

Molecular Cell, Volume 32

Supplemental Data

Posttranscriptional Regulation of miRNAs

Harboring Conserved Terminal Loops

Gracjan Michlewski, Sonia Guil, Colin A. Semple, and Javier F. Cáceres

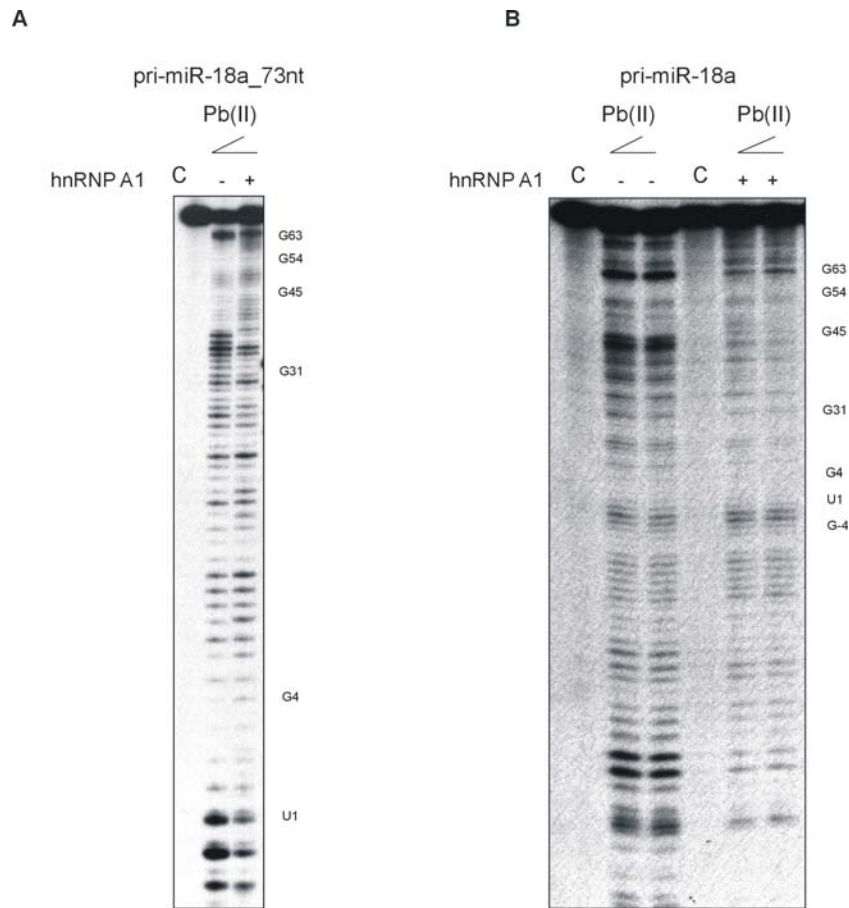


Figure S1. hnRNP A1 primarily binds to the terminal loop of pri-miR-18a transcripts with different length of neighboring sequences (A) Footprint analysis of the pri-miR-18a_73nt/hnRNP A1 complex as shown on Figure 1A. Cleavage patterns were obtained for 5' ³²P-labeled pri-miR-18a_73nt transcript (100×10^3 c.p.m.) incubated in the presence or absence of recombinant hnRNP A1 (100ng), treated with Pb (II) – lead ions (0.5mM). Positions of selected residues are indicated (B) Footprint analysis of the pri-miR-18a/hnRNP A1 complex. Cleavage pattern obtained for 5' ³²P-labeled pri-miR-18a (100×10^3 c.p.m.) incubated in the presence or absence of recombinant hnRNP A1 (100ng), treated with Pb (II) – lead ions (0.25mM and 0.5mM). Analysis and data interpretation was performed as described in the legend to Figure 1.

hnRNP A1 footprint on P³² labeled pri-miR-18a cleaved with Pb(II)

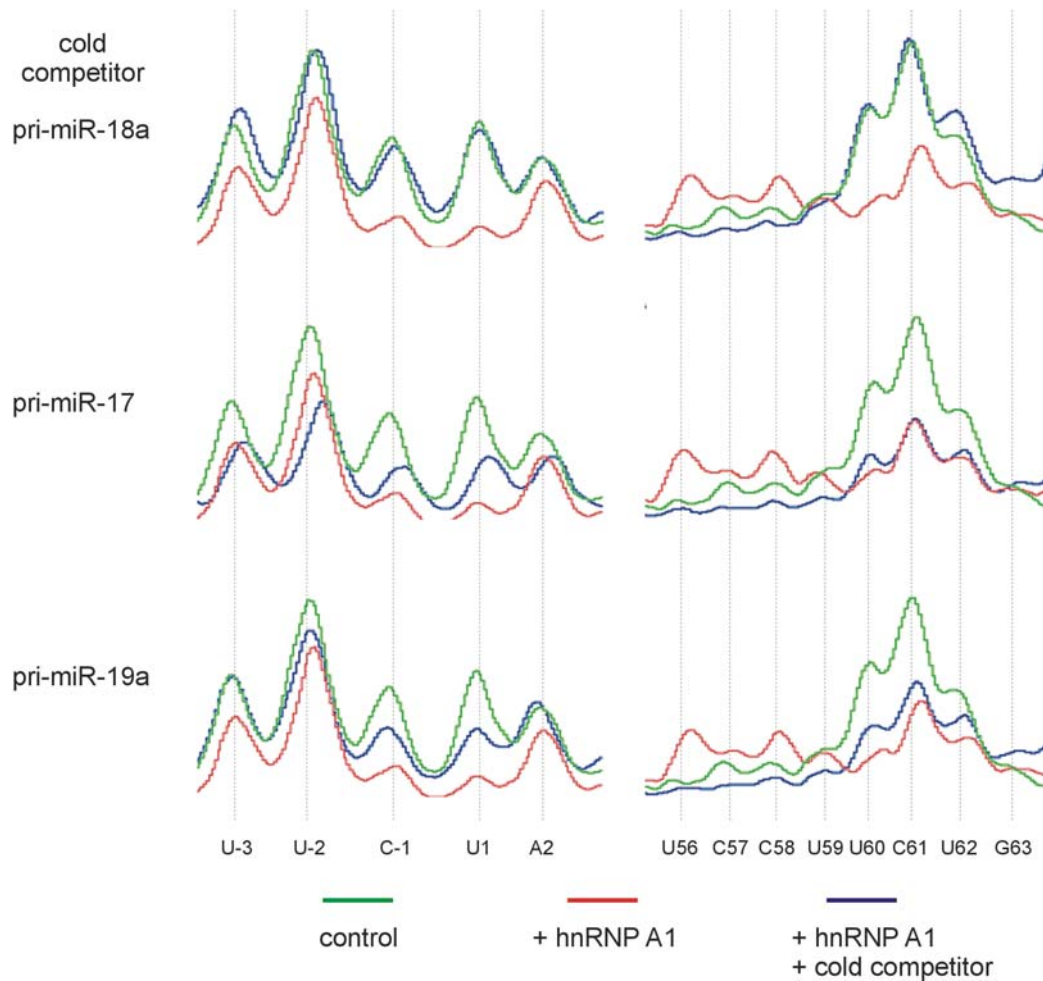


Figure S2. Pri-miR-18a has a high affinity towards hnRNP A1 binding. PhosphorImager peaks represent Pb (II) – lead ions (0.5mM) cleavage patterns of a 5' ³²P-labeled pri-miR-18a transcript (100x10³ c.p.m.) in the absence (green lines) or presence of recombinant hnRNP A1 (100ng) (red lines). When indicated reactions were supplemented with addition of an excess of cold pri-miRNAs competitor (500ng) (blue lines). Nucleotides are numbered as described in the legend to Figure 1.

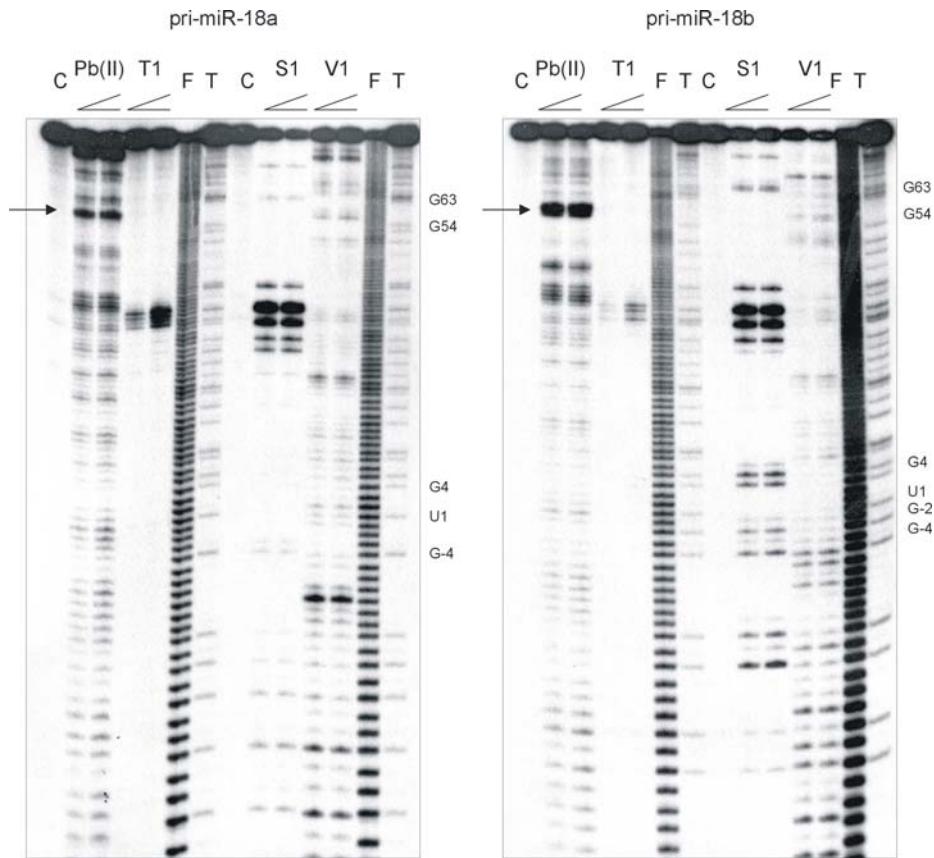


Figure S3. Structure analysis of pri-miR-18a and pri-miR-18b transcripts reveals a fine difference in the stem architecture. Cleavage patterns obtained for the 5' ^{32}P -labeled pri-miR-18a (left panel) and pri-miR-18b (right panel) transcripts (100×10^3 c.p.m.) treated with: Pb (II) – lead ions at increasing concentrations (0.5, 1 mM), T1, ribonuclease T1 (1, 1.5 units/ μl), S1, nuclease S1 (1.25, 2.5 units/ μl ; 1 mM ZnCl_2 was present in each reaction mixture) and V1, ribonuclease V1 (0.037, 0.075 unit/ml). C, incubation control (without probe). F and T identify nucleotide residues subjected to partial digest formamide (every nucleotide) or with ribonuclease T1 (G-specific cleavage), respectively. Electrophoresis was performed in a 10% polyacrylamide gel under denaturing conditions. The positions of selected G residues are shown. The arrows indicate a dinucleotide mismatch in pri-miR-18a and a dinucleotide bulge in pri-miR-18b, respectively.

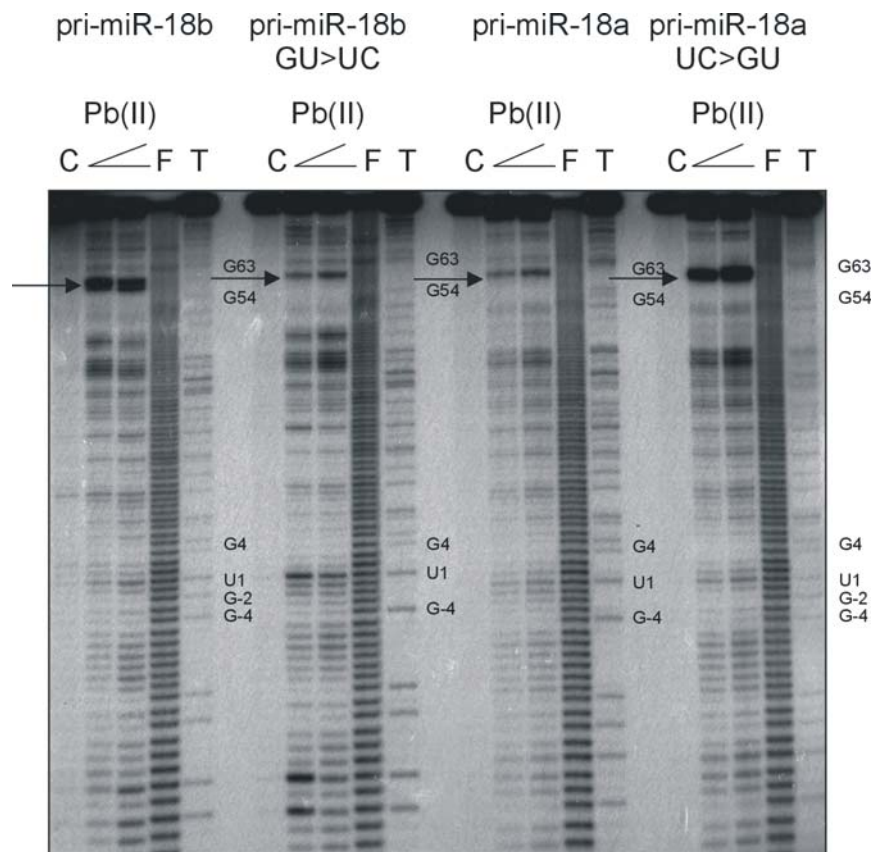


Figure S4. Swapping mutations in the stems of pri-miR-18a and 18b force local changes in the conformation of the stem loops, without perturbing the rest of the structure. Condition of analysis and abbreviations are as described for Figure S3.

