

## Supplementary information

### **Cytotoxic CD8<sup>+</sup> T cells target citrullinated antigens in rheumatoid arthritis**

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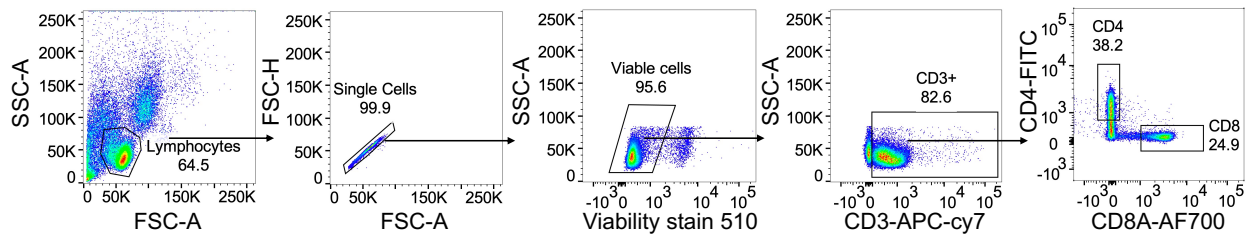
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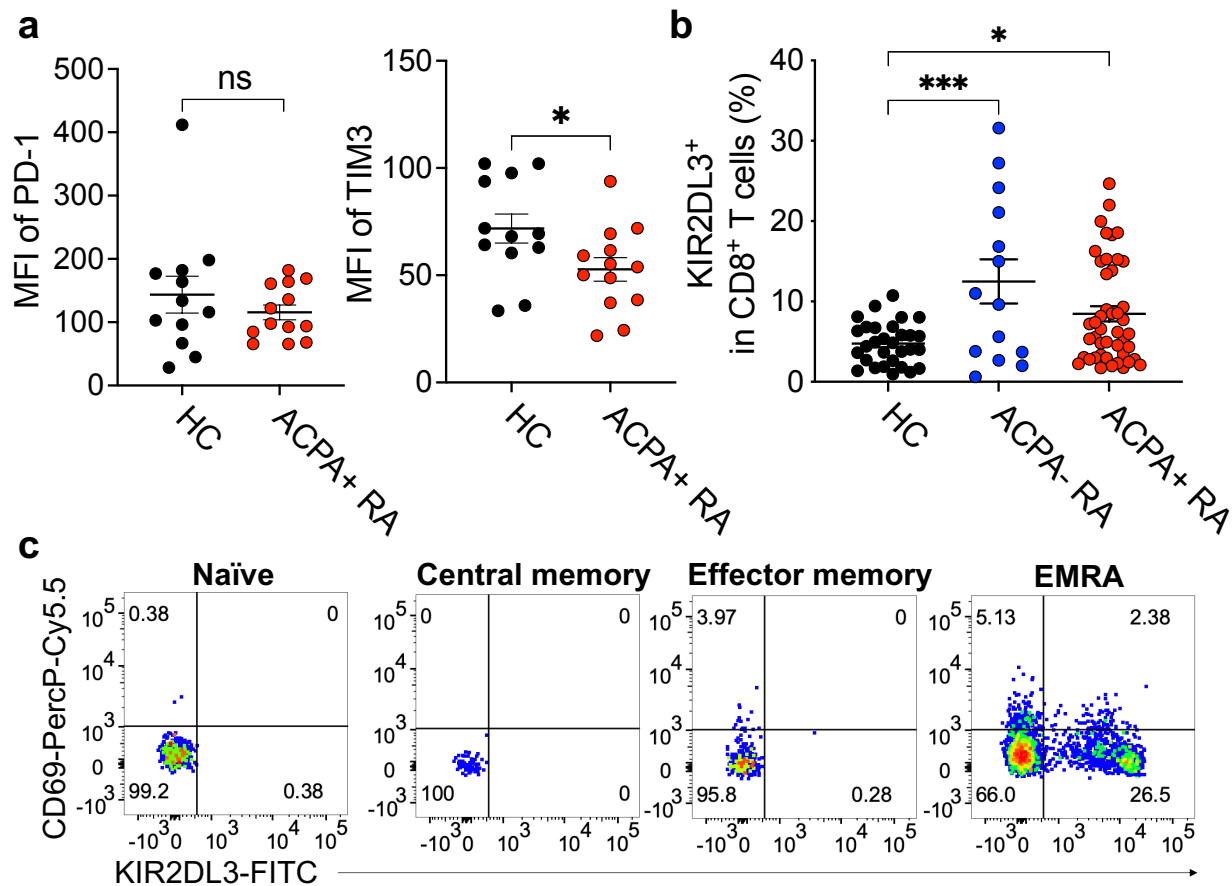
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## Supplementary Figures

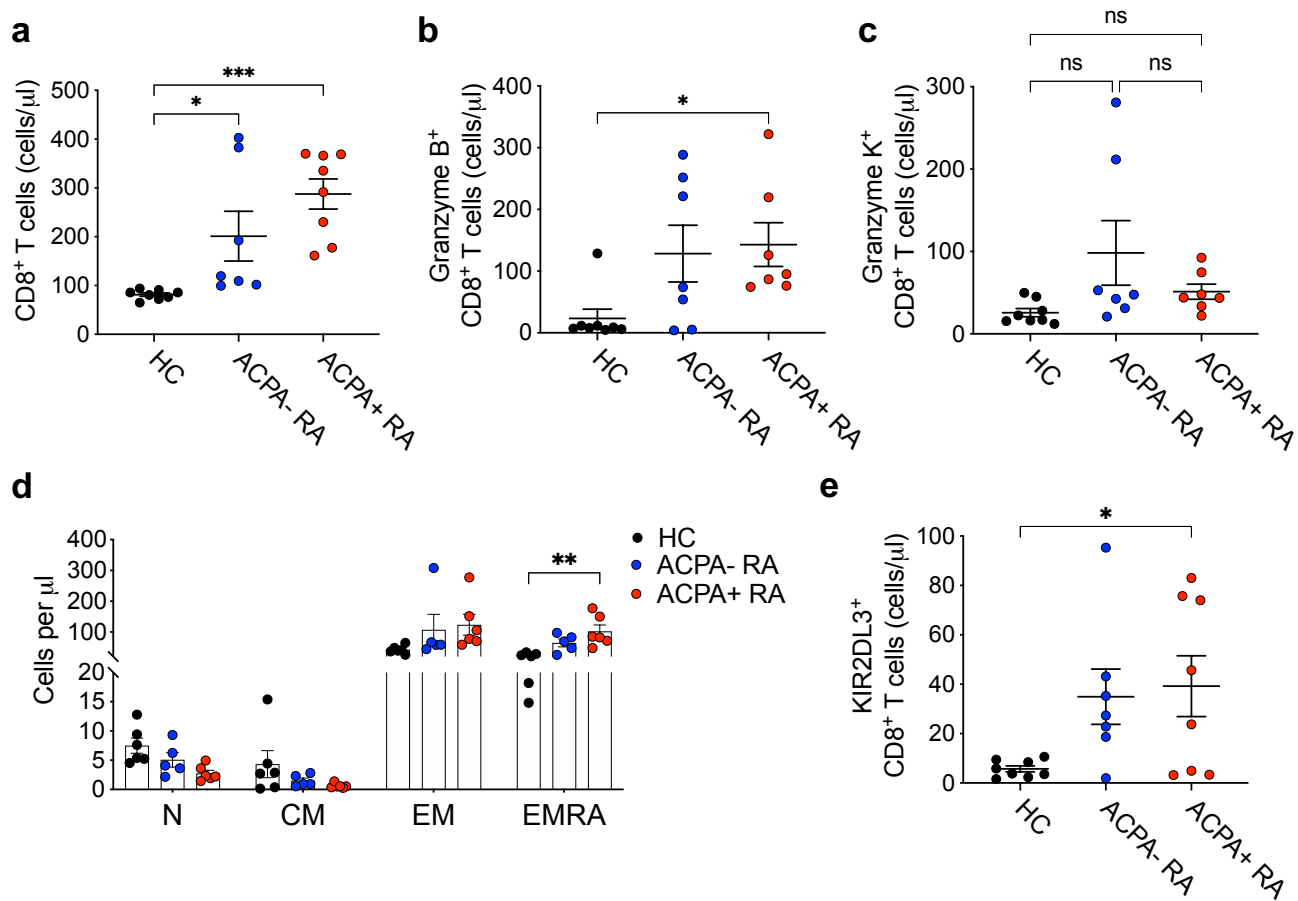


### Supplementary Fig. 1: Flow cytometric analysis of CD8<sup>+</sup> T cells in ACPA<sup>+</sup> RA patient blood.

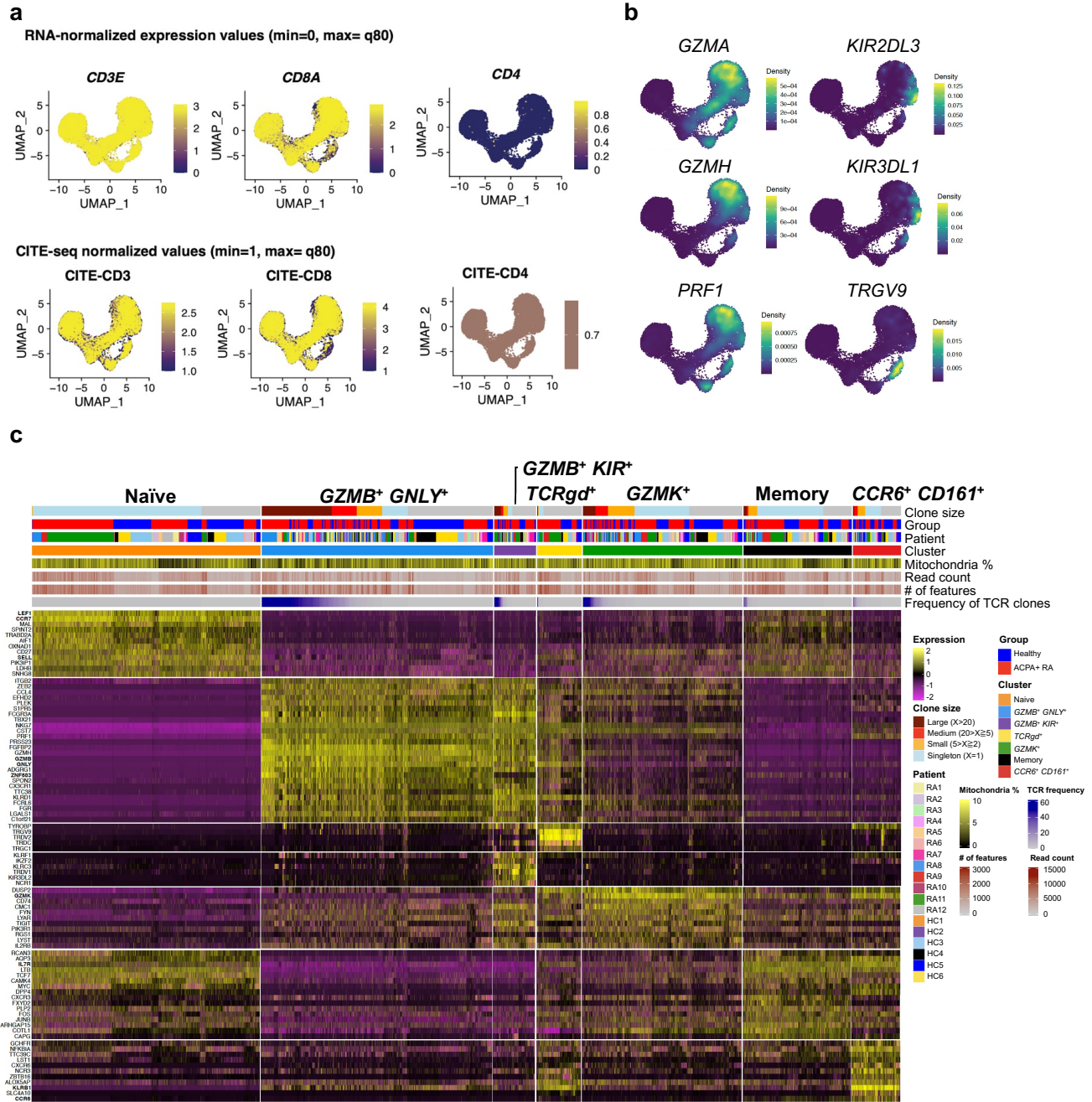
Representative data from ACPA<sup>+</sup> RA PBMCs are displayed as flow cytometric plots, and the gating parameters for the CD8<sup>+</sup> T cell population indicated.



