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Association of CYP2E1 gene polymorphisms with bladder cancer risk

A systematic review and meta-analysis

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Abstract

Background: Human cytochrome P450 (CYP) is an enzyme responsible for the metabolic activation of many carcinogens, including nitrosamines. *CYP2E1* represents a major CYP isoform and is expressed in the human urothelial cells. Recent studies have investigated the association of *CYP2E1* gene polymorphisms with bladder cancer risk but have shown contradictory results. Hence, we performed a systematic literature review and meta-analysis to assess the association between *CYP2E1* gene polymorphisms and bladder cancer.

Methods: Systematic literature searches were conducted with PubMed, Excerpt Medica Database, Science Direct/Elsevier, China National Knowledge Infrastructure, and the Cochrane Library up to January 2018 for studies that involved the association of *CYP2E1* gene polymorphisms with bladder cancer risk. A meta-analysis was performed with Review Manager and Stata software. Combined odds ratios (ORs) were identified with 95% confidence intervals (CIs) in a random or fixed effects model.

Ethics: The protocol was approved by the institutional review board of each study center. Written informed consent will be obtained from all patients before registration, in accordance with the Declaration of Helsinki.

Results: Eight studies were identified, including 1733 cases of bladder cancer and 1814 normal controls. Our results illustrated that there are significant associations between *CYP2E1* gene polymorphisms and bladder cancer in all genetic models (P < .05). The combined ORs and 95% CIs were as follows for each model: additive model [OR 0.56; 95% CI (0.38–0.82)]; dominant model [OR 0.79; 95% CI (0.67–0.93)]; recessive model [OR 0.61; 95% CI (0.41–0.89)]; codominant model [OR 0.80; 95% CI (0.67–0.96)]; allele model [OR 0.75; 95% CI (0.59–0.95)]. A subgroup study showed that there are also significant associations between *CYP2E1* gene polymorphisms and bladder cancer in Asian people. However, there are no significant associations between *CYP2E1* gene polymorphisms and bladder cancer in Caucasian populations.

Conclusions: The present study provides evidence for an association between *CYP2E1* gene polymorphisms and bladder cancer progression, and suggests that *CYP2E1* gene polymorphisms might be a protective factor against bladder cancer in Asian people. However, studies with larger sample sizes are needed to confirm the correlation between *CYP2E1* gene polymorphisms and bladder cancer.

Abbreviations: CI = confidence interval, CYP = cytochrome P450, OR = odds ratio, SNP = single nucleotide polymorphism. **Keywords:** bladder cancer, *CYP2E1* gene polymorphisms, human cytochrome P450, meta-analysis, systematic review

1. Introduction

Globally, bladder cancer is the 10th most common cancer and accounts for 3.3% of all malignancies,^[1,2] with the highest

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Received: 7 April 2018 / Accepted: 24 July 2018 http://dx.doi.org/10.1097/MD.000000000011910 incidence rates reported in Europe, North America, and Australia.^[3,4] Its incidence and mortality is the highest among urinary-system tumors, and the majority (approximately 70%) of cases occurs in men.^[5] The risks of bladder cancer are associated with tobacco smoking, some industrially related carcinogenic compounds amines, and amides, and some anticancer drugs such as phosphoramide mustards.^[6,7] However, not all people exposed to these risk factors are develops bladder cancer, suggests that the variation in individual susceptibility may play an important role to bladder carcinogenesis.

