ORIGINAL ARTICLE

Association study between the polymorphisms of angiogenesisrelated genes and cervical cancer susceptibility in Chinese Uygur population

Lili Han | Sulaiya Husaiyin | Chunhua Ma | Mayinuer Niyazi 🕩

Department of Gynecology, People's Hospital of Xinjiang Uygur Autonomous Region, Urumqi, China

Correspondence

Mayinuer Niyazi, #91 Tianqi Road, Urumqi, Xinjiang 830001, China. Email: mynr68@126.com

Funding information Xinjiang Tianshan Youth Program, Grant/ Award Number: 2017Q008

Abstract

Background: Cervical cancer is the second most common malignant tumor in women, and its invasion and metastasis are regulated by tumor angiogenic growth factors and their cognate receptors. In this study, we explored the relationship between genetic polymorphisms of angiogenesis-related genes (*VEGF-C*, *VEGFR-2*, and *VEGFR-3*) and the risk of cervical cancer in Chinese Uygur population.

Methods: We investigated four single-nucleotide polymorphisms (SNPs) in 342 cervical cancer cases and 498 controls to evaluate their association with the risk of cervical cancer. Their correlations were evaluated by chi-squared test, Fisher's exact test, *t* test, and genetic model analyses. Odds ratios (ORs) and 95% confidence intervals (95% CIs) were calculated using unconditional logistic regression.

Results: We observed that rs12646659 in *VEGF-C* was associated with a lower cervical cancer risk in allele, dominant, and log-additive models (allele: p = .017; dominant: p = .018; log-additive: p = .018). For the individuals older than 43, rs4604006 (*VEGF-C*) was related to an increased cervical cancer risk under codominant model (p = .035), and rs12646659 was significantly associated with a reduced cervical cancer risk in allele, dominant, log-additive models (allele: p = .028; codominant: p = .037; log-additive: p = .037) However, there were no significant correlation of rs1000611 (*VEGFR-2*) and rs1195571 (*VEGFR-3*) with cervical cancer risk in Chinese Uygur population.

Conclusion: Our study firstly provided evidence that rs4604006 and rs12646659 of *VEGF-C* gene were related to the susceptibility of cervical cancer in Chinese Uygur population.

KEYWORDS

cervical cancer, genetics polymorphisms, VEGF-C, VEGFR-2, VEGFR-3

This is an open access article under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

© 2019 The Authors. Molecular Genetics & Genomic Medicine published by Wiley Periodicals, Inc.

WILEY_Molecular Genetics & Genomic Medicine

1 | INTRODUCTION

Cervical cancer is one of the common malignant tumor in women, with 528,000 cases and 266,000 deaths in 2012 (Fang et al., 2017; Niu et al., 2019). The high incidence and mortality of cervical cancer among Uygur women in Xinjiang has become the most important health issue (Lin, Huang, Shen, & Yiming, 2015; Ma, Hong, Lu, Chen, & Ma, 2015). In recent years, the incidence of cervical cancer among women in Xinjiang is on the rise. The chances of the second cure for cervical cancer are very low, and the early detection has a significant impact on the survival of cervical cancer patients (Waggoner, 2003). Increasing growth factors and their homologous receptors have been reported and can regulate the invasion and metastasis of cervical cancer (Tomao et al., 2014). Therefore, the research of these growth factors and their receptors is undoubtedly a great benefit for the treatment of cervical cancer.

Angiogenesis is a pivotal step in tumor formation, growth and metastasis (Wu, 2014). VEGF (vascular endothelial growth factor) is a key angiogenic stimulator, and VEGF signaling pathway has been identified as an important part of angiogenesis (Li, Xu, Gao, Bi, & Huo, 2018). VEGF-C (Vascular Endothelial Growth Factor C; OMIM: 601,528) is an important member of the VEGF family. It encodes proteins that affect angiogenesis, endothelial cell growth, and vascular permeability (Chen et al., 2014). Recent studies have shown that elevated levels of vascular VEGF-C have in many invasive tumors and it is strongly associated with poor prognosis in cancer patients (Cheng, Jiang, Yuan, Liu, & Simoncini, 2018). For example, the level of VEGF-C increases in women with lung carcinoma, and it is significantly associated with lymph node metastasis (Tamura & Ohta, 2003). The high expression of VEGF-C was also observed in cervical cancer (Mitsuhashi et al., 2005). At the same time, VEGF binds to the extracellular receptor domain and promotes the activation of tyrosine kinase in the intracellular receptor domain, thereby phosphorylating tyrosine residues and activating several intracellular signaling pathways (Robinson & Stringer, 2001).

There are three types of VEGF receptors: *VEGFR-1* (vascular endothelial growth factor receptor 1), *VEGFR-2* (vascular endothelial growth factor receptor 2; OMIM: 191,306), and *VEGFR-3* (vascular endothelial growth factor receptor 3; OMIM: 136,352). *VEGFR-2* receptors are predominantly expressed in vascular endothelial cells (Shibuya, 2006), while *VEGFR-3* is especially expressed in endothelial lymphatic cells (Hamrah et al., 2004). The *VEGFR-3* gene, also known as *FLT4* (fms-related tyrosine kinase 4), has a molecular weight of 195 kDa (Takahashi & Shibuya, 2005). *Flt-4* has been found to be expressed in a variety of human malignancies (Su et al., 2006). *VEGFR-2* also known as KDR (kinase insert domain receptor), has a

molecular weight of 230 kDa (Takahashi & Shibuya, 2005). Studies have shown that *VEGF-C* and *VEGF-D* bind to its receptor *VEGFR-2* (*KDR*) and receptor *VEGFR-3* (*Flt-4*), promoting angiogenesis and/or lymphangiogenesis, thus accelerates tumor growth and metastasis (Joukov et al., 1996). These evidences suggested that *VEGF-C*, *VEGFR-2*, and *VEGFR-3* were closely related to the growth and metastasis of cervical cancer.

