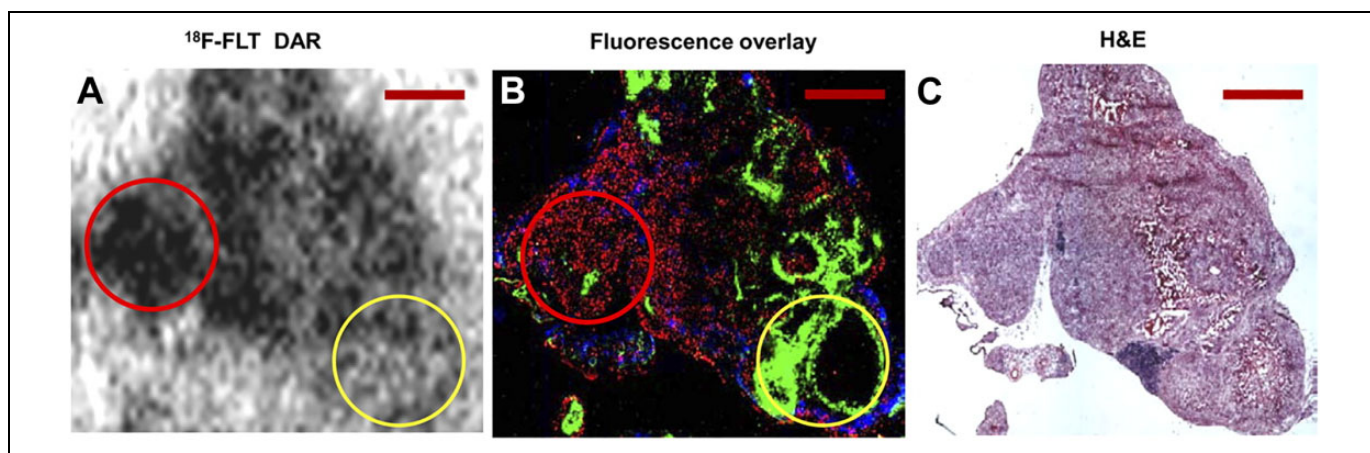


Figure 1. Relationship between hypoxia and cell proliferation. Images derived from a xenograft of HTB177, human lung cancer cell line: (A) ^{18}F -FLT autoradiography images; (B) A fluorescence overlay showing the hypoxic marker pimonidazole (green); proliferation marker bromodeoxyuridine (red); and blood perfusion marker Hoechst 33342 (blue); (C): H&E staining. Red circle: Well oxygenated (pimonidazole-negative) area is co-localized with high cell proliferation (by high levels of ^{18}F -FLT uptake and bromodeoxyuridine binding); Yellow circle: Hypoxic (pimonidazole-positive) area is co-localized with low cell proliferation (by lower ^{18}F -FLT uptake and bromodeoxyuridine binding). This research was originally published in JNM.²⁸ Huang T, Civelek AC, Li J, *et al*. Tumor microenvironment-dependent ^{18}F -FDG, ^{18}F -fluorothymidine, and ^{18}F -misonidazole uptake: a pilot study in mouse models of human non-small cell lung cancer. *J Nucl Med.* 53, 1262-8 (2012) © SNMMI.

cells (ECs), cancer-associated fibroblasts (CAFs), immune cells and extracellular matrix (ECM).¹⁰ Both parenchyma and the mesenchyme are morphologically, phenotypically, and functionally heterogeneous at multiple levels: between patients, between primary tumors and their metastases, and even within a single tumor.^{11,12} As tumor cells proliferate, they recruit nearby non-tumor cells into collaborative processes to foster a specific tumor microenvironment conducive for local growth and metastasis to distant organs. This collection of tumor cells interacting with host cells forms an abnormal organ-like structure.¹³ It is now evident that tumor microenvironment supports tumor growth, causes heterogeneity and contributes to the resistance to systemic therapies.¹⁴

Hypoxia and Proliferation

Oxygen is required for efficient ATP production, and hence hypoxia is unsurprisingly linked to reduced cell proliferation. In the classical Pasteur effect,¹⁵ while yeast growth is reduced under anaerobic conditions, their rate of sugar metabolism is increased. In tumors, as demonstrated by Waki *et al*, well oxygenated cells, which are closely adjacent to functional blood vessels, have a higher proliferation rate, while hypoxic cancer cells, generally located 100-200 μm away from functional blood vessels and/or close to necrotic areas, have a relatively low proliferation rate.¹⁶ Several other studies have also revealed the mutually exclusive pattern between cell proliferation and hypoxia in both microscopic (avascular) and macroscopic (vascularized) tumors.¹⁷⁻²⁰ Further, tumors that are hypovascular may appear dormant, with cells either dividing very slowly or resting in G_0 ,²¹⁻²⁴ though it has also been found that dormancy can result from high proliferation being offset by apoptosis, preventing growth beyond a threshold size.^{25,26}

The picture of an anti-correlation between hypoxia and proliferation has also been confirmed with radiolabeled tracers. Using autoradiography, Fatema *et al* showed that ^{18}F -misonidazole (^{18}F -FMISO, a hypoxia probe) was distributed mostly in the tumor center, whereas ^3H -Fluorothymidine (^3H -FLT, a proliferation probe) was found in the periphery.²⁷ Others also observed a mismatch in the intratumoral distribution of ^{18}F -FLT and ^{18}F -FMISO in lung cancers²⁸⁻³¹ (Figure 1).

Hypoxia and Angiogenesis

Angiogenesis is the process through which new vessels are formed, mainly from existing vessels by means of sprouting.³² ECs lining the luminal surface of blood vessels are normally quiescent but during development, wound healing, and disease, can be activated to enter the angiogenic process.³³

Tumors initially form in a pre-vascular phase: when the tumor diameter is less than 1 mm, there is no vessel formation, and cells mainly depend on permeation to acquire nutrients. As the tumor grows, vessels develop and cells acquire nutrients through angiogenesis. If angiogenesis fails, the absence of oxygen or other nutrients, restricts tumor size to 2-3mm.³⁴⁻³⁶ Studies by Li *et al* have shown that the hypoxic status is closely associated with blood perfusion (Figure 2). In a peritoneal metastatic colorectal cancer animal model, they found that microscopic tumors (< 1 mm in diameter) were avascular and extremely hypoxic, while in those of greater size (1-4 mm in diameter), vessels had developed, and hypoxia was minimal^{19,20} (Figure 3). As tumor volume increased, the rapid expansion of tumor cells exceeded the local blood supply, and the inefficient tumor vascular system could not provide enough oxygen and other nutrients, resulting in hypoxia in macroscopic tumors.