Currently, many studies have reported that genetic factors may play an important role in the risk of developing bladder cancer; many ultimate carcinogens and their genotoxicity functions require enzymatic bioactivation.^[10,11] Therefore, phase I and phase II metabolizing enzymes genes may be an important risk factors of developing cancer. Human cytochrome P450 (CYP) is a phase I enzyme play an important role for the metabolic activation of many procarcinogens.^[12]*CYP2E1* represents a major CYP isoform, and in the liver and to lesser extent in other organs and tissues, including human urothelial cells are constitutively expressed this gene.^[13] Many low-molecular-

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Figure 1. Flow diagram of the selection of eligible studies. CNKI = China National Knowledge Infrastructure, EMBASE = Excerpt Medica Database.

weight carcinogens, such as vinyl chloride, benzene, and tobaccospecific nitrosamines require this critical enzyme to metabolic activation.^[14,15]

However, although recent studies have investigated the association of CYP2E1 gene polymorphisms with bladder cancer risk, the reported results were inconsistent, with some studies reporting that CYP2E1 gene polymorphisms may increase the risk of bladder cancer, but not others.^[16–23] Therefore, we systematically reviewed the available literature and performed a metaanalysis to evaluate the association of CYP2E1 gene polymorphisms with bladder cancer risk, which might shed valuable insights on our understanding of the biology of bladder cancer.

2. Methods

2.1. Literature search

This meta-analysis was restricted to published studies that investigated *CYP2E1* gene polymorphisms and bladder cancer risk. Two independent reviewers searched PubMed, Excerpt Medica Database, Science Direct/Elsevier, MEDLINE, China National Knowledge Infrastructure, and the Cochrane Library from inception to January 2018; the language or study type was not restricted. The search terms combined text words and MeSH terms. For example, the search terms for *CYP2E1* gene were "cytochrome P450 family" or "cytochrome P450 2E1" or "cyp2E1," "cyp2e1" or "CYP2E1";

Table 1				
Characteristics	of	the	included	studi

Characteristics of the included studies.															
		Case							Control						
Author	Country	Ν	C1	C2	C1C1	C1C2	C2C2	n	C1	C2	C1C1	C1C2	C2C2	Genotyping method	Race
Basma, 2013	Lebanon	45	74	16	36	2	7	85	104	66	46	12	27	PCR-RFLP	Asia
Brockmöller, 1996	Germany	372	730	14	358	14	7	348	676	20	328	20	10	PCR-RFLP	Caucasian
Cantor, 2010	Spain	649	1196	127	528	118	3	645	1172	130	533	116	7	PCR-RFLP	Caucasian
Choi, 2003	Korea	214	334	94	124	86	4	194	275	113	93	89	12	PCR-RFLP	Asia
Hao, 2001	China	69	138	0	69	0	0	88	176	0	88	0	0	PCR-RFLP	Asia
Mittal, 2005	India	50	100	0	50	0	0	50	100	0	50	0	0	PCR-RFLP	Asia
Shao, 2008	China	202	324	80	131	62	9	272	431	113	170	91	11	PCR-RFLP	Asia
Zhou, 2007	China	132	210	54	92	26	14	132	198	66	82	34	16	PCR-RFLP	Asia

PCR-RFLP = polymerase chain reaction-restriction fragment length polymorphism.

terms for bladder cancer included "bladder cancer" or "bladder neoplasms" or "cancer of bladder" or "neoplasms, bladder"; search terms for polymorphism included "SNP" or "single nucleotide polymorphism" or "polymorphism" or "variation" or "mutation." All related articles and abstracts were retrieved. In addition, references cited within relevant reviews were retrieved by hand, and only full articles were searched.

2.2. Eligibility criteria

2.2.1. Inclusion criteria. Included studies tested the association of CYP2E1 gene polymorphisms with bladder cancer. The case groups were patients with bladder cancer and the controls were normal people. Genotyping for SNPs of CYP2E1 was conducted using polymerase chain reaction restriction fragment length

polymorphism. Available data were extracted from the article, including eligible and genotyped cases and controls, and number of cases and controls for each *CYP2E1* genotype.

2.2.2. Exclusion criteria. Studies were excluded if they were case reports, meeting reports or review articles, published only in abstract form, included no control population, reported no available genotype frequency, or were a duplication of previous publications.