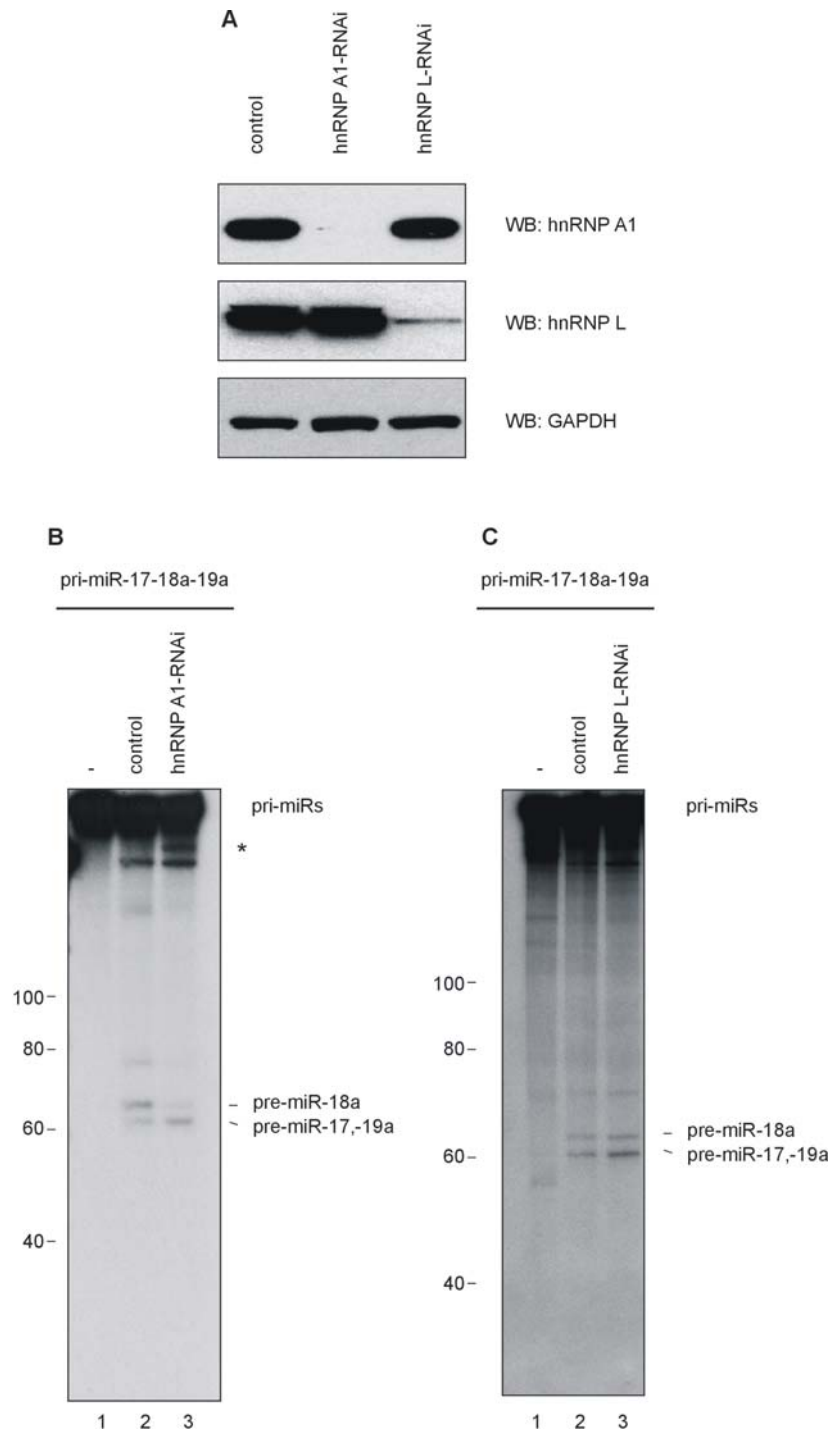


Figure S5. (A) Western blot analysis of HeLa extracts depleted of hnRNP A1 or hnRNP L using specific siRNAs. (B) The processing of pri-miR-18a is dependent on the presence of hnRNP A1. *In vitro* processing of pri-miR-18a in the pri-miR-17-18a-

19a mini cluster is abolished in HeLa cell extracts prepared using different set of siRNAs (hnRNP A1-targeted smart pool of siRNAs from Dharmacon). Primary RNA transcripts were incubated in the presence of either control HeLa extract (lane 2) or hnRNP A1-depleted extract (lane 3). Lane 1 shows negative controls with no extract added. Products were analyzed on an 8% polyacrylamide gel. The asterisk indicates the remaining transcript containing uncleaved pre-miR-18a substrate. (C) The processing of pri-miR-18a is not affected by depletion of hnRNP L. Analysis and data interpretation was performed as described for panel B.

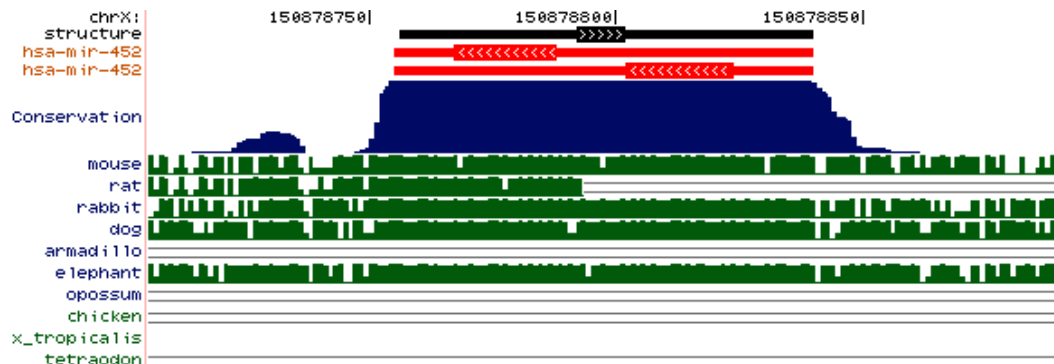
miRNA genes showing significant conservation of predicted loop regions. In each case the miRBase identifier is given followed by the chromosomal position (and strand), the miRBase predicted secondary RNA structure and a diagram displaying overall conservation throughout vertebrate evolution. Each diagram indicates the position of the miRNA (red line), predicted loop position (hatched black line), overall PhastCons conservation score (blue graph) and conservation within available vertebrate species.

hsa-mir-452 chrX:150878756-150878840 (-)

```

      u          AAC G          G A          uuug
gc aagcacuuac  U UUUGCAGA GA ACUGAgac  u
|| |||||      | |||||      || |||||      a
cg uucGUGAAUG  A AAACGUCU CU ugacucug  a
      u          --A G          A C          uauc

```

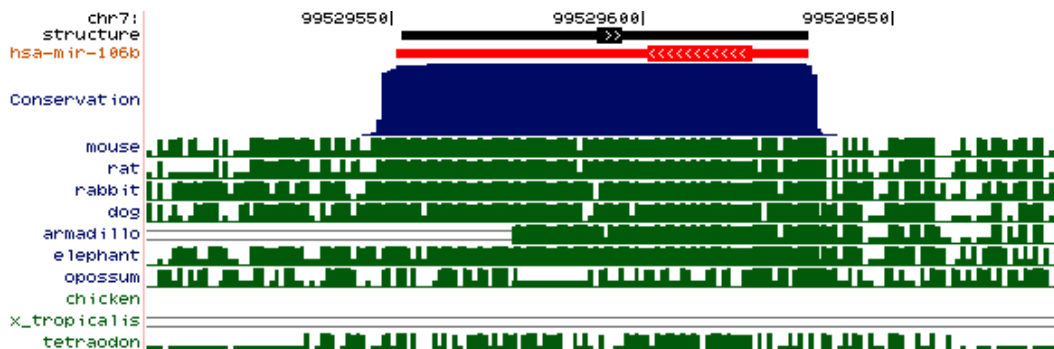


hsa-mir-106b chr7:99529552-99529633 (-)

```

      c      -UA      G      A A      -- uc
ccugc ggggc  AAGUGCU ACAGUGC G Uagu gg c
||| | | | |  ||| | | | | ||| | | | | ||
ggacg ccuCG  UUCAUGG UGUCACG C aucg cc u
      a      UCG      G      C -      ug uc

```

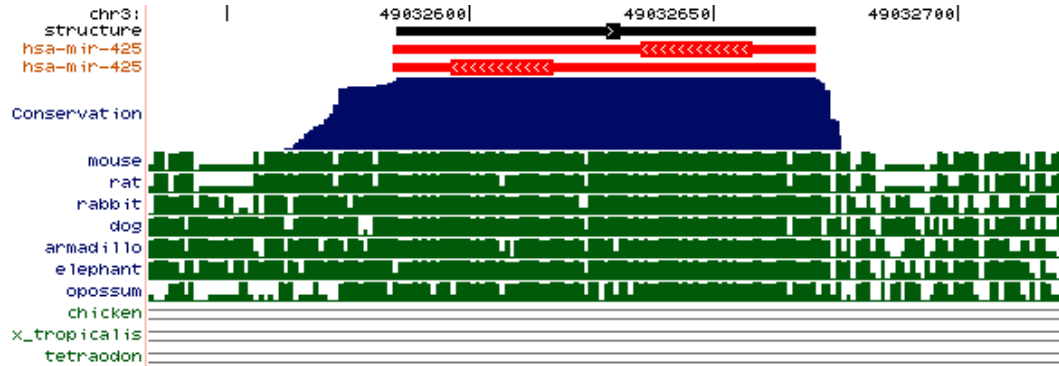


hsa-mir-425 chr3:49032585-49032671 (-)

```

      c u   AAU       U C   U A   ----- g
gaaag gc uugg   GACACGA CA UCCCG UG gu   gg c
||||| ||| ||||   ||||| ||| ||||| ||| |||   ||
cuuuc cg gaCC   CUGUGCU GU AGGGC Ac cg   cc a
      u u   CGC       - A   U   - aagag c

```

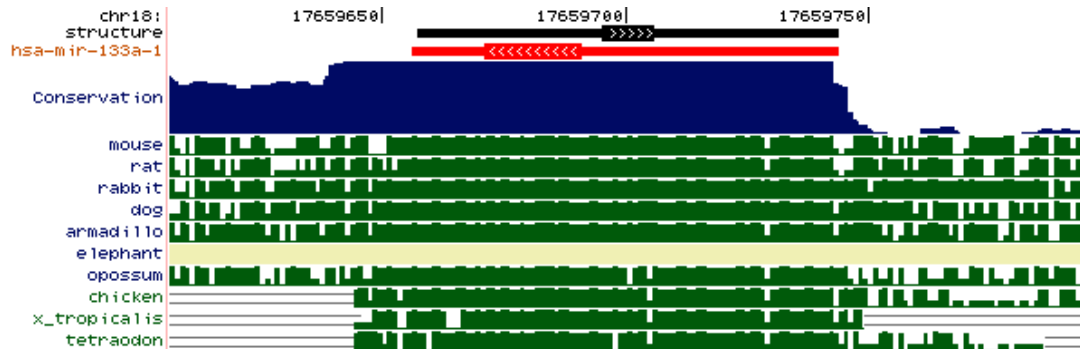


hsa-mir-133a-1 chr18:17659657-17659744 (-)

```

a   uu   g   aa u a   gccuc
caaugc gcua agcuggu aa gg accaaauc   u
||||| ||| ||||| ||| ||| ||||| ||| |||
guuacg cgau UCGACCA UU CC UGGUUUag   u
a   uau   G   AC C C   guaac