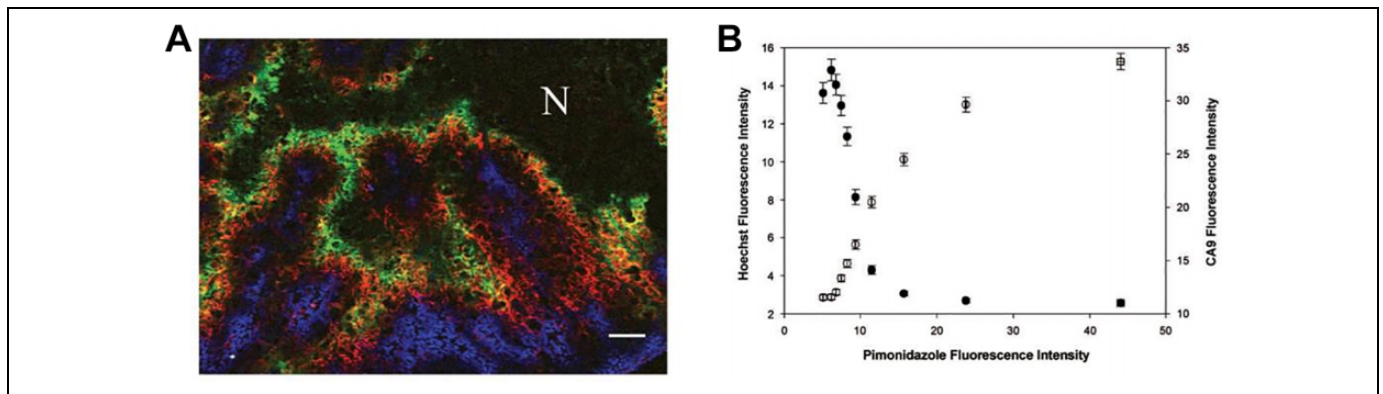


Figure 2. Hypoxia correlates with low blood perfusion. Images derived from a xenograft of HT29, a human colorectal carcinoma cell line. A, A fluorescence overlay, showing the hypoxic markers pimonidazole (green) and CA9 (red); and blood perfusion marker Hoechst 33342 (blue). Necrotic tissue is marked “N.” B, Relationship between hypoxia and blood perfusion. The scattergram represents the decile-binned mean and 95% CIs (confidence interval) of paired fluorescence intensity values acquired on an 8-Ar square pixel-by-pixel basis. ●: Hoechst 33342 versus pimonidazole; ○: CA9 versus pimonidazole. Reproduced with permission from *Cancer Res.*¹⁹ Li XF, Carlin S, Urano M, Russell J, Ling CC, O’Donoghue JA. Visualization of hypoxia in microscopic tumors by immunofluorescent microscopy. *Cancer Res.* 67, 7646-7653 (2007).

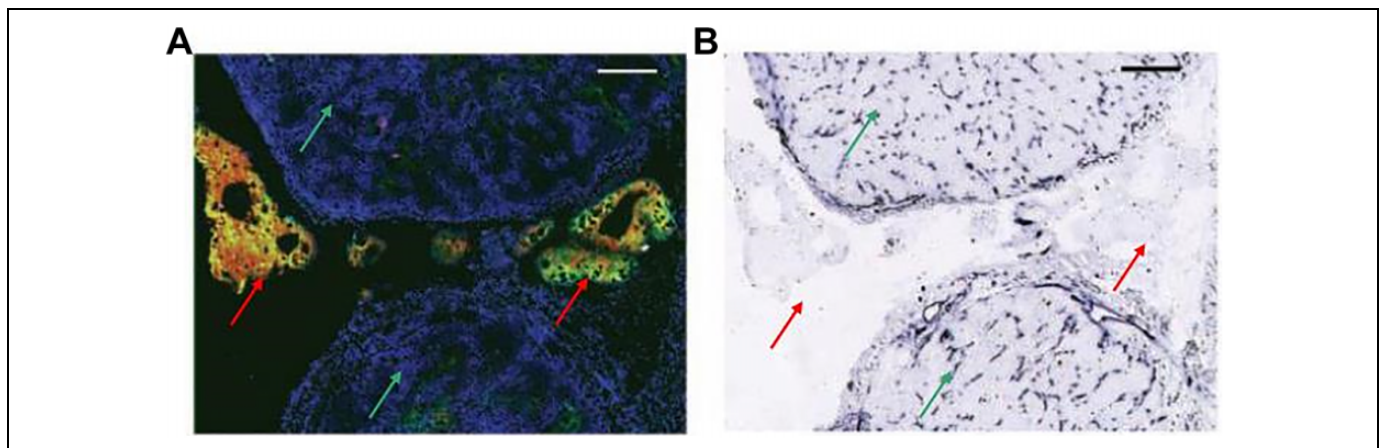


Figure 3. Relationship between hypoxia and angiogenesis. Images derived from a xenograft of HT29, a human colorectal carcinoma cell line. A, A fluorescence overlay, showing the hypoxic markers pimonidazole (green) and CA9 (red); and blood perfusion marker Hoechst 33342 (blue). B, Immunohistochemistry staining for CD31 (blood vessel marker). Hypoxic (high pimonidazole and CA9 binding) areas are avascular with little to no blood perfusion (red arrow), while well oxygenated (little to no pimonidazole or CA9 binding) areas have vasculature with extensive blood perfusion (green arrow). Reproduced with permission from *Cancer Res.*¹⁹ Li XF, Carlin S, Urano M, Russell J, Ling CC, O’Donoghue JA. Visualization of hypoxia in microscopic tumors by immunofluorescent microscopy. *Cancer Res.* 67, 7646-7653 (2007).

