GENOME SEQUENCES

Complete Genome Sequence of Streptomyces sp. Strain SGAir0924, an Actinobacterium Isolated from Outdoor Air in Singapore

Anjali Bansal Gupta,a Akira Uchida,a [Rikky W. Purbojati,](https://orcid.org/0000-0002-2790-4056)a Anthony Wong,a Kavita K. Kushwaha,a Alexander Putra,a Balakrishnan N. V. Premkrishnan,^a Cassie E. Heinle,^a Merrilyn Eng,^a Vineeth Kodengil Vettath,^a D[Ana Carolina M. Junqueira,](https://orcid.org/0000-0003-2382-9842)^b **Daniela I. Drautz-Moses,a Stephan C. Schustera**

aSingapore Centre for Environmental Life Sciences Engineering, Nanyang Technological University, Singapore, Singapore ^bDepartamento de Genética, Instituto de Biologia, Universidade Federal do Rio de Janeiro, Rio de Janeiro, Brazil

ABSTRACT Streptomyces sp. strain SGAir0924 was isolated from outdoor air collected in Singapore. Its genome was assembled using long reads generated by single-molecule real-time sequencing. The final assembly had one chromosome of 7.65 Mb and three plasmids with an average length of 142 kb. The genome contained 6,825 protein-coding genes, 68 tRNAs, and 18 rRNAs.

*S*treptomyces spp. are Gram-positive filamentous bacteria belonging to the phylum Actinobacteria. They are well known for the production of a large variety of natural antibiotics and antifungal and antiparasitic compounds [\(1\)](#page-1-0). More than 600 species of Streptomyces bacteria have been recorded, with the majority of them being recognized as important producers of bioactive compounds [\(2,](#page-1-1) [3\)](#page-1-2).

Streptomyces spp. primarily inhabit soil and water [\(4,](#page-1-3) [5\)](#page-1-4). Here, we report a new strain, Streptomyces sp. strain SGAir0924, isolated from outdoor air in Singapore at global positioning system coordinates 1.35°N, 103.68°E. Air was sampled by impaction onto an electrostatic filter attached on an SASS 3100 dry air sampler (Research International, USA). After sampling, the filter was soaked in phosphate-buffered saline containing 0.1% Triton X-100 to suspend the captured particles. The suspension was then plated onto marine agar (Becton, Dickinson, USA), followed by aerobic incubation at 30°C. Colonies were repeatedly picked and plated onto malt extract agar to obtain clonal colonies. For genomic DNA extraction, a single colony was then inoculated in lysogeny broth (Becton, Dickinson) and incubated at 30°C. DNA extraction was performed with the Wizard genomic DNA purification kit (Promega, USA), following the manufacturer's instructions. The extracted genomic DNA was subjected to library preparation with the SMRTbell template preparation kit 1.0 (Pacific Biosciences, USA), following the manufacturer's 20-kb template preparation using the BluePippin size selection system protocol. The finished library was then sequenced on the RS II single-molecule long-read sequencing platform (Pacific Biosciences). In total, 45,198 subreads with a mean length of 9,337 bp were obtained.

The sequenced genome of Streptomyces sp. strain SGAir0924 was de novo assembled using Hierarchical Genome Assembly Process (HGAP) version 3 [\(6\)](#page-1-5) implemented in the PacBio SMRT Analysis package version 2.3.0 using a seed read length of 500 bp. The consensus assembly generated four contigs with a total length of 8,079,654 bp. This included a linear chromosomal contig (7,653,753 bp, 45.3-fold coverage) and a linear plasmid (377,458 bp, 39.6-fold coverage). Two other plasmids (26,627 bp and 21,816 bp) were able to be circularized using Circlator version 1.1.4 [\(7\)](#page-1-6). The mean G-C content of the chromosome was 72.6%. The average nucleotide identity (ANI) using the Microbial Species Identifier (MiSI) [\(8\)](#page-1-7) method revealed Streptomyces silaceus to be the

Citation Gupta AB, Uchida A, Purbojati RW, Wong A, Kushwaha KK, Putra A, Premkrishnan BNV, Heinle CE, Eng M, Vettath VK, Junqueira ACM, Drautz-Moses DI, Schuster SC. 2019. Complete genome sequence of Streptomyces sp. strain SGAir0924, an actinobacterium isolated from outdoor air in Singapore. Microbiol Resour Announc 8:e00899-19. [https://doi.org/10.1128/MRA.00899-19.](https://doi.org/10.1128/MRA.00899-19)

