



Proteomics and Its Application in Pandemic Diseases



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Pandemics create havoc in human life and are not simply old-world problems. A series of recent viral outbreaks including HIV, influenza, Ebola, Zika, and SARS-CoV-2 have elevated to pandemics. The recent and ongoing coronavirus disease (COVID-19) caused by SARS-CoV-2 has caused more than 1 081 868 deaths and infected more than 37 888 384 humans worldwide, as reported by the World Health Organization (WHO). Prevention, early diagnosis, and early treatment are the keys to pandemic management. Proteomics can play an important role in understanding host-pathogen interactions, diagnosing infections, developing vaccines, and creating therapeutics for pandemic-causing pathogens such as SARS-CoV-2. Notably, COVID-19 emerged in Wuhan, China and was first reported as pneumonia with an unknown cause to a WHO China country office on December 31, 2019. Furthermore, it was elevated to a pandemic by WHO on March 11, 2020. COVID-19 is a serious threat for patients with comorbidities such as compromised immune systems, respiratory diseases, cardiovascular diseases, hypertension, diabetes, and other diseases. This disease has destabilized the healthcare systems and economies worldwide. COVID-19 can infect any age, gender, or ethnicity and can infect healthy people. Body fluids including nasal aerosol and saliva are major modes of transmission for spreading COVID-19. Unfortunately, there are no clinically approved vaccines or curative drugs for it. Efforts are being made to develop vaccines and therapeutics against the deadly SARS-CoV-2 to save lives. There are several emergency drugs for COVID-19, including remdesivir, dexamethasone, and favipiravir. Early treatment with favipiravir and remdesivir may be beneficial during the ongoing COVID-19 pandemic. Both favipiravir and remdesivir are prodrugs that inhibit the RNA-dependent RNA polymerase (RdRp) enzyme and prevent virus replication.

RdRp is one of the favorite targets for antiviral drug discovery. In this “Proteomics in Pandemic Disease” Special Issue, Zhao and Bourne have suggested four classes of binding modes for RdRp binding pockets that can help the *in silico* screening of RdRp inhibitors. Nucleotide analogues such as triphosphates of sofosbuvir, alovudine, AZT, abacavir, lamivudine, and emtricitabine can inhibit the RdRp of SARS-CoV-2, as studied by Chien et al.

Furthermore, Zeng et al. proposed 41 drugs for repurposing, including dexamethasone for COVID-19, using the deep-learning framework and AI methodology. Moreover, by a computational approach including homology modeling, molecular docking, molecular dynamics simulations, and binding affinity calculations, Martin and Cheng found potential targets for toremifene in SARS-CoV-2, such as heptad repeat 1 (HR1) and methyltransferase nonstructural protein (NSP) 14.

Using *in silico* studies, Barros et al. found that the antimalarial drug mequine and anti-HIV antiretroviral saquinavir interact with four SARS-CoV-2 receptors, including Nsp9 replicase, main protease (Mpro), NSP15 endoribonuclease, and spike protein (S protein), interacting with human ACE2; therefore, they may be repurposed for COVID-19 treatment. The bioinformatics analysis of BCG antigens by Glisic et al. suggested that four bacterial proteins, Rv0934, Rv3763, Rv3875, and Rv2997, have similar properties as the S1 protein of SARS-CoV-2; therefore, they might be effective against SARS-CoV-2. A molecular docking and dynamics simulation analysis suggested that noscapine binds with the main protease of SARS-CoV-2 and produces conformational changes (Kumar et al.). Furthermore, Maffucci and Contini used an *in silico* approach to find drug candidates against the main proteinase and spike protein of SARS-CoV-2. This led to the finding that indinavir, polymyxin B, daptomycin, terlipressin, and thymopentin can be repurposed against the SARS-CoV-2 infection. Interestingly, the studies by Stamatakis et al. suggested that the antigenic peptides generated from the S1 spike glycoprotein of SARS-CoV-2 using aminopeptidases ERAP1, ERAP2, and IRAP might be helpful in selecting better epitopes for immunogenic studies and the design of a vaccine for COVID-19.