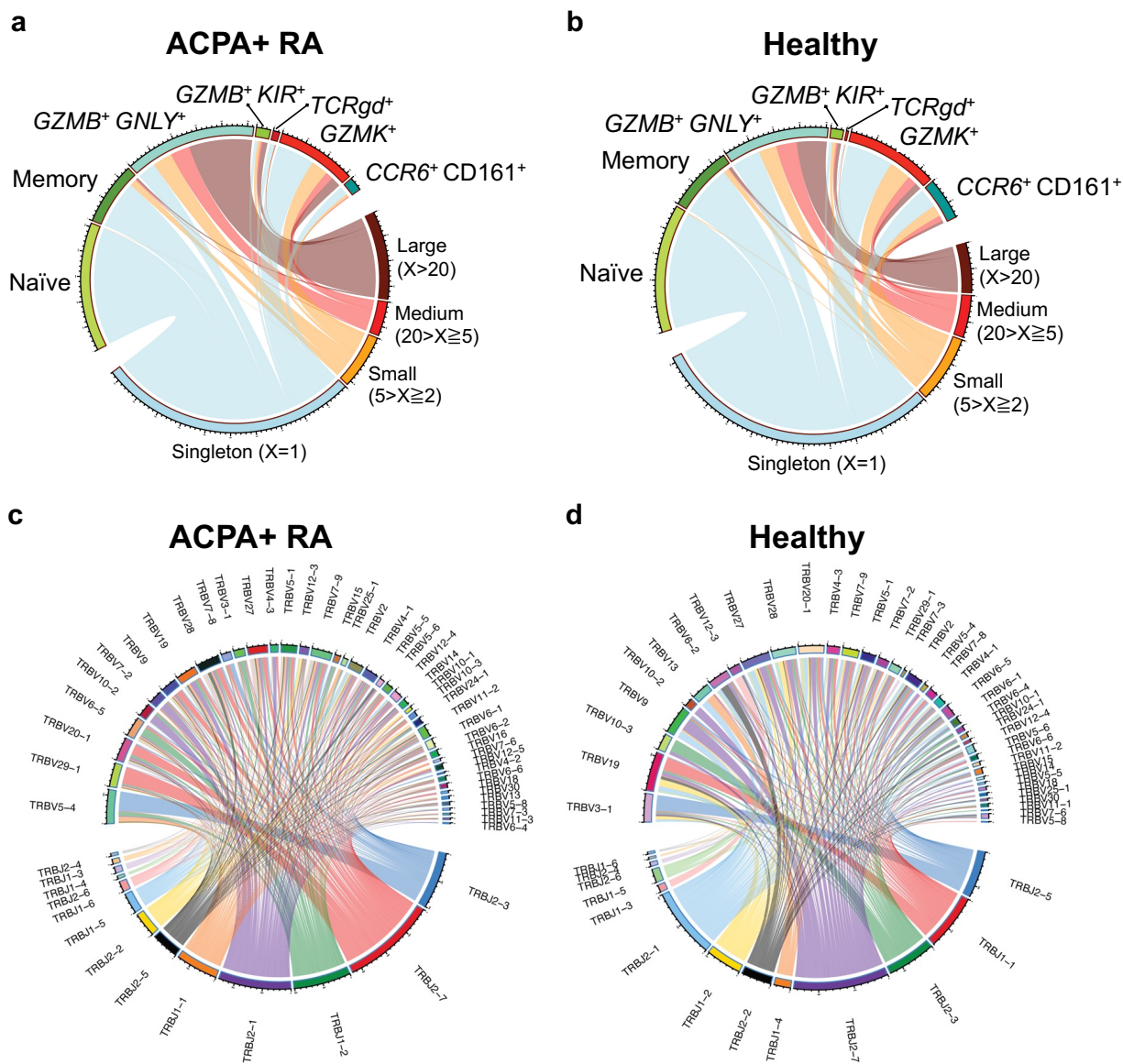
**Supplementary Fig. 2: Phenotypic characterization of CD8<sup>+</sup> T cells in ACPA+ RA patient blood.** **a** Expression of PD-1 or TIM3 in CD8<sup>+</sup> T cells from PBMCs from healthy controls ( $n = 12$ ) or ACPA+ RA patients ( $n = 13$ ) measured by flow cytometry.  $*P=0.0376$ . **b** Percentage of KIR2DL3<sup>+</sup>CD8<sup>+</sup> T cells in HC ( $n = 30$ ), ACPA- RA ( $n = 14$ ), or ACPA+ RA ( $n = 45$ ) analyzed by flow cytometry ( $***P=0.0007$ , or  $*P=0.0355$ ). **c** Representative dot plots showing expression levels of CD69 and KIR2DL3 in CD8<sup>+</sup> T memory subsets (naïve, CM, EM, or EMRA). Data are presented as means  $\pm$  SEM.  $*P < 0.05$  or  $***P < 0.001$  by two-tailed unpaired  $t$ -test (**a**) or one-way ANOVA (**b**) with Tukey's multiple comparisons test. ns, not significant. Source data are provided as a Source Data file. HC healthy control, ACPA anti-citrullinated protein antibodies, RA rheumatoid arthritis, MFI median fluorescence intensity.



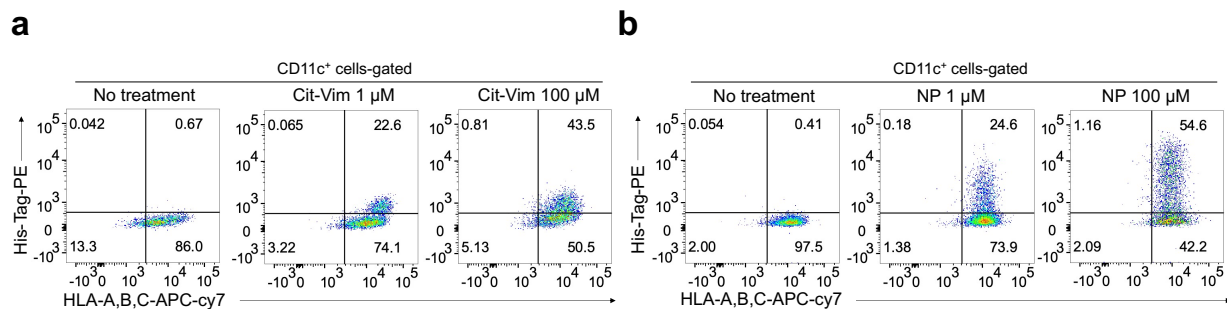
**Supplementary Fig. 3: Absolute cell counts of CD8<sup>+</sup> T cells in ACPA+ RA patients.** Absolute cell numbers of total CD8<sup>+</sup> T cells (**a**, HC n = 8, ACPA- RA n = 7, ACPA+ RA n = 8, \**P*=0.0466, or \*\*\**P*=0.0005), granzyme B-expressing CD8<sup>+</sup> T cells (**b**, HC n = 8, ACPA- RA n = 7, ACPA+ RA n = 7, \**P*=0.0473), or granzyme K (**c**, HC n = 8, ACPA- RA n = 7, ACPA+ RA n = 7)-expressing CD8<sup>+</sup> T cells, memory CD8<sup>+</sup> T cells (**d**, HC n = 6, ACPA- RA n = 5, ACPA+ RA n = 5, \**P*=0.006), or KIR2DL3<sup>+</sup>CD8<sup>+</sup> T cells (**e**, HC n = 8, ACPA- RA n = 7, ACPA+ RA n = 8, \**P*=0.0484) per 1 µl assessed by counting beads in each PBMC. Data are presented as means ± SEM. \**P* < 0.05, \*\**P* < 0.01, or \*\*\**P* < 0.001 by ordinary one-way ANOVA (**a-c**, **e**) or two-way ANOVA (**d**) with Tukey's multiple comparisons test. ns, not significant. Source data are provided as a Source Data file. HC healthy control, ACPA anti-citrullinated protein antibodies, RA rheumatoid arthritis.



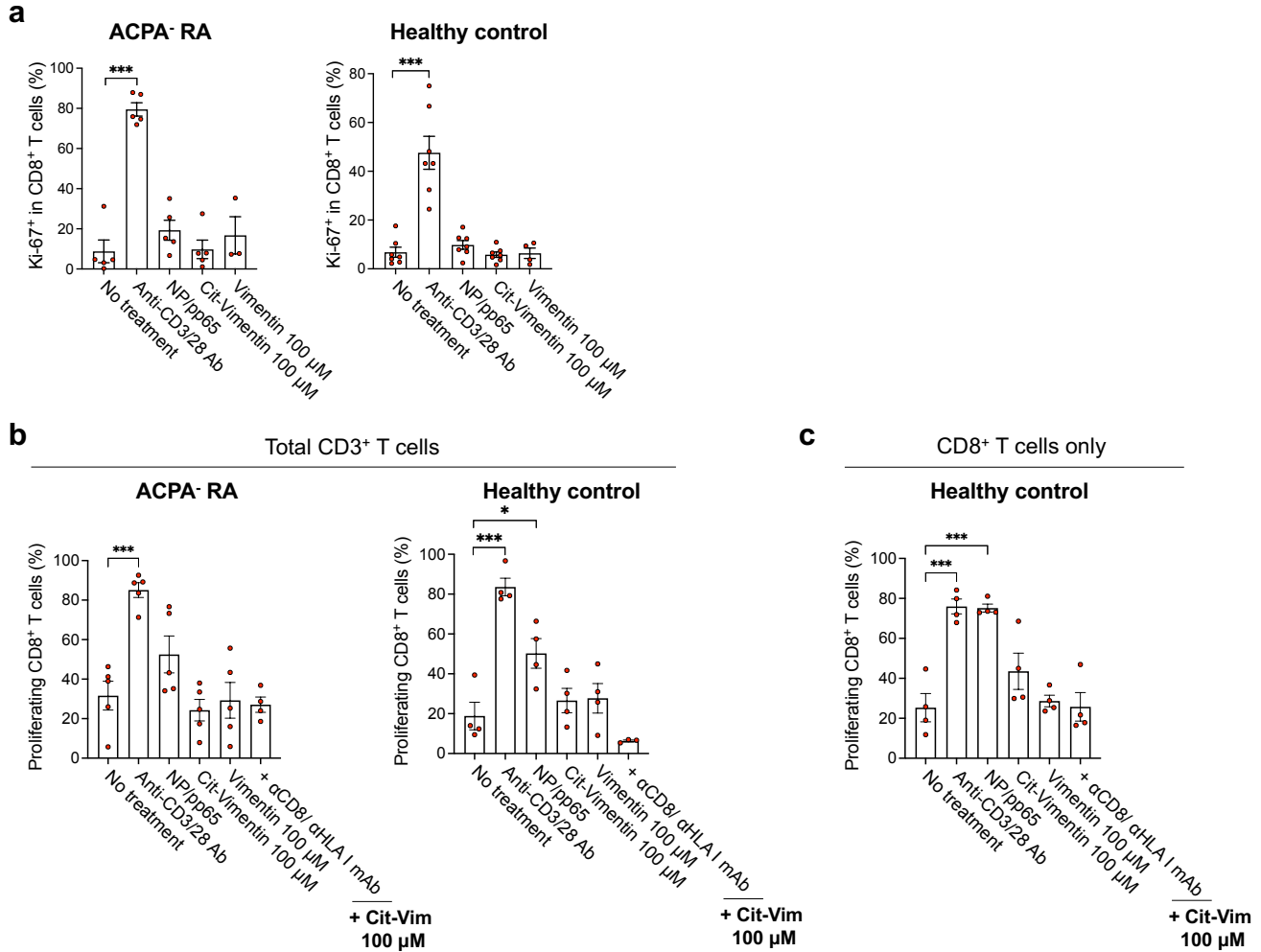
**Supplementary Fig. 4: Expression of cell type marker genes in CD8<sup>+</sup> T cells from ACPA+ RA and healthy control blood. a** mRNA and CITE-seq expression of marker genes (*CD3E*, *CD8A*, and *CD4*) used to identify and confirm analysis of CD8<sup>+</sup> T cells. Color indicates the expression level of each marker gene. **b** Density plots display marker genes including *GZMA*, *KIR2DL3* or *TRGV9*) in CD8<sup>+</sup> T cells. **c** Heatmap of select differentially expressed gene signatures (log2 Fold Change > 0.8) in each cluster with bars; clonal size, patients, clusters, mitochondrial gene percentage, read count, number of features in each cell and frequency of TCR clone. ACPA anti-citrullinated protein antibodies, RA rheumatoid arthritis.



**Supplementary Fig. 5: CD8<sup>+</sup> T cells expressing cytotoxic mediators are clonally expanded in ACPA+ RA blood.** **a, b** Chord diagrams representing the contribution of each cluster to clonal sizes (**a**) and the shared clonotypes between clusters (**b**). **c** TCR beta chain V- and J-gene usage of CD8<sup>+</sup> T cells of ACPA+ RA blood. The width of each band represents the frequency of V- or J- gene usage. ACPA anti-citrullinated protein antibodies, RA rheumatoid arthritis.



**Supplementary Fig. 6: CD11c<sup>+</sup> cells from ACPA+ RA patients endocytose cit-vimentin. a,b,** Representative flow cytometric plots of protein endocytosis by CD11c<sup>+</sup> cells in ACPA+ RA PBMCs show antigen-presenting cells uptake recombinant His-tagged cit-vimentin protein (**a**), or NP protein (**b**) in a dose-dependent manner (1 or 100  $\mu$ M) after 16 hr of culture. Cit-Vim citrullinated vimentin.

