



Soluble Urokinase Plasminogen Activator Receptor (suPAR) as a Biomarker of Systemic Chronic Inflammation

Line Jee Hartmann Rasmussen^{1,2*}, Jens Emil Vang Petersen³ and Jesper Eugen-Olsen¹

¹ Department of Clinical Research, Copenhagen University Hospital Amager and Hvidovre, Hvidovre, Denmark, ² Department of Psychology and Neuroscience, Duke University, Durham, NC, United States, ³ Division of Infectious Diseases, Duke University School of Medicine, Durham, NC, United States

OPEN ACCESS

Edited by:

Andrea Bottaro,
Cooper Medical School of Rowan
University, United States

Reviewed by:

Christopher Sjöwall,
Linköping University, Sweden
Andrea Baragetti,
University of Milan, Italy

*Correspondence:

Line Jee Hartmann Rasmussen
line.jee.hartmann.rasmussen@
regionh.dk

Specialty section:

This article was submitted to
Inflammation,
a section of the journal
Frontiers in Immunology

Received: 21 September 2021

Accepted: 09 November 2021

Published: 02 December 2021

Citation:

Rasmussen LJH, Petersen JEV and
Eugen-Olsen J (2021) Soluble
Urokinase Plasminogen Activator
Receptor (suPAR) as a Biomarker of
Systemic Chronic Inflammation.
Front. Immunol. 12:780641.
doi: 10.3389/fimmu.2021.780641

Systemic chronic inflammation (SCI) is persistent, health-damaging, low-grade inflammation that plays a major role in immunosenescence and in development and progression of many diseases. But currently, there are no recognized standard biomarkers to assess SCI levels alone, and SCI is typically measured by combining biomarkers of acute inflammation and infection, e.g., CRP, IL-6, and TNF α . In this review, we highlight 10 properties and characteristics that are shared by the blood protein *soluble urokinase plasminogen activator receptor* (suPAR) and SCI, supporting the argument that suPAR is a biomarker of SCI: (1) Expression and release of suPAR is upregulated by immune activation; (2) uPAR and suPAR exert pro-inflammatory functions; (3) suPAR is associated with the amount of circulating immune cells; (4) Blood suPAR levels correlate with the levels of established inflammatory biomarkers; (5) suPAR is minimally affected by acute changes and short-term influences, in contrast to many currently used markers of systemic inflammation; (6) Like SCI, suPAR is non-specifically associated with multiple diseases; (7) suPAR and SCI both predict morbidity and mortality; (8) suPAR and SCI share the same risk factors; (9) suPAR is associated with risk factors and outcomes of inflammation above and beyond other inflammatory biomarkers; (10) The suPAR level can be reduced by anti-inflammatory interventions and treatment of disease. Assessing SCI has the potential to inform risk for morbidity and mortality. Blood suPAR is a newer biomarker which may, in fact, be a biomarker of SCI since it is stably associated with inflammation and immune activation; shares the same risk factors as many age-related diseases; is both elevated by and predicts age-related diseases. There is strong evidence that suPAR is a prognostic marker of adverse events, morbidity, and mortality. It is associated with immune activity and prognosis across diverse conditions, including kidney disease, cardiovascular disease, cancer, diabetes, and inflammatory disorders. Thus, we think it likely represents a common underlying disease-process shared by many diseases; that is, SCI. We review the supporting literature and propose a research agenda that can help test the hypothesis that suPAR indexes SCI, with the potential of becoming the new gold standard for measuring SCI.

Keywords: inflammation, biomarkers, inflammation mediators - blood, C-reactive protein, interleukin-6, inflammaging, immunosenescence

INTRODUCTION

Systemic chronic inflammation (SCI) refers to persistent, low-grade inflammation, and it is involved in the pathogenesis of a wide variety of chronic non-communicable diseases that collectively constitute the leading cause of death globally (1). The diseases associated with SCI range from physical health problems, including cardiovascular disease, type 2 diabetes, cancer, and neurodegenerative disorders (2), to mental health disorders, such as depression, anxiety, and schizophrenia (3, 4). Assessing the level of SCI is therefore of utmost importance as it can provide information on disease burden as well as risk of incident disease, disease progression, and ultimately mortality (2).