Metabolites in SARS-CoV-2 patients were analyzed by NMR and mass spectrometry with the observations of an increase in the α -1-acid glycoprotein, an increased kynurenine/tryptophan ratio, low total and HDL apolipoprotein A1, low HDL triglycerides, high LDL and VLDL triglycerides, elevated glutamine/glutamate, and Fischer's ratios that are consistent with diabetes, coronary artery disease, and liver dysfunction risk (Kimhofer et al.). By using NMR spectroscopy, Loo et al. reported that inactivation by heat causes the degradation of lipoproteins and changes in various metabolic information in SARS-CoV-2-infected plasma samples. Saunders et al. described the coronavirus-specific web portal (<https://metatryp-coronavirus.whoi.edu/>) in METATRY V2.0 that can be used for coronavirus proteomics research. This web portal is helpful for finding peptide biomarkers and specific taxonomic groups.

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The prevalence of pandemic diseases has necessitated method development for their detection. A mass-spectrometry-based method was developed by Ihling et al. for gargle solution collected from SARS-CoV-2-infected persons. Gouveia et al. developed a nanoLC-MS/MS method to quantify several virus-specific peptides from COVID-19 patients for proteo-typing of SARS-CoV-2 using nasopharyngeal swabs in a 20 min MS acquisition window. Furthermore, the nanoLC-MS/MS acquisition method for COVID-19 clinical samples reported that the peptides ADETQALPQR and GFYAAQGSR from nucleocapsid proteins were detected within a 3 min window. Nikolaev et al. developed a method for the detection of the SARS-CoV-2 virus using nasopharynx epithelial swabs by the identification of the viral nucleocapsid N protein from the virus. They inactivated the virus by heating, the addition of isopropanol, followed by trypsin digestion and then analyzed it by mass spectrometry. In addition, D'Alessandro et al. reported serum proteomics from COVID-19 patients and observed the up-regulation of IL-6, complement cascade, and inhibitory components of the fibrinolytic cascade in COVID-19 patients compared with the control. Zhou et al. reported the quantitative proteomics of porcine-deltacoronavirus (PDCoV)-infected IPEC-J2 cells (porcine small intestinal epithelial cells) using iTRAQ, which might be helpful to understand its pathogenesis.

Rosa-Fernandes et al. reported a method to study the ocular surface proteome, especially for infants exposed to the Zika virus during the gestation period without early clinical symptoms, and named it the cellular imprinting proteomic assay (CImPA). In the conjunctival epithelium of infants exposed to the Zika virus, neutrophil, eosinophil infiltration, and degranulation were detected by this proteomic assay. Furthermore, virus-like particles (VLPs) have been applied in vaccine therapies. Lavado-García et al. used quantitative proteomics to identify the changes in the secretome of VLPs and the coproduction of extracellular vesicles (EVs) under different conditions, including nontransfected and transfected with and without plasmid coding for HIV-1 Gag polyprotein.

Interestingly, a computational method was used to find an allosteric site on the SARS-CoV-2 spike protein by Di Paola et al., as its detection would weaken the spike–ACE2 interaction and thereby reduce the viral infection. Another study by Verkhivker suggests that the spike protein of SARS-CoV-2 may function as an allosteric regulatory engine fluctuating between dynamic functional states. Hadi-Alijavand and Rouhani reported the higher binding affinity of the closed state of ACE2 for the S1 protein of SARS-CoV-2 compared with the open state of ACE2 by using a computational approach. Furthermore, Nadeau et al. used a computational approach to study the SARS-CoV-2-interacting human proteins using the GoNet algorithm.

Using a ferret model for the H1N1 2009 pandemic influenza A virus, Chen et al. reported the host glycomic response and its age-dependent severity. They suggested that a high level of mannose may be related to the severity of the influenza A virus due to the overactive innate immune system. In the case of the Ebola virus, Banerjee and Mitra proposed the tetrameric assembly model to the VP35 protein and suggested that the C-terminal of VP35 interacts with human protein kinase R to stop its autophosphorylation.