```



hsa-mir-181a-2 chr9:126494542-126494651 (+)

```

agaagggcuaucaaggccagccuua           A U   CU   A   ggga
      gaggacuccaagg ACA UCAACG   GUCGGUG GUuu   u
      ||||| ||||| ||| ||| ||| ||| ||| ||| |||   u
      uuccuggggguuCC UGU AGUUGC   CAGUCAC CAaa   u
-----a           A C   --   -   aaag

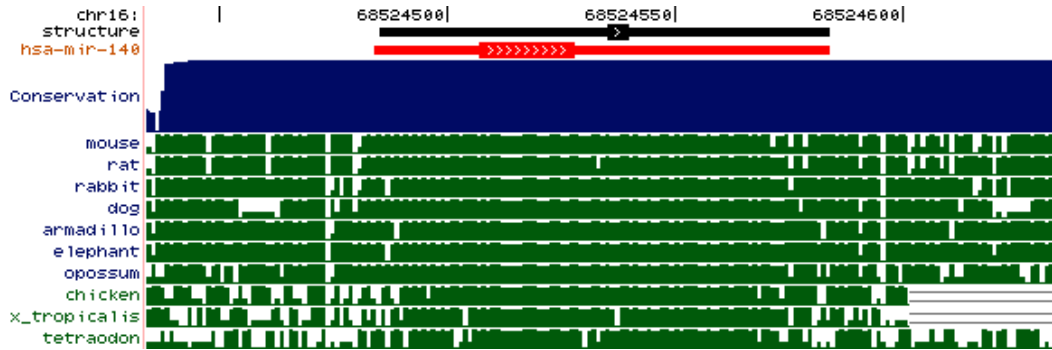
```



```

u      ucucu      - A      A      uu      uc
gugucuc      guguccug      cC      GUGGUUUUACCCU      UGGUAGg      acg      a
|||||      |||||      ||      |||||      |||||      |||||      |||
cacgggg      cauaggac      GG      CACCAAGAUGGGA      ACCAUcu      ugu      u
c      ----c      a      -      C      --      cg

```

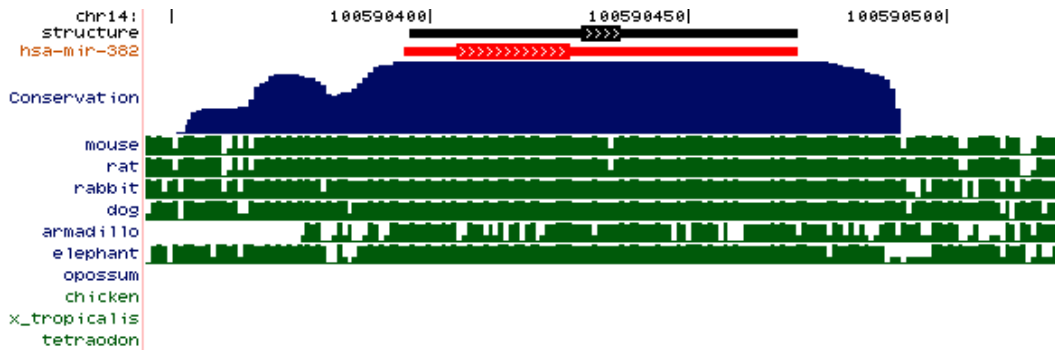


hsa-mir-382 chr14:100590396-100590471 (+)

```

u      -A      UG      - uuu
uacu      gaagaGA      GUUGUUCGUGG      GAUUCG      c      a
||||      |||||      |||||      |||||      |      c
auga      cuuuuu      caacaggcacu      cuaagc      g      u
-      ca      ua      a      uau

```

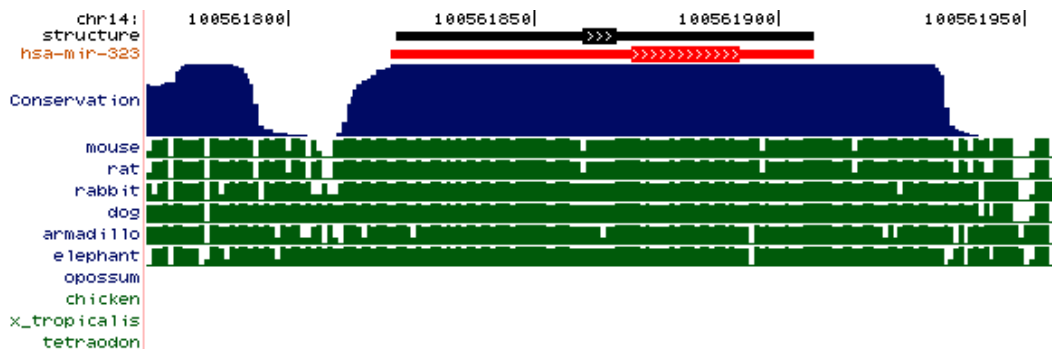


hsa-mir-323 chr14:100561822-100561907 (+)

```

---uu      u      g      G      U      GCGC      U      uua
gguacu      g      agagAGGU      G      CCGUG      GU      CGCu      u
||||      |      |||||      |      |||||      ||      |||||
cuauga      c      uuUCUCCA      C      GGCAC      CA      gcgg      u
cuauu      -      g      G      U      AUUA      C      uau

```

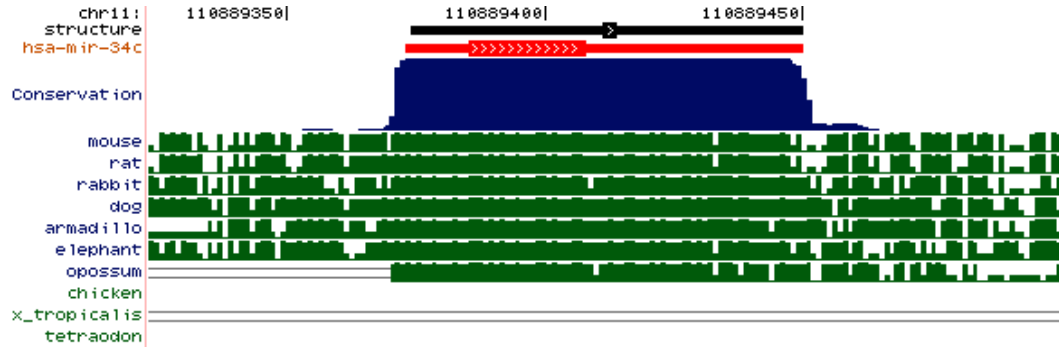


hsa-mir-34c chr11:110889374-110889450 (+)

```

ag   A   A   A   C   C   a
agucu uuacu GGC GUGU GUUAG UGAUUG ua u
||||| ||||| ||| ||||| ||||| ||||| |||
uuaga aaUGG CCG CACA CAAUC ACUAAc au a
aa   A   G   C   -   c   g

```

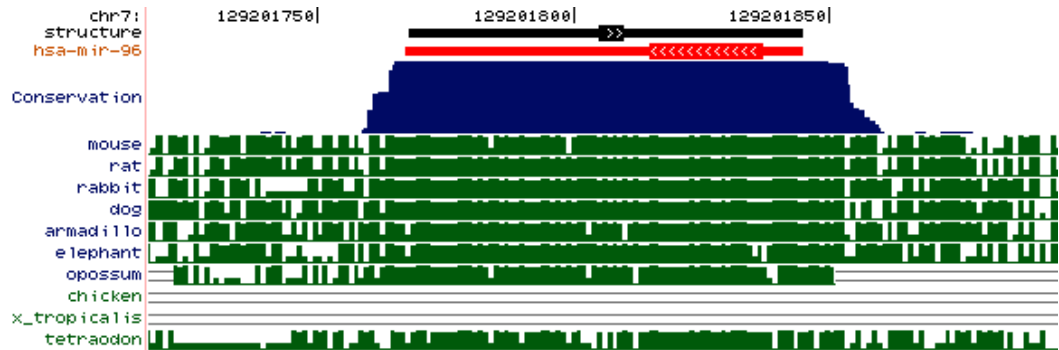


hsa-mir-96 chr7:129201768-129201845 (-)

```

ugg g U   A   UU   --- uc
cc au UUGCACU GCACAU UUGCU gug u
|| || ||||| ||||| ||||| |||
gg UA AACCGUGA CGUGUA AAcgag cgc c
aaa G U   -   CU   ucu cu

```

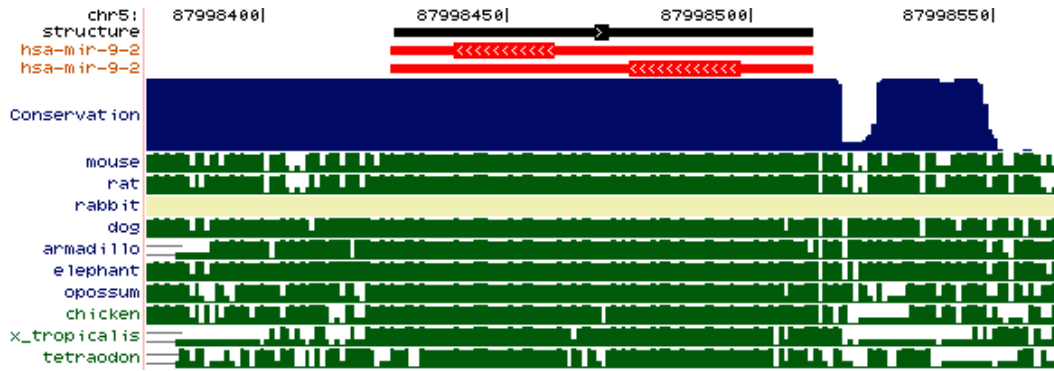


hsa-mir-9-2 chr5:87998427-87998513 (-)

```

g   c   g   UC           G   ug a
gaag gaguu uua UUUGGUUAUCUAGCU UAUGAg u u
|||| | ||| ||||| ||||| ||||| |||
cuuc cucaa aaU AAGCCAAUAGAUCGA AUAcuu g u
a   -   a   GA           A   cu g

```

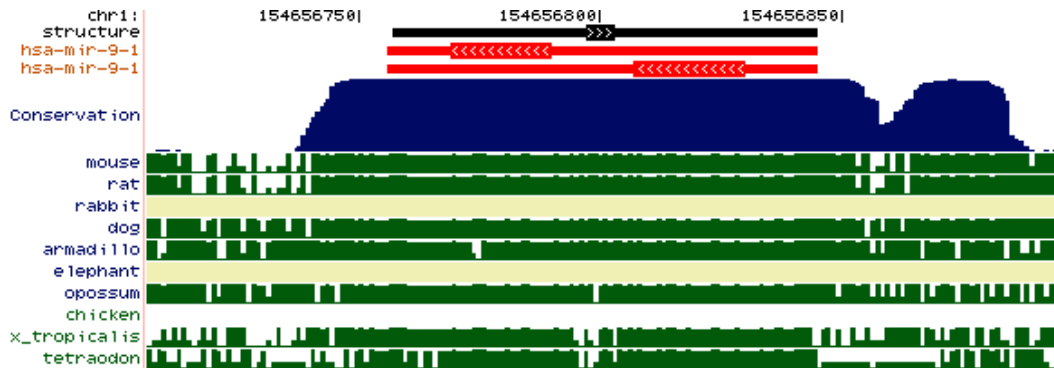


hsa-mir-9-1 chr1:154656757-154656845 (-)

```

c   guug  UC           G   u  ug
gggguug  uua  UUUGGUUAUCUAGCU  UAUGAg  gg  u
|||      |||  |||      |||      |||      |||  ||  g
ccccaau  aaU  AAGCCAAUAGAUCGA  AUAcuu  cu  g
a   -aaa  GA           A   -  ga

```

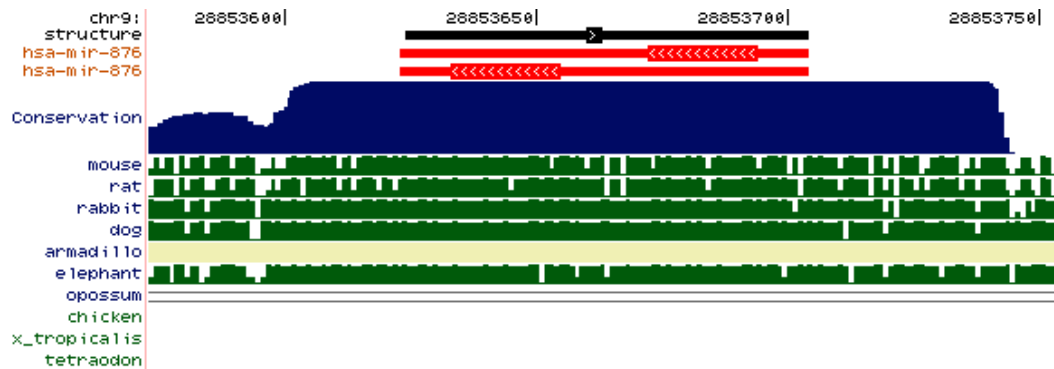


hsa-mir-876 chr9:28853624-28853704 (-)

```

u   ug   U           aucuaa  u
gaag  cugUGGAUU  CUUUGUGAAUCACCAu  gc  a
|||   |||      |||      |||      |||      ||  ||
cuuc  gauACUUA  GAAACAUUUGGUGUGu  ug  a
a   gu   U           -----g  u