The risk of developing SCI can be traced back to childhood development and is promoted by genetic, biological, social, environmental, and lifestyle factors (2). But even though the effects of SCI have been shown to persist throughout life with heightened risk of morbidity and mortality to follow, there are currently no recognized standard biomarkers to indicate and assess the level of SCI (2).

Inflammation

The textbook example of inflammation is an essential immune response aimed at eliminating pathogens, clearing infections, and promoting tissue repair and recovery. This acute inflammatory response is normally a short-term process that serves to protect the host, by coordinating the delivery and activation of immune mediators (plasma proteins and leukocytes, mainly neutrophils) to the site of infection or injury. Under normal circumstances, a successful acute inflammatory response is temporally restricted and results in resolution, repair, and restoration of homeostasis once the threat has been resolved (5). If the acute inflammatory response fails to eliminate the pathogens, foreign bodies, or other causes of tissue damage (for example, persistence of self-antigens in an autoimmune response), the result is a chronic inflammatory state where active inflammation fails to resolve. The chronic inflammatory state is characterized by replacement of the neutrophil infiltrate with macrophages and lymphocytes and in some cases with formation of granulomas and tertiary lymphoid tissues (5).

In contrast to these localized inflammatory responses, *systemic* chronic inflammation is a state of persistent, low-grade immune activation affecting multiple physiological systems. While it can be caused by chronic infections, SCI can also be partly sterile, i.e., it can be triggered in the absence of infectious agents and pathogen-associated molecular patterns (PAMPs). Damage-associated molecular patterns (DAMPs) from excess glucose, cholesterol crystals, or cellular breakdown products are known to trigger the response (2, 5). SCI is characterized by chronically elevated levels of inflammatory cytokines, chemokines, and acute-phase proteins, and this persistent inflammation can end up damaging tissues and organs (2, 5). Thus, the clinical consequences of SCI are linked to a variety of disorders across

organ systems and include increased risk of physical frailty, morbidity, and mortality (2).

SCI increases with age, but also chronic infections, microbiome dysbiosis, classic unhealthy lifestyle behaviors (smoking, physical inactivity, unhealthy diet), and obesity have been linked to the presence and promotion of SCI (2). Recently, social, psychological, and environmental factors including disturbed sleep, social isolation, psychological stress, and exposure to environmental or industrial toxicants (such as air pollutants) have been associated with SCI (2). Additionally, early development and childhood circumstances have been shown to promote SCI in adulthood (2). The long list of risk factors shows that there are many potential intervention targets for reducing SCI.

(Lack of) Biomarkers for Systemic Chronic Inflammation

Despite the established link between SCI and disease, there are no standard biomarkers for SCI (2). The causes and mechanisms of SCI are poorly understood (2, 5)—in part due to the lack of precise consistent diagnostic criteria or ways to measure SCI. At the moment, the measurement of SCI for clinical or research purposes is primarily carried out by assessing biomarkers of infection or acute inflammation, mainly C-reactive protein (CRP) or fibrinogen and other acute-phase proteins, white blood cell count, erythrocyte sedimentation rate, or cytokines, such as interleukin (IL)-1 β , IL-6, and tumor necrosis factor α (TNF α). The current standard for indicating presence of systemic chronic low-grade inflammation is slightly elevated CRP levels (>3 mg/L) measured with high-sensitivity CRP (hs-CRP) tests (6, 7). Alternatively, many studies use composite measures combining canonical biomarkers of acute inflammation when attempting to assess SCI (2). Both approaches have limitations. When using markers that are both sensitive to acute infection and systemic inflammation, acute influences such as undetected infections may be misinterpreted as SCI, or conversely, presence of infection may mask any underlying SCI resulting in a failing to notice actual SCI and misclassifying this as an acute infection; both situations with the result that accurate quantification of the actual level of SCI is obscured. Moreover, many of these inflammatory markers are short-lived and rapidly up- and down-regulated, as their biological function is to tightly control the acute inflammatory response. Their inherent volatility makes their quantification time-sensitive and complicates clinical interpretation. Two high-dimensional measures, the immune aging 'IMM-AGE' and the inflammatory aging clock 'iAge' based on multi-omics cellular immune profiling and deep-learning analysis of 50 inflammatory proteins, respectively, were recently shown to track SCI outcomes, but measures like these are complex and hard to apply broadly in other settings, requiring gene expression data to approximate these measures in cohorts that are less comprehensively phenotyped for immune measures (8, 9).