This Special Issue provides a platform to understand pandemic diseases. Here several topics have been encouraged that connect proteomics and pandemic diseases, including

proteomic technologies, biomarker discovery, pathogenesis, mechanistic details of proteins, protein–protein interactions, signaling pathways, post-translational modifications, computational proteomics, prevention and vaccination, drug repurposing, and therapeutic agents and their mode of action. It will be thrilling to find the bridge between proteomics and pandemic diseases, especially for COVID-19.

Publications in the “Proteomics in Pandemic Disease” Special Issue

1. Whetton, A. D.; Preston, G. W.; Abubeker, S.; Geifman, N. Proteomics and Informatics for Understanding Phases and Identifying Biomarkers in COVID-19 Disease. *J. Proteome Res.* **2020**, [10.1021/acs.jproteome.0c00326](https://doi.org/10.1021/acs.jproteome.0c00326).
2. Tilocca, B.; Britti, D.; Urbani, A.; Roncada, P. Computational Immune Proteomics Approach to Target COVID-19. *J. Proteome Res.* **2020**, [10.1021/acs.jproteome.0c00553](https://doi.org/10.1021/acs.jproteome.0c00553).
3. Yu, H.; Li, C.; Wang, X.; Duan, J.; Yang, N.; Xie, L.; Yuan, Y.; Li, S.; Bi, C.; Yang, B.; Li, Y. Techniques and Strategies for Potential Protein Target Discovery and Active Pharmaceutical Molecule Screening in a Pandemic. *J. Proteome Res.* **2020**, [10.1021/acs.jproteome.0c00372](https://doi.org/10.1021/acs.jproteome.0c00372).
4. Sperk, M.; van Domselaar, R.; Rodriguez, J. E.; Mikaeloff, F.; Vinhas, B. S.; Saccon, E.; Sönnnerborg, A.; Singh, K.; Gupta, S.; Végvári, A.; Neogi, U. Utility of Proteomics in Emerging and Re-Emerging Infectious Diseases Caused by RNA Viruses. *J. Proteome Res.* **2020**, [10.1021/acs.jproteome.0c00380](https://doi.org/10.1021/acs.jproteome.0c00380).
5. Sirpilla, O.; Bauss, J.; Gupta, R.; Underwood, A.; Qutob, D.; Freeland, T.; Bupp, C.; Carillo, J.; Hartog, N.; Rajasekaran, S.; Prokop, J. W. SARS-CoV-2-Encoded Proteome and Human Genetics: From Interaction-Based to Ribosomal Biology Impact on Disease and Risk Processes. *J. Proteome Res.* **2020**, [10.1021/acs.jproteome.0c00421](https://doi.org/10.1021/acs.jproteome.0c00421).
6. Francés-Monerris, A.; Hognon, C.; Miclot, T.; García-Iriepa, C.; Iriepa, I.; Terenzi, A.; Grandemange, S.; Barone, G.; Marazzi, M.; Monari, A. Molecular Basis of SARS-CoV-2 Infection and Rational Design of Potential Antiviral Agents: Modeling and Simulation Approaches. *J. Proteome Res.* **2020**, [10.1021/acs.jproteome.0c00779](https://doi.org/10.1021/acs.jproteome.0c00779).
7. Luan, B.; Huynh, T.; Cheng, X.; Lan, G.; Wang, H.-R. Targeting Proteases for Treating COVID-19. *J. Proteome Res.* **2020**, [10.1021/acs.jproteome.0c00430](https://doi.org/10.1021/acs.jproteome.0c00430).
8. Crisan-Dabija, R.; Pavel, C. A.; Popa, I. V.; Tarus, A.; Burlacu, A. “A Chain Only as Strong as Its Weakest Link”: An Up-to-Date Literature Review on the Bidirectional Interaction of Pulmonary Fibrosis and COVID-19. *J. Proteome Res.* **2020**, [10.1021/acs.jproteome.0c00387](https://doi.org/10.1021/acs.jproteome.0c00387).
9. Heiss, K.; Heidepriem, J.; Fischer, N.; Weber, L. K.; Dahlke, C.; Jaenisch, T.; Loeffler, F. F. Rapid Response to Pandemic Threats: Immunogenic Epitope Detection of Pandemic Pathogens for Diagnostics and Vaccine Development Using Peptide Microarrays. *J. Proteome Res.* **2020**, [10.1021/acs.jproteome.0c00484](https://doi.org/10.1021/acs.jproteome.0c00484).
10. Goh, G. K.-M.; Dunker, A. K.; Foster, J. A.; Uversky, V. N. A Novel Strategy for the Development of Vaccines for SARS-CoV-2 (COVID-19) and Other Viruses Using AI and Viral Shell Disorder. *J. Proteome Res.* **2020**, [10.1021/acs.jproteome.0c00672](https://doi.org/10.1021/acs.jproteome.0c00672).
11. Saei, A. A.; Sharifi, S.; Mahmoudi, M. COVID-19: Nanomedicine Uncovers Blood-Clot Mystery. *J. Proteome Res.* **2020**, [10.1021/acs.jproteome.0c00425](https://doi.org/10.1021/acs.jproteome.0c00425).