```

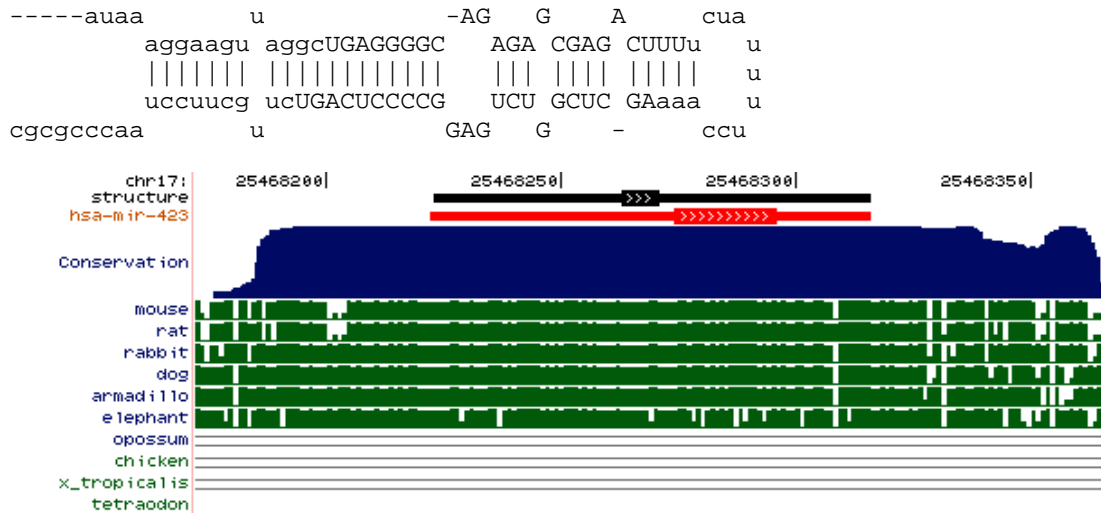


hsa-mir-873 chr9:28878877-28878953 (-)

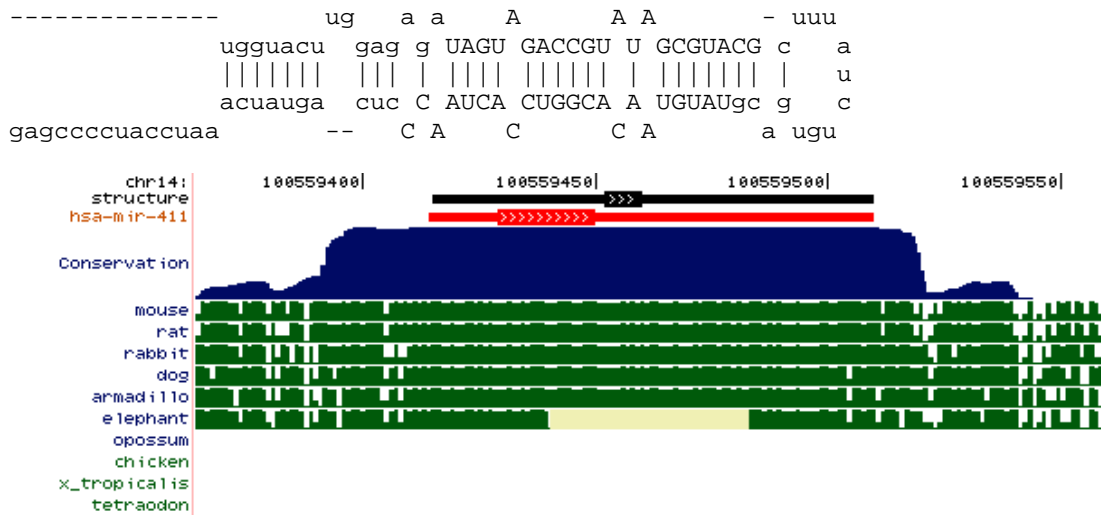
```

--  u  ca  G  A   UG  G   a  ga
gug  g  uuu  C  GGAACU  U  AGUCUCCU  uu  a

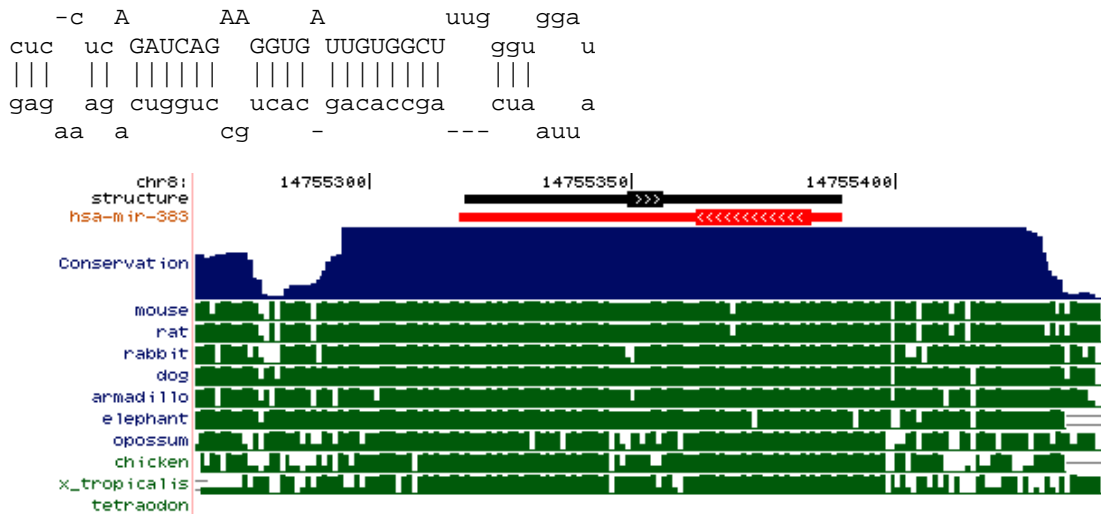
```

hsa-mir-411 chr14:100559415-100559510 (+)

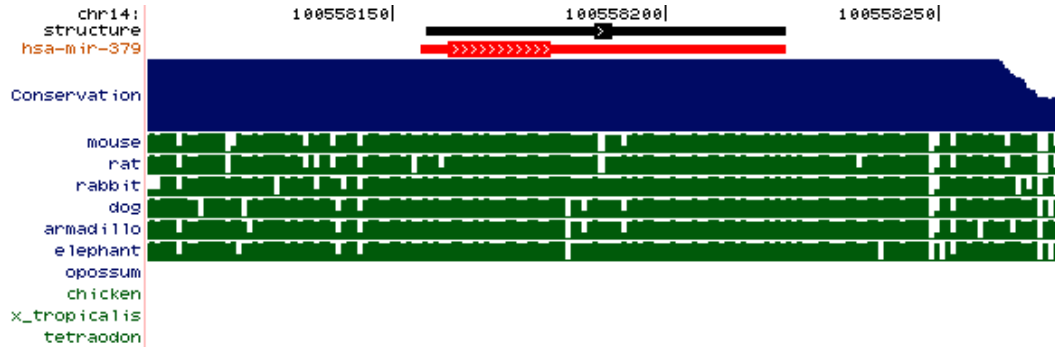


hsa-mir-383 chr8:14755318-14755390 (-)



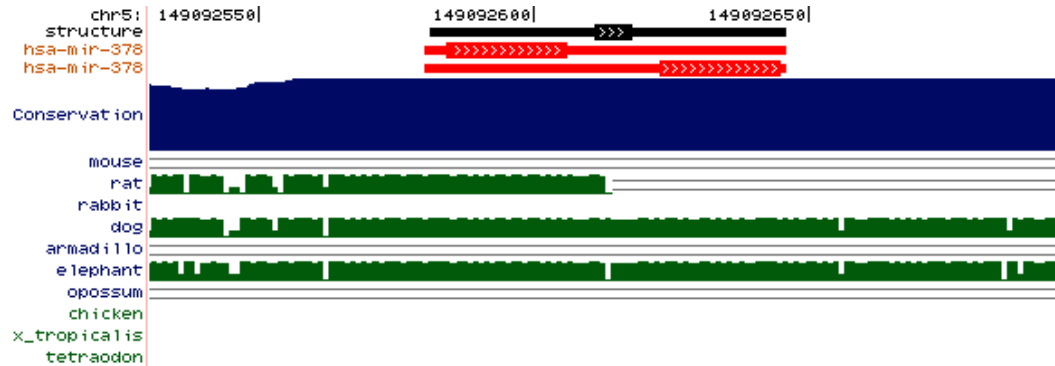
hsa-mir-379 chr14:100558156-100558222 (+)

```
      a  A      GA      - uu u
agag UGGU GACUAUG  ACGUAGG cg a g
||||| ||||| ||||| ||||| ||| |
ucUC AUCA CUGGUAC  UGUAUcc gu u a
      A  C      AA      a cu u
```



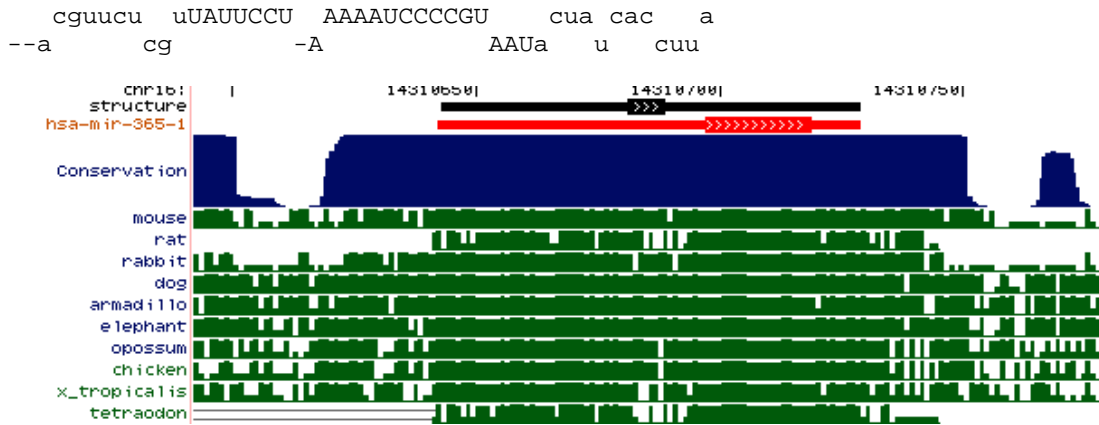
hsa-mir-378 chr5:149092581-149092646 (+)

```
      g  C      UGU      ccu
agg CU CUGACUCCAGGUCC  GUguua a
||| || ||||| ||||| ||||| |||||
ucc GA GACUGAGGUUCAGG  CAcgau g
      G  A      --U      aaa
```

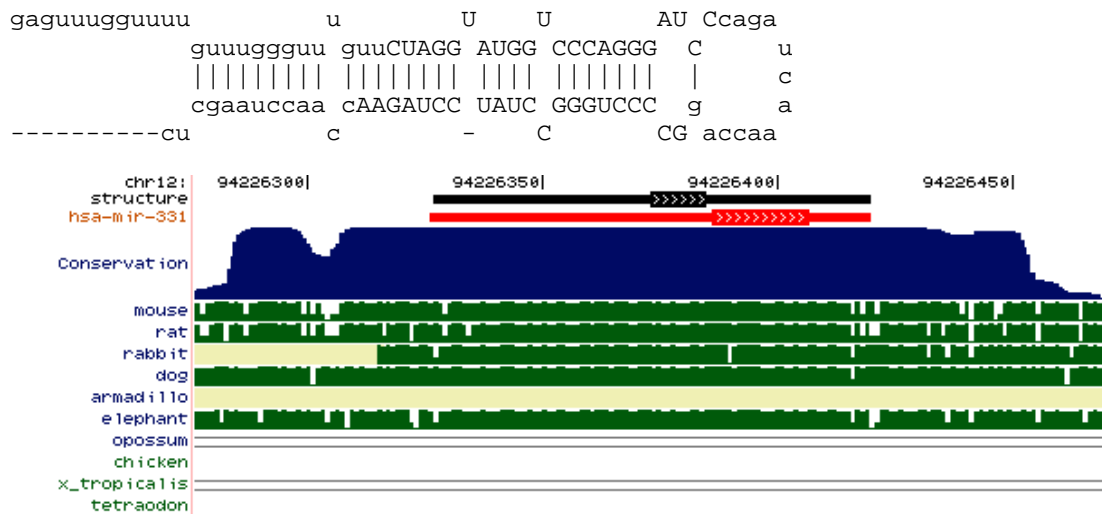


hsa-mir-377 chr14:100598140-100598208 (+)