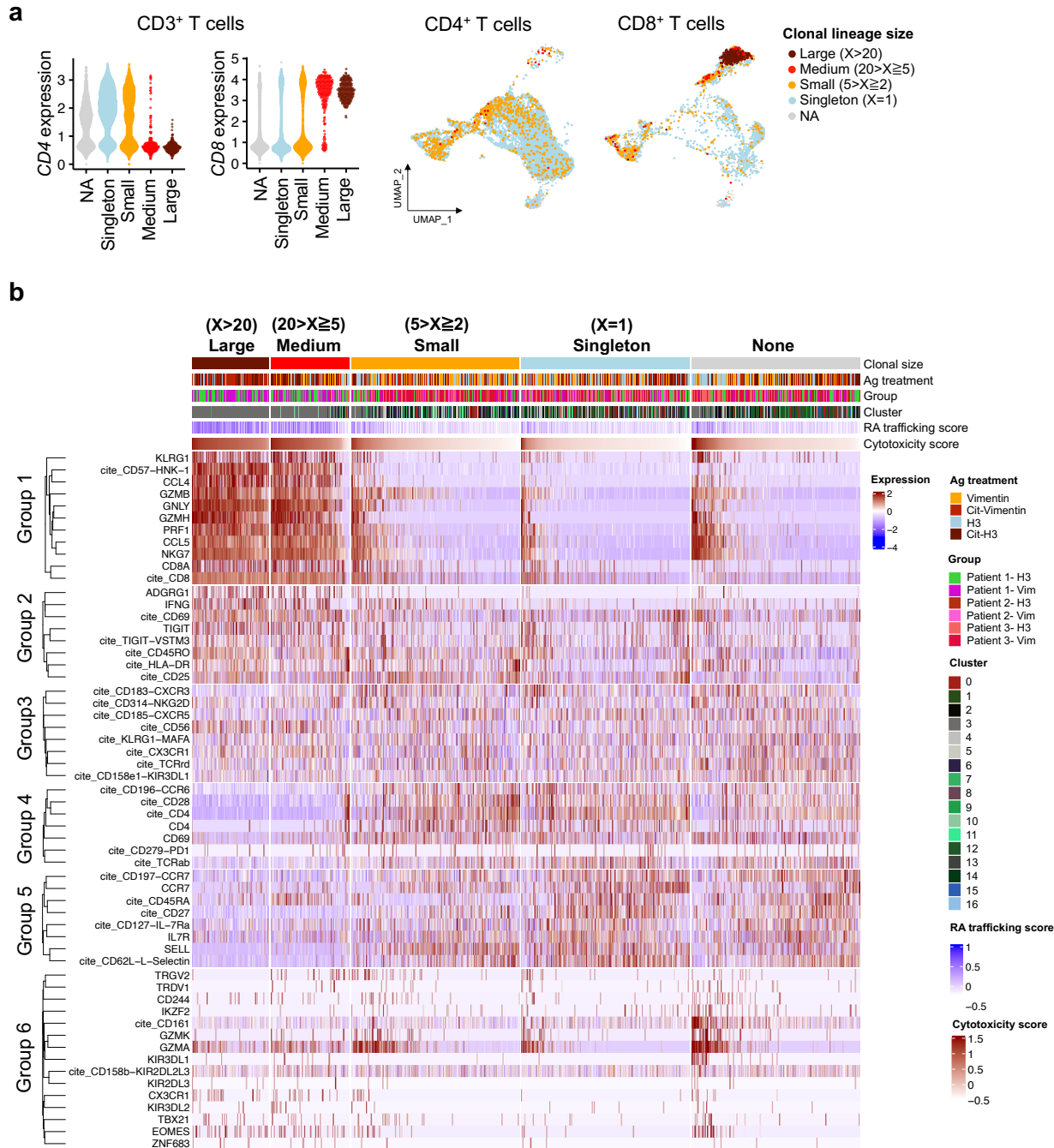


**Supplementary Fig. 7: Citrullinated vimentin does not induce the proliferation of ACPA-RA or HC CD8<sup>+</sup> T cells.** **a** Percentage of Ki-67<sup>+</sup>CD8<sup>+</sup> T cells in ACPA- RA ( $n = 5$ ) or HC ( $n = 7$ ) PBMCs after stimulation with anti-CD3/28 antibodies, NP (Influenza)/ pp65 (CMV) proteins (50  $\mu$ M of each), citrullinated vimentin (100  $\mu$ M), or native vimentin (100  $\mu$ M) for 16 hr. \*\*\* $P < 0.0001$ . **b, c** Quantification of the proliferating CD8<sup>+</sup> T cells in co-culture of monocyte-derived dendritic cells (MoDCs) with ACPA-RA or HC total CD3<sup>+</sup> T cells (**b**, \*\*\* $P=0.0002$  in ACPA- RA, \*\*\* $P=0.0002$ , or \* $P=0.0214$  in HC) or CD8<sup>+</sup> T cells only (**c**,  $n = 4$ , \*\*\* $P=0.0007$ , or \*\*\* $P=0.0005$ ). For **b**, no treatment, anti-CD3/28 Ab, NP/pp65, cit-vimentin or native vimentin  $n = 5$ , cit-vimentin with anti-CD8/ HLA class I antibodies  $n = 4$  in ACPA-RA and no treatment, anti-CD3/28 Ab, NP/pp65, cit-vimentin or native vimentin  $n = 4$  and cit-vimentin with anti-CD8/ HLA class I antibodies  $n = 3$  in HC. Bars represent means  $\pm$  SEM. \* $P < 0.05$  or \*\*\* $P < 0.001$  by ordinary one-way ANOVA with Tukey's multiple comparisons test. Source data are provided as

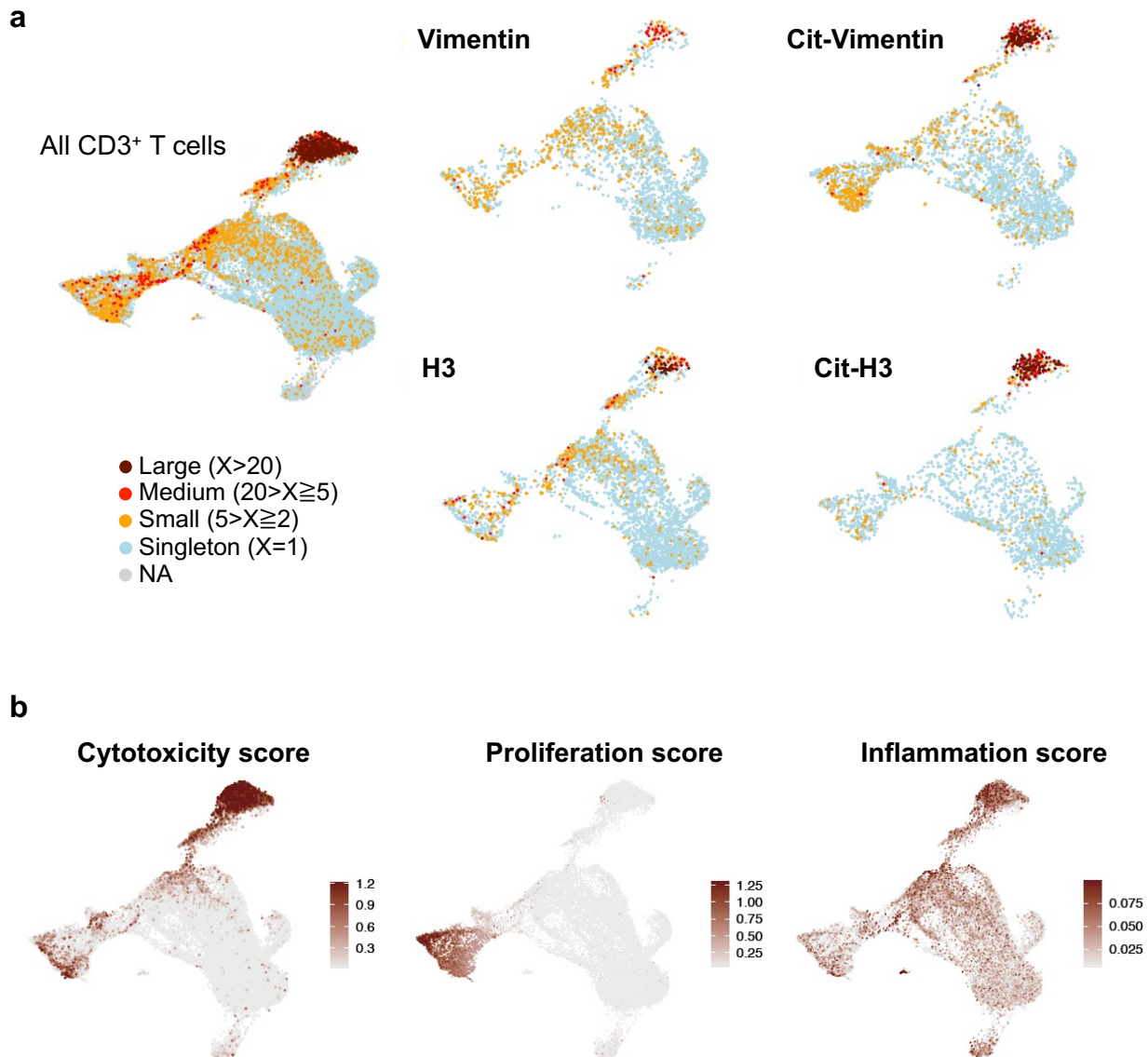
a Source Data file. ACPA anti-citrullinated protein antibodies, RA rheumatoid arthritis. mAb monoclonal antibody.



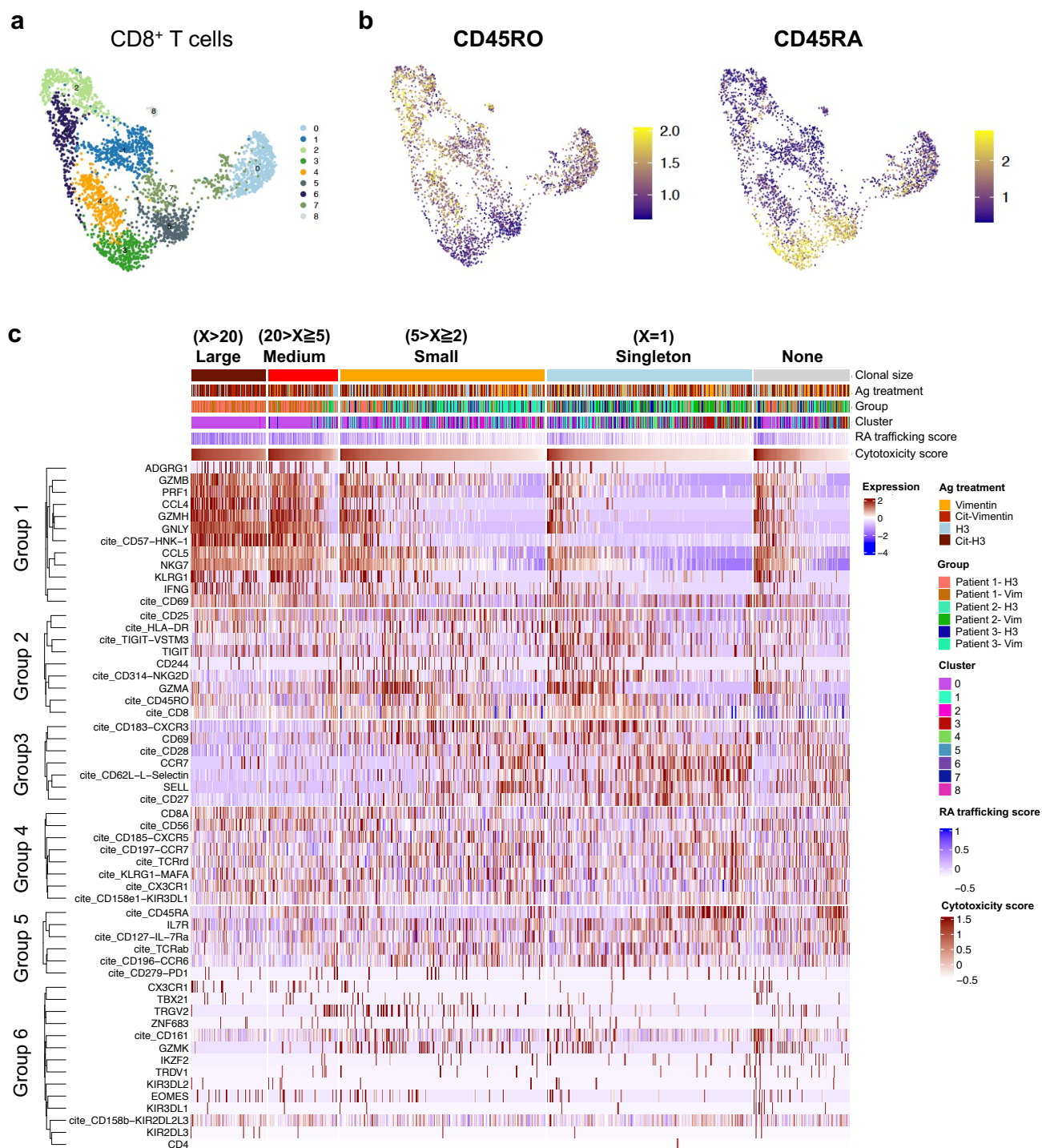
**Supplementary Fig. 8: Single-cell RNA sequencing of CD3<sup>+</sup> T cells stimulated by native or citrullinated antigens.** **a** UMAP plot representing 17 distinct clusters of CD3<sup>+</sup> T cells (n = 13,354) stimulated by native or citrullinated proteins. **b** Expression of *CD8A*, *CD4*, *GZMB*, *GNLY* or *GZMK* in the clusters. Color shows the expression level of each marker. **c** Heatmap showing transcriptional differences in each cluster. Top bars indicate clonal size, Protein treatment, patient group, clusters and scoring scale for RA trafficking and cytotoxicity. Individual T cells are ordered by clonal size in each cluster. RA rheumatoid arthritis, Vim vimentin, Ag antigen.



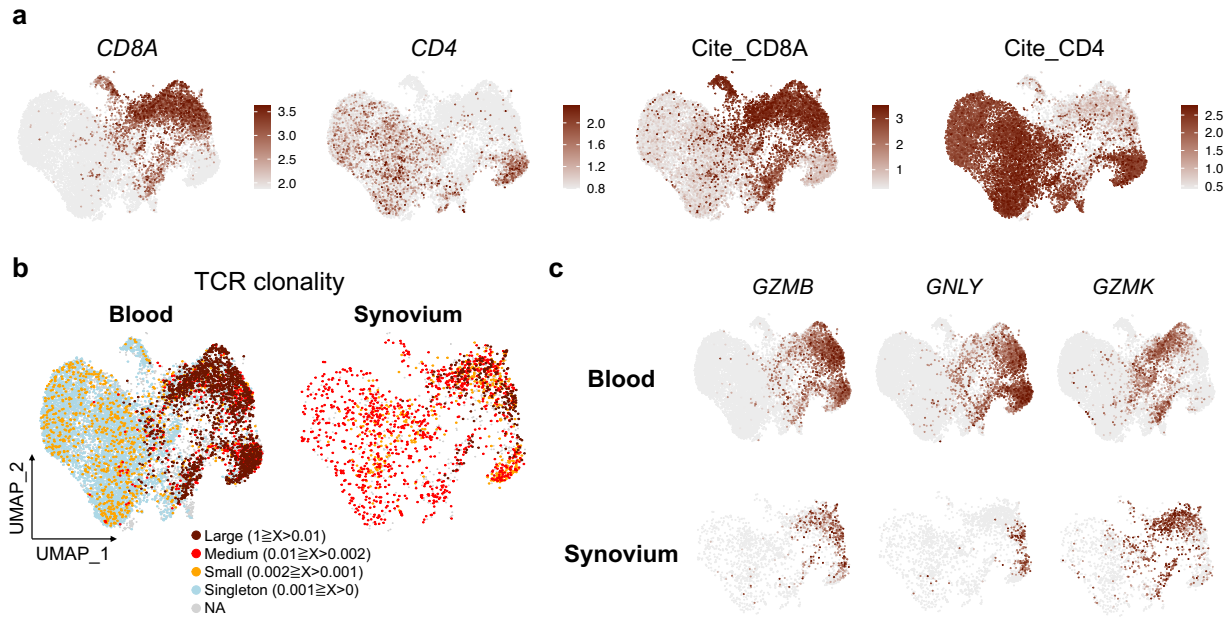
**Supplementary Fig. 9: Clonal lineages of CD4<sup>+</sup> and CD8<sup>+</sup> T cells stimulated by native or citrullinated antigens. a** Distribution of each clonal family in CD4<sup>+</sup> (n = 7,885) or CD8<sup>+</sup> (n = 3,284) T cells. **b** Heatmap representing 6 groups determined by unsupervised clustering based on the level of RNA and CITE-seq antibodies (cite\_). Individual T cells are ordered by cytotoxicity score in each cluster. Ag antigen, Vim vimentin.