12. Zhang, Y.-M.; Wang, L.; Liu, X.-Z.; Zhang, H. The COVID-19 Pandemic from a Human Genetic Perspective. *J. Proteome Res.* **2020**, [10.1021/acs.jproteome.0c00671](https://doi.org/10.1021/acs.jproteome.0c00671).
13. Grossegesse, M.; Hartkopf, F.; Nitsche, A.; Schaade, L.; Doellinger, J.; Muth, T. Perspective on Proteomics for Virus Detection in Clinical Samples. *J. Proteome Res.* **2020**, [10.1021/acs.jproteome.0c00674](https://doi.org/10.1021/acs.jproteome.0c00674).
14. Ihling, C.; Tänzler, D.; Hagemann, S.; Kehlen, A.; Hüttelmaier, S.; Arlt, C.; Sinz, A. Mass Spectrometric Identification of SARS-CoV-2 Proteins from Gargle Solution Samples of COVID-19 Patients. *J. Proteome Res.* **2020**, [10.1021/acs.jproteome.0c00280](https://doi.org/10.1021/acs.jproteome.0c00280).
15. Nikolaev, E. N.; Indeykina, M. I.; Brzhozovskiy, A. G.; Bugrova, A. E.; Kononikhin, A. S.; Starodubtseva, N. L.; Petrotchenko, E. V.; Kovalev, G. I.; Borchers, C. H.; Sukhikh, G. T. Mass-Spectrometric Detection of SARS-CoV-2 Virus in Scrapings of the Epithelium of the Nasopharynx of Infected Patients via Nucleocapsid N Protein. *J. Proteome Res.* **2020**, [10.1021/acs.jproteome.0c00412](https://doi.org/10.1021/acs.jproteome.0c00412).
16. Stamatakis, G.; Samiotaki, M.; Mpakali, A.; Panayotou, G.; Stratikos, E. Generation of SARS-CoV-2 S1 Spike Glycoprotein Putative Antigenic Epitopes in Vitro by Intracellular Aminopeptidases. *J. Proteome Res.* **2020**, [10.1021/acs.jproteome.0c00457](https://doi.org/10.1021/acs.jproteome.0c00457).
17. Gouveia, D.; Miotello, G.; Gallais, F.; Gaillard, J.-C.; Debroas, S.; Bellanger, L.; Lavigne, J.-P.; Sotto, A.; Grenga, L.; Pible, O.; Armengaud, J. Proteotyping SARS-CoV-2 Virus from Nasopharyngeal Swabs: A Proof-of-Concept Focused on a 3 min Mass Spectrometry Window. *J. Proteome Res.* **2020**, [10.1021/acs.jproteome.0c00535](https://doi.org/10.1021/acs.jproteome.0c00535).
18. D'Alessandro, A.; Thomas, T.; Dzieciatkowska, M.; Hill, R. C.; Francis, R. O.; Hudson, K. E.; Zimring, J. C.; Hod, E. A.; Spitalnik, S. L.; Hansen, K. C. Serum Proteomics in COVID-19 Patients: Altered Coagulation and Complement Status as a Function of IL-6 Level. *J. Proteome Res.* **2020**, [10.1021/acs.jproteome.0c00365](https://doi.org/10.1021/acs.jproteome.0c00365).
19. Loo, R. L.; Lodge, S.; Kimhofer, T.; Bong, S.-H.; Begum, S.; Whiley, L.; Gray, N.; Lindon, J. C.; Nitschke, P.; Lawler, N. G.; Schäfer, H.; Spraul, M.; Richards, T.; Nicholson, J. K.; Holmes, E. Quantitative In-Vitro Diagnostic NMR Spectroscopy for Lipoprotein and Metabolite Measurements in Plasma and Serum: Recommendations for Analytical Artefact Minimization with Special Reference to COVID-19/SARS-CoV-2 Samples. *J. Proteome Res.* **2020**, [10.1021/acs.jproteome.0c00537](https://doi.org/10.1021/acs.jproteome.0c00537).