In this case–control study, we genotyped four SNPs (rs10006115 [VEGFR-2], rs4604006 [VEGF-C], rs12646659 [VEGF-C], and rs11955717 [VEGFR-3]) and performed a comprehensive association analysis to identify whether SNPs were associated with cervical cancer risk in Chinese Uygur population.

2 | MATERIALS AND METHODS

2.1 | Ethics statement

The study was approved by the ethics committee of People's Hospital of Xinjiang Uygur Autonomous Region, in accordance with the principles of the Helsinki Declaration. Each participant was informed of the procedures and purpose of our research and signed a written informed consent before donating 5 ml venous blood for further analyses.

2.2 | Research participates

In this case–control study, 342 cervical cancer patients (mean age, 43.27 ± 11.78 years) were recruited from the Department of Gynecology, People's Hospital of Xinjiang Uygur Autonomous Region, between 2016 and 2019. All patients were diagnosed with cervical cancer by histopathological examination according to the International Federation of Gynecology and Obstetrics (FIGO; Du, Wang, Richards, & Wang, 2019).

The healthy control group consisted of 498 individuals $(43.46 \pm 13.03 \text{ years})$, who were recruited from the Health Examination Center of the People's Hospital of Xinjiang Uygur Autonomous Region during the same time with cases. And these control subjects were comparable to the cervical cancer subjects in terms of age and race. The selection criteria for the control group were as follows: no history of cancer or a family history of cancer, no known history of infectious HPV. Written informed consent was obtained from each participant prior to enrollment in the study.

2.3 | SNP selection and genotyping

Genomic DNA was extracted from participant's peripheral venous blood by a Gold Mag Mini Whole Blood Genomic DNA Purification Kit (GoldMag Ltd) following the manufacturer's protocol and then stored at -80°C before genotyping.

SNP_ID	2nd PCRP	1st PCRP	UEP DIR	UEP SEQ
rs10006115	ACGTTGGATGATGGCAGCCATATGGAGTTG	ACGTTGGATGTCCACTGCAGAAGAAATGGC	R	aggaTCTTCTCTGCCAACTC
rs4604006	ACGTTGGATGACATACTCATGTTTGTACCC	ACGTTGGATGGTGTGGCATGAAACAAAAGC	R	ggagGTTTGTACCCAATCCTTTG
rs11955717	ACGTTGGATGCGTCACCAAACAGCATTTCG	ACGTTGGATGGAGGTTTGAATTTACGTGGC	R	CAAGGTAATTAAGTAAAAGGTGTC
rs12646659	ACGTTGGATGAAGCTTGGCTGTCTCTATGG	ACGTTGGATGATGTTCCCACTGAGAACAAC	R	GTCTCTATGGTTATATCTTCAAATA
Abbreviations: PCR,	polymerase chain reaction; UEP, unextended mini-sequencing prin	ter.		

TABLE 1 Primers used for this study

Molecular Genetics & Genomic Medicine _____ F

The concentration and purity of DNAs were determined by the NanoDrop 2000 (Thermo Scientific).

We established the following criteria to identify the target SNPs: (a) MAF (minor allele frequency) of Han Chinese in Beijing (HCB) > 0.05 and disease relevance in 1,000 genome (http://www.internationalgenome.org/); (b) a linkage disequilibrium value of $r^2 < .8$ for each target SNPs. Agena MassARRAY Assay Design 4.0 software was used to design the primers for amplification and extension reactions. Agena MassARRAY RS1000 was used to perform SNP genotyping according to the standard protocol. Two staffs independently operated genotyping assay and randomly selected more than 10% samples for verification, and the results were exactly same in two sets of assays. Then, Agena Typer 4.0 software was applied to analyze and manage our data. The PCR primers for each SNP are shown in Table 1.

2.4 | Statistical analysis

All statistical analysis was done with SPSS version 19.0 software (SPSS) and Microsoft Excel. All analyses were two sided, and statistical significance was set at p < .05. SNP genotype frequencies in the case and controls were calculated by chi-squared test (Hu, Wang, Hu, & Li, 2018; Yang et al., 2018). Deviation from Hardy–Weinberg equilibrium (HWE) was assessed using the chi-squared test to compare the observed and expected genotype frequencies among the control subjects. Logistic regression analysis was used to examine the odds ratios (ORs) and 95% confidence intervals (95% CIs) in order to assess the association between SNPs and cervical cancer (Bland & Altman, 2000; Liu et al., 2017). Four models (codominant, dominant, recessive, and log-additive) were used to test the association between SNPs and Cervical cancer (Jin et al., 2016; Sole, Guino, Valls, & Iniesta, 2006).

3 | RESULTS

3.1 | Demographic characteristics

The general characteristics were listed in Table 2. Among the 840 participants, 342 were patients with cervical cancer and 498 were healthy controls. The mean age and standard deviation were 43.27 ± 11.78 for cases and 43.46 ± 13.03 for control subjects. There were no significant differences between the cases and controls in terms of age.