2.3. Study selection and validity assessment

Two independent reviewers screened the titles and abstracts of all citations from the literature search. All relevant studies that appeared to meet the eligibility criteria were retrieved. If an



Figure 2. Forest plot showing the meta-analysis outcomes of the additive model. Cl = confidence interval.

ambiguous decision was made based on the title and abstract, full texts were needed for the analysis. The final decision of eligible studies was made by reviewing the articles. Disagreements were resolved by consensus or a third reviewer.

2.4. Data extraction and statistical analysis

Data included demographic data (authors, year of publication, country, number, genotyping methods) and outcome data of eligible and genotyped cases and controls, plus number of cases and controls for each *CYP2E1* genotype. Three reviewers extracted data from the studies. Disagreements were resolved by consensus. A quantitative meta-analysis was performed by 2 reviewers using Review Manager software (version 5.2, The

Nordic Cochrane Centre, The Cochrane Collaboration, 2012, Copenhagen, Denmark) and Stata software (version 12.0, College Station, TX). Available data were analyzed in the meta-analysis.

To calculate combined odds ratio (OR) and 95% confidence intervals (CIs), heterogeneity was assessed by the *P* value and the *I* square statistic (I^2) in the pooled analyses, which represented the percentage of total variation across studies. If the *P* value was <.1 or the I^2 value was >50%, the summary estimate was analyzed in a random-effects model. Otherwise, a fixed-effects model was applied. We investigated the association between *CYP2E1* gene polymorphisms and bladder cancer risk in allelic [allele (C2 vs C1)], additive (C2C2 vs C1C1), dominant (C2C2 and C2C1 vs C1C1), recessive (C2C2 vs C2C1



Figure 3. Forest plot showing the meta-analysis outcomes of the codominant model. CI = confidence interval.

and C1C1), and codominant models (C2C1 vs C1C1). In addition, publication biases were detected by visual symmetry of funnel plots, with asymmetry suggesting possible publication bias. If the *P*-value was less than 0.05, publication bias was determined to exist.

3. Results

3.1. Characteristics of the included studies

Figure 1 shows a detailed review process. A total of 701 unduplicated studies were identified, 8 studies were ultimately selected according to eligibility criteria, and all reviewers were in agreement regarding the inclusion of all 6 articles.

Table 1 summarizes general data from the 6 studies. All retrieved studies involved 1733 patients with bladder cancer and 1814 normal controls. All of these studies reported exclusion/inclusion criteria,^[16–23] and all tested for *CYP2E1* gene polymorphisms using restriction fragment-lengthpolymorphism analysis after polymerase chain reaction amplification.

3.2. Meta-analysis

The test of heterogeneity suggested that data of the allele model were analyzed in a random-effects model, and the recessive, additive, dominant, and codominant models were analyzed in a fixed-effects model. Meta-analysis revealed that there were