20. Thomas, T.; Stefanoni, D.; Dzieciatkowska, M.; Issaian, A.; Nemkov, T.; Hill, R. C.; Francis, R. O.; Hudson, K. E.; Buehler, P. W.; Zimring, J. C.; Hod, E. A.; Hansen, K. C.; Spitalnik, S. L.; D'Alessandro, A. Evidence of Structural Protein Damage and Membrane Lipid Remodeling in Red Blood Cells from COVID-19 Patients. *J. Proteome Res.* **2020**, [10.1021/acs.jproteome.0c00606](https://doi.org/10.1021/acs.jproteome.0c00606).
21. Zhou, X.; Zhou, L.; Ge, X.; Guo, X.; Han, J.; Zhang, Y.; Yang, H. Quantitative Proteomic Analysis of Porcine Intestinal Epithelial Cells Infected with Porcine Deltacoronavirus Using iTRAQ-Coupled LC-MS/MS. *J. Proteome Res.* **2020**, [10.1021/acs.jproteome.0c00592](https://doi.org/10.1021/acs.jproteome.0c00592).
22. Chen, S.; Kasper, B.; Zhang, B.; Lashua, L. P.; Ross, T. M.; Ghedin, E.; Mahal, L. K. Age-Dependent Glycomic Response to the 2009 Pandemic H1N1 Influenza Virus and Its Association with Disease Severity. *J. Proteome Res.* **2020**, [10.1021/acs.jproteome.0c00455](https://doi.org/10.1021/acs.jproteome.0c00455).
23. Rosa-Fernandes, L.; Barbosa, R. H.; dos Santos, M. L. B.; Angeli, C. B.; Silva, T. P.; Melo, R. C. N.; de Oliveira, G. S.; Lemos, B.; Van Eyk, J. E.; Larsen, M. R.; Cardoso, C. A.; Palmisano, G. Cellular Imprinting Proteomics Assay: A Novel Method for Detection of Neural and Ocular Disorders Applied to Congenital Zika Virus Syndrome. *J. Proteome Res.* **2020**, [10.1021/acs.jproteome.0c00320](https://doi.org/10.1021/acs.jproteome.0c00320).
24. Lavado-García, J.; González-Domínguez, I.; Cervera, L.; Jorge, I.; Vázquez, J.; Gòdia, F. Molecular Characterization of the Coproduced Extracellular Vesicles in HEK293 during Virus-Like Particle Production. *J. Proteome Res.* **2020**, [10.1021/acs.jproteome.0c00581](https://doi.org/10.1021/acs.jproteome.0c00581).
25. Banerjee, A.; Mitra, P. Ebola Virus VP35 Protein: Modeling of the Tetrameric Structure and an Analysis of Its Interaction with Human PKR. *J. Proteome Res.* **2020**, [10.1021/acs.jproteome.0c00473](https://doi.org/10.1021/acs.jproteome.0c00473).
26. Goh, G. K.-M.; Dunker, A. K.; Foster, J. A.; Uversky, V. N. Shell Disorder Analysis Suggests That Pangolins Offered a Window for a Silent Spread of an Attenuated SARS-CoV-2 Precursor among Humans. *J. Proteome Res.* **2020**, [10.1021/acs.jproteome.0c00460](https://doi.org/10.1021/acs.jproteome.0c00460).