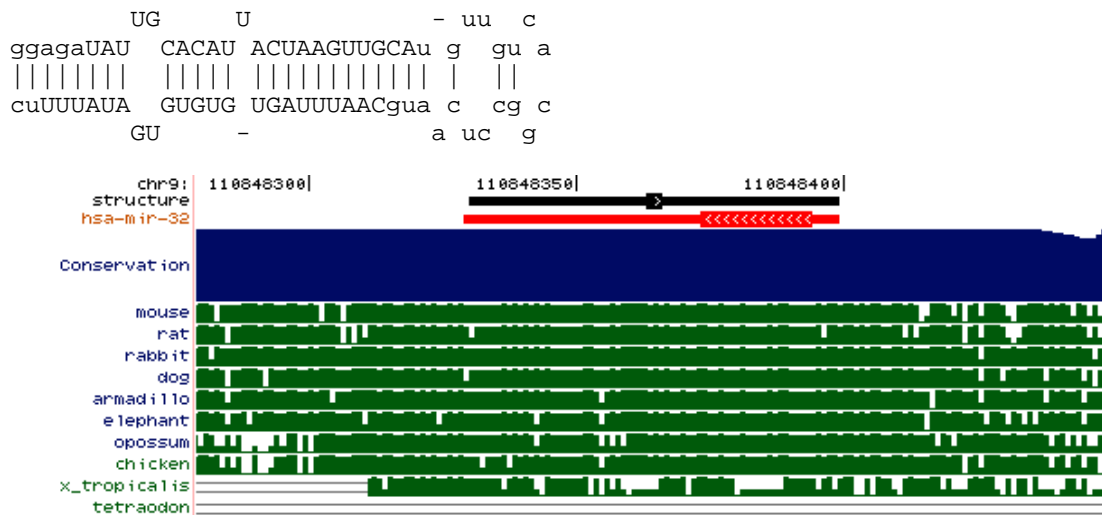
```
uu      C  -  A  - uuu
gagcAGAGGUUGCC UUG GUGA UUCg c a
||||| ||||| ||||| ||||| ||| | u
uuUGUUUUCAACGG AAC CACU Aagu g u
-g      A  A  -  u uau
```

hsa-mir-331 chr12:94226327-94226420 (+)



hsa-mir-32 chr9:110848330-110848399 (-)



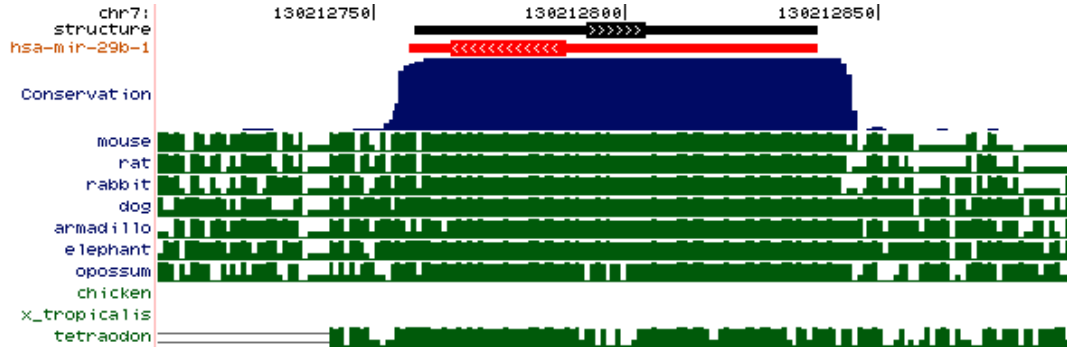
hsa-mir-31 chr9:21502114-21502184 (-)

gA G C -U gaa


```

-          -          U          GU          uuaaa
cuucagga GCUGGUUUCA AUGGUG UUAGAu      u
|||||    |||||    |||||    |||||    |||||    a
ggguucUU  UGACUAAAGU UACCAC GAUcug      g
g          G          U          --          uuagu

```

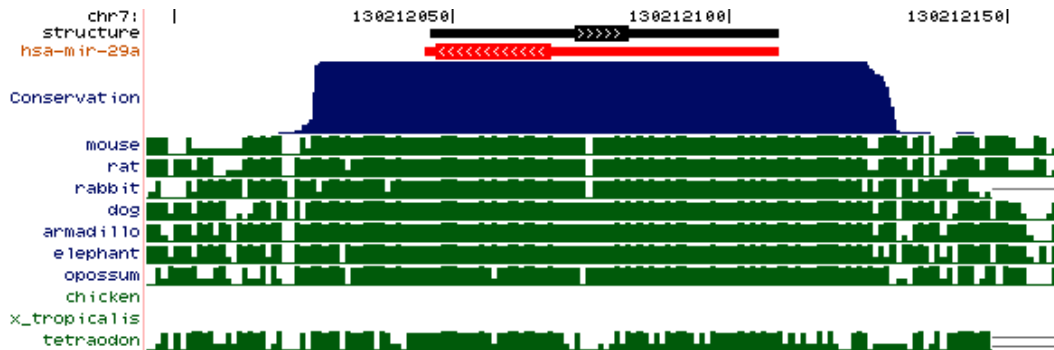


hsa-mir-29a chr7:130212046-130212109 (-)

```

UUU          C          ucaa
augACUGAUUUC UGGUGUU AGag      u
|||||    |||||    |||||    a
uAUUGGCUAAAG ACCACGA Ucuu      u
UCU          -          uuaa

```

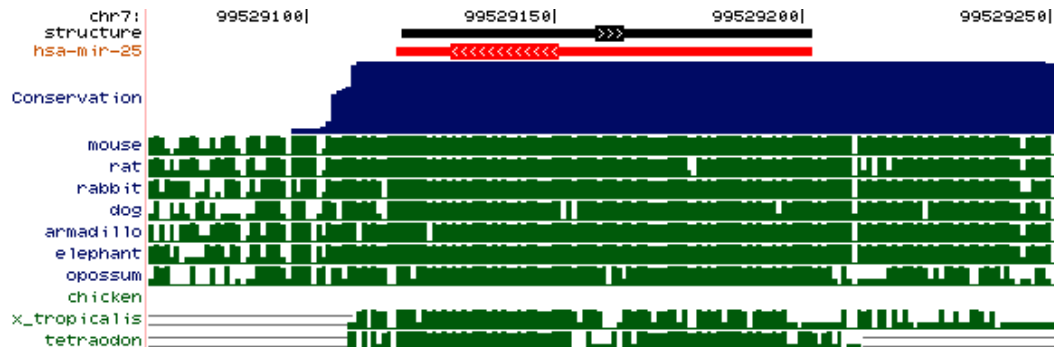


hsa-mir-25 chr7:99529119-99529202 (-)

```

a ug ag G UU G U -- ac
ggcc g uug AGGC GAGAC G GCAAU Gcu gg g
||||| | ||| ||||| ||||| | ||||| ||| || c
ccgg c gac UCUG CUCUG C CGUUA Cgg cc u
c gu AG G UU A - gu cg

```

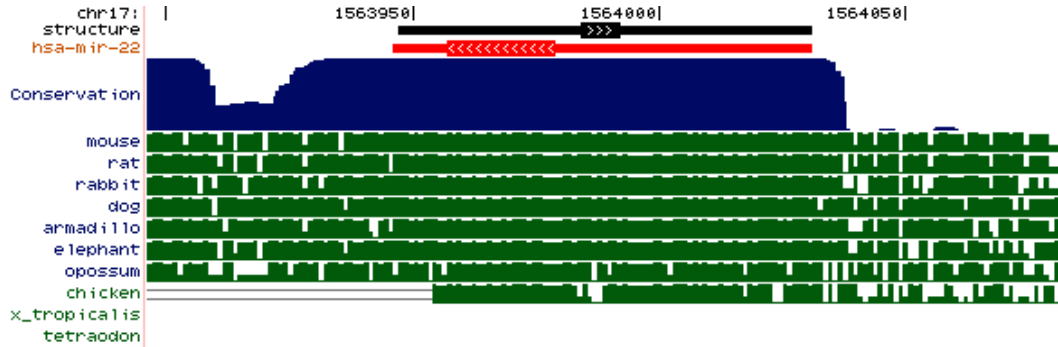


hsa-mir-22 chr17:1563947-1564031 (-)

```

u   cc           -   A   u   ccu
ggc gag gcaguAGUUCUUCAG UGGCA GCUUUA gu   g
||| ||| ||||||||||||||| ||||| ||||| ||   a
ccg cuc cguUGUCAAGAAGUU ACCGU CGAAau cg   c
u   -c           G   -   -   acc

```

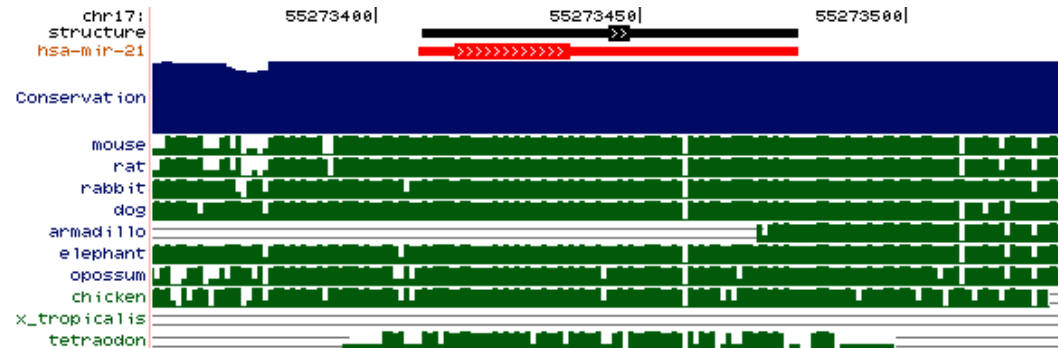


hsa-mir-21 chr17:55273409-55273480 (+)

```

u   gU           A   A   A   u   a
gucgg AGCUUAUC GACUG UGUUG cugu g a
||| ||| ||||||||||| ||||| ||||| |   u
caguc UCGGGUAG CUGAC ACAAC ggua c c
a   UG           -   C   -   -   u

```



hsa-mir-204 chr9:72614711-72614820 (-)

```

ggcuacagucuuucu - -   ucg   U           A   U   gagaau
          uca ug ugac   uggac UCCCUUUGUC UCCUA GCCU   a
          ||| ||| ||||| ||||| ||||| ||||| |||||   u
          ggu ac acug   acuug agggaaacgg agggg cggg   a
-----c   c   u   uua   c           a   -   ggaagu

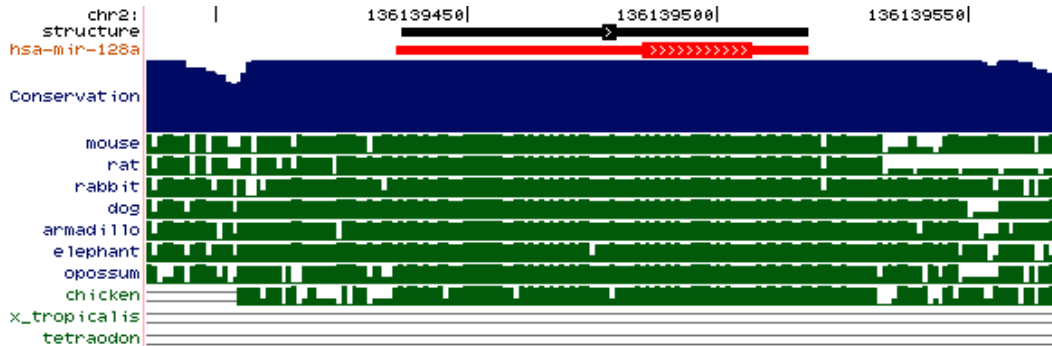
```



```

u   u   uuc   uag   cu   u
gagc guugga   ggggccg   cacugu   gagaggu u
||| | |||||   ||||| |||   ||||| |||||
uucg cgacuu   CUCUGGC   GUGACA   CUuuua a
c   u   UUU   CAA   --   c

```

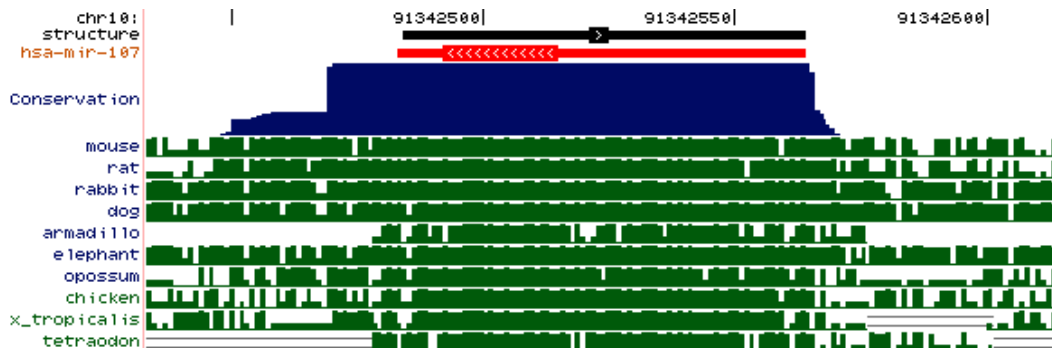


hsa-mir-107 chr10:91342484-91342564 (-)

```

c   c   --c   u   u   c   u   a
ucu ugcuuu   agcu cu uacaguguugc uug ggc u
||| | |||||   ||||| || | ||||| ||||| ||| ||| g
aga acgaaA   UCGG GA AUGUUACGACG Aac uug g
-   c   CUA   -   C   -   -   a