	Case Control		ol		Odds Ratio	Odds Ratio			
Study or Subgroup	Events	Total	Events	Total	Weight	M-H, Fixed, 95% Cl	M-H, Fixed, 95% Cl		
Basma HA et al. 2013	9	45	39	85	7.1%	0.29 [0.13, 0.69]	· · · · · · · · · · · · · · · · · · ·		
Brockmoller J et al. 1996	21	372	30	348	9.6%	0.63 [0.36, 1.13]			
Cantor KP et al. 2010	121	649	123	645	33.0%	0.97 [0.74, 1.29]			
Choi JY et al. 2003	90	214	101	194	20.2%	0.67 [0.45, 0.99]			
Hao YG et al.2001	0	69	0	88		Not estimable			
Mittal RD et al. 2005	0	50	0	50		Not estimable			
Shao J et al. 2008	71	202	102	272	18.6%	0.90 [0.62, 1.32]			
Zhou CH et al. 2007	40	132	50	132	11.5%	0.71 [0.43, 1.19]			
Total (95% CI)		1733		1814	100.0%	0.79 [0.67, 0.93]	•		
Total events	352		445						
Heterogeneity: Chi ² = 9.25	df = 5 (P)	= 0.10)	I ² = 46%	2					
Test for overall effect: $Z = 2$	77 (P = 0)	006)		2			0.2 0.5 1 2 5		
		,					Case Control		
6	Cas	e	Contr	ol		Odds Ratio	Odds Ratio		
Study or Subaroup	Events	Total	Events	Total	Weight	M-H. Fixed, 95% Cl	M-H. Fixed, 95% Cl		
2.2.1 Asia									
Basma HA et al. 2013	9	45	39	85	7.1%	0.29 (0.13, 0.69)			
Choi JY et al. 2003	90	214	101	194	20.2%	0.67 [0.45, 0.99]			
Hao YG et al 2001	0	69	0	88		Not estimable			
Mittal RD et al. 2005	Ő	50	Ő	50		Not estimable			
Shan Jet al. 2008	71	202	102	272	18.6%	0.90 (0.62, 1.32)			
Zhou CH et al. 2007	40	132	50	132	11.5%	0 71 10 43 1 191			
Subtotal (95% CI)		712		821	57.3%	0.71 [0.56, 0.89]	•		
Total events	210		292						
Heterogeneity: Chi ² = 5 79	df = 3/P	= 0 1 2)	· 12 = 48%	<u>(</u>					
Test for overall effect: $Z = 2$	2.96 (P = 0	.003)							
2.2.2 Caucasian									
Brockmoller J et al. 1996	21	372	30	348	9.6%	0.63 [0.36, 1.13]			
Cantor KP et al. 2010	121	649	123	645	33.0%	0.97 [0.74, 1.29]	+		
Subtotal (95% CI)		1021		993	42.7%	0.90 [0.70, 1.15]	+		
Total events	142		153						
Heterogeneity: Chi ² = 1.71	df = 1 (P)	= 0.19)	: I ² = 41%						
Test for overall effect: Z = 0).86 (P = 0	.39)							
Total (95% CI)		1733		1814	100.0%	0.79 [0.67. 0.93]	•		
Total events	352		445	Gentle (CB)					
Heterogeneity: Chi ² = 9.25	, df = 5 (P = 0	= 0.10) 006)	; I ^z = 46%	6					
Test for subaroup differen	ces: Chi ² =	1.87.	df = 1 (P :	= 0.17)	² = 46.6	%	Case Control		
3									

Figure 4. Forest plot showing the meta-analysis outcomes of the dominant model. Cl = confidence interval.

significant associations between *CYP2E1* gene polymorphisms and bladder cancer in all genetic models (P < .05). The combined OR and its 95% CIs were as follows for each model: additive model [OR 0.56; 95% CI (0.38–0.82)] (Fig. 2A); codominant model [OR 0.80, 95% CI (0.67–0.96)] (Fig. 3A); dominant model [OR 0.79; 95% CI (0.67–0.93)] (Fig. 4A); recessive model [OR 0.61; 95% CI (0.41–0.89)] (Fig. 5A); allele model [ORs 0.75; 95% CI (0.59–0.95)] (Fig. 6A). Subgroup study showed that there were also significant associations between *CYP2E1* gene polymorphisms and bladder cancer in Asian people (P < .05). However, there were no significant associations between *CYP2E1* gene polymorphisms and bladder cancer in Caucasian individuals (P > .05) (Fig. 2B, 3B, 4B, 5B, 6B). Begg funnel plots were largely symmetric (Fig. 7A, 8A, 9A, 10A, 11A), suggesting that there were no publication biases in the meta-analysis. In order to evaluate the stability and reliability of the meta-analysis, we conducted a sensitivity analysis. We sequentially omitted 1 study, and calculated the combined ORs of the remaining studies, but the final conclusions were not changed [there were significant associations between *CYP2E1* gene polymorphisms and bladder cancer in all genetic models (P < .05)], which suggest that the results were statistically stable and reliable (Fig. 7B, 8B, 9B, 10B, 11B).



Figure 5. Forest plot showing the meta-analysis outcomes of the recessive model. CI = confidence interval.