Angiogenesis is activated in response to hypoxia via the HIF1 pathway, and the consequent production of vascular endothelial growth factor (VEGF) both tumor and stromal cells. Tumor-activated ECs have distinctive gene expression profiles and luminal cell-surface receptors, resulting in abnormalities in tumor vasculature.³⁷ This abnormal vascular network is characterized by leaky and compressed blood and lymphatic vessels leads to further regional hypoxia, creating a vicious cycle. As a result, tumor cells move through periods of hypoxia-reoxygenation that produce a more aggressive phenotype.³⁸

Hypoxia and Metabolism

Low extracellular pH is another feature of solid tumors and hypoxia leads to acidosis in the tumor microenvironment.^{39,40} Most cancers are characterized by a high rate of glycolysis, as low oxygen levels force tumor cells to shift from oxidative phosphorylation to glycolysis, with a resultant production of metabolic acids.⁴¹⁻⁴³ Like increased angiogenesis, these changes are mediated by hypoxia-inducible factor 1 α (HIF-1 α) driven transcription, which leads to upregulation of both glucose transporters and genes involved in glycolysis.⁴⁴ Li *et al* have observed a strong association between hypoxia, glucose

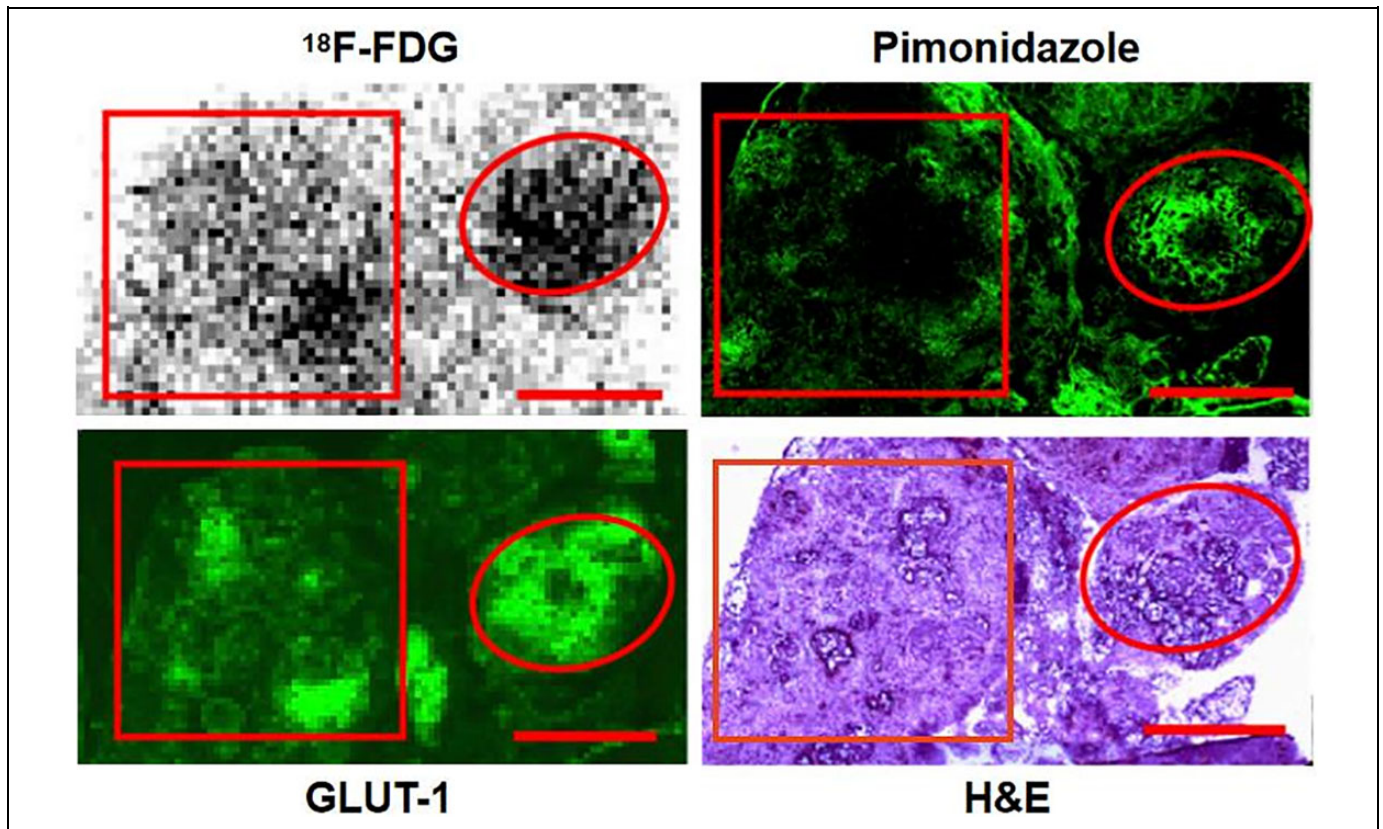


Figure 4. Relationship between glucose metabolism, hypoxia and cell proliferation. Circle: Hypoxic (high pimonidazole binding) area is co-localized with high ^{18}F -FDG uptake and high GLUT-1 expression. Square: Relatively non-hypoxic (lower pimonidazole binding) area is co-localized with lower ^{18}F -FDG uptake and lower GLUT-1 expression. This research was originally published in *JNM*.⁴³ Li XF, Ma Y, Sun X, Humm JL, Ling CC, O'Donoghue JA. High ^{18}F -FDG uptake in microscopic peritoneal tumors requires physiologic hypoxia. *J Nucl Med.* 51, 632-38 (2010). © SNMMI.

transporter-1 (GLUT-1) and ^{18}F -fluorodeoxyglucose (^{18}F -FDG) uptake^{43,45} (Figure 4).