```

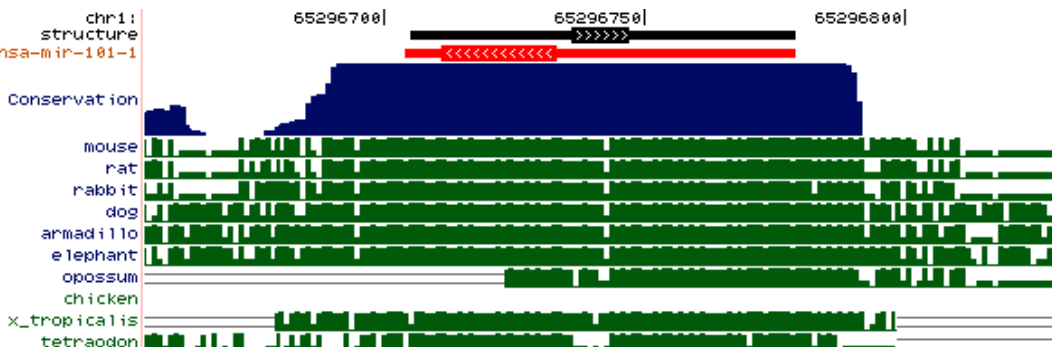


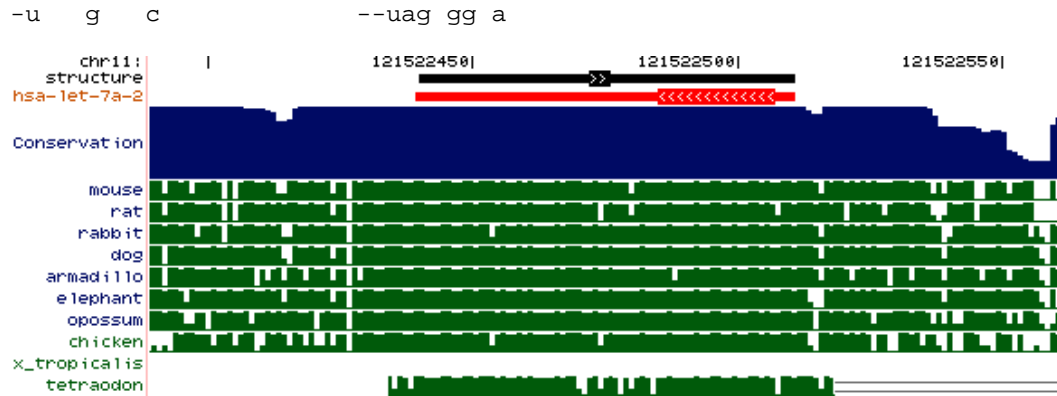
hsa-mir-101-1 chr1:65296705-65296779 (-)

```

u   cuggc   A   gucua
gcc   uCAGUUUAUCACAGUGCUG UGCU   u
|||   ||||| ||||| ||||| ||||| |||||
cgg   AGUCAAUAGUGUCAUGAC AUgg   u
a   uaggA   -   aaauc

```

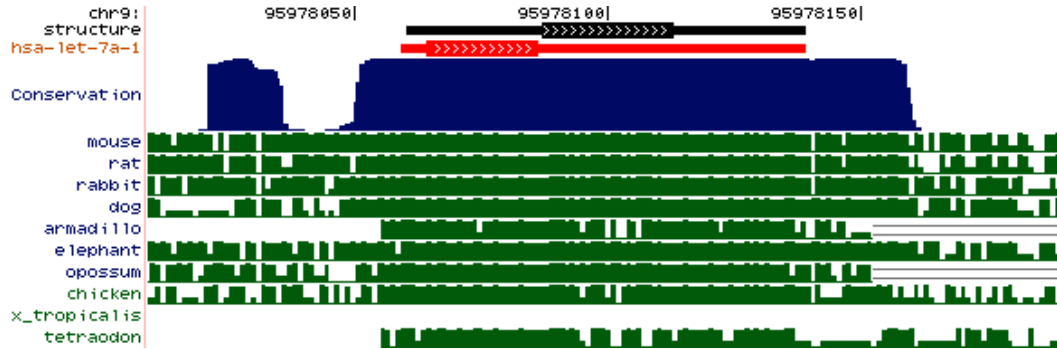




hsa-let-7a-1 chr9:95978060-95978139 (+)

```

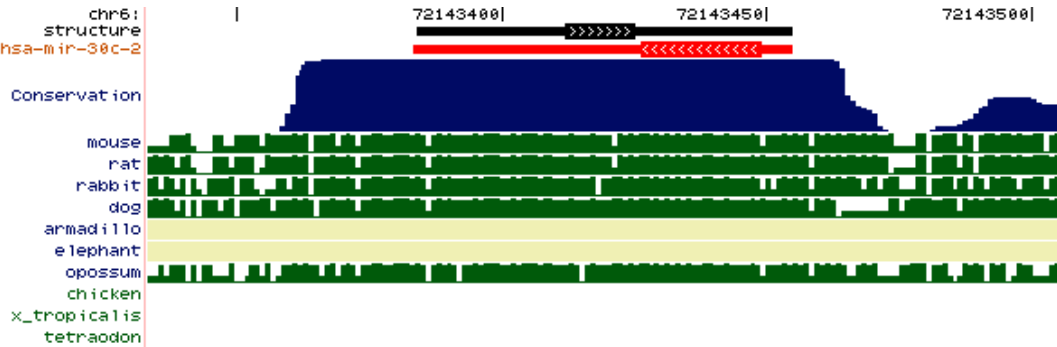
      U  GU                uuagggucacac
uggga GAG AGUAGGUUGUAUAGUU          c
|||   |||   |||   |||   |||   |||   |||   |||   |||   |||   |||   |||   |||
aucCU UUC UCAUCUAACAUAUCaa          a
-   UG                uagagggucacc
  
```



hsa-mir-30c-2 chr6:72143384-72143455 (-)

```

uacU      U  ACA      guggaa
aga  GUAAACA CCU  CUCUCAGCu  a
|||   |||   |||   |||   |||   |||   |||   |||   |||   |||   |||   |||
ucu  CAUUUGU GGA  GAGGGUCga  g
uUCU      C  --A      aagaau
  
```



hsa-mir-628 chr15:53452430-53452524 (-)

```

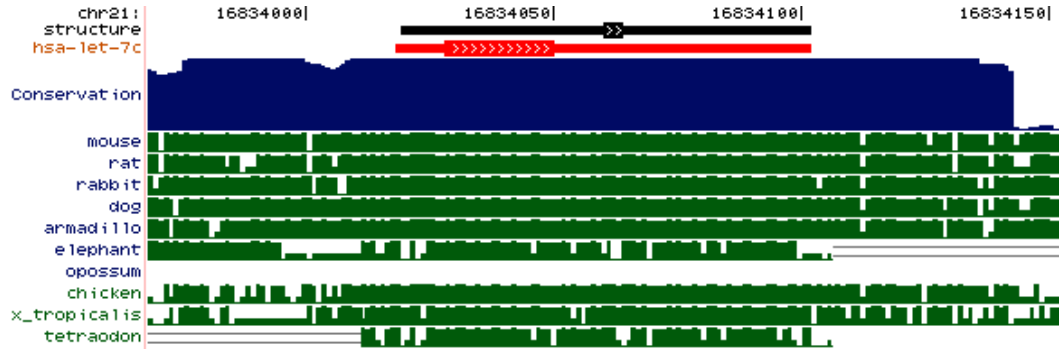
auagcugu u  a  cA - A A      aaaa
      ug  guc cuuccu  UG CUG CAU UUUACUAGAGGgu  u
  
```


hsa-let-7c chr21:16834019-16834102 (+)

```

a      uU  G  U              UA  G  UA  A
gc uccggg GAG UAG AGGUUGUAUGGUU GA U  C  C
|| ||||| ||| ||| ||||| ||||| ||| | | C
cg agguuc uuc auc uccaacaugucaA UU A  G  C
-      cu  g  u              -- G  GG  U

```

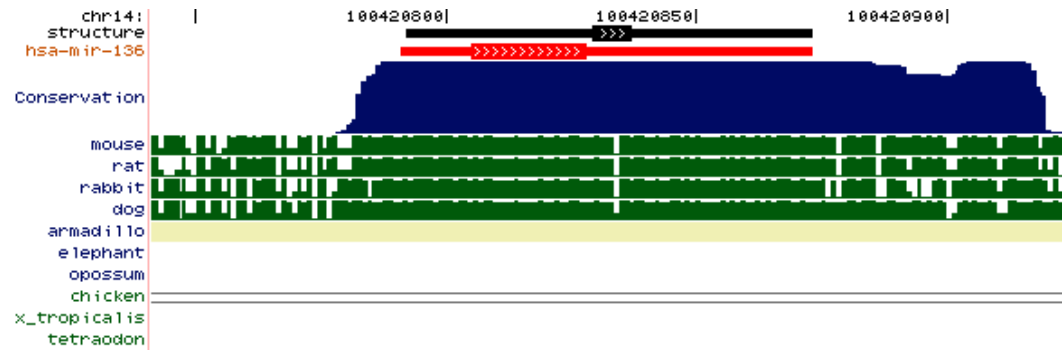


hsa-mir-136 chr14:100420792-100420873 (+)

```

u      g      C      UUU      uuc
gagccuc gaggACUC AUUUG  UGAUGAUGGA u
||||||| ||||| ||||| ||||| ||||| u
cuugggag cuUCUGAG UAAAC  GCUACUACcu a
u      a      -      UCU      cgu

```



hsa-mir-361 chrX:85045297-85045368 (-)

```

UU      --U  A  U      auaa
ggagc  AUCAGAAUC  CC GGGG ACuuu  u
|||||  ||||| ||| ||| ||| |||
cuucg  UAGUCUUAG  GG CCCC Ugaaa  u
UU      UGU  A  C      aacu

```


Figure S6. Multiple sequence alignments for the human genome with 16 other vertebrate genomes of human 74 pri-miRNAs that show high levels of conservation in their terminal loop regions.

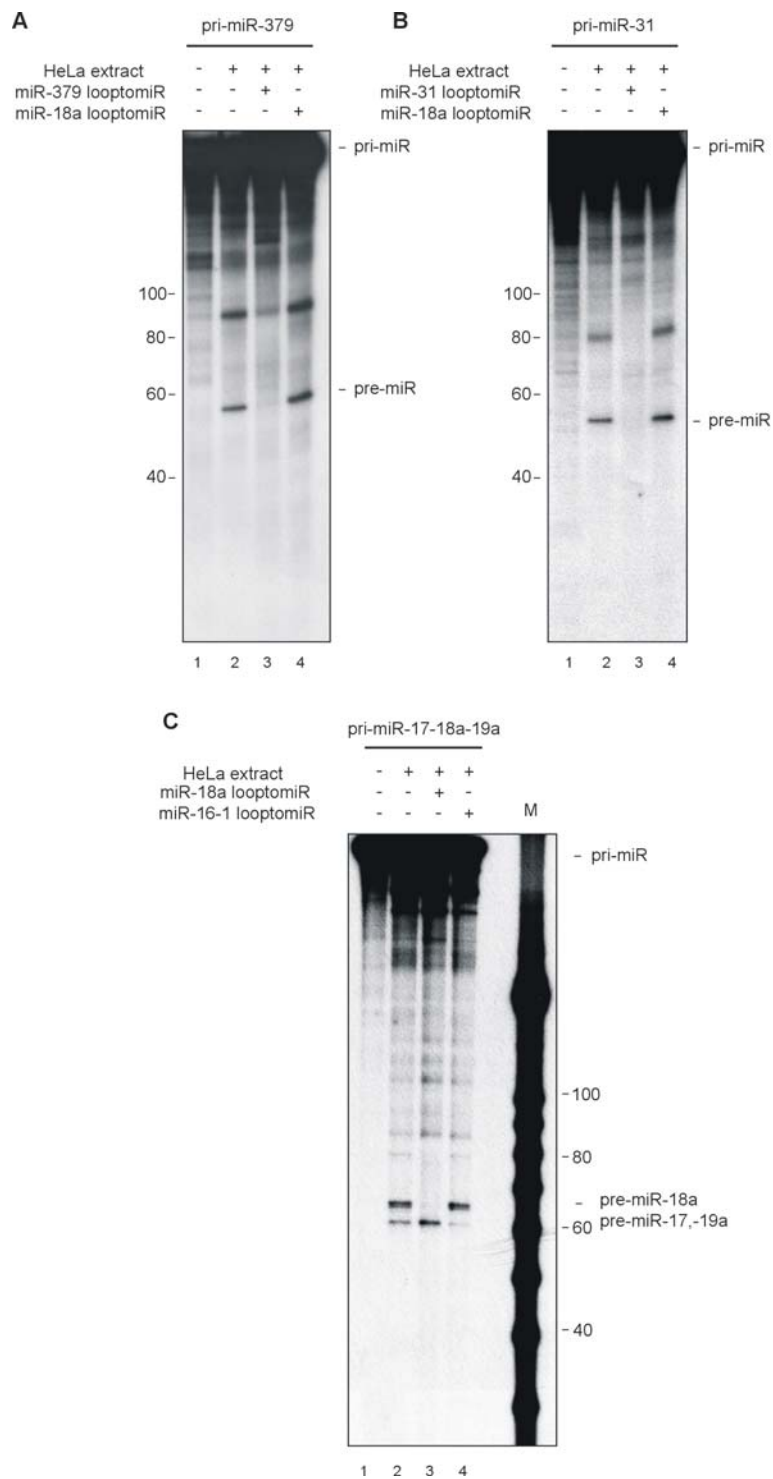


Figure S7. (A, B) The *in vitro* processing of pri-miRNAs-379 and pri-miR-31 is sensitive to the presence of specific looptomiRs. Radiolabeled pri-miR-379 and pri-

miR-31 (50×10^3 c.p.m.) were processed in HeLa extracts with the addition of a specific looptomiR (lanes 3) or with a control looptomiR specific for miR-18a (lanes 4). As a control, the reaction was also carried out in the absence of any looptomiR (lanes 2). Lanes 1 represent control reaction without extract. M, RNA size marker. (C) A miR-18a looptomiR selectively abolishes the processing of pri-miR-18a in the context of the miR-17-18a-19a mini cluster. The RNA substrate comprising pri-miR-17, pri-miR-18a and pri-miR-19a (50×10^3 c.p.m.) was processed *in vitro* in the presence of a miR-18a looptomiR (lane 3), or a control looptomiR (lane 4). Lane 1 shows negative controls with no extract added.