	Case		Control			Odds Ratio	Odds Ratio	
Study or Subgroup	Events	Total	Events	Total	Weight	M-H, Random, 95% Cl	M-H, Random, 95% C	
Basma HA et al. 2013	16	90	66	170	10.2%	0.34 [0.18, 0.63]		
Brockmoller J et al. 1996	14	744	20	696	8.8%	0.65 [0.32, 1.29]		
Cantor KP et al. 2010	127	1323	130	1302	23.4%	0.96 [0.74, 1.24]		
Choi JY et al. 2003	94	428	113	388	20.6%	0.68 [0.50, 0.94]		
Hao YG et al.2001	0	138	0	176		Not estimable		
Mittal RD et al. 2005	0	100	0	100		Not estimable		
Shao J et al. 2008	80	404	113	544	20.4%	0.94 [0.68, 1.30]		
Zhou CH et al. 2007	54	264	66	264	16.6%	0.77 [0.51, 1.16]		
						8. 15 14	3	
Total (95% CI)		3491		3640	100.0%	0.75 [0.59, 0.95]	•	
Total events	385		508					
Heterogeneity: Tau ² = 0.05	; Chi ² = 11	.41, df	= 5 (P = 1	0.04); P	² = 56%			-
Test for overall effect: Z = 2	.37 (P = 0	.02)					0.2 0.5 1 2 5	1
A							Case Control	
	Cas	е	Contr	ol		Odds Ratio	Odds Ratio	
Study or Subgroup	Events	Total	Events	Total	Weight	M-H, Random, 95% Cl	M-H, Random, 95% Cl	
2.5.1 Asia								
Basma HA et al. 2013	16	90	66	170	10.2%	0.34 [0.18, 0.63]		
Choi JY et al. 2003	94	428	113	388	20.6%	0.68 [0.50, 0.94]		
Hao YG et al.2001	0	138	0	176		Not estimable		
Mittal RD et al. 2005	0	100	0	100		Not estimable		
Shao J et al. 2008	80	404	113	544	20.4%	0.94 [0.68, 1.30]		
Zhou CH et al. 2007	54	264	66	264	16.6%	0.77 [0.51, 1.16]		
Subtotal (95% CI)		1424		1642	67.8%	0.69 [0.50, 0.96]	•	
Total events	244		358					
Heterogeneity: Tau ² = 0.07	; Chi ² = 8.	41, df =	3 (P = 0.	.04); 17:	= 64%			
Test for overall effect: Z = 2	.20 (P = 0	.03)						
2.5.2 Caucasian								
Brockmoller J et al. 1996	14	744	20	696	8.8%	0.65 [0.32, 1.29]		
Cantor KP et al. 2010	127	1323	130	1302	23.4%	0.96 [0.74, 1.24]		
Subtotal (95% CI)		2067		1998	32.2%	0.90 [0.69, 1.19]	•	
Total events	141		150					
Heterogeneity: Tau ² = 0.01	; Chi ² = 1.	07, df=	1 (P = 0	30); I ^z :	= 7%			
Test for overall effect: $Z = 0$.73 (P = 0	.46)						
Total (95% CI)		3491		3640	100.0%	0.75 [0.59, 0.95]	•	
Total events	385		508				(2 c) (2)	
Heterogeneity: Tau ² = 0.05; Chi ² = 11.41, df = 5 (P = 0.04); l ² = 56%								
Test for overall effect: Z = 2	Case Control	1						
Test for subaroup difference	es: Chi ² =	1.54.	df = 1 (P :	= 0.22)	I ² = 34.9	%	0436 001401	
В								

Figure 6. Forest plot showing the meta-analysis outcomes of the allele model. CI = confidence interval.