3.2 | Hardy–Weinberg equilibrium and SNPs alleles

Basic information containing SNP ID, alleles, role, MAF distribution, p-HWE value, ORs, 95% CIs of all candidate SNPs were presented in Table 3. The call rate for all SNPs was above 95% in cases and controls, which was considered

4 of 10

VII FV_Molecular Genetics & Genomic Medicine

T.	A	B 1	LΕ	2	General characteristics the of this study population	
----	---	------------	----	---	--	--

Variables	Case	Control	Total	<i>p</i> -value
Total	342	498		
Age				
≤43	166 (49%)	235 (47%)	401	>.05
>43	176 (51%)	263 (53%)	439	
Mean age $\pm SD$	43.27 ± 11.78	43.46 ± 13.03		
T stage				
III + IV	80 (23%)			
I + II	132 (39%)			
Unavailable	130 (38%)			

Note: p-values were calculated from two-sided chi-squared test/Fisher's exact test; p < .05 indicates statistical significance.

as high quality to perform association analyses. None of the candidate SNPs significantly deviated from HWE. OR = 1 indicates that the factor had no effect on the disease; OR > 1 means it is a risk factor; and OR < 1 means it is a protective factor. The comparison of allele distributions between the cervical cancer patients and the control subjects with the χ^2 -test revealed that there was a statistical correlation between the rs12646659 polymorphism of *VEGF-C* and the risk reduction of cervical cancer (OR = 0.41, 95% CI = 0.20–0.87; p = .017).

3.3 | Associations between genotype frequencies and cervical cancer risk

Furthermore, we analyzed the association between the SNPs and the risk of cervical cancer under multiple inheritance models (codominant, dominant, recessive, log-additive models; Table 4). Our analyses showed that rs12646659 in *VEGF-C* gene was correlated with a decrease the risk of cervical cancer in the dominant model (OR = 0.40, 95% CI = 0.19–0.86, p = .018 for the "C/G-G/G" genotype) and log-additive model (OR = 0.40, 95% CI = 0.19–0.86, p = .018) before and after adjustment for age, respectively. In addition, we failed to find any significant association between other polymorphisms and the risk of cervical cancer.

3.4 | Stratification analysis by age

To further explore the potential effect of age on the relationship of *VEGF-C*, *VEGFR-2*, and *VEGFR-3* gene polymorphisms with the susceptibility to cervical cancer, we performed the same statistical analysis on the recruited population after stratification of age (Table 5). Among the individuals older than 43, rs4604006 of *VEGF-C* gene was correlated with a 1.55-fold increased the risk of cervical cancer in the codominant model (adjusted, OR = 1.55, 95%

						Frequency (MAF)			
SNP	Chr	Position	Gene(s)	Role	Alleles	Cases	Controls	<i>p</i> - HWE	OR (95% CI)	<i>p</i> -value
rs10006115	4	55,080,187	VEGFR-2	ncRNA_intronic	G/T	0.056	0.049	.107	1.14 (0.74–1.76)	.563
rs4604006	4	176,687,621	VEGF-C	ncRNA_intronic	С/T	0.342	0.365	.067	0.90 (0.74–1.11)	.326
rs12646659	4	176,764,117	VEGF-C	Intronic	C/G	0.013	0.031	1.000	0.41 (0.20-0.87)	.017*
rs11955717	5	180,606,639	VEGFR-3	Intronic	С/T	0.329	0.336	.070	0.97 (0.79–1.19)	.752
hbraviations: 05% C	T 050% confider	nca intervol: Allalae A/B	Minor/Major allelee	". UWF Uardy Wainbara and	uilibrium: MAE	ninor ollala frac	to odde main	o. CND single mole	actide notwornhiem	

Basic information about candidate SNPs and association with the risk of cervical cancer in allele model

c

TABLE

porymorpmsm. *p*-values were calculated with Pearson's χ^2 tests Abb

 $^{*}p$ < .05 indicates statistical significance

Gene SNP Model VEGFR-2 rs10006115 Codominant Dominant			Coso			OD WEEK OD	
<i>VEGFR–2</i> rs10006115 Codominant Dominant	Genotype	Control	Labe	UN (32% CI)	<i>p</i> -value	UK (%% CI)	<i>p</i> -value
Dominant	G/G	452(90.8%)	305 (89.2%)	1.00		1.00	
Dominant .	G/T	43 (8.6%)	36 (10.5%)	1.24 (0.78–1.98)	.364	1.24(0.78 - 1.98)	.363
Dominant .	T/T	3 (0.6%)	1(0.3%)	0.49 (0.05–4.77)	.542	0.49 (0.05–4.78)	.543
	G/G	452(90.8%)	305 (89.2%)	1.00	.451	1.00	.449
	G/T-T/T	46 (9.2%0	37 (10.8)	1.19 (0.76–1.88)		1.19(0.76 - 1.88)	
Kecessive	G/G-G/T	495 (99.4%)	341 (99.7%)	1.00	.530	1.00	.531
	T/T	3 (0.6%)	1(0.3%)	0.48 (0.05-4.67)		0.48(0.05 - 4.68)	
Log-additive	I			1.13 (0.74–1.73)	.572	1.13(0.74 - 1.73)	.570
VEGF-C rs4604006 Codominant	C/C	210 (42.2%)	146 (42.7%)	1.00		1.00	
	C/T	212 (42.6%)	158 (46.2%)	1.07 (0.80–1.44)	.644	1.07 (0.80–1.44)	.640
	T/T	76 (15.3%)	38 (11.1%)	0.72 (0.46–1.12)	.145	0.72 (0.46–1.12)	.143
Dominant	C/C	210 (42.2%)	146 (42.7%)	1.00	.881	1.00	.882
	T/C-C/C	288 (57.8%)	196 (57.3%)	0.98 (0.74–1.29)		0.98 (0.74–1.29)	
Recessive	C/C -T/C	422 (84.7%)	304 (88.9%)	1.00	.086	1.00	.084
	T/T	76 (15.3%)	38 (11.1%)	0.69 (0.46–1.05)		0.69(0.46 - 1.05)	
Log-additive	I			0.91 (0.74–1.11)	.336	0.91 (0.74–1.11)	.335
rs12646659 Codominant	C/C	464 (93.5%)	333 (97.4%)	1.00		1.00	
	G/C	31 (6.3%)	9 (2.6%)	I		I	
	G/G	0(0.0%)	0 (0.0%)	1			
Dominant	C/C	464(93.5%)	333 (97.4%)	1.00	.019*	1.00	.018*
	C/G-G/G	31 (6.3%)	9 (2.6%)	0.40 (0.19–0.86)		0.40(0.19 - 0.86)	
Recessive	C/C-G/C	495(100.0%)	342 (100.0%)	1.00		1.00	
	G/G	0(0.0%)	0 (0.0%)				
Log-additive	I			0.40 (0.19–0.86)	.019*	0.40(0.19 - 0.86)	.018*
VEGFR-3 rs11955717 Codominant	T/T	227 (45.9%)	157 (45.9%)	1.00		1.00	
	T/C	203(41.0%)	145 (42.4%)	1.03 (0.77–1.39)	.830	1.03 (0.77–1.39)	.828
	C/C	65 (13.1%)	40 (11.7%)	0.89 (0.57–1.39)	.606	0.89 (0.57–1.39)	609.
Dominant	T/T	227 (45.9%)	157 (45.9%)	1.00	.989	1.00	.992
	T/C-C/C	268 (54.1%)	185 (54.1%)	1.00 (0.76–1.32)		1.00 (0.76–1.32)	
Recessive	T/T-T/C	430(86.9%)	302 (88.3%)	1.00	.538	1.00	.541
	C/C	65 (13.1%)	40 (11.7%)	0.88 (0.58–1.33)		0.88 (0.58–1.34)	
Log-additive	Ι			0.97 (0.79–1.18)	.759	0.97 (0.79–1.19)	.763