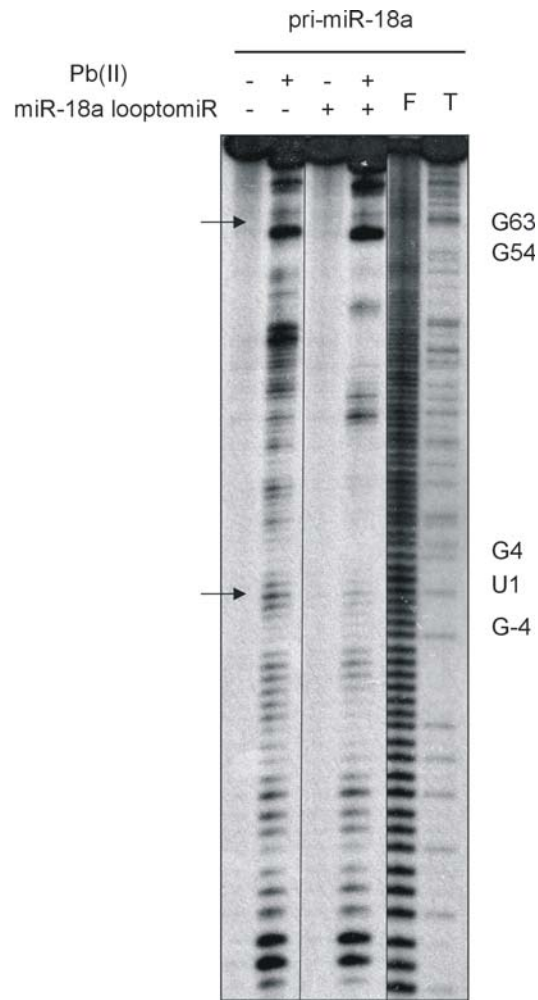


Figure S8. A LooptomiR targeting pri-miR-18a binds to the terminal loop region without disturbing RNA structure of the region surrounding the Drosha cleavage site. Cleavage patterns obtained for the 5' ³²P-labeled pri-miR-18a transcript (100x10³ c.p.m.) treated with Pb (II) – lead ions (0.5mM) with or without addition of specific looptomiR (50μM) are shown. Drosha cleavage sites are indicated with black arrows.

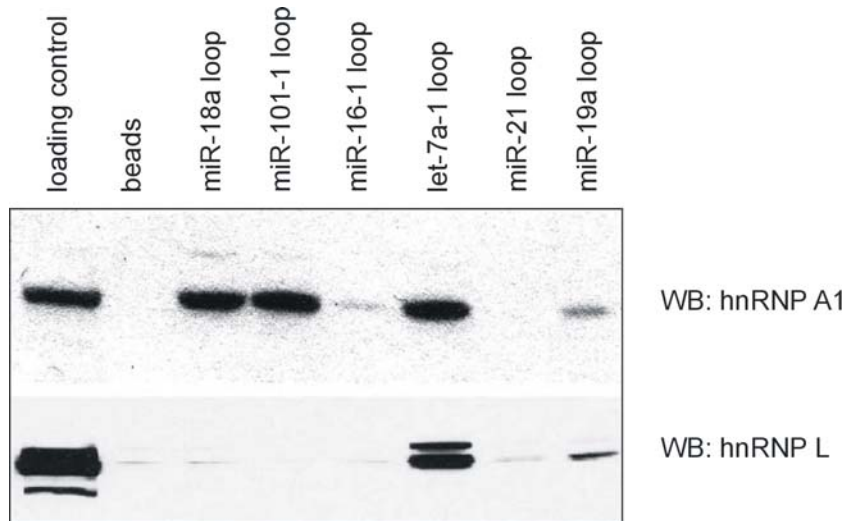


Figure S9. RNA chromatography of selected pri-miRNAs harboring conserved (miR-18a, miR-101-1 and let-7a-1) or not conserved (miR-16-1, miR-21, miR-19a) loops was performed in HeLa nuclear extracts. Samples resulting from RNA chromatography were resolved on SDS gel and visualized by Western blot analysis with specific antibodies against hnRNP A1 and hnRNP L.

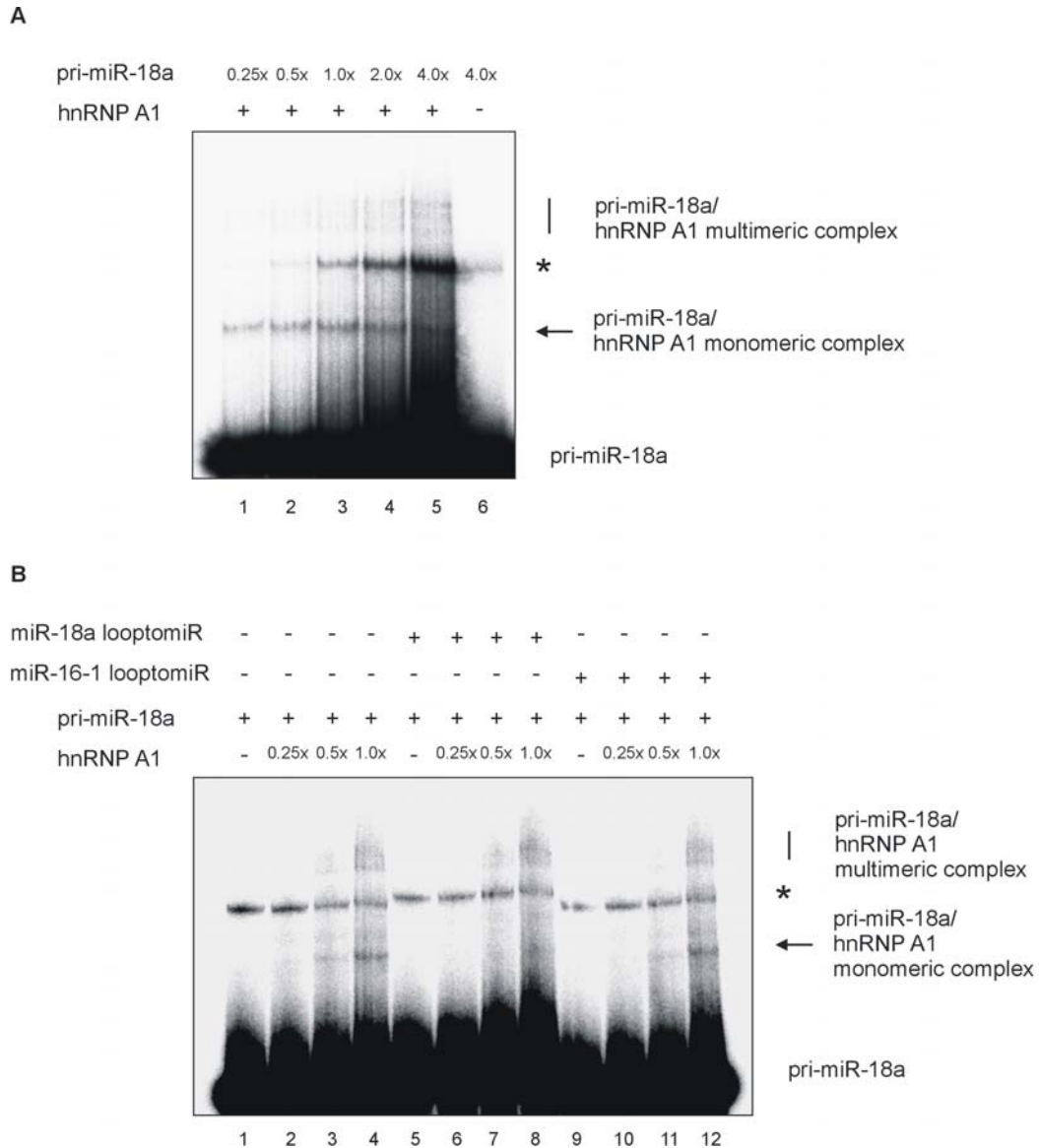


Figure S10. Several hnRNP A1 molecules bind pri-miR-18a and an specific looptomiR blocks the formation of one of the complexes. (A) EMSA analysis of pri-miR-18a transcript with recombinant hnRNP A1 protein. Native gel electrophoresis with increasing amounts of 5' ³²P-labeled transcript (25x10³ c.p.m. up to 400x10³ c.p.m./ 0.5 pmol up to 8.0 pmol) incubated in the presence of constant amounts of recombinant hnRNP A1 protein (the molar ratio indicated) reveals several, stoichiometrically different, complexes. The asterisk indicates the minor structural

conformer of pri-miR-18a transcript. (B). EMSA analysis of pri-miR-18a/hnRNP A1 complexes formed in the presence of looptomiRs. Equal amounts of 5' ³²P-labeled pri-miR-18a (100x10³ c.p.m/ 2 pmol) incubated with increasing amounts of hnRNP A1 (0.5 pmol up to 2 pmol) were programmed with a specific LooptomiR targeting pri-miR-18a specific or acontrol looptomiR (50μM) and assayed as described above.

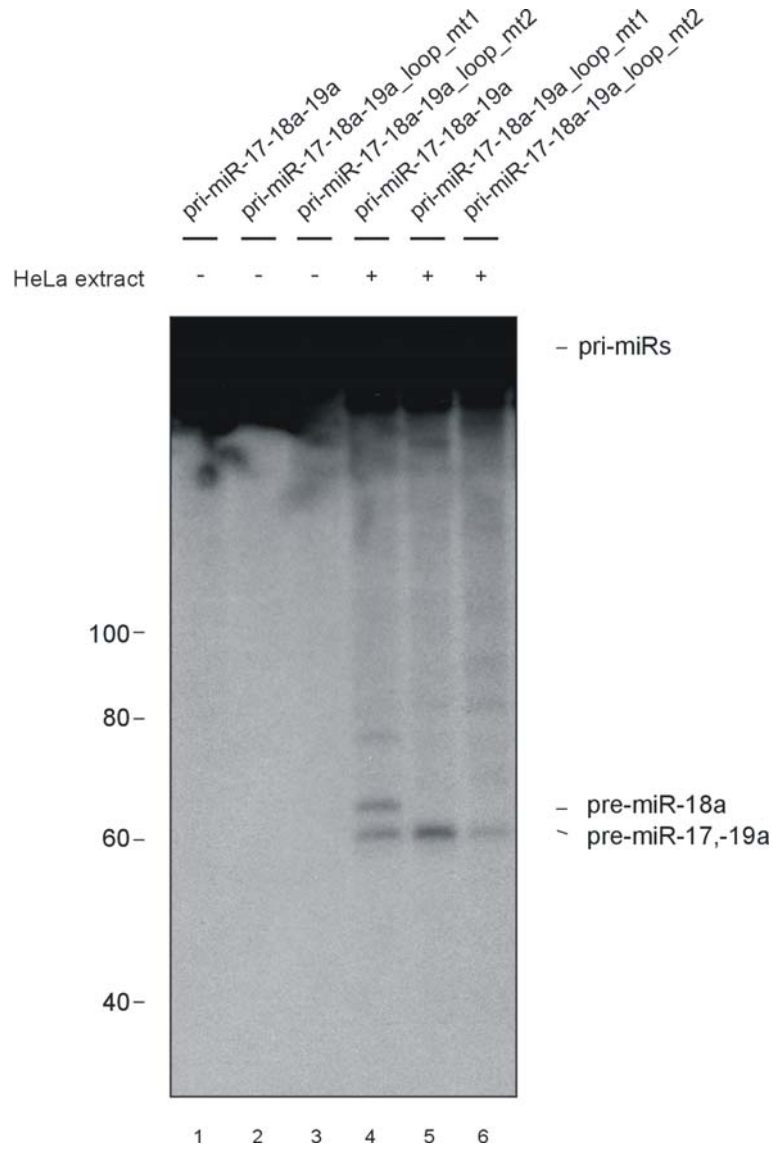


Figure S11. The conserved terminal loop of *pri-miR-18a* is required for its efficient cleavage by Droscha in the natural context of other *pri-miRNAs*. *In vitro* processing of *pri-miR-17-18a-19a* and *pri-18a* loop mutants in the mini cluster. Radiolabeled *pri*-RNAs (50×10^3 c.p.m.) were incubated in HeLa cell extracts (lanes 4, 5 and 6). Lanes 1, 2 and 3 show negative controls with no extract added. Products were analyzed on an 8% polyacrylamide gel.