The lack of good stable biomarkers for SCI has the implication that there is no operational definition of SCI, and

thus an individual cannot be diagnosed with *systemic chronic inflammation*. This poses a serious problem, as there is a consensus that SCI is both a major risk factor and causally involved in the pathogenesis of many diseases, in addition to being a hallmark of *immunosenescence*, the age-related decline in immune function. Ultimately, emerging pathological processes may fly under the radar and go unnoticed for too long, wasting potential windows of opportunity for treatment or interventions that could slow the course of disease—or completely prevent disease development.

But what constitutes a good biomarker of SCI? First, a biomarker of SCI needs to accurately and reliably capture the level of SCI; it should be correlated with other measures of inflammation, without being overshadowed by any acute inflammation or other short-term influences. Second, it should be easily measured; stable over long time periods *in vitro* and *in vivo* (i.e., high test-retest reliability and temporal stability); and independent of smaller day-to-day variations, such as circadian/diurnal rhythm and fasting, while still being sensitive to significant contributions of psychosocial, environmental, and lifestyle factors as well as onset—or resolution—of chronic pathological processes.

suPAR as a Biomarker for Systemic Chronic Inflammation

We propose that the blood levels of the protein *soluble urokinase plasminogen activator receptor* (suPAR) is a robust biomarker of SCI, with potential to be the new gold standard for measuring SCI. suPAR has been found to be a broad prognostic biomarker associated with incident disease and adverse clinical outcomes across both general and patient populations. suPAR has been reviewed as a marker of kidney disease, sepsis, cardiovascular disease, and inflammatory disorders (10–13)—but given its nonspecific associations with immune activity and prognosis in very diverse diseases and conditions, it is not a disease-specific diagnostic marker. We think it represents a common underlying disease-process shared by many diseases; that is, SCI (14).

We bring forward 10 properties and characteristics of suPAR supporting the argument that suPAR is a biomarker of SCI: (i) Expression and release of suPAR is upregulated by immune activation; (ii) urokinase plasminogen activator receptor (uPAR) and suPAR exert pro-inflammatory functions; (iii) suPAR is associated with the amount of circulating immune cells; (iv) Blood suPAR levels correlate with the levels of established inflammatory biomarkers; (v) suPAR is minimally affected by acute changes and short-term influences; (vi) Like SCI, suPAR is non-specifically associated with multiple diseases; (vii) suPAR and SCI both predict morbidity and mortality; (viii) suPAR and SCI share the same risk factors; (ix) suPAR is associated with risk factors and outcomes of inflammation above and beyond other inflammatory biomarkers; (x) The suPAR level can be reduced by anti-inflammatory interventions and treatment of disease.

Since no clear definition or criterion has previously been established for the state of SCI, we review the supporting literature, integrating evidence from many different sources

and studies (including experimental, population-based, and clinical research), and propose a research agenda that can help test the hypothesis that suPAR indexes SCI.

What Is suPAR?

The protein suPAR is found in blood [plasma, serum (15, 16)] and other body fluids [cerebrospinal fluid (17), saliva (18), urine (15)] and is the soluble form of the membrane-bound receptor uPAR. When expressed on the cell surface membrane, uPAR is a central mediator of plasminogen activation and fibrinolysis, and is involved in several critical cellular processes by regulating extracellular matrix degradation (**Figure 1**). As such, uPAR is involved in proliferation, migration, adhesion, angiogenesis, and in the inflammatory response (20). Proteolytic cleavage of uPAR releases the soluble form, suPAR, to the bloodstream (21).

Release of suPAR from immune cells is increased upon an inflammatory stimulus (**Figure 1**); thus, the blood suPAR level is thought to reflect an individual's level of inflammation and immune activation (13). suPAR is indeed positively correlated with several inflammatory biomarkers, including CRP, IL-6, and TNF α (22–24), see **Appendix I**. While the suPAR level is elevated by a variety of pro-inflammatory conditions, it is generally low—although still detectable—in healthy individuals. In blood donors, the median suPAR level is around 2 ng/mL (25), and women generally have slightly higher suPAR than men (25–27). However, suPAR seems to increase more with age in men compared to women, and suPAR is similar among men and women in older adults (≥ 74 years of age) (28). suPAR concentrations are higher in serum than in plasma within individuals (27, 29).