TABLE 4 Logistic regression analysis of the association between prominent SNPs and cervical cancer risk

5 of 10

p-values were calculated with Pearson's χ^2 tests. *p < .05 indicates statistical significance.

	1	T			,)					
				Age ≤ 43				Age > 43		
Gene	SNP	Model	Genotype	Control	Case	OR (95% CI)	<i>p</i> -value	Control	Case	OR (95% CI)
VEGFR-2	rs10006115	Codominant	G/G	215 (91.5%)	149 (89.8%)	1.00		237 (90.1%)	156 (88.6%)	1.00
			G/T	19 (8.1%)	17 (10.2%)	1.28 (0.64–2.54)	.466	24 (9.1%)	$19\ (10.8\%)$	1.19 (0.63–2.25)

Relationship of prominent SNPs with the cervical cancer risk stratified by age TABLE 5

6 of 1	W	ILEY	<u>N</u>	/lole	cular Gene	etics &	Genom	C Medici Open Ac	ine	
	lue	.586	.795	.644	.784	.723	.684	.035*	.425	.141

0.71 (0.06–7.95) 1.11 (0.63-1.95)

1 (0.6%)

2 (0.8%)

.670 .668

1.14 (0.59-2.2)

1.00

315 (94.9%)

449 (95.5%)

ΰ

Allele

Log-additive

1.55 (1.03-2.34)

0.77 (0.4–1.46)

17 (9.7%)

39 (14.8%)

.178

0.66 (0.36-1.22)

128

0.7 (0.46–1.09)

67 (40.4%) 21 (12.6%) 78 (47.0%)

108 (46.0%)

CJ T/T

37 (15.7%) 90 (38.3%)

1.00

68 (38.6%)

120 (45.6%) 143 (54.4%) 224 (85.2%)

.083

1.00

0.69 (0.46-1.04)

88 (53.0%)

145 (61.7%) 198 (84.3%)

T/C-C/C

C/C

Dominant

1.13 (0.63-2.02)

1.00

68 (38.6%) 91 (51.7%)

120 (45.6%) 104 (39.6%)

21 (6.0%)

28 (5.3%)

1.15 (0.6–2.22)

1.00

78 (47.0%) 17 (5.1%)

90 (38.3%)

C/C

Codominant

rs4604006

VEGF-C

21 (4.5%)

1.00

331 (94.0%)

498 (94.7%)

0.73 (0.07-8.11)

1 (0.6%)

2 (0.8%)

999 .556

1.00

156 (88.6%)

237 (90.1%)

1.16 (0.62-2.15)

20 (11.4%)

26 (9.9%)

1.21 (0.61-2.4)

17 (10.2%)

20 (8.5%)

G/T-T/T G/G

1.00

166 (100.0%)

234 (99.6%)

G/G-G/T

Recessive

0 (0.0%)

1 (0.4%)

ΓŢ

1.00

49 (89.8%)

215 (91.5%)

Dominant

0 (0.0%)

1 (0.4%)

ΓŢ

1.00

175 (99.4%)

261 (99.2%)

999

p-value

0.27 (0.08-0.94)

3 (0.8%)

16 (3.0%)

0.55 (0.21-1.44)

6(1.8%)

15 (3.2%)

1.00

349 (99.2%)

508 (97.0%)

.220

1.00

326 (98.2%)

451 (96.8%)

H υ

Allele

Log-additive

.219

0.56 (0.21-1.48)

.028*

.037*

0.26 (0.08-0.92)

.037*

1.00

173 (98.3%)

246 (93.9%)

.219

1.00

160 (96.4%)

218 (93.6%)

Dominant

0.26 (0.08-0.92)

3 (1.7%)

[6 (6.1%)

0.56 (0.21-1.48)

5 (3.6%)

15 (6.4%)

C/G-G/G

1.00

166 (100.0%)

233 (100.0%)

C/C-G/C

Recessive

0 (0.0%)

0 (0.0%)

G/G

1.00

176 (100.0%)

262 (100.0%)

0~(0.0%)

0(0.0%)

.112

1.34 (0.91-1.98)

108 (61.4%)

1.00

159 (90.3%)

.386

0.61 (0.33-1.12)

17 (9.7%)

39 (14.8%)

.775 .782

1.04 (0.79-1.38)

1.04 (0.78-1.38)

125 (35.5%)

182 (34.6%) 246 (93.9%)

0.77 (0.58-1.04))

109 (32.8%)

223 (67.2%)

288 (61.3%) 182 (38.7%) 218 (93.6%)

1.00

160 (96.4%)

6 (3.6%)

15 (6.4%)

G/C G/G C/C

C/C

Codominant

rs12646659

υ

F

Allele

Log-additive

0 (0.0%)

0 (0.0%)

227 (64.5%)

344 (65.4%)

.087

790.