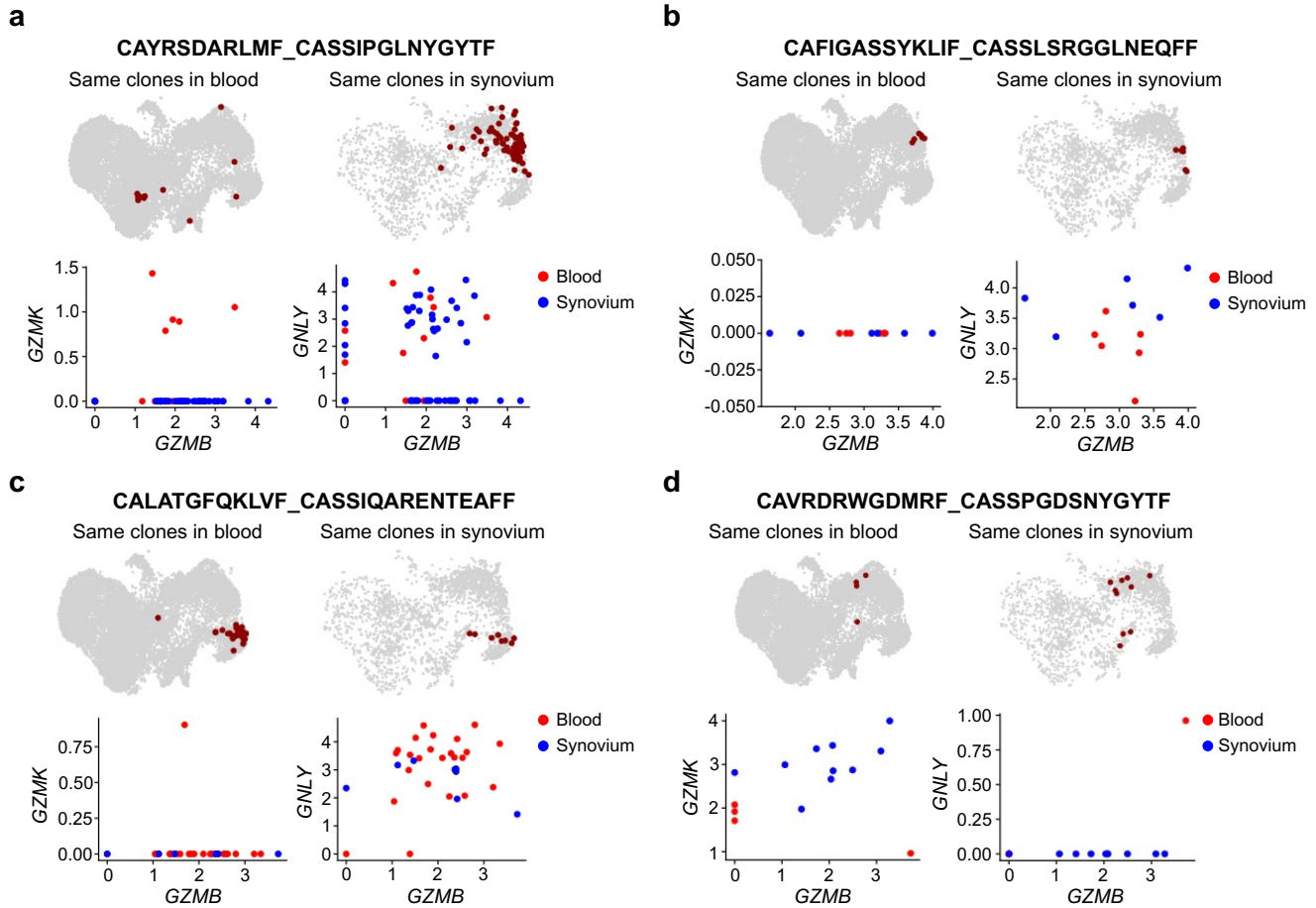
**Supplementary Fig. 10: Citrullinated antigens stimulate clonal expansion of CD3<sup>+</sup> T cells with high cytotoxicity scores.** **a** UMAP plots of all CD3<sup>+</sup> T cells stimulated by native or citrullinated Vimentin or H3 integrated with TCR clonality. **b** UMAP plots showing cytotoxicity, proliferation, or inflammation score. The score was calculated by the expression level of canonical marker genes. Cit- citrullinated.



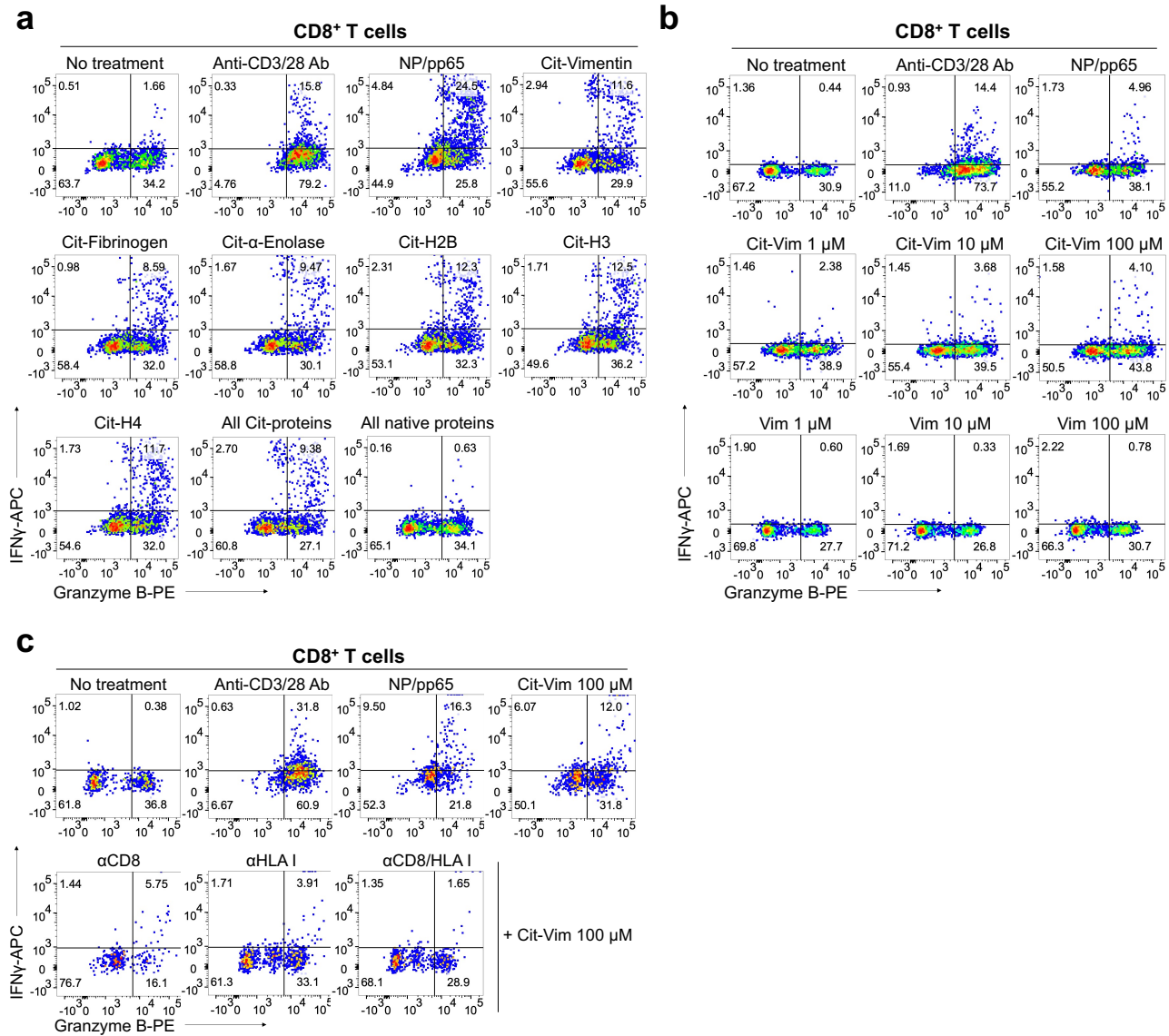
**Supplementary Fig. 11: Phenotype of CD8<sup>+</sup> T cells stimulated by native or citrullinated antigens.** **a** UMAP plot of CD8<sup>+</sup> T cells having 9 transcriptionally different clusters. **b** Expression level of CD45RO and CD45RA CITE-seq antibodies in CD8<sup>+</sup> T cells. **c** Heatmap of 6 groups by unsupervised clustering using the level of RNA and CITE-seq antibodies (cite\_). RA rheumatoid arthritis, Vim vimentin, Cit-citrullinated.



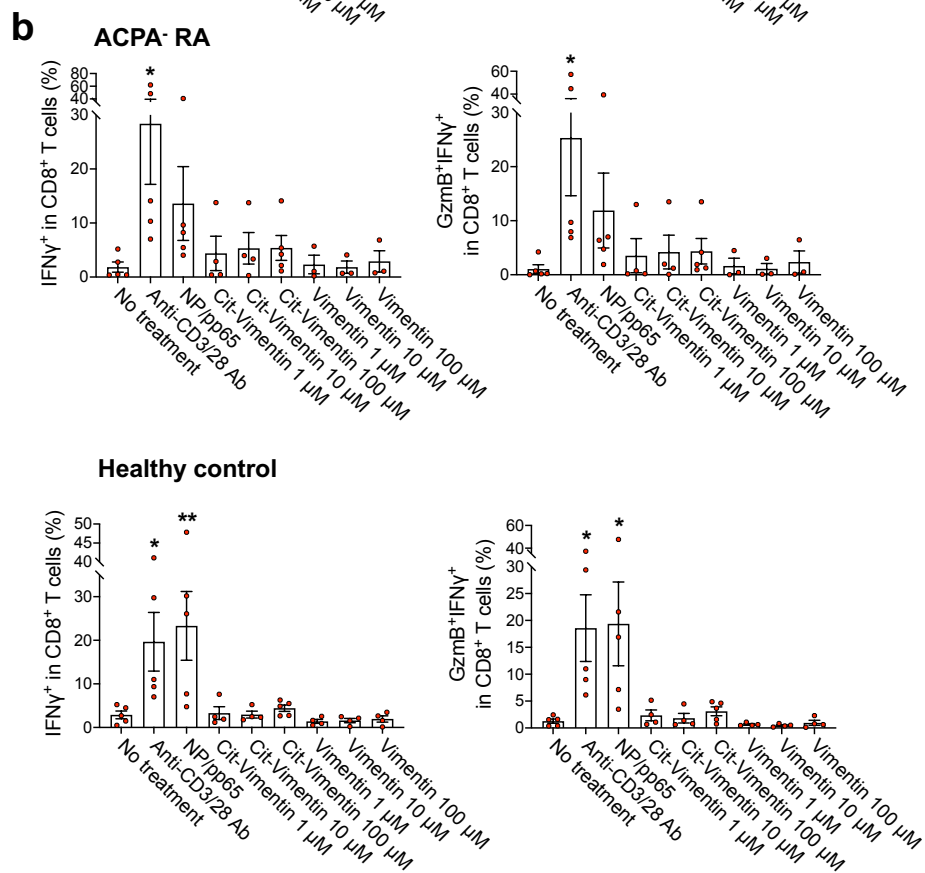
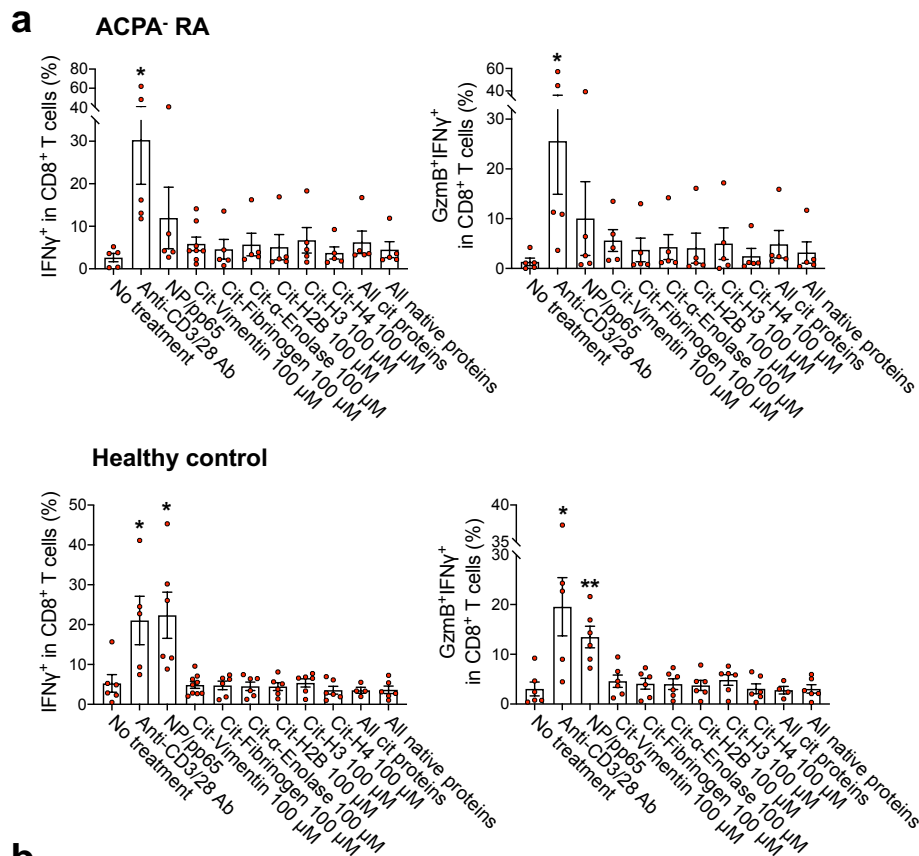
**Supplementary Fig. 12: Integrated analysis of single cell RNA-seq data from paired RA blood and synovium CD8<sup>+</sup> T cells.** **a** UMAP plots representing the expression levels of CD8A and CD4 in mRNA and CITE-seq Abs. **b** UMAP plots of ACPA+ RA blood ( $n = 3$ ) and synovium ( $n = 4$ ) CD8<sup>+</sup> T cells integrated with TCR clonality ( $n = 9,360$  paired  $TCR\alpha\beta$  sequences). Color indicates the groups by the ratio of clonotypes in total cells. **c** UMAP plots showing the expression of *GZMB*, *GNLY* and *GZMK* in integrated blood and synovial CD8<sup>+</sup> T cells. Scale bars represent the normalized expression values. TCR T cell receptor.