The mechanistic/mammalian target of rapamycin (mTOR) pathway is a central signaling pathway that controls metabolic processes. Multiple points along the glycolytic pathway are influenced by mTOR via regulation of critical transcription factors such as HIF-1 α .⁴⁶ The expression of HIF-1 α is dependent on mTORC1 and mTORC2.⁴⁷ Both mTORC1 and mTORC2 play important roles in glucose uptake and glycolytic metabolism of tumor cells.^{48,49} Thus, application of mTOR inhibitors to control tumor metabolism and tumor microenvironment will become innovative anti-tumor strategy.⁵⁰

Even in the absence of hypoxia, oncogene activation may also promote glycolysis through upregulation of enzymes and cell surface nutrient transporters.³⁹ Therefore, tumor acidity depends on both physiological and genetic factors. Sengupta *et al* revealed a more complex tumor metabolism which has significant heterogeneity. However, they found little correlation between FDG uptake and lactate release.⁵¹ This suggests that tumor cells have metabolic plasticity: They can use a variety of anabolic and catabolic pathways to adapt to energetic needs and availability of nutrients.⁵² Pathways available for

tumor cells to produce adenosine triphosphate (ATP) include oxidative phosphorylation, glutaminolysis, and fatty acid metabolism. Investigating and monitoring tumor metabolism, therefore, can be an essential step leading to a deeper understanding, better diagnostics, and more effective therapeutic strategies for tumor treatment.

Hypoxia and the Immune System

Cancer-associated inflammation is involved in all stages of tumorigenesis and development, which may cause genomic instability, cell proliferation, angiogenesis, enhancement of anti-apoptotic pathways and cancer dissemination.⁵³⁻⁵⁶ Hypoxia can enhance the resistance of tumor cells to immune attack and evade immunosurveillance.⁵⁷ Hypoxia-inducible factors (HIFs), the key mediator of hypoxia signaling, regulate a series of genes that are responsible for tumor immunity under hypoxic condition.⁵⁸

Studies indicate that intratumoral hypoxic regions are infiltrated by high levels of myeloid-derived suppressor cells (MDSCs), tumor-associated macrophages (TAMs), and T-regulatory (Treg) cells.^{57,59} Under long-term exposure to hypoxia, activated HIF-1 α may reduce and inactivate

cytotoxic lymphocytes, including NK cells and CD8⁺ T cells, and result in the inhibition of their anti-tumor efficacy.⁶⁰⁻⁶² Furthermore, increased production of lactate in TME can inactivate cytotoxic lymphocytes, such as NK cells and CD8⁺ T cells, thus resulting in the reduction of their anti-tumor effect.^{63,64} Mahiddine *et al*⁶⁵ found that relief of tumor hypoxia can unleash the tumoricidal potential of neutrophils. Furthermore, there is robust evidence that hypoxia has a strong impact on the induction of HIF-1 α -dependent PD-L1 expression in tumor cells, macrophages, and dendritic cells, which is a crucial mechanism to block antitumor immunity by cancer cells.⁶⁶⁻⁶⁹

Hypoxia increases glycolytic rate of B cells, which reduces proliferation, increases apoptosis, and impairs class-switching. HIF will be activated at key points in the life of B cells, which may shape and control B cell fate and function. Constitutive HIF stabilization via deletion of Vhl reduces GC B cells and impairs affinity maturation, class-switching to the pro-inflammatory IgG2c isotype, recall response and early memory B cell formation. Thus, hypoxia induced over activation of HIF is detrimental to B cell function.^{70,71}

The effect of hypoxia on Dendritic cells (DCs) in the tumor microenvironment has not been well described. Current reports suggest that hypoxia may be beneficial to the ability of DC to induce immune response.^{72,73} Given that the effect of hypoxia was mediated by HIF1 α expression, HIF1 α modulates survival, migration, antigen presentation, interferon synthesis and differentiation of DCs.⁷⁴ HIF1 α has also been found to positively regulate the migratory capability of human monocyte-derived DCs.⁷⁵

Detecting Tumor Hypoxia

Several methods can be used for hypoxia detection:

1. Polarographic oxygen electrodes: the gold standard for hypoxia detection, this is an invasive method which directly measures tissue oxygen pressure. This method is only used for scientific research, and can not be applied to clinical.
2. Positron emission tomography (PET): a widely used imaging technology in clinic, which is a non-invasive, three-dimensional, direct assessment of intratumoral oxygen levels. By employing different radiotracer probes to directly report cell oxygen levels, rather than through hypoxia mediated phenotype changes,⁷⁶ PET has high sensitivity and specificity toward regional hypoxia and also can monitor the whole tumors. However, its spatial resolution is relatively low.⁷⁷⁻⁷⁹

At present, widely used radiotracer probes for hypoxia imaging include 18F-FDG, 18F-FMISO, 18F-Fluoroazomycin-arabinofuranoside (18F-FAZA), 18F-2-(2-nitro-1H-imidazol-1-yl)-N-(2,2,3,3,3-pentafluoropropyl)-acetamide (18F-EF5), ¹³¹I-labeled iodo-azomycin galactopyranoside (¹³¹I-IAZGP).

3. Immunofluorescence and immunohistochemical methods: an indirect manner to measure hypoxia in tissue sections by assessing hypoxia-related protein expression, such as exogenous hypoxic markers (pimonidazole, EF-5) and endogenous cell markers (HIF-1, VEGF, CA9, GLUT-1/3). This method is only used to evaluate the hypoxic state of tissue *in vitro*.

Potential Treatment Exploiting Tumor Hypoxia

Several therapeutic strategies targeting hypoxia have been developed. It may be possible to directly deliver O₂ into the tumor through different methods such as the catalytic decomposition of endogenous hydrogen peroxide (H₂O₂) and light-triggered water splitting.⁸⁰

There are also many ongoing hypoxia-targeted therapy trials involving several kinds of drugs with different mechanisms. HIF inhibitors are an important type of hypoxia targeted therapy drugs and a large number of studies have shown that HIF inhibitors (such as PX-478, LW6), either as a single agent or in combination with other therapeutic agents, exhibited promising anti-tumor effects.⁸¹⁻⁸⁴

Hypoxia-activated prodrugs (HAPs) are bioreductive drugs that are selectively activated under hypoxic conditions and that can accurately target the hypoxic regions of solid tumors. Researches on HAPs is currently underway. Representative drugs include tirapazamine, evofosfamide. A large number of pre-clinical and clinical studies have reported exciting anti-tumor effects, regardless of single-agent application or combination therapy.⁸⁵⁻⁸⁷