A person's suPAR level is determined by various factors, including genetics, lifestyle, and chronic- and acute disease. How much each of these factors contributes to suPAR has not been fully elucidated, but suPAR seems to capture the overall impact of these. In addition to adaptable contributions by lifestyle, underlying chronic disease, and acute conditions, genetic predispositions also affect an individual's tissue uPAR expression and blood suPAR level. In a recent genome-wide association study, we found that blood suPAR levels were under substantial genetic influence (30), with a heritability estimate of 60% and 13 independently genome-wide significant sequence variants associated with suPAR across 11 distinct loci. Associated variants were found in and around *PLAUR* as well as the gene encoding the uPAR ligand urokinase plasminogen activator (uPA, or urokinase) *PLAU*, the kidney-disease-associated gene *PLA2R1*, and genes with relations to glycosylation, glycoprotein biosynthesis, and the immune response (30). This indicates that a combination of polymorphisms in different genes may affect the immune system and cause a higher basal level of suPAR. In studies of genetic polymorphisms in *PLAUR*, the polymorphisms rs344781 and rs4251923 have been associated with various clinical conditions (31–34), but none of these studies examined the corresponding suPAR levels and do therefore not offer a genetic explanation for increased suPAR levels in disease in general.

suPAR is removed from the circulation *via* renal excretion and cardiac clearance (27). Elevated suPAR levels are strongly associated with decline in kidney function, and as a result of poor