**Supplementary Fig. 13: Shared T cell clonotypes between ACPA+ RA blood and matched synovium. a-d** Mapping of shared clonotypes in the matched blood and synovium from ACPA+ RA patients. The frequent clonotypes are represented as CDR3 alpha-beta sequences. Dot plots show the expression level of *GZMB*, *GZMK*, and/or *GNLY* in the same clonotypes located in either blood (red) and synovium (blue). Source data are provided as a Source Data file.

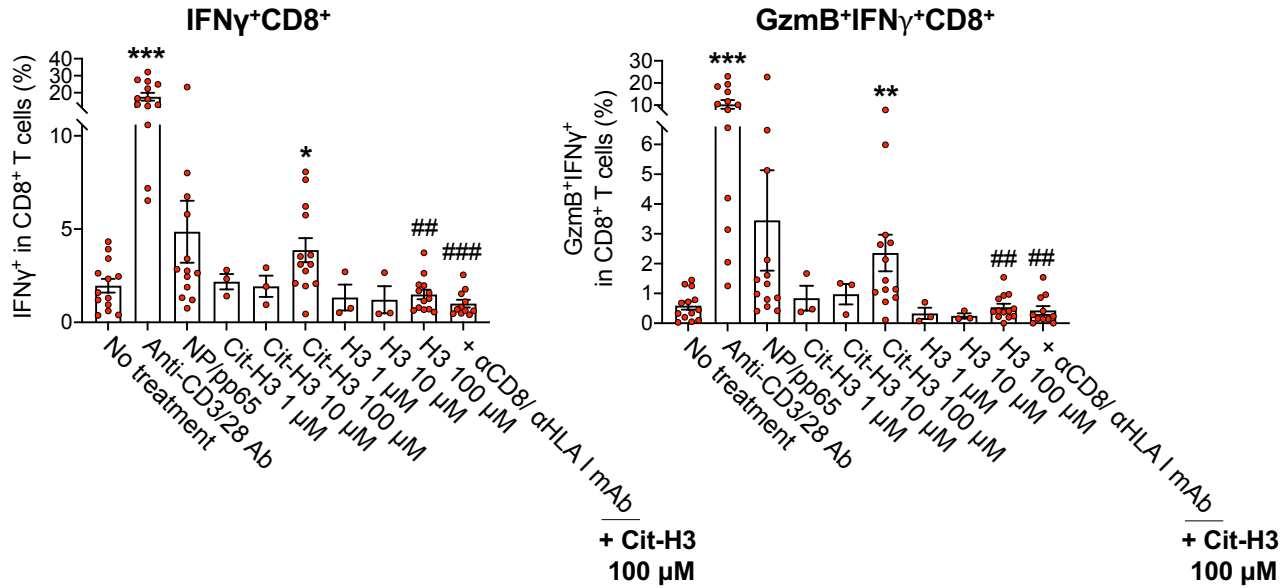


**Supplementary Fig. 14: ACPA<sup>+</sup> RA patient blood CD8<sup>+</sup> T cells are activated by citrullinated proteins in a CD8:HLA class I-dependent manner. a-c** Representative flow cytometric plots of ACPA<sup>+</sup> RA blood CD8<sup>+</sup> T cells stimulated with citrullinated proteins (a), a range of concentrations of cit-vimentin or native vimentin (b), and in the presence of anti-CD8/ HLA class I-blocking antibodies (c). Ab antibody, Vim vimentin, Cit- citrullinated.



**Supplementary Fig. 15: ACPA- RA patient CD8<sup>+</sup> T cells react with viral but not citrullinated**

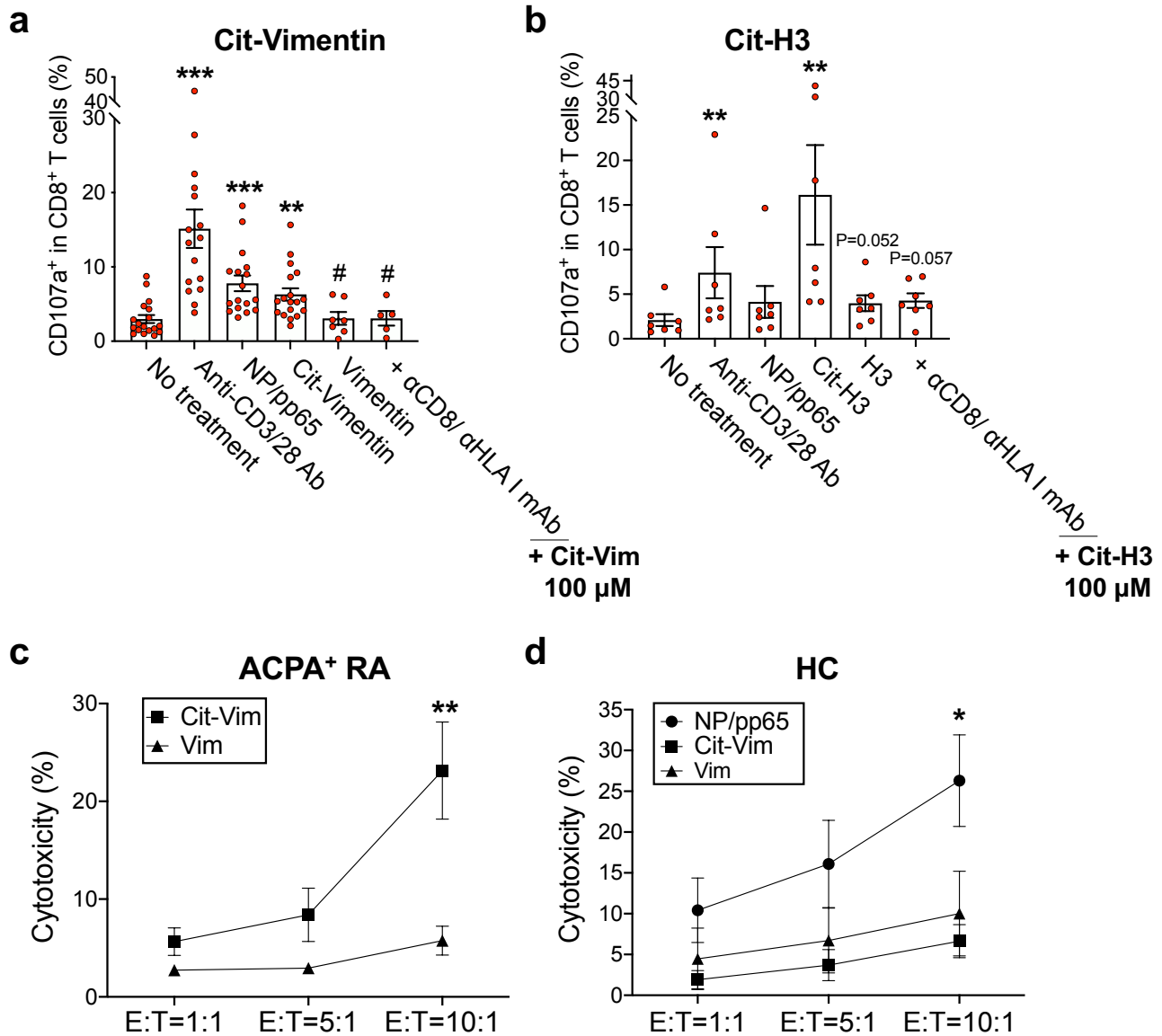
**antigens. a** Percentages of IFN $\gamma$  and/ or GzmB-expressing CD8<sup>+</sup> T cells from PBMCs of ACPA- RA or HCs were measured by intracellular staining after stimulation with anti-CD3/28 antibodies, NP (Influenza)/ pp65 (CMV) proteins (50  $\mu$ M of each), citrullinated proteins (vimentin, fibrinogen,  $\alpha$ -enolase, and histones H2B, H3 or H4) (100  $\mu$ M), all citrullinated proteins (20  $\mu$ M each), or all native proteins (20  $\mu$ M each) for 16 hr. For ACPA- RA, no stimulation (n = 5), anti-CD3/28 Abs (n = 5), NP/pp65 (n = 5), cit-vimentin (IFN $\gamma$ <sup>+</sup> n = 8, GzmB<sup>+</sup>IFN $\gamma$ <sup>+</sup> n = 5), cit-fibrinogen,  $\alpha$ -enolase, and histones H2B, H3 or H4 (n = 5), all cit proteins (n = 5), or all native proteins (n = 5); \**P*=0.0296, or \**P*=0.0498. For HC, no stimulation (n = 6), anti-CD3/28 Abs (n = 5), NP/pp65 (n = 6), cit-vimentin (IFN $\gamma$ <sup>+</sup> n = 9, GzmB<sup>+</sup>IFN $\gamma$ <sup>+</sup> n = 6), cit-fibrinogen,  $\alpha$ -enolase, and histones H2B, H3 or H4 (n = 6), all cit proteins (n = 4), or all native proteins (n = 6); \**P*=0.0274, \**P*=0.0201, \**P*=0.0150, or \*\**P*=0.0023. **b** Quantification of IFN $\gamma$ <sup>+</sup> or IFN $\gamma$ <sup>+</sup>GzmB<sup>+</sup> CD8<sup>+</sup> T cells in ACPA- RA or HC PBMCs stimulated for 16 hr with cit-vimentin or native vimentin in a concentration-dependent manner. No stimulation (ACPA- RA and HC n = 5), anti-CD3/28 Abs (ACPA- RA and HC n = 5), NP/pp65 (ACPA- RA and HC n = 5), cit-vimentin 1 or 10  $\mu$ M (ACPA- RA and HC n = 4), cit-vimentin 100  $\mu$ M (ACPA- RA and HC n = 5), vimentin 1,10 or 100  $\mu$ M (ACPA- RA n = 3 and HC n = 4). In ACPA- RA, \**P*=0.0462, or \**P*=0.05. In HC, \**P*=0.0385, \*\**P*=0.033, \**P*=0.024, or \**P*=0.0491. For **a, b**, data are presented as means  $\pm$  SEM. By two-tailed unpaired *t*-test: \**P* < 0.05 or \*\**P* < 0.01 versus no treatment. Source data are provided as a Source Data file. ACPA anti-citrullinated protein antibodies, RA rheumatoid arthritis, Ab antibody.



**Supplementary Fig. 16: ACPA+ RA blood CD8<sup>+</sup> T cells react with cit-H3.**

Percentage of IFN $\gamma$  or GzmB expressing CD8<sup>+</sup> T cells in ACPA+ RA blood measured by intracellular staining after stimulation with a range of concentrations of citrullinated histone H3 (cit-H3) or native H3 protein in presence or absence of anti-CD8/HLA class I-blocking antibodies for 16 hrs. No stimulation (n = 13), anti-CD3/28 Abs (n = 13), NP/pp65 (n = 13), cit-H3 or native H3 1 or 10  $\mu$ M (n = 3), cit-H3 or native H3 100  $\mu$ M (n = 13), cit-H3 with anti-CD8/ HLA class I antibodies (n = 11). For IFN $\gamma$ <sup>+</sup>CD8<sup>+</sup>, \*\*\* $P$ <0.0001, \* $P$ =0.0172, ## $P$ =0.0023, or #### $P$ =0.0007. For GzmB<sup>+</sup>IFN $\gamma$ <sup>+</sup>CD8<sup>+</sup>, \*\*\* $P$ <0.0001, \*\* $P$ =0.0098, ## $P$ =0.0077, or ## $P$ =0.0099. Data are presented as means  $\pm$  SEM. By two-tailed unpaired  $t$ -test: \* $P$  < 0.05, \*\* $P$  < 0.01 or \*\*\* $P$  < 0.001 versus no treatment, and ## $P$  < 0.01 or #### $P$  < 0.001 versus cit-H3 100  $\mu$ M.

Source data are provided as a Source Data file. Ab antibody.