Other strategies targeting hypoxia are also under study. As mentioned above, mTOR pathway plays a key role in tumor metabolism, mTOR targeted drugs are essential for regulating the tumor microenvironment under hypoxia.⁸⁸ Jain *et al.*^{13,89-91} have proposed 2 strategies to alleviate hypoxia: vascular normalization and vessel decompression. They conducted a lot of research to improve the efficacy of anti-vascular treatment (anti VEGF/VEGFR drugs, such as bevacizumab, sunitinib), and suggested that strategies aimed at alleviating tumor hypoxia while improving perfusion may enhance the outcome of radiotherapy, chemotherapy, and immunotherapy. Since hypoxia has a strong impact on the induction of HIF-1 α -dependent PD-L1 expression in tumor cells, blockade of PD-L1 may thus represent a novel approach for cancer immunotherapy.^{69,92}

These kinds of studies are still in their early phase and need to be further investigated to overcome challenges for successful clinical translation. Further studies of hypoxia in tumors are of utmost importance in both biological and therapeutic implications.

Conclusion

Hypoxia is an important property of solid malignancies, and is widely involved in all aspects of tumorigenesis and development. Lack of vascularity is the main cause of hypoxia, which in turn leads to low cell proliferation, increased glucose

metabolism, immunosuppression and resistance to immune attack. At present, a variety of methods are developed to detect tumor hypoxia, among which, PET imaging technology using radiotracer probes has great advantages due to its non-invasive, three-dimensional assessment and high sensitivity and specificity and has been widely applied in clinic. For therapeutic purposes, hypoxia has been used as a target to develop a variety of therapeutic strategies. Hypoxia-activated prodrugs (HAPs) have been developed and validated since 1960s. Many types of HAPs are already in phase 3 clinical trials. The application of hypoxia targeted therapy will open up a new way for tumor treatment.

Authors' Note

Yue Li performed the literature search and wrote the manuscript; Long Zhao performed the literature search and figure editing; Xiao-Feng Li contributed to write and revise the manuscript; all authors had approved the final manuscript to be submitted. This is a review article with no original animal and human studies.

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Declaration of Conflicting Interests

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References

- Brown JM, Wilson WR. Exploiting tumour hypoxia in cancer treatment. *Nat Rev Cancer*. 2004;4(6):437-447. doi:10.1038/nrc1367.
- Evans SM, Koch CJ. Prognostic significance of tumor oxygenation in humans. *Cancer Lett*. 2003;195(1):1-16. doi:10.1016/s0304-3835(03)00012-0
- Höckel M, Vaupel P. Tumor hypoxia: definitions and current clinical, biologic, and molecular aspects. *J Natl Cancer Inst*. 2001;93(4):266-276. doi:10.1093/jnci/93.4.266.
- Vaupel P, Mayer A. Hypoxia in cancer: significance and impact on clinical outcome. *Cancer Metastasis Rev*. 2007;26(2):225-239. doi:10.1007/s10555-007-9055-1
- Casazza A, Di Conza G, Wenes M, Finisguerra V, Deschoemaker S, Mazzone M. Tumor stroma: a complexity dictated by the hypoxic tumor microenvironment. *Oncogene*. 2014;33(14):1743-1754. doi:10.1038/onc.2013.121
- Vaupel P. The role of hypoxia-induced factors in tumor progression. *Oncologist*. 2004;9(Suppl 5):10-17. doi:10.1634/theoncologist.9-90005-10
- Cardenas-Navia LI, Richardson RA, Dewhirst MW. Targeting the molecular effects of a hypoxic tumor microenvironment. *Front Biosci*. 2007;12:4061-4578. doi:10.2741/2372
- Carmeliet P. VEGF as a key mediator of angiogenesis in cancer. *Oncology*. 2005;69(Suppl 3):4-10. doi:10.1159/000088478
- Challapalli A, Carroll L, Aboagye EO. Molecular mechanisms of hypoxia in cancer. *Clin Transl Imaging*. 2017;5(3):225-253. doi:10.1007/s40336-017-0231-1
- Arneth B. Tumor microenvironment. *Medicina (Kaunas)*. 2019;56(1):15. doi:10.3390/medicina56010015
- Naxerova K, Brachtel E, Salk JJ, et al. Hypermutable DNA chronicles the evolution of human colon cancer. *Proc Natl Acad Sci U S A*. 2014;111(18):E1889-1898. doi:10.1073/pnas.1400179111
- Naxerova K, Jain RK. Using tumour phylogenetics to identify the roots of metastasis in humans. *Nat Rev Clin Oncol*. 2015;12(5):258-272. doi:10.1038/nrclinonc.2014.238
- Martin JD, Fukumura D, Duda DG, Boucher Y, Jain RK. Reengineering the tumor microenvironment to alleviate hypoxia and overcome cancer heterogeneity. *Cold Spring Harb Perspect Med*. 2016;6(12):a027094. doi:10.1101/cshperspect.a027094
- Qin Y, Roszik J, Chattopadhyay C, et al. Hypoxia-driven mechanism of vemurafenib resistance in melanoma. *Mol Cancer Ther*. 2016;15(10):2442-2454. doi:10.1158/1535-7163.MCT-15-0963
- Hook SV. *Louis Pasteur: Groundbreaking Chemist & Biologist*. ABDO Publishing Company; 2011:8-112.
- Waki A, Kato H, Yano R, et al. The importance of glucose transport activity as the rate-limiting step of 2-deoxyglucose uptake in tumor cells in vitro. *Nucl Med Biol*. 1998;25(7):593-597. doi:10.1016/s0969-8051(98)00038-9
- Kennedy AS, Raleigh JA, Perez GM, et al. Proliferation and hypoxia in human squamous cell carcinoma of the cervix: first report of combined immunohistochemical assays. *Int J Radiat Oncol Biol Phys*. 1997;37(4):897-905. doi:10.1016/s0360-3016(96)00539-1
- Durand RE, Raleigh JA. Identification of nonproliferating but viable hypoxic tumor cells in vivo. *Cancer Res*. 1998;58(16):3547-3550.
- Li XF, Carlin S, Urano M, Russell J, Ling CC, O'Donoghue JA. Visualization of hypoxia in microscopic tumors by immunofluorescent microscopy. *Cancer Res*. 2007;67(16):7646-7653. doi:10.1158/0008-5472.CAN-06-4353
- Li XF, O'Donoghue JA. Hypoxia in microscopic tumors. *Cancer Lett*. 2008;264(2):172-180. doi:10.1016/j.canlet.2008.02.037
- Barnhill RL, Piepkorn MW, Cochran AJ, Flynn E, Karaoli T, Folkman J. Tumor vascularity, proliferation, and apoptosis in human melanoma micrometastases and macrometastases. *Arch Dermatol*. 1998;134(8):991-994. doi:10.1001/archderm.134.8.991
- Aguirre Ghiso JA, Kovalski K, Ossowski L. Tumor dormancy induced by downregulation of urokinase receptor in human