0.78 (0.59-1.04)

0.78 (0.44–1.4)

37 (15.7%)

T/T

1.00

145 (87.4%) 21 (12.6%)

C/C -T/C

Recessive

1.00

173 (98.3%)

3 (1.7%)

16 (6.1%)

0 (0.0%)

0 (0.0%)

(Continued
S
E
Γ
B
Ē

				Age ≤ 43							
Gene	SNP	Model	Genotype	Control	Case	OR (95% CI)	<i>p</i> -value	Control	Case	OR (95% CI)	<i>p</i> -value
VEGFR-3	rs11955717	Codominant	T/T	106 (45.3%)	82 (49.4%)	1.00		121 (46.4%)	75 (42.6%)	1.00	
			T/C	98 (41.9%)	68 (41.0%)	0.91 (0.59–1.38)	.614	105 (40.2%)	77 (43.8%)	1.19 (0.79–1.8)	.39
			C/C	30 (12.8%)	16 (9.6%)	0.68 (0.35–1.34)	.278	35 (13.4%)	24 (13.6%)	1.11 (0.61–2.01)	.73
		Dominant	T/T	106 (45.3%)	82 (49.4%)	1.00	.419	121 (46.4%)	75 (42.6%)	1.00	.41
			T/C-C/C	128 (54.7%)	84 (50.6%)	0.85 (0.57–1.27)		140 (53.6%)	101 (57.4%)	1.17 (0.8–1.73)	
		Recessive	T/T-T/C	204 (87.2%)	150 (90.4%)	1.00	.327	226 (86.6%)	152 (46.4%)	1.00	.95
			C/C	30 (12.8%)	16(9.6%)	0.72 (0.38–1.36)		35 (13.4%)	24 (86.4%)	1.02 (0.58–1.78))	
		Log-additiv e				0.85 (0.63–1.15)	.290			1.09 (0.83–1.43)	.54
		Allele	Т	310 (66.2%)	232 (69.9%)	1.00	.278	347 (66.5%)	227 (64.5%)	1.00	.54
			C	158 (33.8%)	100 (30.1%)	0.85 (0.62–1.15)		175 (33.5%)	125 (35.5%)	1.09(0.82 - 1.45)	

Molecular Genetics & Genomic Medicine

7 of 10

CI = 1.03–2.34, p = .035 for the "C/T" genotype). While rs1264665 of *VEGF-C* was related to a decreased the risk of cervical cancer under the allele model (OR = 0.27, 95% CI, 0.08–0.94; p = .028 for the "C" allele) and dominant model (adjusted, OR = 0.26, 95% CI = 0.08–0.92, p = .037 for the "C/G-G/G" genotype). The variant rs1264665 was also observed to decrease the risk of cervical cancer in the logadditive model (adjusted, OR = 0.26, 95% CI = 0.08–0.92, p = .037). However, no significant association between candidate polymorphisms and cervical cancer risk was found in populations at age ≤ 43 .

4 | DISCUSSION

In this hospital-based case–control study, we investigated the association of four important polymorphisms (rs10006115, rs4604006, rs12646659, and rs11955717) with the risk of cervical cancer in Chinese Uygur population, and we observed a significant association between the *VEGF-C* rs12646659 polymorphism and the risk of cervical cancer. The presence of the *VEGF-C* rs12646659 conferred a lower risk of developing cervical cancer. Further stratified analysis revealed that rs4604006 of *VEGF-C* gene was related to a higher risk of cervical cancer at age >43, while rs12646659 of *VEGF-C* was associated with a lower risk of cervical cancer at age >43. Our present study is the first to provide substantial basic evidence that gene polymorphisms in *VEGF-C* were corrected with cervical cancer risk in Chinese Uygur population.

Due to tumor growth and metastasis require the formation of new blood vessels, blocking tumor angiogenesis can be used to treat cancer. Meanwhile, genes related to angiogenesis also become potential target molecules for tumor treatment (Hajari Taheri et al., 2019). VEGF family proteins are specific and potent angiogenic factors that increase vessel permeability, endothelial cell growth, proliferation, migration, and differentiation (Keck et al., 1989). VEGF-C, a member of the VEGF family, is a protein-coding gene (Olofsson et al., 1996). VEGF-C associated with Diseases, including Lymphedema, Hereditary, and Lymphedema. Recently, several reports have confirmed the correlation between VEGF-C expression in tumor tissue specimens and lymph node metastasis. In cervical cancer, VEGF-C mRNA expression in tumor tissue samples has been shown to be associated with lymph node metastasis (Niki et al., 2000). VEGF-C tissue status also was an important independent factor for poor prognosis, and serum VEGF and VEGF-C levels can also be used as biomarkers for cervical SCC (Mitsuhashi et al., 2005). In addition, VEGF-C can binds to receptor Flt-4 and promotes angiogenesis and/or lymphangiogenesis, thus accelerating cancer metastasis (Joukov et al., 1996; Lohela, Bry, Tammela, & Alitalo, 2009). VEGF-C accelerated cervical cancer invasiveness via regulation of galectin-3 or moesin

p < .05 indicates statistical significance.

"/"means no data.

protein expression (Liu, Cheng, He, & Yao, 2014). *VEGF-C* can also reduce the expression of miR-326 and increase the expression of cortactin through c-Src signaling, leading to enhanced cervical cancer invasiveness (Cheng et al., 2018).