**Supplementary Fig. 17: Citrullinated antigens stimulate ACPA<sup>+</sup> RA blood CD8<sup>+</sup> T cells to exhibit cytotoxic activity and mediate cell killing.** **a, b** Flow cytometry analysis of CD107a<sup>+</sup> expression by CD8<sup>+</sup> T cells was used as a measure of degranulation activity. ACPA<sup>+</sup> RA blood samples were stimulated with citrullinated or native forms of the RA autoantigens vimentin and histone H3 (100 μM each), with or without anti-CD8/ HLA class I-blocking antibodies, and flow cytometry analysis performed to quantitate CD107a<sup>+</sup>CD8<sup>+</sup> T cells. For **(a)**, no stimulation (n = 18), anti-CD3/28 Abs (n = 16), NP/pp65 (n = 17), cit-vimentin (n = 18), vimentin (n = 7), cit-vimentin with anti-CD8/ HLA class I antibodies (n = 5); \*\*\**P* < 0.0001, \*\*\**P* = 0.0003, \*\**P* = 0.0021, #*P* = 0.0364, or #*P* = 0.0494. For **(b)**, n = 7; \*\*\**P* < 0.0001, or \**P* = 0.028. **c, d** Antigen-independent cytotoxic activity of cit-vimentin, native vimentin, or viral protein

stimulated CD8<sup>+</sup> T cells from ACPA+ RA (**c**) or HC (**d**) blood against the DLD-1 cell line was determined using a lactate dehydrogenase (LDH) release assay. For (**c**), cit-vimentin (n = 7) and vimentin (n = 5); \*\**P*=0.0025. For (**d**), NP/pp65 (n = 14), cit-vimentin or vimentin (n = 3). E:T ratio means effector/target cell ratio. Data are presented as means ± SEM. For (**a, b**), using two-tailed unpaired *t*-test: \**P* < 0.05, \*\**P* < 0.01, or \*\*\**P* < 0.001 versus no treatment; #*P* < 0.05 versus cit-vimentin; *P* value represented on the graph was compared with cit-vimentin. For (**c, d**), using two-way ANOVA with Tukey's multiple comparisons test: \*\**P* < 0.01. Source data are provided as a Source Data file. HC healthy control, ACPA anti-citrullinated protein antibodies, RA rheumatoid arthritis, mAb monoclonal antibody, Cit-Vim citrullinated vimentin.

## Supplementary Tables

**Supplementary Table 1. Demographic characteristics of RA patients.**

	<b>RA patients</b>
Total Number	59
Gender, Female/Male	4/55
Age (years) - mean $\pm$ SD	66.15 $\pm$ 12.24
Dis duration (years) - mean $\pm$ SD	11.35 $\pm$ 10.69
Disease activity	
ACPA positive (%)	76.27
RF positive (%)	73.33
CDAI score - mean $\pm$ SD	15.17 $\pm$ 15.95
MTX use (%)	51.11
MTX duration (years)	4.96 $\pm$ 4.9

**Supplementary Table 2 Differentially expressed genes in each cluster identified in ACPA+ RA as compared to HC blood.** Differentially expressed genes in each cluster were selected with Log2 Fold change > 0.8. Wilcoxon Rank Sum test and the Benjamini–Hochberg method to adjust the *P* values for multiple testing. Avg average, Pct percentage, p\_val\_adj adjusted *P* value.

<b>Gene</b>	<b>Avg_log2FC</b>	<b>Pct.1</b>	<b>Pct.2</b>	<b>p_val_adj</b>	<b>Cluster</b>
<i>CCR7</i>	2.17188736	0.952	0.528	0	Naïve
<i>LEF1</i>	2.00017763	0.954	0.652	0	Naïve
<i>ACTN1</i>	1.50463023	0.94	0.711	0	Naïve
<i>NELL2</i>	1.42062488	0.889	0.612	0	Naïve
<i>MAL</i>	1.32861183	0.804	0.643	0	Naïve
<i>LTB</i>	1.29286009	0.964	0.78	0	Naïve
<i>SELL</i>	1.25489209	0.951	0.817	0	Naïve
<i>NOSIP</i>	1.2525314	0.947	0.812	0	Naïve
<i>OXNAD1</i>	1.2385171	0.956	0.681	0	Naïve
<i>PIK3IP1</i>	1.1523528	0.96	0.805	0	Naïve
<i>AIF1</i>	1.14401643	0.867	0.767	0	Naïve
<i>TRABD2A</i>	1.07210543	0.915	0.757	0	Naïve
<i>LDLRAP1</i>	1.05680921	0.928	0.822	0	Naïve
<i>CD27</i>	1.03820024	0.934	0.718	0	Naïve
<i>SPINT2</i>	1.02349056	0.967	0.724	0	Naïve
<i>SNHG8</i>	0.99485116	0.993	0.924	0	Naïve
<i>NDFIP1</i>	0.97624462	0.986	0.869	0	Naïve
<i>ABLIM1</i>	0.96524899	0.787	0.402	0	Naïve
<i>CD8B</i>	0.90637784	0.98	0.9	0	Naïve
<i>CAMK4</i>	0.85677228	0.958	0.737	0	Naïve
<i>ARMH1</i>	0.82194778	0.793	0.738	0	Naïve
<i>HSPB1</i>	0.79107008	0.972	0.821	0	Naïve
<i>PCED1B</i>	0.76851258	0.867	0.737	0	Naïve
<i>RCAN3</i>	0.74887121	0.782	0.717	0	Naïve
<i>LDHB</i>	0.74089477	0.961	0.907	0	Naïve
<i>TCF7</i>	0.95014837	0.841	0.758	3.76E-301	Naïve
<i>CD55</i>	0.86560341	0.88	0.786	3.17E-286	Naïve
<i>SATB1</i>	0.76575809	0.749	0.593	1.77E-205	Naïve
<i>FLT3LG</i>	0.71486197	0.879	0.791	1.39E-199	Naïve
<i>SI00B</i>	0.81117364	0.72	0.75	1.09E-184	Naïve

<i>FGFBP2</i>	2.64092332	0.994	0.896	0	<i>GZMB+ GNLY+</i>
<i>GZMH</i>	2.48081498	0.982	0.899	0	<i>GZMB+ GNLY+</i>
<i>GNLY</i>	2.2873411	0.956	0.666	0	<i>GZMB+ GNLY+</i>
<i>GZMB</i>	2.20826241	0.949	0.829	0	<i>GZMB+ GNLY+</i>
<i>ZNF683</i>	1.93895227	0.954	0.888	0	<i>GZMB+ GNLY+</i>
<i>NKG7</i>	1.86436384	0.999	0.888	0	<i>GZMB+ GNLY+</i>
<i>PRF1</i>	1.71556813	0.988	0.773	0	<i>GZMB+ GNLY+</i>
<i>ADGRG1</i>	1.69693841	0.956	0.796	0	<i>GZMB+ GNLY+</i>
<i>LGALS1</i>	1.62412354	0.905	0.789	0	<i>GZMB+ GNLY+</i>
<i>SPON2</i>	1.56725113	0.933	0.694	0	<i>GZMB+ GNLY+</i>
<i>PRSS23</i>	1.46520941	0.945	0.865	0	<i>GZMB+ GNLY+</i>
<i>FCGR3A</i>	1.40710564	0.927	0.738	0	<i>GZMB+ GNLY+</i>
<i>CX3CR1</i>	1.31747937	0.874	0.788	0	<i>GZMB+ GNLY+</i>
<i>ZEB2</i>	1.29236084	0.834	0.797	0	<i>GZMB+ GNLY+</i>
<i>PLEK</i>	1.27834466	0.87	0.707	0	<i>GZMB+ GNLY+</i>
<i>CST7</i>	1.25537403	0.993	0.952	0	<i>GZMB+ GNLY+</i>
<i>CCL4</i>	1.25062717	0.856	0.876	0	<i>GZMB+ GNLY+</i>
<i>ITGB1</i>	1.21597628	0.94	0.911	0	<i>GZMB+ GNLY+</i>
<i>KLRD1</i>	1.21207058	0.915	0.787	0	<i>GZMB+ GNLY+</i>
<i>EFHD2</i>	1.17334611	0.876	0.717	0	<i>GZMB+ GNLY+</i>
<i>FCRL6</i>	1.15172947	0.935	0.93	0	<i>GZMB+ GNLY+</i>
<i>TBX21</i>	1.15137019	0.938	0.891	0	<i>GZMB+ GNLY+</i>
<i>GZMA</i>	1.14945792	0.986	0.847	0	<i>GZMB+ GNLY+</i>
<i>SI00A4</i>	1.13366146	0.995	0.937	0	<i>GZMB+ GNLY+</i>
<i>ITGB2</i>	1.07810461	0.995	0.959	0	<i>GZMB+ GNLY+</i>
<i>FLNA</i>	1.06917519	0.974	0.941	0	<i>GZMB+ GNLY+</i>
<i>SIPR5</i>	1.01688766	0.906	0.847	0	<i>GZMB+ GNLY+</i>
<i>SLC9A3R1</i>	0.98919573	0.974	0.894	0	<i>GZMB+ GNLY+</i>
<i>CCL5</i>	0.97731527	0.994	0.97	0	<i>GZMB+ GNLY+</i>
<i>HOPX</i>	0.97559354	0.913	0.871	0	<i>GZMB+ GNLY+</i>
<i>FCGR3A</i>	1.78521332	0.98	0.784	3.47E-253	<i>GZMB+ KIR+</i>
<i>TTC38</i>	1.06213688	0.961	0.761	3.68E-205	<i>GZMB+ KIR+</i>
<i>KIR3DL2</i>	0.78538508	0.816	0.57	4.05E-177	<i>GZMB+ KIR+</i>
<i>TRDV1</i>	1.75850042	0.823	0.758	3.73E-166	<i>GZMB+ KIR+</i>
<i>NKG7</i>	1.08540583	0.999	0.916	6.13E-161	<i>GZMB+ KIR+</i>
<i>SIPR5</i>	0.96269525	0.953	0.86	1.03E-154	<i>GZMB+ KIR+</i>
<i>GZMB</i>	1.12077486	0.971	0.859	1.27E-147	<i>GZMB+ KIR+</i>
<i>KLRC3</i>	1.17386824	0.793	0.594	2.35E-140	<i>GZMB+ KIR+</i>
<i>ADGRG1</i>	0.93451484	0.941	0.837	1.72E-135	<i>GZMB+ KIR+</i>

<i>CST7</i>	0.93584796	0.991	0.962	1.76E-130	<i>GZMB+ KIR+</i>
<i>PRF1</i>	1.01856434	0.989	0.828	4.97E-124	<i>GZMB+ KIR+</i>
<i>TBX21</i>	0.78939386	0.944	0.903	1.23E-123	<i>GZMB+ KIR+</i>
<i>C1orf21</i>	0.7426384	0.904	0.855	7.30E-122	<i>GZMB+ KIR+</i>
<i>KLRD1</i>	1.06721614	0.936	0.819	3.51E-117	<i>GZMB+ KIR+</i>
<i>FGR</i>	0.87854003	0.909	0.886	2.99E-115	<i>GZMB+ KIR+</i>
<i>FCRL6</i>	0.8177497	0.964	0.93	9.00E-114	<i>GZMB+ KIR+</i>
<i>FGFBP2</i>	0.8305155	0.983	0.922	9.59E-108	<i>GZMB+ KIR+</i>
<i>CCL5</i>	0.78234757	0.995	0.976	9.86E-99	<i>GZMB+ KIR+</i>
<i>PLEK</i>	0.84156567	0.912	0.747	7.33E-93	<i>GZMB+ KIR+</i>
<i>GZMH</i>	0.76278171	0.967	0.921	9.21E-89	<i>GZMB+ KIR+</i>
<i>CMC1</i>	0.83244954	0.816	0.632	8.14E-75	<i>GZMB+ KIR+</i>
<i>CCL4</i>	0.8534083	0.916	0.868	2.86E-68	<i>GZMB+ KIR+</i>
<i>IKZF2</i>	0.77393067	0.693	0.574	4.91E-65	<i>GZMB+ KIR+</i>
<i>TIGIT</i>	1.00714799	0.741	0.696	9.89E-58	<i>GZMB+ KIR+</i>
<i>ZEB2</i>	0.80594977	0.82	0.807	3.56E-55	<i>GZMB+ KIR+</i>
<i>CD63</i>	0.7813272	0.859	0.812	1.37E-53	<i>GZMB+ KIR+</i>
<i>GNLY</i>	0.80162342	0.881	0.744	2.43E-53	<i>GZMB+ KIR+</i>
<i>KLRF1</i>	0.93276105	0.648	0.531	5.53E-38	<i>GZMB+ KIR+</i>
<i>NCR1</i>	0.82343017	0.592	0.417	5.63E-19	<i>GZMB+ KIR+</i>
<i>TYROBP</i>	0.84125194	0.66	0.688	3.20E-12	<i>GZMB+ KIR+</i>
<i>TRDV2</i>	1.99128693	0.913	0.407	0	<i>TCRgd+</i>
<i>TRGV9</i>	1.92133086	0.852	0.666	1.24E-221	<i>TCRgd+</i>
<i>TYROBP</i>	1.33879925	0.875	0.677	2.75E-189	<i>TCRgd+</i>
<i>MAP3K8</i>	0.63507239	0.974	0.865	1.50E-171	<i>TCRgd+</i>
<i>SI00B</i>	0.62382483	0.867	0.736	5.13E-162	<i>TCRgd+</i>
<i>DUSP2</i>	1.05992088	0.996	0.965	4.05E-146	<i>TCRgd+</i>
<i>KLRB1</i>	1.04734785	0.882	0.763	1.68E-121	<i>TCRgd+</i>
<i>MATK</i>	0.71312879	0.949	0.881	6.32E-108	<i>TCRgd+</i>
<i>KLRG1</i>	0.85459333	0.913	0.73	2.58E-103	<i>TCRgd+</i>
<i>KLRD1</i>	0.77247908	0.933	0.819	1.04E-102	<i>TCRgd+</i>
<i>ID2</i>	0.92916918	0.965	0.78	3.99E-97	<i>TCRgd+</i>
<i>ALOX5AP</i>	0.80492512	0.915	0.815	1.02E-71	<i>TCRgd+</i>
<i>GNLY</i>	0.74442669	0.879	0.743	5.58E-71	<i>TCRgd+</i>
<i>CST7</i>	0.58130969	0.99	0.962	1.93E-64	<i>TCRgd+</i>
<i>NKG7</i>	0.57142342	0.998	0.916	3.11E-63	<i>TCRgd+</i>
<i>DUSP1</i>	0.82391136	0.98	0.889	2.01E-61	<i>TCRgd+</i>
<i>KRT10</i>	0.55492149	0.988	0.941	6.52E-59	<i>TCRgd+</i>
<i>TRDC</i>	1.1402084	0.663	0.562	4.61E-51	<i>TCRgd+</i>