- carcinoma involves integrin and MAPK signaling. *J Cell Biol.* 1999;147(1):89-104. doi:10.1083/jcb.147.1.89
23. Aguirre-Ghiso JA, Estrada Y, Liu D, Ossowski L. ERK(MAPK) activity as a determinant of tumor growth and dormancy; regulation by p38(SAPK). *Cancer Res.* 2003;63(7):1684-1695.
24. Aguirre-Ghiso JA, Liu D, Mignatti A, Kovalski K, Ossowski L. Urokinase receptor and fibronectin regulate the ERK(MAPK) to p38(MAPK) activity ratios that determine carcinoma cell proliferation or dormancy in vivo. *Mol Biol Cell.* 2001;12(4):863-879. doi:10.1091/mbc.12.4.863
25. Naumov GN, Bender E, Zurakowski D, et al. A model of human tumor dormancy: an angiogenic switch from the nonangiogenic phenotype. *J Natl Cancer Inst.* 2006;98(5):316-325. doi:10.1093/jnci/djj068
26. Udagawa T, Fernandez A, Achilles EG, Folkman J, D'Amato RJ. Persistence of microscopic human cancers in mice: alterations in the angiogenic balance accompanies loss of tumor dormancy. *FASEB J.* 2002;16(11):1361-1370. doi:10.1096/fj.01-0813com
27. Fatema CN, Zhao S, Zhao Y, et al. Dual tracer evaluation of dynamic changes in intratumoral hypoxic and proliferative states after radiotherapy of human head and neck cancer xenografts using radiolabeled FMISO and FLT. *BMC Cancer.* 2014;14:692. doi:10.1186/1471-2407-14-692
28. Huang T, Civelek AC, Li J, et al. Tumor microenvironment-dependent 18F-FDG, 18F-fluorothymidine, and 18F-misonidazole uptake: a pilot study in mouse models of human non-small cell lung cancer. *J Nucl Med.* 2012;53(8):1262-1268. doi:10.2967/jnumed.111.098087
29. Huang T, Civelek AC, Zheng H, et al. 18F-misonidazole PET imaging of hypoxia in micrometastases and macroscopic xenografts of human non-small cell lung cancer: a correlation with autoradiography and histological findings. *Am J Nucl Med Mol Imaging.* 2013;3(2):142-153.
30. Cui YL, Wang X, Li XF. (18)F-fluoromisonidazole PET reveals spatial and temporal heterogeneity of hypoxia in mouse models of human non-small-cell lung cancer. *Future Oncol.* 2015;11(20):2841-2849. doi:10.2217/fo.15.205
31. Wang X, He Y, Zhou W, et al. Mismatched intratumoral distribution of [18F] fluorodeoxyglucose and 3'-deoxy-3'-[18F] fluorothymidine in patients with lung cancer. *Oncol Lett.* 2017;14(5):5279-5284. doi:10.3892/ol.2017.6840
32. Viillard C, Larrivée B. Tumor angiogenesis and vascular normalization: alternative therapeutic targets. *Angiogenesis.* 2017;20(4):409-426. doi:10.1007/s10456-017-9562-9
33. Li T, Kang G, Wang T, Huang H. Tumor angiogenesis and anti-angiogenic gene therapy for cancer. *Oncol Lett.* 2018;16(1):687-702. doi:10.3892/ol.2018.8733
34. Ferrara N, Chen H, Davis-Smyth T, et al. Vascular endothelial growth factor is essential for corpus luteum angiogenesis. *Nat Med.* 1998;4(3):336-340. doi:10.1038/nm0398-336
35. Scappaticci FA. Mechanisms and future directions for angiogenesis-based cancer therapies. *J Clin Oncol.* 2002;20(18):3906-3927. doi:10.1200/JCO.2002.01.033
36. Folkman J. Tumor angiogenesis: therapeutic implications. *N Engl J Med.* 1971;285(21):1182-1186. doi:10.1056/NEJM19711182852108
37. St Croix B, Rago C, Velculescu V, et al. Genes expressed in human tumor endothelium. *Science.* 2000;289(5482):1197-1202. doi:10.1126/science.289.5482.1197
38. Kunz M, Ibrahim SM. Molecular responses to hypoxia in tumor cells. *Mol Cancer.* 2003;2:23. doi:10.1186/1476-4598-2-23
39. Chiche J, Brahimi-Horn MC, Pouysselgur J. Tumour hypoxia induces a metabolic shift causing acidosis: a common feature in cancer. *J Cell Mol Med.* 2010;14(4):771-794. doi:10.1111/j.1582-4934.2009.00994.x
40. Ward PS, Thompson CB. Metabolic reprogramming: a cancer hallmark even Warburg did not anticipate. *Cancer Cell.* 2012;21(3):297-308. doi:10.1016/j.ccr.2012.02.014
41. Pugachev A, Ruan S, Carlin S, et al. Dependence of FDG uptake on tumor microenvironment. *Int J Radiat Oncol Biol Phys.* 2005;62(2):545-553. doi:10.1016/j.ijrobp.2005.02.009
42. Zhang G, Li J, Wang X, et al. The reverse Warburg effect and 18F-FDG uptake in non-small cell lung cancer A549 in mice: a pilot study. *J Nucl Med.* 2015;56(4):607-612. doi:10.2967/jnumed.114.148254
43. Shen B, Huang T, Sun Y, Jin Z, Li XF. Revisit 18F-fluorodeoxyglucose oncology positron emission tomography: "systems molecular imaging" of glucose metabolism. *Oncotarget.* 2017;8(26):43536-43542. doi:10.18632/oncotarget.16647
44. Semenza GL. Targeting HIF-1 for cancer therapy. *Nat Rev Cancer.* 2003;3(10):721-732. doi:10.1038/nrc1187
45. Li XF, Ma Y, Sun X, Humm JL, Ling CC, O'Donoghue JA. High 18F-FDG uptake in microscopic peritoneal tumors requires physiologic hypoxia. *J Nucl Med.* 2010;51(4):632-638. doi:10.2967/jnumed.109.071233
46. Magaway C, Kim E, Jacinto E. Targeting mTOR and metabolism in cancer: lessons and innovations. *Cells.* 2019;8(12):1584. doi:10.3390/cells8121584
47. Toschi A, Lee E, Gadir N, Ohh M, Foster DA. Differential dependence of hypoxia-inducible factors 1 alpha and 2 alpha on mTORC1 and mTORC2. *J Biol Chem.* 2008;283(50):34495-34499. doi:10.1074/jbc.C800170200
48. Thomas GV, Tran C, Mellinshoff IK, et al. Hypoxia-inducible factor determines sensitivity to inhibitors of mTOR in kidney cancer. *Nat Med.* 2006;12(1):122-127. doi:10.1038/nm1337
49. Masui K, Tanaka K, Akhavan D, et al. mTOR complex 2 controls glycolytic metabolism in glioblastoma through FoxO acetylation and upregulation of c-Myc. *Cell Metab.* 2013;18(5):726-739. doi:10.1016/j.cmet.2013.09.013
50. Li Y, He ZC, Liu Q, et al. Large intergenic non-coding RNA-RoR inhibits aerobic glycolysis of glioblastoma cells via Akt pathway. *J Cancer.* 2018;9(5):880-889. doi:10.7150/jca.20869
51. Sengupta D, Mongersun A, Kim TJ, et al. Multiplexed single-cell measurements of FDG uptake and lactate release using droplet microfluidics. *Technol Cancer Res Treat.* 2019;18:1533033819841066. doi:10.1177/1533033819841066
52. DeBerardinis RJ, Chandel NS. Fundamentals of cancer metabolism. *Sci Adv.* 2016;2(5):e1600200. doi:10.1126/sciadv.1600200