VEGF binds to the receptor tyrosine kinase (VEGFR) by transphosphatase, and then activates VEGFR. VEGFR-2 and VEGFR-3 are belongs to the tyrosine kinase receptor family (Masabumi, 2006). VEGFR-3 is activated by two proteins (VEGF-C and VEGF-D), and plays an essential role in the morphogenesis of the lymphatic vessel network during embryonic development, being involved in formation of new lymphatic vessels in the life. Binding of VEGF-C to VEGFR-3 is responsible for mostly biological effects of VEGFR-3 (Olsson, Dimberg, Kreuger, & Claesson-Welsh, 2006). Some studies have reported that VEGFR-2 may also interfere with lymphangiogenesis by binding VEGF-C and VEGF-D, which was crucial for the normal process of vasculogenesis during embryonic development (Ferrara, 2004; Vokes & Krieg, 2015). VEGFR-2 and VEGFR-3 are involved in normal and pathological angiogenesis through different mechanisms, such as: activation of MAPK extracellular signal-regulated kinases (ERK1/2) through the PKC and Ras pathways (Shibuya, 2011; important pathways in cell proliferation), as well as the PI3K Akt/PKB pathway (involved especially in survival of lymphatic endothelial cells; Olsson et al., 2006). Moreover, it activates certain integrins, which disrupt cell to cell cohesion and initiate cellular migration (Takahashi & Shibuya, 2005).

In this study, we explored the relationship of VEGF-C, VEGFR-2, and VEGFR-3 polymorphisms with cervical cancer risk. There were significant associations between their polymorphisms and the risk of various diseases (hepatocellular carcinoma, oral cancer, gastric cancer, lymphedema, etc.; Chien et al., 2013; Debrah et al., 2017; Hsieh et al., 2014; Li, Yu, et al., 2018), except cervical cancer. We found that the rs4604006 of VEGF-C gene was associated with an increased risk of cervical cancer, while rs12646659 was significantly associated with a decreased risk of cervical cancer for the first time. Meanwhile, there are few reports focused on the polymorphisms of rs4604006 and rs12646659. Only one study aimed to assess the role of VEGF and VEGFR polymorphisms in clinical outcomes of HCC patients receiving sorafenib therapy, and it found that rs4604006 (VEGF-C) T allele was significantly associated with patients' progressionfree survival and overall survival (Scartozzi et al., 2014). In addition, rs4604006 and rs12646659 were located in the intron region of VEGF-C gene and may be involved in mRNA processing, and hence regulate posttranscriptional modification, protein translation, or promoter/enhancer cluster activity (Rose, 2008; Zhang et al., 2018). Besides, we will expand the sample size to verify our results and conduct further functional studies so as to provide more evidence for the effect of VEGF-C polymorphism on cervical cancer risk.

Inevitably, this study had some limitations that should be accounted for when interpreting the results. First of all, the inherent selection bias and information bias were inevitable problems, because all participants were recruited from the identical hospitals. Second, the number of cases in our study was limited and our study population was all Chinese Uygur population, which cannot preclude false-negative results and extrapolated to other populations. Hence, larger prospective studies are necessary to fully elucidate the role of these polymorphisms in cervical cancer. Despite these limitations, our current findings provide scientific evidence of *VEGFR-2*, *VEGF-C*, and *VEGFR-3* with the risk of cervical cancer in the future studies.

5 | CONCLUSION

To sum up, our study firstly provided evidence that the variants of *VEGF-C* gene had a significant effect on the risk of cervical cancer in Chinese Uygur population, especially individuals older than 43. These results may contribute to further elucidate the potential role of *VEGF-C* in cervical cancer susceptibility among Chinese Uygur population.

ACKNOWLEDGMENTS

This study was supported by Xinjiang Tianshan Youth Program in 2017 years (No. 2017Q008). We were grateful to the individuals for their participation in this study. We also thank the clinicians and hospital staff who contributed to the sample and data collection for this study. We thank all participants for this manuscript.

CONFLICT OF INTEREST

The authors declare that they have no competing interests.

ORCID

Mayinuer Niyazi D https://orcid.org/0000-0002-6686-1650

REFERENCES

- Bland, J. M., & Altman, D. G. (2000). Statistics notes: The odds ratio. *Bmj*, 320(7247),1468–1468.
- Chen, Y. H., Pan, S. L., Wang, J. C., Kuo, S. H., Cheng, C. H., & Teng, C. M. (2014). Radiation-induced VEGF-C expression and endothelial cell proliferation in lung cancer. *Strahlentherapie Und Onkologie*, 190(12), 1154–1162. https://doi.org/10.1007/ s00066-014-0708-z
- Cheng, Y., Jiang, S., Yuan, J., Liu, J., & Simoncini, T. (2018). Vascular endothelial growth factor C promotes cervical cancer cell invasiveness via regulation of microRNA-326/cortactin expression. *Gynecological Endocrinology*, 34(10), 853–858. https://doi. org/10.1080/09513590.2018.1458304