<i>PRELID1</i>	0.56047116	0.969	0.858	2.82E-46	<i>TCRgd+</i>
<i>ANXA1</i>	0.52941704	0.973	0.894	4.07E-45	<i>TCRgd+</i>
<i>APOBEC3G</i>	0.53168543	0.853	0.825	6.76E-42	<i>TCRgd+</i>
<i>ZFP36</i>	0.52644306	0.981	0.916	3.04E-38	<i>TCRgd+</i>
<i>IL2RB</i>	0.53117476	0.928	0.861	1.33E-31	<i>TCRgd+</i>
<i>CD300A</i>	0.51766821	0.7	0.544	1.28E-24	<i>TCRgd+</i>
<i>NCR3</i>	0.5182753	0.65	0.648	2.80E-12	<i>TCRgd+</i>
<i>TRGC1</i>	0.68535161	0.565	0.393	4.71E-10	<i>TCRgd+</i>
<i>KLRC1</i>	0.58682728	0.394	0.33	7.14E-06	<i>TCRgd+</i>
<i>GZMK</i>	1.99717379	0.895	0.768	0	<i>GZMK+</i>
<i>DUSP2</i>	1.35848465	0.991	0.96	0	<i>GZMK+</i>
<i>CD74</i>	1.07464734	0.946	0.907	0	<i>GZMK+</i>
<i>COTL1</i>	0.66638748	0.93	0.872	7.40E-248	<i>GZMK+</i>
<i>RGS1</i>	0.83275317	0.705	0.643	9.78E-207	<i>GZMK+</i>
<i>FYN</i>	0.81771733	0.904	0.784	1.04E-193	<i>GZMK+</i>
<i>CMC1</i>	1.0803568	0.754	0.615	1.22E-184	<i>GZMK+</i>
<i>CCL4</i>	0.63132847	0.892	0.866	3.33E-149	<i>GZMK+</i>
<i>CCL5</i>	0.52091684	0.989	0.975	5.05E-143	<i>GZMK+</i>
<i>LYAR</i>	0.58186799	0.882	0.839	3.17E-104	<i>GZMK+</i>
<i>IL10RA</i>	0.51340213	0.945	0.897	4.31E-87	<i>GZMK+</i>
<i>TIGIT</i>	0.68709491	0.686	0.701	1.23E-81	<i>GZMK+</i>
<i>LYST</i>	0.56801848	0.693	0.615	2.91E-77	<i>GZMK+</i>
<i>PIK3R1</i>	0.57399294	0.74	0.748	1.04E-46	<i>GZMK+</i>
<i>LTB</i>	1.07119384	0.977	0.804	0	Memory
<i>IL7R</i>	1.03381631	0.981	0.791	0	Memory
<i>COTL1</i>	0.76448942	0.959	0.871	0	Memory
<i>CAPG</i>	0.55036677	0.898	0.714	0	Memory
<i>AQP3</i>	0.54135708	0.898	0.798	5.71E-169	Memory
<i>LGALS3</i>	0.58233773	0.816	0.783	2.94E-168	Memory
<i>JUNB</i>	0.65509324	0.997	0.977	3.32E-153	Memory
<i>ARHGAP15</i>	0.76563726	0.913	0.809	2.03E-151	Memory
<i>PTGER2</i>	0.55948452	0.912	0.815	5.78E-145	Memory
<i>LDHB</i>	0.60480366	0.963	0.915	1.60E-143	Memory
<i>CXCR3</i>	0.64707736	0.868	0.738	4.90E-143	Memory
<i>FXD2</i>	0.72697369	0.762	0.702	1.39E-142	Memory
<i>FOS</i>	0.59514237	0.921	0.784	6.91E-139	Memory
<i>TIMP1</i>	0.619823	0.838	0.698	3.64E-115	Memory
<i>PLP2</i>	0.68495292	0.86	0.759	9.64E-105	Memory
<i>TCF7</i>	0.53829795	0.884	0.763	1.00E-100	Memory

<i>KLRB1</i>	2.46140509	0.967	0.757	0	<i>CCR6+ CD161+</i>
<i>LST1</i>	0.65603783	0.865	0.596	1.93E-208	<i>CCR6+ CD161+</i>
<i>IL7R</i>	1.23562461	0.984	0.806	4.41E-207	<i>CCR6+ CD161+</i>
<i>DPP4</i>	0.91774681	0.888	0.701	1.12E-188	<i>CCR6+ CD161+</i>
<i>TYROBP</i>	1.19564119	0.884	0.675	1.58E-184	<i>CCR6+ CD161+</i>
<i>MYC</i>	0.7221775	0.942	0.667	4.50E-178	<i>CCR6+ CD161+</i>
<i>TTC39C</i>	0.78485329	0.964	0.854	2.13E-173	<i>CCR6+ CD161+</i>
<i>GZMK</i>	1.10947754	0.925	0.783	1.66E-162	<i>CCR6+ CD161+</i>
<i>NFKB1A</i>	1.35943949	0.936	0.831	2.11E-147	<i>CCR6+ CD161+</i>
<i>NCR3</i>	1.36753518	0.838	0.637	1.26E-143	<i>CCR6+ CD161+</i>
<i>DUSP1</i>	1.12077918	0.982	0.888	3.22E-142	<i>CCR6+ CD161+</i>
<i>AQP3</i>	1.08914582	0.927	0.804	1.39E-133	<i>CCR6+ CD161+</i>
<i>LGALS3</i>	0.95770062	0.805	0.786	3.32E-125	<i>CCR6+ CD161+</i>
<i>SLAMF1</i>	0.66166051	0.811	0.609	1.59E-108	<i>CCR6+ CD161+</i>
<i>ALOX5AP</i>	0.97222512	0.922	0.813	6.24E-106	<i>CCR6+ CD161+</i>
<i>GCHFR</i>	0.84474297	0.805	0.628	2.54E-105	<i>CCR6+ CD161+</i>
<i>KLRG1</i>	0.91738742	0.904	0.73	1.29E-93	<i>CCR6+ CD161+</i>
<i>CXCR6</i>	0.89331155	0.719	0.547	7.54E-90	<i>CCR6+ CD161+</i>
<i>CCR6</i>	0.81160558	0.728	0.758	1.09E-88	<i>CCR6+ CD161+</i>
<i>LTB</i>	0.80620585	0.967	0.817	1.07E-75	<i>CCR6+ CD161+</i>
<i>ZBTB16</i>	0.90784986	0.697	0.641	6.60E-71	<i>CCR6+ CD161+</i>
<i>CD69</i>	0.83337076	0.925	0.84	3.02E-52	<i>CCR6+ CD161+</i>
<i>GBP5</i>	0.60826196	0.851	0.675	5.81E-48	<i>CCR6+ CD161+</i>
<i>FOS</i>	0.65314205	0.928	0.794	1.06E-43	<i>CCR6+ CD161+</i>
<i>ERN1</i>	0.83350715	0.73	0.697	4.33E-36	<i>CCR6+ CD161+</i>
<i>SLC4A10</i>	0.89645098	0.615	0.378	8.03E-31	<i>CCR6+ CD161+</i>
<i>JAML</i>	0.6162617	0.669	0.581	7.01E-29	<i>CCR6+ CD161+</i>
<i>ZFP36L1</i>	0.69958413	0.82	0.735	5.53E-26	<i>CCR6+ CD161+</i>
<i>TMIGD2</i>	0.72239382	0.697	0.714	9.21E-26	<i>CCR6+ CD161+</i>
<i>DDIT4</i>	0.7047064	0.984	0.947	1.59E-17	<i>CCR6+ CD161+</i>

**Supplementary Table 3. HLA typing of ACPA+ RA patients. 12 ACPA+ RA patients were randomly selected and sequenced to get HLA types.**

Sample ID	A/1	A/2	B/1	B/2	C/1	C/2
JG330	A*29:02:01:01	A*32:01:01:01	B*35:03:01:01	B*44:02:01:01	C*12:03:01:01	C*05:01:01:02
JG331	A*24:07:01	A*11:02:01	B*15:35	B*55:02:01:03	C*07:02:01:01	C*01:02:01:01
JG332	A*24:02:01:01	A*01:01:01:01	B*27:05:02:05	B*08:01:01:01	C*02:02:02:01	C*07:01:01:01
JG127	A*03:01:01:01	A*01:01:01:01	B*35:01:01:05	B*52:01:01:02	C*04:01:01:11	C*12:02:02:01
JG128	A*23:01:01:01	A*11:01:01:01	B*07:02:01:01	B*35:01:01:05	C*07:02:01:03	C*04:01:01:01
JG129	A*23:01:01:01	A*68:01:01:02	B*49:01:01:01	B*15:220:01:01	C*07:01:01:16	C*04:01:01:01
JG143	A*02:01:01:01	A*74:01:01:01	B*42:01:01	B*27:05:02:01	C*17:01:01:02	C*15:02:01:01
JG147	A*11:01:01:01	A*02:06:01:01	B*40:01:02:01	B*51:01:01:05	C*03:04:01:02	C*15:02:01:03
JG149	A*24:02:01:01	A*31:01:02:01	B*08:01:01:01	B*57:01:01:01	C*07:01:01:01	C*06:02:01:01
JG159	A*29:02:01:02	A*29:02:01:01	B*45:01:01:01	B*44:03:01:01	C*06:02:01:01	C*16:01:01:01
JG262	A*03:01:01:01	A*01:01:01:01	B*40:01:02:01	B*08:01:01:01	C*03:04:01:01	C*07:01:01:01
JG272	A*30:01:01:01	A*30:01:01:01	B*53:01:01:01	B*44:03:01:10	C*04:01:01:14	C*17:01:01:02
Sample ID	DPA1/1	DPA1/2	DPB1/1	DPB1/2	DQA1/1	DQA1/2
JG330	DPA1*02:02:02:01	DPA1*01:03:01:05	DPB1*05:01:01:01	DPB1*04:02:01:02	DQA1*03:01:01	DQA1*05:05:01:02
JG331	DPA1*02:02:02:01	X	DPB1*05:01:01:01	DPB1*01:01:01:04	DQA1*03:03:01:02	DQA1*06:01:01:02
JG332	DPA1*01:03:01:01	DPA1*1:03:01:05	DPB1*02:01:02:05/416:01	DPB1*04:02:01:02/105:01/665:01	DQA1*03:01:01	DQA1*01:01:01:05
JG127	DPA1*01:03:01:04	DPA1*01:03:01:02	DPB1*04:01:01:06	DPB1*04:01:01:05	DQA1*01:01:01:05	DQA1*03:01:01
JG128	DPA1*02:06	DPA1*01:03:01:04	DPB1*04:01:01:06	DPB1*04:01:01:05	DQA1*01:02:01:03	DQA1*03:01:01
JG129	DPA1*02:01:08:02	DPA1*02:01:01:01	DPB1*01:01:01:01	DPB1*11:01:01:01	DQA1*03:03:01:01	DQA1*01:02:01:04
JG143	DPA1*02:01:08:02	DPA1*01:03:01:01	DPB1*01:01:01:01	DPB1*02:01:02:05	DQA1*04:01:01:04	DQA1*04:01:01:02
JG147	DPA1*02:02:02:01	DPA1*02:02:02:01	DPB1*05:01:01:01	DPB1*03:01:01:01	DQA1*03:03:01:03	DQA1*03:03:01:03
JG149	DPA1*01:03:01:04	DPA1*01:03:01:02	DPB1*04:01:01:06	DPB1*04:01:01:05	DQA1*05:01:01:02	DQA1*03:03:01:01
JG159	DPA1*01:03:01:02	DPA1*01:03:01:01	DPB1*16:01:01:01	DPB1*04:01:01:05	DQA1*03:03:01:01	DQA1*02:01:01:01
JG262	DPA1*01:03:01:03	DPA1*01:03:01:02	DPB1*03:01:01:01	DPB1*04:01:01:05	DQA1*03:01:01	DQA1*05:01:01:02
JG272	DPA1*01:03:01:02	DPA1*02:01:08:02	DPB1*104:01:01:01	DPB1*01:01:01:01	DQA1*01:02:01:03	DQA1*05:05:01:01
Sample ID	DQB1/1	DQB1/2	DRB1/1	DRB1/2	DRB345/1	DRB345/2
JG330	DQB1*03:02:01:01	DQB1*03:01:01:03	DRB1*04:01:01:03	DRB1*11:04:01	DRB4*01:03:01:10	DRB3*02:02:01:04
JG331	DQB1*04:02:01:09	DQB1*03:01:01:12	DRB1*04:05:01:01	DRB1*12:02:01:01	DRB4*01:03:01:01	DRB3*03:01:03
JG332	DQB1*03:02:01:01	DQB1*05:01:01:03	DRB1*04:01:01:03	DRB1*01:01:01	DRB4*01:03:01:10	Absent

<b>JG127</b>	DQB1*05:01:0 1:03	DQB1*03:02:0 1:02	DRB1*01:01:01	DRB1*04:04:01	DRB4*01:03:0 3	Absent
<b>JG128</b>	DQB1*06:02:0 1:01	DQB1*03:02:0 1:02	DRB1*15:01:01:05	DRB1*04:04:01	DRB5*01:01:0 1:01	DRB4*01:03:0 1:01
<b>JG129</b>	DQB1*02:02:0 1:02	DQB1*05:01:0 1:03	DRB1*09:01:02:01	DRB1*13:02:01:02	DRB4*01:01:0 1:01	DRB3*03:01:0 1:01
<b>JG143</b>	DQB1*04:02:0 1:08	DQB1*04:02:0 1:04	DRB1*08:11	DRB1*03:02:01	DRB3*01:62	Absent
<b>JG147</b>	DQB1*04:01:0 1:02	DQB1*04:01:0 1:02	DRB1*04:05:01:03	DRB1*04:05:01:02	DRB4*01:03:0 1:01	DRB4*01:03:0 1:01
<b>JG149</b>	DQB1*02:01:0 1	DQB1*03:01:0 1:01	DRB1*03:01:01:01	DRB1*04:01:01:01	DRB3*01:01:0 2:01	DRB4*01:03:0 1:10
<b>JG159</b>	DQB1*03:01:0 1:01	DQB1*02:02:0 1:01	DRB1*04:01:01:03	DRB1*07:01:01:04	DRB4*01:03:0 1:10	DRB4*01:01:0 1:01
<b>JG262</b>	DQB1*03:02:0 1:02	DQB1*02:01:0 1	DRB1*04:04:01	DRB1*03:01:01:01	DRB4*01:03:0 1:10	DRB3*01:01:0 2:01
<b>JG272</b>	DQB1*06:02:0 1:01	DQB1*03:01:0 1:03	DRB1*15:03:01:02	DRB1*13:03:01	DRB5*01:01:0 1:01	DRB3*01:01:0 2:03