53. Hanahan D, Weinberg RA. Hallmarks of cancer: the next generation. *Cell*. 2011;144(5):646-674. doi:10.1016/j.cell.2011.02.013
54. Teng MW, Galon J, Fridman WH, Smyth MJ. From mice to humans: developments in cancer immunoeediting. *J Clin Invest*. 2015;125(9):3338-3346. doi:10.1172/JCI80004
55. Gonzalez H, Hagerling C, Werb Z. Roles of the immune system in cancer: from tumor initiation to metastatic progression. *Genes Dev*. 2018;32(19-20):1267-1284. doi:10.1101/gad.314617.118
56. Palucka AK, Coussens LM. The basis of oncoimmunology. *Cell*. 2016;164(6):1233-1247. doi:10.1016/j.cell.2016.01.049
57. Li Y, Patel SP, Roszik J, Qin Y. Hypoxia-driven immunosuppressive metabolites in the tumor microenvironment: new approaches for combinational immunotherapy. *Front Immunol*. 2018;9:1591. doi:10.3389/fimmu.2018.01591
58. Palazon A, Goldrath AW, Nizet V, Johnson RS. HIF transcription factors, inflammation, and immunity. *Immunity*. 2014;41(4):518-528. doi:10.1016/j.immuni.2014.09.008
59. Noman MZ, Hasmin M, Messai Y, et al. Hypoxia: a key player in antitumor immune response. A review in the theme: cellular responses to hypoxia. *Am J Physiol Cell Physiol*. 2015;309(9):C569-579. doi:10.1152/ajpcell.00207.2015
60. Parodi M, Raggi F, Cangelosi D, et al. Hypoxia modifies the transcriptome of human NK cells, modulates their immunoregulatory profile, and influences NK cell subset migration. *Front Immunol*. 2018;9:2358. doi:10.3389/fimmu.2018.02358
61. Velásquez SY, Killian D, Schulte J, Sticht C, Thiel M, Lindner HA. Short term hypoxia synergizes with interleukin 15 priming in driving glycolytic gene transcription and supports human natural killer cell activities. *J Biol Chem*. 2016;291(25):12960-12977. doi:10.1074/jbc.M116.721753
62. Balsamo M, Manzini C, Pietra G, et al. Hypoxia downregulates the expression of activating receptors involved in NK-cell-mediated target cell killing without affecting ADCC. *Eur J Immunol*. 2013;43(10):2756-2764. doi:10.1002/eji.201343448
63. Husain Z, Huang Y, Seth P, Sukhatme VP. Tumor-derived lactate modifies antitumor immune response: effect on myeloid-derived suppressor cells and NK cells. *J Immunol*. 2013;191(3):1486-1495. doi:10.4049/jimmunol.1202702
64. Piñeiro Fernández J, Luddy KA, Harmon C, O'Farrelly C. Hepatic tumor microenvironments and effects on NK cell phenotype and function. *Int J Mol Sci*. 2019;20(17):4131. doi:10.3390/ijms20174131
65. Mahiddine K, Blaisdell A, Ma S, Créquer-Grandhomme A, Lowell CA, Erlebacher A. Relief of tumor hypoxia unleashes the tumoricidal potential of neutrophils. *J Clin Invest*. 2020;130(1):389-403. doi:10.1172/JCI130952
66. Topalian SL, Drake CG, Pardoll DM. Immune checkpoint blockade: a common denominator approach to cancer therapy. *Cancer Cell*. 2015;27(4):450-461. doi:10.1016/j.ccell.2015.03.001
67. Barsoum IB, Smallwood CA, Siemens DR, Graham CH. A mechanism of hypoxia-mediated escape from adaptive immunity in cancer cells. *Cancer Res*. 2014;74(3):665-674. doi:10.1158/0008-5472.CAN-13-0992
68. Noman MZ, Desantis G, Janji B, et al. PD-L1 is a novel direct target of HIF-1 α , and its blockade under hypoxia enhanced MDSC-mediated T cell activation. *J Exp Med*. 2014;211(5):781-790. doi:10.1084/jem.20131916
69. Palsson-McDermott EM, Dyck L, Zasłona Z, et al. Pyruvate kinase M2 Is required for the expression of the immune checkpoint PD-L1 in immune cells and tumors. *Front Immunol*. 2017;8:1300. doi:10.3389/fimmu.2017.01300
70. Cho SH, Raybuck AL, Stengel K, et al. Germinal centre hypoxia and regulation of antibody qualities by a hypoxia response system. *Nature*. 2016;537(7619):234-238. doi:10.1038/nature19334
71. Burrows N, Maxwell PH. Hypoxia and B cells. *Exp Cell Res*. 2017;356(2):197-203. doi:10.1016/j.yexcr.2017.03.019
72. Bosco MC, Pierobon D, Blengio F, et al. Hypoxia modulates the gene expression profile of immunoregulatory receptors in human mature dendritic cells: identification of TREM-1 as a novel hypoxic marker in vitro and in vivo. *Blood*. 2011;117(9):2625-2639. doi:10.1182/blood-2010-06-292136
73. Pierobon D, Bosco MC, Blengio F, et al. Chronic hypoxia reprograms human immature dendritic cells by inducing a proinflammatory phenotype and TREM-1 expression. *Eur J Immunol*. 2013;43(4):949-966. doi:10.1002/eji.201242709
74. Hammami A, Charpentier T, Smans M, Stäger S. IRF-5-mediated inflammation limits CD8 $^{+}$ T cell expansion by inducing HIF-1 α and impairing dendritic cell functions during leishmania infection. *PLoS Pathog*. 2015;11(6):e1004938. doi:10.1371/journal.ppat.1004938
75. Filippi I, Morena E, Aldinucci C, Carraro F, Sozzani S, Naldini A. Short-term hypoxia enhances the migratory capability of dendritic cell through HIF-1 α and PI3K/Akt pathway. *J Cell Physiol*. 2014;229(12):2067-2076. doi:10.1002/jcp.24666
76. Chia K, Fleming IN, Blower PJ. Hypoxia imaging with PET: which tracers and why? *Nucl Med Commun*. 2012;33(3):217-222. doi:10.1097/MNM.0b013e32834eacb7
77. Li XF, Sun X, Ma Y, et al. Detection of hypoxia in microscopic tumors using 131I-labeled iodo-azomycin galactopyranoside (131I-IAZGP) digital autoradiography. *Eur J Nucl Med Mol Imaging*. 2010;37(2):339-348. doi:10.1007/s00259-009-1310-y
78. Bussink J, van Herpen CM, Kaanders JH, Oyen WJ. PET-CT for response assessment and treatment adaptation in head and neck cancer. *Lancet Oncol*. 2010;11(7):661-669. doi:10.1016/S1470-2045(09)70353-5
79. Fleming IN, Manavaki R, Blower PJ, et al. Imaging tumour hypoxia with positron emission tomography. *Br J Cancer*. 2015;112(2):238-2350. doi:10.1038/bjc.2014.610
80. Sahu A, Kwon I, Tae G. Improving cancer therapy through the nanomaterials-assisted alleviation of hypoxia. *Biomaterials*. 2020;228:119578. doi:10.1016/j.biomaterials.2019.119578
81. Fallah J, Rini BI. HIF inhibitors: status of current clinical development. *Curr Oncol Rep*. 2019;21(1):6. doi:10.1007/s11912-019-0752-z
82. Albadari N, Deng S, Li W. The transcriptional factors HIF-1 and HIF-2 and their novel inhibitors in cancer therapy. *Expert Opin Drug Discov*. 2019;14(7):667-682. doi:10.1080/17460441.2019.1613370
83. Parczyk J, Ruhnau J, Pelz C, et al. Dichloroacetate and PX-478 exhibit strong synergistic effects in a various number of cancer