Molecular Genetics & Genomic Medicine

- hien M-H Liu V-F Hsin C-H
- Chien, M.-H., Liu, Y.-F., Hsin, C.-H., Lin, C.-H., Shih, C.-H., Yang, S.-F., ... Lin, C.-W. (2013). Impact of VEGF-C gene polymorphisms and environmental factors on oral cancer susceptibility in Taiwan. *PLoS ONE*, 8(4), e60283. https://doi.org/10.1371/journ al.pone.0060283
- Debrah, L. B., Albers, A., Debrah, A. Y., Brockschmidt, F. F., Becker, T., Herold, C., ... Pfarr, K. (2017). Single nucleotide polymorphisms in the angiogenic and lymphangiogenic pathways are associated with lymphedema caused by Wuchereria bancrofti. *Human Genomics*, 11(1), 26. https://doi.org/10.1186/s40246-017-0121-7
- Du, G. H., Wang, J. K., Richards, J. R., & Wang, J. J. (2019). Genetic polymorphisms in tumor necrosis factor alpha and interleukin-10 are associated with an increased risk of cervical cancer. *International Immunopharmacology*, 66, 154–161. https://doi.org/10.1016/j. intimp.2018.11.015
- Fang, J., Li, Y., Zhang, J., Yan, M., Li, J., Bao, S., & Jin, T. (2017). Correlation between polymorphisms in microRNA-regulated genes and cervical cancer susceptibility in a Xinjiang Uygur population. *Oncotarget*, 8(19), 31758–31764. https://doi.org/10.18632/oncotarget.15970
- Ferrara, N. (2004). Vascular endothelial growth factor: Basic science and clinical progress. *Endocrine Reviews*, 25(4), 581–611. https:// doi.org/10.1210/er.2003-0027
- Hajari Taheri, F., Hassani, M., Sharifzadeh, Z., Behdani, M., Arashkia, A., & Abolhassani, M. (2019). T cell engineered with a novel nanobody-based chimeric antigen receptor against VEGFR2 as a candidate for tumor immunotherapy. *IUBMB Life*, https://doi. org/10.1002/iub.2019
- Hamrah, P., Chen, L., Cursiefen, C., Zhang, Q., Joyce, N. C., & Dana, M. R. (2004). Expression of vascular endothelial growth factor receptor-3 (VEGFR-3) on monocytic bone marrow-derived cells in the conjunctiva. *Experimental Eye Research*, 79(4), 553–561. https ://doi.org/10.1016/j.exer.2004.06.028
- Hsieh, M. C., Hsu, H. T., Hsiao, P. C., Yang, S. F., Yeh, C. B., Bien, M. Y., ... Chien, M. H. (2014). Role of VEGF-C gene polymorphisms in susceptibility to hepatocellular carcinoma and its pathological development. *Journal of Clinical Laboratory Analysis*, 28(3), 237– 244. https://doi.org/10.1002/jcla.21672
- Hu, N., Wang, J., Hu, P., & Li, Z. (2018). Investigation of Renalase gene rs2576178 polymorphism in patients with coronary artery disease. *Bioscience Reports*, 38(5), https://doi.org/10.1042/bsr20180839
- Jin, T., Wu, X., Yang, H., Liu, M., He, Y., He, X., ... Yuan, D. (2016). Association of the miR-17-5p variants with susceptibility to cervical cancer in a Chinese population. *Oncotarget*, 7(47), 76647–76655. https://doi.org/10.18632/oncotarget.12299
- Joukov, V., Pajusola, K., Kaipainen, A., Chilov, D., Lahtinen, I., Kukk, E., ... Alitalo, K. (1996). A novel vascular endothelial growth factor, VEGF-C, is a ligand for the Flt4 (VEGFR-3) and KDR (VEGFR-2) receptor tyrosine kinases. *EMBO Journal*, 15(7), 1751. https://doi. org/10.1002/j.1460-2075.1996.tb00521.x
- Keck, P. J., Hauser, S. D., Krivi, G., Sanzo, K., Warren, T., Feder, J., & Connolly, D. T. (1989). Vascular permeability factor, an endothelial cell mitogen related to PDGF. *Science*, 246(4935), 1309–1312.
- Li, S., Xu, G., Gao, F., Bi, J., & Huo, R. (2018). Erratum: Expression and association of VEGF-Notch pathways in infantile hemangiomas. *Experimental and Therapeutic Medicine*, 16(4), 3752. https:// doi.org/10.3892/etm.2018.6529
- Li, T., Yu, J., Luo, X., Ren, W., Zhang, Y., & Cao, B. (2018). VEGFR-2 as a novel predictor of survival in gastric cancer: A systematic review and meta-analysis. *Pathology, Research and Practice*, 214(4), 560–564. https://doi.org/10.1016/j.prp.2018.02.005