**Supplementary Table 4. Antibodies used in flow cytometry experiments.**

<b>Marker</b>	<b>Clone</b>	<b>Fluorescence</b>	<b>Company</b>	<b>Dilution</b>
<b>CD3</b>	SK7	APC-H7	BD Biosciences	1:200
<b>CD4</b>	RPA-T4	FITC	BD Biosciences	1:200
		Pacific Blue	BioLegend	1:200
<b>CD8</b>	RPA-T8	Alexa Fluor 700	BD Biosciences	1:200
<b>CD69</b>	FN50	PerCP-Cy5.5	BD Biosciences	1:200
<b>GPR56</b>	CG4	PE	BioLegend	1:200
<b>TCR<math>\gamma\delta</math></b>	B1	FITC	BD Biosciences	1:200
<b>Granzyme K</b>	G3H69	Alexa Fluor 647	BD Biosciences	1:150
<b>Granzyme B</b>	GB11	PE	BD Biosciences	1:200
		FITC		1:200
<b>CCR7</b>	150503	PE	BD Biosciences	1:150
<b>CD45RA</b>	HI100	Brilliant Violet 421	BioLegend	1:200
		FITC	BD Biosciences	1:200
<b>PD-1</b>	NAT105	PerCP-Cy5.5	BioLegend	1:200
<b>TIM3</b>	F38-2E2	APC	BioLegend	1:200
<b>CD158b (KIR2DL2/L3)</b>	DX27	FITC	BioLegend	1:200
<b>His-tag</b>	J095G46	PE	BioLegend	1:100
<b>HLA-A,B,C</b>	W6/32	APC-Cy7	BioLegend	1:200
<b>Ki-67</b>	Ki-67	Brilliant Violet 421	BioLegend	1:100
<b>IFN<math>\gamma</math></b>	4S.B3	APC-Cy7	BioLegend	1:200
	B27	PE	BioLegend	1:200
	B27	APC	BD Biosciences	1:150
<b>CD107a</b>	H4S3	APC-Cy7	BioLegend	1:200

**Supplementary Table 5. CITE-seq antibodies used in 10X single cell RNA sequencing.**

<b>TotalSeqC-Barcode</b>	<b>Specificity</b>	<b>Clone</b>	<b>Reactivity</b>	<b>Barcode sequence</b>
34	CD3	UCHT1	Human	CTCATTGTAACCTCCT
45	CD4	SK3	Human	GAGGTTAGTGATGGA
46	CD8	SK1	Human	GCGCAACTTGATGAT
50	CD19	HIB19	Human	CTGGGCAATTACTCG
63	CD45RA	HI100	Human	TCAATCCTTCCGCTT
84	CD56	QA17A16	Human	TTCGCCGCATTGAGT
85	CD25	BC96	Human	TTTGTCTGTACGCC
87	CD45RO	UCHL1	Human	CTCCGAATCATGTTG
88	PD-1	EH12.2H7	Human	ACAGCGCCGTATTTA
89	TIGIT	A15153G	Human	TTGCTTACCGCCAGA
139	TCRrd	B1	Human	CTTCCGATTTCATTCA
140	CXCR3	G025H7	Human	GCGATGGTAGATTAT
143	CCR6	G034E3	Human	GATCCCTTTGTCACT
144	CXCR5	J252D4	Human	AATTCAACCGTCGCC
146	CD69	FN50	Human	GTCTCTTGGCTTAAA
147	CD62L	DREG-56	Human	GTCCCTGCAACTTGA
148	CD197/CCR7	G043H7	Human	AGTTCAGTCAACCGA
149	CD161	HP-3G10	Human	GTACGCAGTCCTTCT
154	CD27	O323	Human	GCACTCCTGCATGTA
159	HLA-DR	L243	Human	AATAGCGAGCAAGTA
165	NKG2D	1D11	Human	CGTGTTTGTTCCTCA
168	CD57	QA17A04	Human	AACTCCCTATGGAGG
179	CX3CR1	K0124E1	Human	AGTATCGTCTCTGGG
224	TCRab	IP26	Human	CGTAACGTAGAGCGA
246	CD122	TU27	Human	TCATTTTCCTCCGATT
250	KLRG1	2F1/KLRG1	Human/Mouse	GTAGTAGGCTAGACC
386	CD28	CD28.2	Human	TGAGAACGACCCTAA
389	CD38	HIT2	Human	TGTACCCGCTTGTGA
390	IL-7Ra	A019D5	Human	GTGTGTTGTCCTATG
592	CD158b/KIR2DL2/L3	DX27	Human	GACCCGTAGTTTGAT
599	CD158e1/KIR3DL1	DX9	Human	GGACGCTTTCCTTGA

**Supplementary Table 6. Gene list used to measure a score of synovial trafficking, cytotoxicity, proliferation or inflammation.**

<b>RA trafficking</b>	<b>Cytotoxicity</b>	<b>Proliferation</b>	<b>Inflammation</b>
<i>CXCR1</i>	<i>GNLY</i>	<i>HMGB2</i>	<i>AGER</i>
<i>CXCR2</i>	<i>GZMA</i>	<i>CDK1</i>	<i>ALOX12</i>
<i>CXCR3</i>	<i>GZMB</i>	<i>NUSAP1</i>	<i>ALOX15</i>
<i>CXCR4</i>	<i>GZMH</i>	<i>UBE2C</i>	<i>ALOX5</i>
<i>CXCR5</i>	<i>GZMM</i>	<i>BIRC5</i>	<i>AREG</i>
<i>CCR1</i>	<i>GZMK</i>	<i>TPX2</i>	<i>ATF2</i>
<i>CCR2</i>	<i>NKG7</i>	<i>TOP2A</i>	<i>BCL2L1</i>
<i>CCR3</i>	<i>KLRB1</i>	<i>NDC80</i>	<i>BCL6</i>
<i>CCR4</i>	<i>KLRD1</i>	<i>CKS2</i>	<i>BIRC2</i>
<i>CCR5</i>	<i>KLRK1</i>	<i>NUF2</i>	<i>C1QA</i>
<i>CCR6</i>	<i>NCR1</i>	<i>CKS1B</i>	<i>C1QB</i>
<i>CCR10</i>	<i>SLAMF6</i>	<i>MKI67</i>	<i>C1R</i>
<i>CX3CR1</i>	<i>SLAMF7</i>	<i>TMPO</i>	<i>C1S</i>
<i>CCL2</i>	<i>TYROBP</i>	<i>CENPF</i>	<i>C2</i>
<i>CCL3</i>	<i>KIR3DL1</i>	<i>TACC3</i>	<i>C3</i>
<i>CCL4</i>	<i>KIR3DL2</i>	<i>SMC4</i>	<i>C3AR1</i>
<i>CCL5</i>		<i>CCNB2</i>	<i>C5</i>
<i>CCL19</i>		<i>CKAP2L</i>	<i>CCL17</i>
<i>CCL20</i>		<i>CKAP2</i>	<i>CCL19</i>
<i>CXCL10</i>		<i>AURKB</i>	<i>CCL2</i>
<i>CXCL16</i>		<i>BUB1</i>	<i>CCL20</i>
		<i>KIF11</i>	<i>CCL22</i>
		<i>ANP32E</i>	<i>CCL23</i>
		<i>TUBB4B</i>	<i>CCL24</i>
		<i>GTSE1</i>	<i>CCL3</i>
		<i>KIF20B</i>	<i>CCL4</i>
		<i>HJURP</i>	<i>CCL5</i>
		<i>CDCA3</i>	<i>CCR1</i>
		<i>CDC20</i>	<i>CCR2</i>
		<i>TTK</i>	<i>CCR3</i>
		<i>CDC25C</i>	<i>CCR4</i>
		<i>KIF2C</i>	<i>CCR7</i>
		<i>RANGAP1</i>	<i>CD4</i>

		<i>NCAPD2</i>	<i>CD40</i>
		<i>DLGAP5</i>	<i>CD40LG</i>
		<i>CDCA2</i>	<i>CD55</i>
		<i>CDCA8</i>	<i>CD86</i>
		<i>ECT2</i>	<i>CDC42</i>
		<i>KIF23</i>	<i>CEBPB</i>
		<i>HMMR</i>	<i>CFB</i>
		<i>AURKA</i>	<i>CFD</i>
		<i>PSRC1</i>	<i>CFL1</i>
		<i>ANLN</i>	<i>CREB1</i>
		<i>LBR</i>	<i>CSF1</i>
		<i>CKAP5</i>	<i>CSF2</i>
		<i>CENPE</i>	<i>CXCL10</i>
		<i>CTCF</i>	<i>CXCL3</i>
		<i>NEK2</i>	<i>CXCL9</i>
		<i>G2E3</i>	<i>CXCR1</i>
		<i>GAS2L3</i>	<i>CXCR2</i>
		<i>CBX5</i>	<i>CXCR4</i>
		<i>CENPA</i>	<i>CYSLTR1</i>
			<i>CYSLTR2</i>
			<i>DAXX</i>
			<i>DDIT3</i>
			<i>ELK1</i>
			<i>FASLG</i>
			<i>FLT1</i>
			<i>FOS</i>
			<i>FXRD2</i>
			<i>GNAQ</i>
			<i>GNAS</i>
			<i>GNB1</i>
			<i>GRB2</i>
			<i>HDAC4</i>
			<i>HIF1A</i>
			<i>HLA-DRA</i>
			<i>HLA-DRB1</i>
			<i>HMGB1</i>
			<i>HMGB2</i>
			<i>HMGN1</i>
			<i>HRAS</i>

			<i>HSH2D</i>
			<i>HSPB1</i>
			<i>IFI44</i>
			<i>IFIT1</i>
			<i>IFIT2</i>
			<i>IFIT3</i>
			<i>IFNG</i>
			<i>IL10</i>
			<i>IL10RB</i>
			<i>IL12A</i>
			<i>IL13</i>
			<i>IL15</i>
			<i>IL17A</i>
			<i>IL18</i>
			<i>IL18RAP</i>
			<i>IL1A</i>
			<i>IL1B</i>
			<i>IL1R1</i>
			<i>IL1RAP</i>
			<i>IL1RN</i>
			<i>IL2</i>
			<i>IL21</i>
			<i>IL22</i>
			<i>IL22RA2</i>
			<i>IL23A</i>
			<i>IL23R</i>
			<i>IL3</i>
			<i>IL4</i>
			<i>IL5</i>
			<i>IL6</i>
			<i>IL6R</i>
			<i>IL7</i>
			<i>IRF1</i>
			<i>IRF3</i>
			<i>IRF5</i>
			<i>IRF7</i>
			<i>ITGB2</i>
			<i>JUN</i>
			<i>KEAP1</i>

			<i>LIMK1</i>
			<i>LTA</i>
			<i>LTB</i>
			<i>LTB4R</i>
			<i>LTB4R2</i>
			<i>LY96</i>
			<i>MAFF</i>
			<i>MAFG</i>
			<i>MAFK</i>
			<i>MAP2K1</i>
			<i>MAP2K4</i>
			<i>MAP2K6</i>
			<i>MAP3K1</i>
			<i>MAP3K5</i>
			<i>MAP3K7</i>
			<i>MAP3K9</i>
			<i>MAPK1</i>
			<i>MAPK14</i>
			<i>MAPK3</i>
			<i>MAPK8</i>
			<i>MAPKAPK2</i>
			<i>MAPKAPK5</i>
			<i>MASP2</i>
			<i>MAX</i>
			<i>MEF2A</i>
			<i>MEF2C</i>
			<i>MEF2D</i>
			<i>MKNK1</i>
			<i>MMP9</i>
			<i>MRC1</i>
			<i>MX1</i>
			<i>MX2</i>
			<i>MYC</i>
			<i>MYD88</i>
			<i>NFATC3</i>
			<i>NFE2L2</i>
			<i>NFKB1</i>
			<i>NLRP3</i>
			<i>NOD1</i>

			<i>NOD2</i>
			<i>NR3C1</i>
			<i>OAS2</i>
			<i>OASL</i>
			<i>PDGFA</i>
			<i>PLA2G4A</i>
			<i>PLCB1</i>
			<i>PPP1R12B</i>
			<i>PRKCA</i>
			<i>PRKCB</i>
			<i>PTGDR2</i>
			<i>PTGER2</i>
			<i>PTGER4</i>
			<i>PTGIR</i>
			<i>PTGS1</i>
			<i>PTGS2</i>
			<i>PTK2</i>
			<i>RAC1</i>
			<i>RAF1</i>
			<i>RAPGEF2</i>
			<i>RELA</i>
			<i>RELB</i>
			<i>RHOA</i>
			<i>RIPK1</i>
			<i>RIPK2</i>
			<i>ROCK2</i>
			<i>RPS6KA5</i>
			<i>SHC1</i>
			<i>SMAD7</i>
			<i>STAT1</i>
			<i>STAT2</i>
			<i>STAT3</i>
			<i>TBXA2R</i>
			<i>TCF4</i>
			<i>TGFB1</i>
			<i>TGFB3</i>
			<i>TGFBR1</i>
			<i>TLR1</i>
			<i>TLR2</i>

			<i>TLR3</i>
			<i>TLR4</i>
			<i>TLR5</i>
			<i>TLR6</i>
			<i>TLR8</i>
			<i>TLR9</i>
			<i>TNF</i>
			<i>TNFAIP3</i>
			<i>TNFSF14</i>
			<i>TOLLIP</i>
			<i>TRADD</i>
			<i>TRAF2</i>
			<i>TREM2</i>
			<i>TSLP</i>
			<i>TYROBP</i>