

NUCLEIC ACID AND PROTEIN SEQUENCES OF
PHOSPHOCHOLINE-BINDING LIGHT CHAINS*

By S.-P. KWAN, S. RUDIKOFF, J. G. SEIDMAN, P. LEDER, AND M. D. SCHARFF

*From the Department of Cell Biology, Albert Einstein College of Medicine, Bronx, New York 10461;
and the Laboratory of Cell Biology, National Cancer Institute, and the Laboratory of Molecular
Genetics, National Institute of Child Health and Development, National Institutes of Health,
Bethesda, Maryland 20205*

Recent studies on the control of the immune response have revealed the importance of idiotypes and their recognition by both B and T cells. Most of these studies have focused on a few well-studied idiotypes, one of which is T15. This is the major idotype elicited when BALB/c mice are immunized with the hapten phosphocholine (PC), either in its natural form as part of bacterial or parasite cell walls or artificially attached to protein and other carriers (1). Other strains of mice produce lesser amounts of T15 idotype. The heavy-chain variable region of the T15 myeloma, an IgA κ protein that is the prototype for these antibodies, has been completely sequenced (2, 3). The amino acid sequence of the variable region of the S107 heavy chain, which is also an IgA κ myeloma, is identical to T15 (2, 4). The heavy chain of HOPC8, a third PC-binding IgA κ myeloma bearing the T15 idotype, has also been completely sequenced and differs by only one amino acid in its third hypervariable region from T15 and S107 (2, 4). Only partial sequences through residues 30-40 of the light chains of these proteins have been reported, and the peptide containing the third hypervariable region has been especially difficult to purify, precluding the immediate completion of the protein sequences (2). It has become necessary to determine the sequence of the third hypervariable region and J segment of the S107 light chain to study hybridomas producing PC-binding antibodies and somatic variants of S107 with changes in antigen binding (5). We have therefore cloned and sequenced the rearranged light-chain variable region gene that codes for the secreted S107 light chain. Here we report the base sequence that codes for amino acid residues 1 through 108 and compare it to the amino acid sequence of the T15 light chain, which has been determined through amino acid residue 88.

Materials and Methods

Cloning of the Light-Chain Gene. The S107 cell line was obtained from the Salk cell bank (The Salk Institute, La Jolla, Calif.). S107 DNA was isolated from cultured cells and the genomic DNA was digested with Eco RI restriction enzyme. Fragments containing the recombinated κ gene DNA were identified by Southern blot analysis of S107 compared with embryo DNA with a κ -chain constant-region probe (6). Fragments were then further enriched by preparative agarose gel electrophoresis as described (7). Two distinct fragments of ~18 kilobase (kb) contained rearranged κ -chain DNA. Both were analyzed, but only one, named S107A, is

* Supported by grants AI-10702 and AI-15231 from the National Institutes of Health, grant PLM75-13609 from the National Science Foundation, and grant AP-317 from the American Cancer Society. S.-P. Kwan was supported by a fellowship from the Cancer Research Institute, Inc.

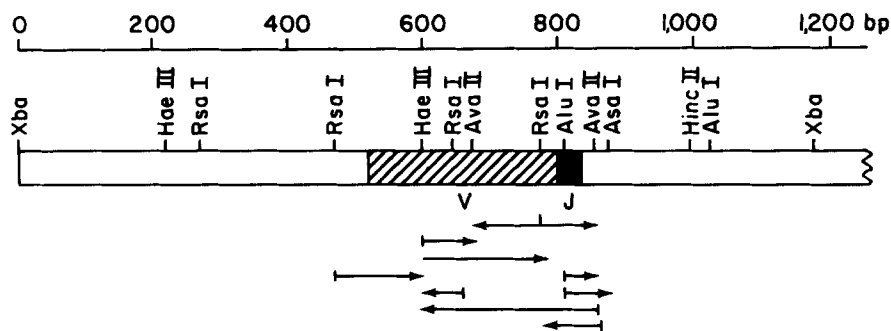


FIG. 1. Physical map of the rearranged S107A light chain DNA. The arrows denote the DNA sequencing strategy. The coding region for the V region is stippled; the J region is solid.

reported here. The DNA from this band was pooled, ligated into the Charon 4A vector, packaged, and transfected into *Escherichia coli* (8). Hybrid phages containing κ -chain DNA were identified by *in situ* hybridization of nitrocellulose filter blots (9) using the C region-specific probe (6) and grown preparatively on *E. coli* LE392. For sequence determination, the S107A clone was restricted with Bam HI generating a 7-kb fragment which was shown to contain both variable and constant sequences by R loop analysis with S107 light chain messenger RNA. This was subcloned in pBR322 (10). The plasmid was grown and the 7-kb insert was purified as described by Seidman et al. (6). These experiments were carried out in compliance with the National Institutes of Health guidelines on recombinant DNA research.