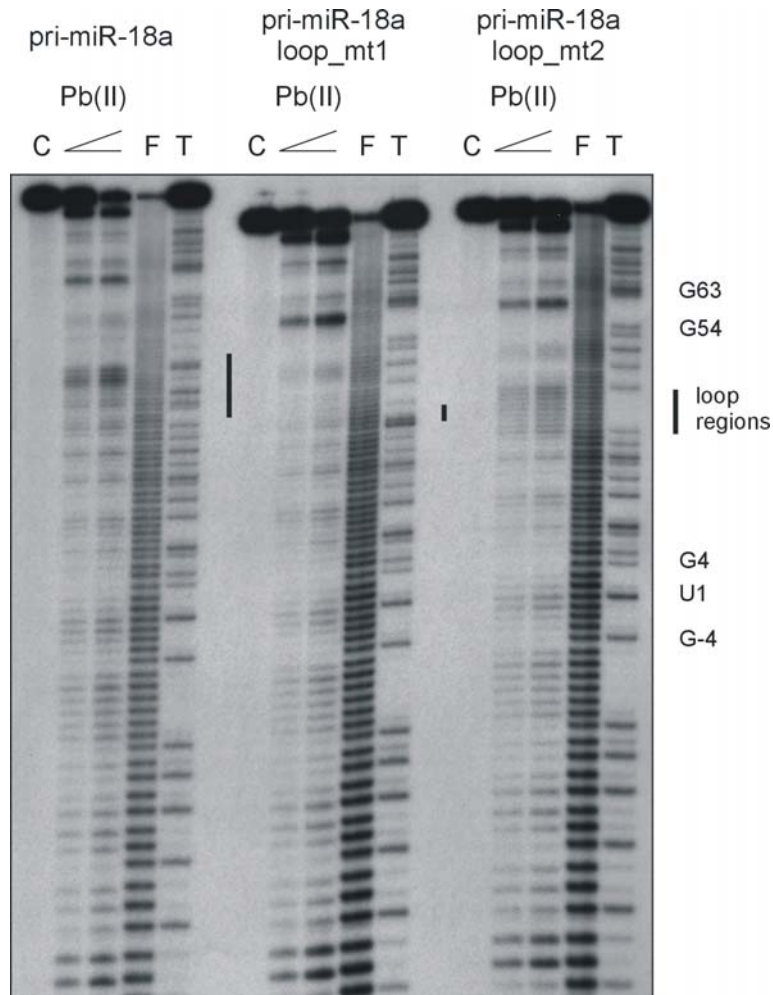


Figure S12. Structure analysis of wild-type and loop mutants of pri-miR-18a (A)
 Cleavage patterns obtained for the 5' ³²P-labeled pri-miR-18a (left panel) and pri-miR-18a loop mutant 1 (middle panel) and loop mutant 2 (right panel) transcripts (100x10³ c.p.m.) treated with: Pb (II) – lead ions at increasing concentrations (0.5, 1 mM). C, incubation control (without probe). F and T identify nucleotide residues subjected to partial digest with formamide (every nucleotide) or ribonuclease T1 (G-specific cleavage), respectively. Electrophoresis was performed in a 10% polyacrylamide gel under denaturing conditions. The positions of selected G residues are shown. The loop regions are indicated by bars.

SUPPLEMENTAL EXPERIMENTAL PROCEDURES

Materials and Methods

Preparation of plasmids and DNA templates

For the structural analysis, the RNA substrates were transcribed *in vitro* from a DNA fragment containing the T7 promoter sequence directly upstream of the pri-miRNA precursor sequence. The DNA fragments were amplified by PCR from human genomic DNA using the following oligonucleotides: pri-miR-17: T7-1018for-G (5'-TTAATACGACTCACTATAGGCCATCACCTTGTA AAACTGAAG-3') and 80rev (5'-TAAAGCCCAACTTGGCTTCCCG-3'); pri-miR-19a: T7-236for-G (5'-TTAATACGACTCACTATAGGCGCCAAGCAAGTATATAGGTG-3') and 1446rev (5'-TTAGTAAAATCATTTCATTTG-3'); pri-miR-18a: T7-103for-G (5'-TTAATACGACTCACTATAGGCGTGCAGGGCCTGCTGATGTTGAGTGC-3') and 233rev (5'-GAATTATTGGATGAATACATAAC-3'); pri-miR-18b: T7-18bfor-G (5'-TTAATACGACTCACTATAGGCGCTTTTGAGCTGCTTCTTATAATG-3') and 18brev (5'-CTTTTAAAATGCTGTATATTAGGC-3'); pri-miR-18a_73nt: T7-1for-G (5'-TTAATACGACTCACTATAGGTGTTCTAAGGTGCATCTAGT-3') and 71rev (5'-TGCCAGAAGGAGCACTTAG-3').

The sequences used for *in vitro* transcription and pri-miRNA processing assays were obtained by PCR from human genomic DNA, cloned in pGEM-T-easy (Promega) and linearized before transcription. The oligonucleotides used were as follows: pri-106a-18b-20b: clusterXfor (5'-CAGGAATATTA ACTAGTAG-3') and clusterXrev (5'-ACGCTGAAATGCAAACCTGC-3'); pri-miR-17-18a-19a: 844for (5'-GAATTCTTAAGGCATAAATACG-3') and 1553rev (5'-GTAGATAACTAAACTACC-3'); pri-miR-16_1: miR16for (5'-TGATAGCAATGTCAGCAGTGCC-3') and miR16rev (5'-

GTAGAGTATGGTCAACCTTAC-3'); pri-miR-27a: cl23-143for (5'-GGCAGAGAGGCCCGAAGCC-3') and cl23-279rev (5'-CAGGCGGCAAGGCCAGAGGAGG-3'); pri-let-7a-1: let-7a-1for (5'-GGAGCGGATTCAGATAACCA-3') and let-7a-1rev (5'-CCAGGCCATAAACAAATGCT-3'); pri-miR-101-1: miR-101-1for (5'-GGGGAGCCTTCAGAGAGAGT-3') and miR-101-1rev (5'-AGCCACCTGTTTCACATTCC-3') pri-miR-31: miR-31for (5'-TCATCCCTGTGGTTTCACAA-3') and miR-31revbis (5'-GGAAATCCACATCCAAGGAA-3'); pri-miR-379: miR-379for (5'-CAAATCCAGCCTCAGAAAGC-3') and miR-379rev (5'-AGCCCAAGTTGCATCACTTC-3'). Mutants 18aUC>GU and 18bGU>CU were derived from the corresponding wild-type templates with oligonucleotides 18a_TC/GTfor (5'-TGAGTGCTTTTTGTGTTAAGGTGCATCTAG-3') together with 18a_TC/GTrev (5'-CTAGATGCACCTTAACACAAAAAGCACTCA-3') and 18b_GT/TCfor (5'-AATGTGTCTCTTGTCTAAGGTGCATCTAG-3') together with 18b_GT/TCrev (5'-CTAGATGCACCTTAGAACAAGAGACACATT-3'), respectively by site-directed mutagenesis (QuickChange, Stratagene). Templates for terminal loop mutants for pri-miR-18a, pri-miR-16-1, pri-miR-101-1 and pri-let-7a-1 were derived from the corresponding wild-type plasmids by PCR amplification with oligonucleotides 18a_loop_mt1for (5'-AATACTGCCCTAAGTGCTCCTT-3') together with 18a_loop_mt1rev (5'-GCTATCTGCACTAGATGCACCT-3'); 18a_loop_mt2for (5'-CTCTACTGCCCTAAGTGCTCCTT-3') together with 18a_loop_mt2rev (5'-AGATATCTGCACTAGATGCACCT-3'); 16_1_loop_mt1for (5'-AATTATCTCCAGTATTAAGTGT-3') together with 16_1_loop_mt1rev (5'-GCTTAACGCCAATATTTACGTG-3'); 16_1_loop_mt2for (5'-

CTCTTATCTCCAGTATTAAGTGT-3') together with 16_1_loop_mt2rev (5'-AGATTAACGCCAATATTTACGTG-3'); 101-1_loop_mt1for (5'-AAGGTACAGTACTGTGATAACTGA -3') together with 101-1_loop_mt1rev (5'-GCAGCATCAGCACTGTGATAACTG -3'); 101-1_loop_mt2for (5'-CTCGGTACAGTACTGTGATAACTGA -3') together with 101-1_loop_mt2rev (5'-AGAAGCATCAGCACTGTGATAACTG -3'); let-7a-1_loop_mt1for (5'-AAAAGTATACAATCTACTGTCTTTC -3') together with let-7a-1_loop_mt1rev (5'-GCAACTATACAACCTACTACCTCA -3'); let-7a-1_loop_mt2for (5'-CTCAACTATACAATCTACTGTCTTTC -3') together with let-7a-1_loop_mt2rev (5'-AGAACTATACAACCTACTACCTCA -3'), using Pfu polymerase (Promega), followed by T4 ligation (Roche) and transformation of DH5alpha competent cells (Invitrogen).

RNA chromatography and western blotting.

Substrate RNAs let-7a-1 loop (5'-GUUGUAUAGUUUAGGGUCACACCCACCACUGGGAGAUAAACUAUACAAUC-3'); miR-101-1 loop (5'-AGUGCUGAUGCUGUCUAUUCUAAAGGUACAGUACUG -3'); miR-379 loop (5'-AACGUAGGCGUUAUGAUUUCUGACCUAUGUA-3'), miR-21 loop (5'-GACUGAUGUUGACUGUUGAAUCUCAUGGCAACACCAGUCGA-3'); miR-19a loop (5'-UUGCACUACAAGAAGAAUGUAGUUGUGCAAU-3'); miR-16-1 loop (5'-AAUAUUGGCGUUAAGAUUCUAAAUAUCUCCAGUAUUA-3') purchased from Sigma-Aldrich were coupled to the agarose beads (adipic acid dihydrazide - Sigma-Aldrich). First, 1 μmol of specific RNA was incubated for 1h in room

temperature in the mix containing 100 mM NaAc pH 5.0 and 5 mM of freshly prepared m-sodium periodate. Then, RNA was precipitated and resuspended in 100 mM NaAc pH 5.0. Meanwhile the agarose beads were washed extensively and resuspended in enough volume of the 100 mM NaAc pH 5.0 to have a final mix of 50% beads. One volume of agarose mix was added to two volumes of the m-periodate treated RNAs and left rocking over night at 4°C (All steps mentioned above were performed in eppendorf tubes wrapped in alu foil to protect the oxidation of RNA from light). Non-bound RNA was washed once with the 2 M KCl, twice with the 1M KCl and three times with Roeder D (KCl 100mM, glicerol 20% p/v, EDTA 0,2mM, Hepes.KOH 100 mM pH 7,9, DTT 0,5 mM, protease inhibitors-Complete EDTA free (Roche)). To purify the factors from the nuclear extracts beads-coupled RNAs were incubated for 30min. at 37°C in the mixture containing 250µl (final 40% v/v) HeLa nuclear extract (Cilbiotech), 1.5 mM MgCl₂, 0.5 mM ATP and 20 mM creatine phosphate. To analyze the factors bound to the RNA, -agarose beads were washed four times with 1ml Roeder D supplemented with 1.5 mM MgCl₂. Proteins were separated using NuPAGE Novex Bis-Tris system (Invitrogen). Protein bands visualized by GelCode Blue (Pierce) were subjected to Mass Spectroscopy (Dr Jim Creanor's service – Edinburgh University). Alternatively western blot analysis was carried out with hnRNP A1 – specific monoclonal antibody 4B10 (Abcam), with hnRNP L – specific monoclonal antibody 4D11 (Sigma-Aldrich) and hnRNP I/PTB – specific monoclonal antibody SH54 (Abcam).

EMSA analysis of pri-miR-18a/hnRNP A1 complexes. 5' ³²P-labeled pri-miR-18a transcripts (in the range from 20,000 c.p.m. to 400,000 c.p.m.) were incubated with recombinant hnRNP A1 (in the range from 50 ng to 200 ng) for 30 min in RT in the

buffer containing 0.5 mM ATP, 50 mM creatine phosphate, 5 mM MgCl₂ and 20 mg/ml heparin. When protein was not present the reactions were supplemented with corresponding volume of BC100. Where indicated 50µM looptomiR was added prior addition of the protein. The samples with added loading buffer (10mM Tris-HCl, 3% sucrose and dyes) were loaded on the 8% (w/v) non-denaturing gel and run in the constant power of 9W.