4. Discussion

In our study, 8 reports that studied the association of *CYP2E1* gene polymorphisms with bladder cancer risk were analyzed. For the additive, dominant, recessive, codominant, and allele models, 1, 2, 2, 1, and 2 studies, respectively, reported a significant association between *CYP2E1* gene polymorphisms and bladder cancer. Our results revealed that, on the whole, significant associations between *CYP2E1* gene polymorphisms and bladder cancer were found in all genetic models. The subgroup study showed that there are also significant associations between *CYP2E1* gene polymorphisms and bladder cancer in Asian populations. However, no significant associations between

CYP2E1 gene polymorphisms and bladder cancer were identified in Caucasian populations. Thus, these data indicate that CYP2E1 gene polymorphisms might be a protective factor against bladder cancer in Asian individuals.

The CYP superfamily is an important enzyme and plays a considerable role in the metabolism of endogenous and exogenous compounds, such as numerous carcinogens such as nitrosamines. The enzyme also produce reactive free radicals through oxidation of other compounds, such as ethanol, and may initiate lipid peroxidation and consequently carcinogenesis.^[24,25] CYP2E1 represents a major CYP isoform, in the liver, kidney, and other organs and tissues, such as urothelial cells are













constitutively expressed this gene.^[13] It participates in phase I metabolic oxidization and activates many carcinogenic compounds, such as alkanes, alkenes, and aromatic and halogenated hydrocarbons.^[26,27]

There is strong experimental evidence that CYP2E1 is an important activator of carcinogenic compounds.^[28,29]

Several restriction fragment length polymorphisms of the *CYP2E1* have been identified^[30] and a variant C2 allele recognized by *RsaI* digestion in the 5'-flanking region of the gene appears to be associated with decreased enzyme activity or noninducibility,^[31] some in vitro studies showed that the C2 allele decreased the expression of a reporter gene construct.^[32,33] Our results showed that the C2 allele distribution frequency in the control group was significantly higher than that in patients with bladder cancer, suggesting that *CYP2E1* gene polymorphisms might be a protective factor against bladder cancer.

The potential molecular basis for the association between the *CYP2E1* C1/C1 genotype and bladder cancer risk may be related to the expression levels of the gene. The *CYP2E1* C1/C1 genotype shows higher transcriptional activity compared with the *CYP2E1* C2/C2 genotype.^[34] Therefore, upon exposure, the enzymatic

activity of bladder procarcinogens in C2/C2 subjects may compete poorly compared with C1/C1,^[35–37] leading to increased cancer susceptibility. In our subanalysis, there were no significant associations between *CYP2E1* gene polymorphisms and bladder cancer in Caucasian people. This may be because of racial differences in polymorphism distribution.^[38,39] Another study found that the frequency of the C2 allele in Asian populations is higher than that in Caucasians.^[40] However, this hypothesis requires further investigation before drawing conclusions.

There are some limitations in our study, which need to be taken into consideration when interpreting the results of this metaanalysis. First, the sample size of each study was relatively small, with a total of 1733 patients with bladder cancer and 1814 normal controls investigated in all 8 studies; furthermore, only 2 studies were selected in our subanalysis study. Second, several studies related to the subject were excluded because of a lack of control data. As such, it is hard to make definitive conclusions about the clinical value of *CYP2E1* gene variants and bladder cancer.

In summary, the results of this meta-analysis suggest that the current article adds to the evidence of an association between *CYP2E1* gene polymorphisms and bladder cancer progression. These data suggest that *CYP2E1* gene polymorphisms might be a protective factor against bladder cancer in Asian people. The possible mechanism may occur via *CYP2E1* gene C2 allele mutations, weakening the expression of the gene and enzymatic bioactivation procarcinogens to become ultimate carcinogens and cause genotoxicity. However, studies with larger sample sizes are needed to definitively determine the correlation between *CYP2E1* gene polymorphisms and bladder cancer.

Author contributions

Conceptualization: Xiangrui Yin, Shengqiang Qian, Yu Guo.