27. Nadeau, R.; Fard, S. S.; Scheer, A.; Hashimoto-Roth, E.; Nygard, D.; Abramchuk, I.; Chung, Y.-E.; Bennett, S. A. L.; Lavallée-Adam, M. Computational Identification of Human Biological Processes and Protein Sequence Motifs Putatively Targeted by SARS-CoV-2 Proteins Using Protein–Protein Interaction Networks. *J. Proteome Res.* **2020**, [10.1021/acs.jproteome.0c00422](https://doi.org/10.1021/acs.jproteome.0c00422).
28. Barros, R. O.; Junior, F. L. C. C.; Pereira, W. S.; Oliveira, N. M. N.; Ramos, R. M. Interaction of Drug Candidates with Various SARS-CoV-2 Receptors: An in Silico Study to Combat COVID-19. *J. Proteome Res.* **2020**, [10.1021/acs.jproteome.0c00327](https://doi.org/10.1021/acs.jproteome.0c00327).
29. Di Paola, L.; Hadi-Alijanvand, H.; Song, X.; Hu, G.; Giuliani, A. The Discovery of a Putative Allosteric Site in the SARS-CoV-2 Spike Protein Using an Integrated Structural/Dynamic Approach. *J. Proteome Res.* **2020**, [10.1021/acs.jproteome.0c00273](https://doi.org/10.1021/acs.jproteome.0c00273).
30. Verkhivker, G. M. Molecular Simulations and Network Modeling Reveal an Allosteric Signaling in the SARS-CoV-2 Spike Proteins. *J. Proteome Res.* **2020**, [10.1021/acs.jproteome.0c00654](https://doi.org/10.1021/acs.jproteome.0c00654).
31. Hadi-Alijanvand, H.; Rouhani, M. Studying the Effects of ACE2 Mutations on the Stability, Dynamics, and Dissociation Process of SARS-CoV-2 S1/hACE2 Complexes. *J. Proteome Res.* **2020**, [10.1021/acs.jproteome.0c00348](https://doi.org/10.1021/acs.jproteome.0c00348).
32. Zeng, X.; Song, X.; Ma, T.; Pan, X.; Zhou, Y.; Hou, Y.; Zhang, Z.; Li, K.; Karypis, G.; Cheng, F. Repurpose Open Data to Discover Therapeutics for COVID-19 Using Deep Learning. *J. Proteome Res.* **2020**, [10.1021/acs.jproteome.0c00316](https://doi.org/10.1021/acs.jproteome.0c00316).
33. Maffucci, I.; Contini, A. In Silico Drug Repurposing for SARS-CoV-2 Main Proteinase and Spike Proteins. *J. Proteome Res.* **2020**, [10.1021/acs.jproteome.0c00383](https://doi.org/10.1021/acs.jproteome.0c00383).
34. Glisic, S.; Perovic, V. R.; Sencanski, M.; Paessler, S.; Veljkovic, V. Biological Rationale for the Repurposing of BCG Vaccine against SARS-CoV-2. *J. Proteome Res.* **2020**, [10.1021/acs.jproteome.0c00410](https://doi.org/10.1021/acs.jproteome.0c00410).
35. Chatterjee, D.; Priyadarshini, P.; Das, D. K.; Mushtaq, K.; Singh, B.; Agrewala, J. N. Deciphering the Structural Enigma of HLA Class-II Binding Peptides for Enhanced Immunoinformatics-based Prediction of Vaccine Epitopes. *J. Proteome Res.* **2020**, [10.1021/acs.jproteome.0c00405](https://doi.org/10.1021/acs.jproteome.0c00405).