- cell lines. *BMC Cancer*. 2021;21(1):481. doi:10.1186/s12885-021-08186-9
84. Lee JY, Lee K, Lee K, et al. Pharmacokinetic characterization of LW6, a novel hypoxia-inducible factor-1 α (HIF-1 α) inhibitor in mice. *Molecules*. 2021;26(8):2226. doi:10.3390/molecules26082226
85. Li Y, Zhao L, Li XF. The hypoxia-activated prodrug TH-302: exploiting hypoxia in cancer therapy. *Front Pharmacol*. 2021; 12:636892. doi:10.3389/fphar.2021.636892
86. Baran N, Konopleva M. Molecular pathways: hypoxia-activated prodrugs in cancer therapy. *Clin Cancer Res*. 2017;23(10):2382-2390. doi:10.1158/1078-0432.CCR-16-0895
87. Phillips RM. Targeting the hypoxic fraction of tumours using hypoxia-activated prodrugs. *Cancer Chemother Pharmacol*. 2016;77(3):441-457. doi:10.1007/s00280-015-2920-7
88. Tian T, Li X, Zhang J. mTOR signaling in cancer and mTOR inhibitors in solid tumor targeting therapy. *Int J Mol Sci*. 2019; 20(3):755. doi:10.3390/ijms20030755
89. Jain RK. Normalizing tumor vasculature with anti-angiogenic therapy: a new paradigm for combination therapy. *Nat Med*. 2001;7(9):987-989. doi:10.1038/nm0901-987
90. Jain RK. Normalization of tumor vasculature: an emerging concept in antiangiogenic therapy. *Science*. 2005;307(5706):58-62. doi:10.1126/science.1104819
91. Jain RK. Antiangiogenesis strategies revisited: from starving tumors to alleviating hypoxia. *Cancer Cell*. 2014;26(5): 605-622. doi:10.1016/j.ccell.2014.10.006
92. Saleh R, Taha RZ, Sasidharan Nair V, Alajezi NM, Elkord E. PD-L1 blockade by atezolizumab downregulates signaling pathways associated with tumor growth, metastasis, and hypoxia in human triple negative breast cancer. *Cancers (Basel)*. 2019;11(8):1050. doi:10.3390/cancers11081050