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References

- Ferlay J, Shin H, Bray F, et al. Estimates of worldwide burden of cancer in 2008: GLOBOCAN 2008. Int J Cancer 2010;127:2893–917.
- [2] Ploeg M, Aben KK, Kiemeney LA. The present and future burden of urinary bladder cancer in the world. World J Urol 2009;27:289–93.
- [3] Ploeg M, Aben K, Kiemeney L. The present and future burden of urinary bladder cancer in the world. World J Urol 2009;27:289–93.
- [4] Jemal R, Bray F, Center M, et al. Global cancer statistics. CA Cancer J Clin 2011;61:69–90.
- [5] Parkin DM. International variation. Oncogene 2004;23:6329-40.
- [6] Cohen SM, Shirai T, Steineck G. Epidemiology and etiology of premalignant and malignant urothelial changes. Scand J Urol Nephrol Suppl 2000;205:105–15.
- [7] Kiriluk KJ, Prasad SM, Patel AR, et al. Bladder cancer risk from occupational and environmental exposures. Urol Oncol 2012;30:199–211.
- [8] Martiny VY, Miteva MA. Advances in molecular modeling of human cytochrome P450 polymorphism. J Mol Biol 2013;425:3978–92.
- [9] Stiborova M, Levova K, Barta F, et al. Bioactivation versus detoxication of the urothelial carcinogen aristolochic acid I by human cytochrome P450 1A1 and 1A2. Toxicol Sci 2012;125:345–58.
- [10] Sato M, Sato T, Izumo T, et al. Genetic polymorphism of drugmetabolizing enzymes and susceptibility to oral cancer. Carcinogenesis 1999;20:1927–31.
- [11] Yue J, Peng R, Chen J, et al. Effects of rifampin on CYP2E1-dependent hepatotoxicity of isoniazid in rats. Pharmacol Res 2009;59:112–9.
- [12] Guengerich FP. Molecular advances for the cytochrome P450 superfamily. Trends Pharmacol Sci 1991;12:281–3.
- [13] Sheweita SA, Abu El-Maati MR, El-Shahat FG, et al. Changes in the expression of cytochrome P450 2E1 and the activity of carcinogen metabolizing enzymes in Schistosoma haematobium-infected human bladder tissues. Toxicology 2001;162:43–52.
- [14] Hou DF, Wang SL, He ZM, et al. Expression of CYP2E1 in human nasopharynx and its metabolic effect in vitro. Mol Cell Biochem 2007;298:93–100.
- [15] Feng J, Pan X, Yu J, et al. Functional Pstl/RsaI polymorphism in CYP2E1 is associated with the development, progression and poor outcome of gastric cancer. PLoS One 2012;7:e44478.