DNA Sequencing. S107A DNA was digested with Xba I and a 1.2-kb-long fragment was isolated by electrophoresis in 5% polyacrylamide gel. The 1.2-kb fragment was digested with different restriction enzymes as shown in Fig. 1, treated with bacterial alkaline phosphatase (ATP), and labeled at the 5' end with $\gamma^{32}\text{P}$ -ATP using polynucleotide kinase. These fragments were subjected to second restriction enzyme and single end-labeled DNA were isolated by polyacrylamide gel electrophoresis. Sequencing was performed according to the method of Maxam and Gilbert (11).

Protein Sequencing. The N-terminal sequences of the T15 and S107 κ chains have previously been reported (2). The amino acid sequence of T15 from Trp-35 to Phe-62 (Fig. 2) was determined from a peptide obtained by cleavage at Trp-35 with BNPS-skatole as described (12). This peptide extends from Trp-35 into the C region. The sequence from Phe-62 to Cys-88 was obtained from a tryptic peptide extending from Phe-62 to Lys-103.¹ Sequencing procedures employed have previously been described in considerable detail (12). Positions 61 (Arg) and 85 (His) were not identified in the amino acid sequencing experiments of T15, and are assigned only by nucleotide sequence of S107A.

Results and Discussion

When DNA from the S107 cell line was digested with Eco RI and analyzed for rearranged κ -chain constant-region genes, two bands of ~ 18 kb were identified and cloned in Charon 4A. A total of 120,000 recombinant phages were analyzed, 10 of which contained κ C-region sequences. On the basis of restriction endonuclease analysis and heteroduplex mapping (not shown), these clones represent two distinct genes. Both genes have been sequenced. One, which we have named S107B, has an unusual V-J recombination and shares only 50% of its coding sequence with the light chain secreted by S107. This will be described in detail elsewhere.² A second recombined gene, which we have named S107A, is the subject of the investigations

¹ Rudikoff, S., Y. Satow, E. Padlan, P. Davies, and M. Potter. 1981. Kappa chain structure from a captalized murine Fab': role of joining segment in hapten binding. Manuscript submitted.

² Kwan, S.-P., E. Max, J. G. Seidman, P. Leder, and M. D. Scharff. Manuscript in preparation.

ATG TAT ATA TGT TTG TTG TCT ATT TCT TAT TGT AGG TGC CTC GTG CAC	1	GAC ATT GTG ATG
		ASP ILE VAL MET
ACT CAG TCT CCA ACT TTC CTT GCT GTG ACA GCA AGT AAG AAG GTC ACC ATT AGT TGC ACT	20	
THR GLN SER PRO THR PHE LEU ALA VAL THR ALA SER LYS LYS VAL THR ILE SER CYS THR		
GCX TCT GAG AGC CTT TAT TCA AGC AAA CAC AAG GTG CAC TAC TTG GCT TGG TAC CAG AAG	30	
ALA SER GLU SER LEU TYR SER SER LYS HIS LYS VAL HIS TYR LEU ALA TRP TYR GLN LYS		
AAA CCA GAG CAA TCT CCT AAA CTG CTG ATA TAC GGG GCA TCC AAC CGA TAC ATT GGG GTC	40	50
LYS PRO GLU GLN SER PRO LYS LEU LEU ILE TYR GLY ALA SER ASN ARG TYR ILE GLY VAL		
CCT GAT CGC TTC ACA GGC AGT GGA TCT GGG ACA GAT TTC ACT CTG ACC ATC AGC AGT GTA	60	70
PRO ASP ARG PHE THR GLY SER GLY SER GLY THR ASP PHE THR LEU THR ILE SER SER VAL		
CAG GTT GAA GAC CTC ACA CAT TAT TAC TGT GCA CAG TTT TAC AGC TAT CCT CTC ACG TTC	80	90
GLN VAL GLU ASP LEU THR HIS TYR TYR CYS ALA GLN PHE TYR SER TYR PRO LEU THR PHE		
GGT GCT GGG ACC AAG CTG GAG CTG AAA CGT AAG TAC ACT TTT CTC ATC TTTT	100	
GLY ALA GLY THR LYS LEU GLU LEU LYS ARG		

Fig. 2. Nucleotide sequence of the active S107A light chain gene and the light chain for which it codes. The numbers indicate the amino acid residue using the Kabat numbering system. The nucleotide sequence was determined as described in the text. The amino acids represent the amino acid sequence deduced from the nucleotide sequence. The solid line represents the identity of amino acid sequence of T15 determined as described in the text.

reported in the present paper. This clone codes for the light chain polypeptide employed in the secreted PC-binding S107 protein. Through the restriction analysis illustrated in Fig. 1, a 1.2-kb Xba-Xba fragment containing all of the V region was obtained and its base sequence was determined. Bases 49-171 code for an amino acid sequence that is identical to that reported for the N-terminal 35 amino acids of the light chain secreted by both the T15 and S107 tumors (2). The sequence of an additional 50 amino acids of the T15 light chain has been determined. As indicated by the continuous line in Fig. 2, this sequence is also identical with that determined from the DNA sequence.