Editor Steven R. Gill, University of Rochester School of Medicine and Dentistry

Copyright © 2019 Gupta et al. This is an openaccess article distributed under the terms of the [Creative Commons Attribution 4.0](https://creativecommons.org/licenses/by/4.0/) [International license.](https://creativecommons.org/licenses/by/4.0/)

Address correspondence to Stephan C. Schuster, [SCSchuster@ntu.edu.sg.](mailto:SCSchuster@ntu.edu.sg)

Received 29 July 2019 **Accepted** 4 August 2019 **Published** 29 August 2019 closest relative, with an identity of 83% and an alignment fraction value of 0.22. However, based on the 16S rRNA gene sequence, the closest known species is Streptomyces sp. strain NEAU-L11, with 100% identity. As such, strain SGAir0924 can only be identified up to the genus but not the species level. Default parameters were used for all programs unless otherwise specified.

The assembled genome was annotated using the NCBI Prokaryotic Genome Annotation Pipeline (PGAP) version 4.2 [\(9\)](#page-1-8). A total of 7,166 genes were predicted, including 6,825 protein-coding genes (PCGs), 18 rRNA operons (5S, 16S, and 23S rRNAs), 68 tRNAs, 3 noncoding RNAs, and 252 pseudogenes. Functional annotation with the Rapid Annotations using Subsystems Technology (RAST) server [\(10](#page-1-9)[–](#page-1-10)[12\)](#page-1-11), using the classic RAST annotation scheme with an option to fix frameshifts, identified a total of 7,054 coding sequences and 85 RNAs covering 445 subsystems. Of those, 49 genes were related to sigmaB stress response regulators. SigmaB is a transcription factor that is switched on under physical stress conditions [\(13,](#page-1-12) [14\)](#page-1-13), which may be related to desiccation in air and the survival of this strain under arid conditions. Furthermore, the biosynthetic potential of this strain predicted with antiSMASH [\(15\)](#page-1-14) resulted in 33 secondary metabolite biosynthetic gene clusters; many of them are related to antimicrobial compounds, such as streptothricin and candicidin.

Data availability. The complete genome sequences of Streptomyces sp. strain SGAir0924 and its plasmids have been deposited in DDBJ/EMBL/GenBank under accession numbers [CP027296,](https://www.ncbi.nlm.nih.gov/nuccore/CP027296) [CP027297,](https://www.ncbi.nlm.nih.gov/nuccore/CP027297) [CP027298,](https://www.ncbi.nlm.nih.gov/nuccore/CP027298) and [CP027299](https://www.ncbi.nlm.nih.gov/nuccore/CP027299) and in the SRA database under accession number [SRR8948646.](https://www.ncbi.nlm.nih.gov/Traces/study/?acc=SRR8948646&go=go)

ACKNOWLEDGMENT

The work was supported by a Singapore Ministry of Education Academic Research Fund tier 3 grant (MOE2013-T3-1-013).