- Lin, C., Huang, F., Shen, G., & Yiming, A. (2015). MicroRNA-101 regulates the viability and invasion of cervical cancer cells. *International Journal of Clinical & Experimental Pathology*, 8(9), 10148–10155.
- Liu, D. I., Wang, M., Tian, T., Wang, X.-J., Kang, H.-F., Jin, T.-B., ... Dai, Z.-J. (2017). Genetic polymorphisms (rs10636 and rs28366003) in metallothionein 2A increase breast cancer risk in Chinese Han population. *Aging*, 9(2), 547–555. https://doi.org/10.18632/ aging.101177
- Liu, J., Cheng, Y., He, M., & Yao, S. (2014). Vascular endothelial growth factor C enhances cervical cancer cell invasiveness via upregulation of galectin-3 protein. *Gynecological Endocrinology*, 30(6), 461–465. https://doi.org/10.3109/09513590.2014.898054
- Lohela, M., Bry, M., Tammela, T., & Alitalo, K. (2009). VEGFs and receptors involved in angiogenesis versus lymphangiogenesis. *Current Opinion in Cell Biology*, 21(2), 154–165. https://doi.org/10.1016/j. ceb.2008.12.012
- Ma, L., Hong, Y., Lu, C., Chen, Y., & Ma, C. (2015). The occurrence of cervical cancer in Uygur women in Xinjiang Uygur Autonomous Region is correlated to microRNA-146a and ethnic factor. *International Journal of Clinical and Experimental Pathology*, 8(8), 9368–9375.
- Masabumi, S. (2006). Differential roles of vascular endothelial growth factor receptor-1 and receptor-2 in angiogenesis. *Journal of Biochemistry and Molecular Biology*, 39(5), 469–478.
- Mitsuhashi, A., Suzuka, K., Yamazawa, K., Matsui, H., Seki, K., & Sekiya, S. (2005). Serum vascular endothelial growth factor (VEGF) and VEGF-C levels as tumor markers in patients with cervical carcinoma. *Cancer*, 103(4), 724–730. https://doi.org/10.1002/cncr. 20819
- Niki, T., Iba, S., Tokunou, M., Yamada, T., Matsuno, Y., & Hirohashi, S. (2000). Expression of vascular endothelial growth factors A, B, C, and D and their relationships to lymph node status in lung adenocarcinoma. *Clinical Cancer Research*, 6(6), 2431–2439.
- Niu, F., Wang, T., Li, J., Yan, M., Li, D., Li, B., & Jin, T. (2019). The impact of genetic variants in IL1R2 on cervical cancer risk among Uygur females from China: A case-control study. *Molecular Genetics & Genomic Medicine*, 7(1), e00516. https:// doi.org/10.1002/mgg3.516
- Olofsson, B., Pajusola, K., Kaipainen, A., Von Euler, G., Joukov, V., Saksela, O., ... Eriksson, U. (1996). Vascular endothelial growth factor B, a novel growth factor for endothelial cells. *Proceedings of the National Academy of Sciences of the United States of America*, 93(6), 2576–2581. https://doi.org/10.1073/pnas.93.6.2576
- Olsson, A. K., Dimberg, A., Kreuger, J., & Claesson-Welsh, L. (2006). VEGF receptor signalling - In control of vascular function. *Nature Reviews Molecular Cell Biology*, 7(5), 359–371. https://doi. org/10.1038/nrm1911
- Robinson, C. J., & Stringer, S. E. (2001). The splice variants of vascular endothelial growth factor (VEGF) and their receptors. *Journal of Cell Science*, 114(5), 853–865.
- Rose, A. B. (2008). Intron-mediated regulation of gene expression. Current Topics in Microbiology and Immunology, 326(6), 277–290.
- Scartozzi, M., Faloppi, L., Svegliati Baroni, G., Loretelli, C., Piscaglia, F., Iavarone, M., ... Cascinu, S. (2014). VEGF and VEGFR genotyping in the prediction of clinical outcome for HCC patients receiving sorafenib: The ALICE-1 study. *International Journal of Cancer*, 135(5), 1247–1256. https://doi.org/10.1002/ijc.28772
- Shibuya, M. (2006). Differential roles of vascular endothelial growth factor receptor-1 and receptor-2 in angiogenesis. *Journal of*

Biochemistry and Molecular Biology, 39(5), 469–478. https://doi. org/10.5483/BMBRep.2006.39.5.469

- Shibuya, M. (2011). Vascular Endothelial Growth Factor (VEGF) and its Receptor (VEGFR) signaling in angiogenesis: A crucial target for anti- and pro-angiogenic therapies. *Genes & Cancer*, 2(12), 1097– 1105. https://doi.org/10.1177/1947601911423031
- Sole, X., Guino, E., Valls, J., Iniesta, R., & Moreno, V. (2006). SNPStats: A web tool for the analysis of association studies. *Bioinformatics*, 22(15), 1928–1929. https://doi.org/10.1093/bioinformatics/btl268
- Su, J.-L., Yang, P.-C., Shih, J.-Y., Yang, C.-Y., Wei, L.-H., Hsieh, C.-Y., ... Kuo, M.-L. (2006). The VEGF-C/Flt-4 axis promotes invasion and metastasis of cancer cells. *Cancer Cell*, 9(3), 209–223. https:// doi.org/10.1016/j.ccr.2006.02.018
- Takahashi, H., & Shibuya, M. (2005). The vascular endothelial growth factor (VEGF)/VEGF receptor system and its role under physiological and pathological conditions. *Clinical Science*, 109(3), 227–241. https://doi.org/10.1042/cs20040370
- Tamura, M., & Ohta, Y. (2003). Serum vascular endothelial growth factor-C level in patients with primary nonsmall cell lung carcinoma: A possible diagnostic tool for lymph node metastasis. *Cancer*, 98(6), 1217–1222. https://doi.org/10.1002/cncr.11529
- Tomao, F., Di Tucci, C., Imperiale, L., Boccia, S. M., Marchetti, C., Palaia, I., ... Panici, P. B. (2014). Cervical cancer: Are there potential new targets? An update on preclinical and clinical results. *Current Drug Targets*, 15(12), 1107–1120.

- Vokes, S. A., & Krieg, P. A. (2015). Vascular molecular embryology. In *PanVascular medicine* (2nd ed., pp. 27–51). Heidelberg, Berlin: Springer. https://doi.org/10.1007/978-3-642-37078-6_2
- Waggoner, S. E. (2003). Cervical cancer. Lancet, 361(9376), 2217– 2225. https://doi.org/10.1016/s0140-6736(03)13778-6
- Wu, H. (2014). The effect of ZnMoS4 on tumor angiogenesis. (Electronic Thesis or Dissertation). Retrieved from https://etd.ohiolink.edu/
- Yang, P., Wang, M., Tian, T., Feng, Y., Zheng, Y., Yang, T., ... Dai, Z. (2018). CYP17 polymorphisms are associated with decreased risk of breast cancer in Chinese Han women: A case-control study. *Cancer Management and Research*, 10, 1791–1798. https://doi. org/10.2147/cmar.s167503
- Zhang, Q., Wang, W., Xu, W., Du, M., Ma, G., Liu, H., ... Shao, J. (2018). Genetic variations in the 3'-untranslated regions of genes involved in the cell cycle and apoptosis pathways affect bladder cancer risk. *Cancer Genomics & Proteomics*, 15(1), 67–72.

How to cite this article: Han L, Husaiyin S, Ma C, Niyazi M. Association study between the polymorphisms of angiogenesis-related genes and cervical cancer susceptibility in Chinese Uygur population. *Mol Genet Genomic Med.* 2019;7:e899. https://doi.org/10.1002/mgg3.899