The S107 light chain is of particular interest in that this chain, as well as the idiotype of the native molecule, is highly conserved in all inbred mouse strains (13), which suggests a strong selective value for these gene products. The genetic rearrangement producing the active κ -chain gene involves recombination of the V gene with the J1 gene segment, which is the first J found 5' from the κ C-region gene (14). It is intriguing that all κ chains analyzed to date from PC-binding proteins employ the J1 sequence, although a number of these light chains (i.e., M603 and M167) differ considerably in amino acid sequence (2, 12).¹ Three-dimensional X-ray analysis of the PC-binding myeloma protein M603 has demonstrated that Leu 96, the first amino acid of the J segment, is a hapten-contacting residue (15).² Thus J1, the only J segment encoding Leu at position 96 (15), may be required for PC binding, or these V genes may, due to genetic restrictions, only pair with J1. The nucleotide structure of the S107 κ chain has thus provided important information on the third hypervariable region and J segment from this molecule as well as demonstrating that the T15

and S107 κ chains are identical for at least their N-terminal 88 amino acids. It is therefore probable that these κ chains are identical throughout this entire V region, as has been demonstrated for the heavy-chain V regions.

Summary

An 18-kilobase DNA fragment containing the sequence coding for both the variable and constant regions of the S107 mouse immunoglobulin light chain was cloned from total cellular DNA. The complete nucleotide sequence of the κ -chain variable-region gene is reported. Determination of the amino acid sequence encoded by the DNA is found to be identical to the protein sequence of the T15 light chain through residue 88. Direct sequence analysis confirmed that the J1 joining segment is used in the recombination event producing the active κ light chain gene.

We would like to thank Marion Nau for technical assistance.

Received for publication 9 February 1981.

References

1. Kohler, H. 1975. The response to phosphorylcholine: dissecting an immune response. *Transplant. Rev.* **27**:24.
2. Barstad, P., S. Rudikoff, M. Potter, M. Cohn, W. Konigsberg, and L. Hood. 1974. Immunoglobulin structure: amino terminal sequences of mouse myeloma proteins that bind phosphorylcholine. *Science (Wash. D. C.)*. **183**:962.
3. Rudikoff, S., and M. Potter. 1978. Size differences among immunoglobulin heavy chains from phosphorylcholine-binding proteins. *Proc. Natl. Acad. Sci. U. S. A.* **73**:2109.
4. Barstad, P., J. Hubert, M. Hunkapiller, A. Goetz, J. Schilling, B. Black, B. Eaton, J. Richards, M. Wigert, and L. Hood. 1978. Immunoglobulins with hapten-binding activity: structure-function correlations and genetic implications. *Eur. J. Imm.* **8**:497.
5. Cook, W., and M. D. Scharff. 1977. Antigen-binding mutants of mouse myeloma cells. *Proc. Natl. Acad. Sci. U. S. A.* **74**:5687.
6. Seidman, J. G., A. Leder, M. Nau, B. Norman, and P. Leder. 1978. Antibody diversity. *Science (Wash. D. C.)*. **202**:11.
7. Polsky, F., M. H. Edgell, J. G. Seidman, and P. Leder. 1978. High capacity gel preparative electrophoresis for purification of fragments of genomic DNA. *Anal. Biochem.* **87**:397.
8. Blattner, F. R., B. G. Williams, A. E. Blechl, K. D. Thompson, H. E. Faber, L. A. Furlong, D. J. Grunwald, D. O. Kiefer, D. D. Moore, J. W. Schuman, E. L. Sheldon, and O. Smithies. 1977. Charon pages: safer derivative of bacteriophage lambda for DNA cloning. *Science (Wash. D. C.)*. **196**:161.
9. Benton, W. D., and R. W. Davis. 1977. Screening λ gt recombinant clones by hybridization to single plaques in situ. *Science (Wash. D. C.)*. **196**:180.
10. Bolivar, E., R. L. Rodriguez, P. J. Green, M. C. Behach, H. L. Heyneker, H. W. Boyer, J. H. Crosa, and S. Falkow. 1977. Construction and characterization of new cloning vehicles II. A multipurpose cloning system. *Gene (Amst.)*. **2**:95.
11. Maxam, A., and W. Gilbert. 1977. A new method for sequencing DNA. *Proc. Natl. Acad. Sci. U. S. A.* **74**:560.
12. Rudikoff, S., and M. Potter. 1978. κ chain variable region from M167, a phosphorylcholine binding myeloma protein. *Biochemistry*. **17**:2703.
13. Clafin, J. L., and J. M. Davie. 1974. Clonal nature of the immune response to phospho-

- rylcholine TV. Idiotypic uniformity of binding site associated antigenic determinants among mouse antiphosphorylcholine antibodies. *J. Exp. Med.* **140**:673.
14. Max, E., J. G. Seidman, and P. Leder. 1979. Sequences of five potential recombination sites encoded close to an immunoglobulin κ constant region gene. *Proc. Natl. Acad. Sci. U. S. A.* **76**:3450.
 15. Segal, D. M., E. A. Padlan, G. H. Cohen, S. Rudikoff, M. Potter, and D. R. Davies. 1974. The three dimensional structure of a phosphorylcholine binding mouse immunoglobulin Fab and the nature of the antigen binding site. *Proc. Natl. Acad. Sci. U. S. A.* **71**:4298.