REFERENCES

- 1. Harir M, Bendif H, Bellahcene M, Fortas Z, Pogni R. 2018. Streptomyces secondary metabolites, p 99-122. In Enany S (ed), Basic biology and applications of actinobacteria. IntechOpen, London, United Kingdom.
- 2. Bérdy J. 2005. Bioactive microbial metabolites. J Antibiot (Tokyo) 58: 1–26. [https://doi.org/10.1038/ja.2005.1.](https://doi.org/10.1038/ja.2005.1)
- 3. Chater KF. 2016. Recent advances in understanding Streptomyces. F1000Res 5:2795–2795. [https://doi.org/10.12688/f1000research.9534.1.](https://doi.org/10.12688/f1000research.9534.1)
- 4. Kharel MK, Shepherd MD, Nybo SE, Smith ML, Bosserman MA, Rohr J. 2010. Isolation of Streptomyces species from soil. Curr Protoc Microbiol 19: 10E.4.1–10E.4.5. [https://doi.org/10.1002/9780471729259.mc10e04s19.](https://doi.org/10.1002/9780471729259.mc10e04s19)
- 5. Hakvåg S, Fjærvik E, Josefsen K, Ian E, Ellingsen T, Zotchev S. 2008. Characterization of Streptomyces spp. isolated from the sea surface microlayer in the Trondheim Fjord, Norway. Mar Drugs 6:620 – 635. [https://doi.org/10.3390/md6040620.](https://doi.org/10.3390/md6040620)
- 6. Chin C-S, Alexander DH, Marks P, Klammer AA, Drake J, Heiner C, Clum A, Copeland A, Huddleston J, Eichler EE, Turner SW, Korlach J. 2013. Nonhybrid, finished microbial genome assemblies from long-read SMRT sequencing data. Nat Methods 10:563–569. [https://doi.org/10.1038/](https://doi.org/10.1038/nmeth.2474) [nmeth.2474.](https://doi.org/10.1038/nmeth.2474)
- 7. Hunt M, Silva ND, Otto TD, Parkhill J, Keane JA, Harris SR. 2015. Circlator: automated circularization of genome assemblies using long sequencing reads. Genome Biol 16:294. [https://doi.org/10.1186/s13059-015-0849-0.](https://doi.org/10.1186/s13059-015-0849-0)
- 8. Varghese NJ, Mukherjee S, Ivanova N, Konstantinidis KT, Mavrommatis K, Kyrpides NC, Pati A. 2015. Microbial species delineation using whole genome sequences. Nucleic Acids Res 43:6761– 6771. [https://doi.org/10](https://doi.org/10.1093/nar/gkv657) [.1093/nar/gkv657.](https://doi.org/10.1093/nar/gkv657)
- 9. Tatusova T, DiCuccio M, Badretdin A, Chetvernin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI Prokaryotic Genome Annotation Pipeline. Nucleic Acids Res 44: 6614 – 6624. [https://doi.org/10.1093/nar/gkw569.](https://doi.org/10.1093/nar/gkw569)
- 10. Overbeek R, Olson R, Pusch GD, Olsen GJ, Davis JJ, Disz T, Edwards RA, Gerdes S, Parrello B, Shukla M, Vonstein V, Wattam AR, Xia F, Stevens R. 2014. The SEED and the Rapid Annotation of microbial genomes using Subsystems Technology (RAST). Nucleic Acids Res 42:D206 –D214. [https://](https://doi.org/10.1093/nar/gkt1226) [doi.org/10.1093/nar/gkt1226.](https://doi.org/10.1093/nar/gkt1226)
- 11. Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O. 2008. The RAST server: Rapid Annotations using Subsystems Technology. BMC Genomics 9:75–75. [https://doi.org/10.1186/1471-2164-9-75.](https://doi.org/10.1186/1471-2164-9-75)
- 12. Brettin T, Davis JJ, Disz T, Edwards RA, Gerdes S, Olsen GJ, Olson R, Overbeek R, Parrello B, Pusch GD, Shukla M, Thomason JA, III, Stevens R, Vonstein V, Wattam AR, Xia F. 2015. RASTtk: a modular and extensible implementation of the RAST algorithm for building custom annotation pipelines and annotating batches of genomes. Sci Rep 5:8365– 8365. [https://doi.org/10.1038/srep08365.](https://doi.org/10.1038/srep08365)
- 13. Boylan SA, Redfield AR, Brody MS, Price CW. 1993. Stress-induced activation of the sigma B transcription factor of Bacillus subtilis. J Bacteriol 175:7931. [https://doi.org/10.1128/jb.175.24.7931-7937.1993.](https://doi.org/10.1128/jb.175.24.7931-7937.1993)
- 14. Guldimann C, Boor KJ, Wiedmann M, Guariglia-Oropeza V. 2016. Resilience in the face of uncertainty: sigma factor B fine-tunes gene expression to support homeostasis in Gram-positive bacteria. Appl Environ Microbiol 82:4456. [https://doi.org/10.1128/AEM.00714-16.](https://doi.org/10.1128/AEM.00714-16)
- 15. Blin K, Weber T, Kim HU, Lee SY, Takano E, Breitling R, Shelest E, Wolf T, Chevrette MG, Suarez Duran HG, Kautsar SA, Lu X, Medema MH, Schwalen CJ, Mitchell DA, de los Santos ELC, Nave M, Dickschat JS. 2017. antiSMASH 4.0—improvements in chemistry prediction and gene cluster boundary identification. Nucleic Acids Res 45:W36 –W41. [https://doi](https://doi.org/10.1093/nar/gkx319) [.org/10.1093/nar/gkx319.](https://doi.org/10.1093/nar/gkx319)