- [16] Cantor KP, Villanueva CM, Silverman DT. Polymorphisms in GSTT1, GSTZ1, and CYP2E1, disinfection by-products, and risk of bladder cancer in Spain. Environ Health Perspect 2010;118:1545–50.
- [17] Mittal RD, Srivastava DS. Genetic polymorphism of drug metabolizing enzymes (CYP2E1, GSTP1) and susceptibility to bladder cancer in North India. Asian Pac J Cancer Prev 2005;6:6–9.
- [18] Zhou CH. The research on the polymorphism of CYP2E1, GSTM1 and the susceptibility of bladder cancer. J Changchun Jilin Univ 2007;12:221–6.
- [19] Hao YG. The research on bladder tumor metabolic enzyme gene polymorphism and the cell cycle protein. J Jinan 2001;14:1216–23.
- [20] Shao J, Gu M, Zhang Z. Genetic variants of the cytochrome P450 and glutathione S-transferase associated with risk of bladder cancer in a south-eastern Chinese population. Int J Urol 2008;15:216–21.
- [21] Choi JY, Lee KM, Cho SH, et al. CYP2E1 and NQO1 genotypes, smoking and bladder cancer. Pharmacogenetics 2003;13:349–55.
- [22] Brockmöller J, Cascorbi I, Kerb R, et al. Combined analysis of inherited polymorphisms in arylamine N-acetyltransferase 2, glutathione S-transferases M1 and T1, microsomal epoxide hydrolase, and cytochrome P450 enzymes as modulators of bladder cancer risk. Cancer Res 1996;56:3915–25.
- [23] Basma HA, Kobeissi LH, Jabbour ME, et al. CYP2E1 and NQO1 genotypes and bladder cancer risk in a Lebanese population. Int J Mol Epidemiol Genet 2013;4:207–17.
- [24] Yamazaki H, Inui Y, Yun CH, et al. Cytochrome P450 2E1 and 2A6 enzymes as major catalysts for metabolic activation of N-nitrosodialkylamines and tobacco-related nitrosamines in human liver microsomes. Carcinogenesis 1992;13:1789–94.
- [25] Wolf CR. Cytochrome P450s: a multigene family involved in carcinogen metabolism. Trends Genet 1986;2:209–14.
- [26] Wolbold R, Klein K, Burk O, et al. Sex is a major determinant of CYP3A4 expression in human liver. Hepatology 2003;38:978–88.
- [27] Guengerich FP. Characterization of the roles of human cytochrome P450 enzymes in carcinogen metabolism. Asia Pacific J Pharmacol 1990;5:327–45.
- [28] Guengerich FP, Kim DH, Iwasaki M. Role of human cytochrome P450 IIEI in the oxidation of many low molecular weight cancer suspects. Chem Res Toxicol 1991;4:168–79.
- [29] Mannervik B, Awasthi YC, Board PG, et al. Nomenclature for human glutathione transferases. Biochem J 1992;282:305–6.
- [30] Hu Y, Oscarson M, Johansson I, et al. Genetic polymorphism of human CYP2E1: characterization of two variant alleles. Mol Pharmacol 1997;51:370–6.
- [31] Lin D, Tang Y, Peng Q. Genetic polymorphisms of cytochrome P450 2E1 and glutathione S-transferase P1 and susceptibility to esophageal cancer. Chin J Oncol 1998;20:14–7.
- [32] Watanabe J, Hayashi S, Kawajiri K. Different regulation and expression of the human CYP2E1 gene due to the RsaI polymorphism in the 5'flanking region. J Biochem (Tokyo) 1994;116:321–6.
- [33] Tan W, Song N, Wang GQ. Impact of genetic polymorphisms in cytochrome P450 2E1 and glutathione S-transferases M1, T1, and P1 on susceptibility to esophageal cancer among high-risk individuals in China. Cancer Epidemiol Biomarkers Prev 2000;9:551–6.
- [34] Hayashi S, Watanabe J, Kawajiri K. Genetic polymorphisms in the 5'flanking region change transcriptional regulation of the human cytochrome P450IIE1 gene. J Biochem 1991;110:559–65.
- [35] Hayashi SI, Watanabe J, Kawajiri K. Genetic polymorphisms in the 5'flanking region change translational regulation of the human cytochrome P45011E1 gene. J Biochem 1991;110:559–65.
- [36] Marchand LL, Wilkinson GR, Wilkens LR. Genetic and dietary predictors of CYP2E1 activity: a phenotyping study in Hawaii Japanese using chlorzoxazone. Cancer Epidemiol Biomarkers Prev 1999;8:495–500.
- [37] Kim RB, Yamazaki H, Chiba K, et al. In vivo and in vitro characterization of CYP2E1 activity in Japanese and Caucasians. J Pharmacol Exp Ther 1996;279:4–11.
- [38] Wang J, Liu Z, Lu X, et al. Cytochrome P450 1A1 and cytochrome P450 2E1 gene polymorphisms in Guangzhou Hans [in Chinese]. Zhonghua Yi Xue Yi Chuan Xue Za Zhi 2000;17:259–61.
- [39] Kato S, Shields PG, Caporaso NE, et al. Cytochrome P450 2E1 genetic polymorphisms, racial variation, and lung cancer risk. Cancer Res 1992;52:6712–5.
- [40] Stephens EA, Taylor JA, Kaplan N, et al. Ethnic variation in the CYP2E1 gene: polymorphism analysis of 695 African-Americans, European-Americans and Taiwanese. Pharmacogenetics 1994;4:185–92.

Uncited References

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