36. Martin, W. R.; Cheng, F. Repurposing of FDA-Approved Toremifene to Treat COVID-19 by Blocking the Spike Glycoprotein and NSP14 of SARS-CoV-2. *J. Proteome Res.* **2020**, [10.1021/acs.jproteome.0c00397](https://doi.org/10.1021/acs.jproteome.0c00397).

37. Kumar, N.; Sood, D.; van der Spek, P. J.; Sharma, H. S.; Chandra, R. Molecular Binding Mechanism and Pharmacology Comparative Analysis of Noscapine for Repurposing against SARS-CoV-2 Protease. *J. Proteome Res.* **2020**, [10.1021/acs.jproteome.0c00367](https://doi.org/10.1021/acs.jproteome.0c00367).

38. Chien, M.; Anderson, T. K.; Jockusch, S.; Tao, C.; Li, X.; Kumar, S.; Russo, J. J.; Kirchdoerfer, R. N.; Ju, J. Nucleotide Analogues as Inhibitors of SARS-CoV-2 Polymerase, a Key Drug Target for COVID-19. *J. Proteome Res.* **2020**, [10.1021/acs.jproteome.0c00392](https://doi.org/10.1021/acs.jproteome.0c00392).

39. Zhao, Z.; Bourne, P. E. Structural Insights into the Binding Modes of Viral RNA-Dependent RNA Polymerases Using a Function-Site Interaction Fingerprint Method for RNA Virus Drug Discovery. *J. Proteome Res.* **2020**, [10.1021/acs.jproteome.0c00623](https://doi.org/10.1021/acs.jproteome.0c00623).

40. Kimhofer, T.; Lodge, S.; Whiley, L.; Gray, N.; Loo, R. L.; Lawler, N. G.; Nitschke, P.; Bong, S.-H.; Morrison, D. L.; Begum, S.; Richards, T.; Yeap, B. B.; Smith, C.; Smith, K. G. C.; Holmes, E.; Nicholson, J. K. Integrative Modeling of Quantitative Plasma Lipoprotein, Metabolic, and Amino Acid Data Reveals a Multiorgan Pathological Signature of SARS-CoV-2 Infection. *J. Proteome Res.* **2020**, [10.1021/acs.jproteome.0c00519](https://doi.org/10.1021/acs.jproteome.0c00519).

41. Achutha, A. S.; Pushpa, V. L.; Suchitra, S. Theoretical Insights into the Anti-SARS-CoV-2 Activity of Chloroquine and Its Analogs and In Silico Screening of Main Protease Inhibitors. *J. Proteome Res.* **2020**, [10.1021/acs.jproteome.0c00683](https://doi.org/10.1021/acs.jproteome.0c00683).

42. Saunders, J. K.; Gaylord, D. A.; Held, N. A.; Symmonds, N.; Dupont, C. L.; Shepherd, A.; Kinkade, D. B.; Saito, M. A. METATRYP v 2.0: Metaproteomic Least Common Ancestor Analysis for Taxonomic Inference Using Specialized Sequence Assemblies—Standalone Software and Web Servers for Marine Microorganisms and Coronaviruses. *J. Proteome Res.* **2020**, [10.1021/acs.jproteome.0c00385](https://doi.org/10.1021/acs.jproteome.0c00385).

Suman S. Thakur  orcid.org/0000-0001-5928-8836

■ AUTHOR INFORMATION

Complete contact information is available at:

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