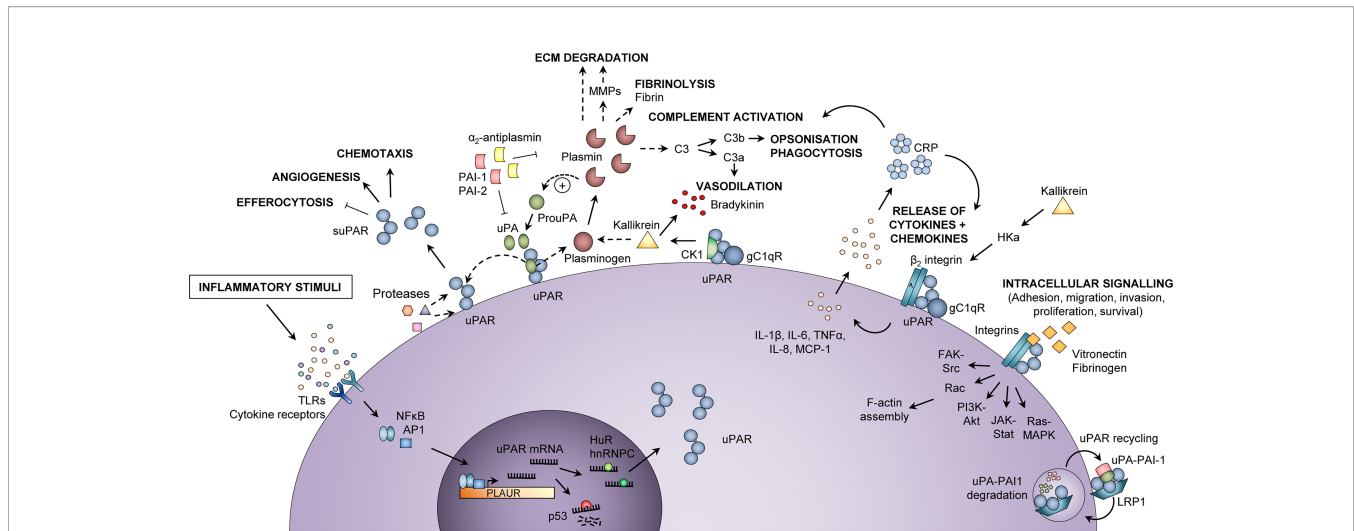


FIGURE 1 | Inflammatory functions of uPAR and suPAR. Upon an inflammatory stimulus, e.g., stimulation of toll-like receptors (TLRs) or cytokine receptors, the expression of urokinase plasminogen activator receptor (uPAR) in immunologically active cells is increased *via* activation of transcription factors, such as nuclear factor kappa-light-chain-enhancer of activated B cells (NF- κ B) and activator protein 1 (AP1), which bind to the promoter region of the *PLAUR* gene. The uPAR mRNA is either degraded (by p53) or stabilized for translation (by HuR or hnRNPC), after which uPAR is expressed at the cell surface, bound to the membrane *via* a glycosyl phosphatidylinositol (GPI) anchor. At the cell surface, uPAR can become cleaved by various proteases or its own ligand urokinase plasminogen activator (uPA), thus generating suPAR, which plays a role in inflammation by impairing neutrophil efferocytosis and stimulating angiogenesis and chemotaxis. Active uPA cleaves plasminogen to plasmin, which in turn cleaves and activates uPA. Plasmin activates matrix metalloproteases (MMPs), cleaves extracellular matrix (ECM) components, degrades fibrin, and activates the classical complement pathway, thereby promoting migration and invasion of cells, fibrinolysis, vasodilation, opsonization, and phagocytosis of foreign pathogens. Co-localization of uPAR with the proteins cytokeratin-1 (CK1) and globular C1q receptor (gC1qR) on the surface of endothelial cells also promotes vasodilation through release of bradykinin *via* activation of kallikrein. In a complex with β_2 integrin and gC1qR, uPAR also induces release of cytokines (IL-1 β , IL-6, TNF α) and chemokines (IL-8, MCP-1), upon stimulation by cleaved high molecular weight kininogen (HKa). Cytokines stimulate the production of C-reactive protein (CRP) from the liver, and CRP itself functions as an opsonin and also activates the classical complement pathway. Furthermore, uPAR interacts with vitronectin, fibrinogen, and integrins, mainly $\alpha_v\beta_2$ integrin (Mac-1) but also β_1 and β_3 integrin complexes, activating intracellular signaling pathways that facilitate cell adhesion, migration, invasion, proliferation, and survival by affecting F-actin assembly and gene transcription. The activity of uPA and plasmin is inhibited by plasminogen activator inhibitor (PAI)-1, PAI-2, and α_2 -antiplasmin. Binding of PAI-1 and low-density lipoprotein receptor-related protein 1 (LRP1) mediates endocytosis of uPAR-uPA-PAI-1 complexes, followed by lysosomal degradation of uPA and PAI-1 and recycling of uPAR back to the membrane. In endothelial cells, co-localization of uPAR with CK1 and gC1qR activates kallikrein and promotes the release of the vasodilator bradykinin. hnRNPC, heterogeneous nuclear ribonucleoprotein C; HuR, Hu antigen R; IL, interleukin; MCP-1, monocyte chemoattractant protein-1; TNF α , tumor necrosis factor α . Adapted from Rasmussen, LJH (2018) (19) with permission.

filtration patients on dialysis have consistently been shown to have very high suPAR levels (35, 36). However, suPAR retains its prognostic value even at low glomerular filtration rates (37), indicating that it is not just a marker of kidney function.

Structure of suPAR

uPAR and suPAR share the same overall structure, aside from the glycosyl phosphatidylinositol (GPI) anchor that tethers uPAR to the cell membrane. Both have three homologous domains, D1–D3, connected by a linker region between D1 and D2 (**Figure 2**). uPAR has cleavage sites for several proteases in the linker region (chymotrypsin, elastase, matrix metalloproteases, cathepsin G, plasmin, uPA) and the GPI anchor (phospholipase C and D, cathepsin G, plasmin), which upon cleavage can result in three suPAR isoforms (suPAR_{I-III} [full-length isoform], suPAR_I, suPAR_{II-III}) (21), **Figure 2**. The molecular weight of suPAR varies between 24–66 kDa due to variations in posttranslational glycosylation (21, 27). Additional isoforms generated by alternative splicing have been described on the RNA level, but whether these are transcribed and their possible roles remain unclear (38).

10 PROPERTIES AND CHARACTERISTICS INDICATING THAT suPAR IS A MARKER OF SCI

1. Expression and Release of suPAR Is Upregulated by Increased Immune Activation

uPAR is mainly expressed on the cell membrane of immune cells, such as monocytes, macrophages, neutrophils, and activated T-lymphocytes, but also on endothelial cells, fibroblasts, vascular smooth muscle cells, megakaryocytes, keratinocytes, and some cancer cells (21). Expression of uPAR is normally low, but expression is increased during activation and differentiation of leukocytes, extracellular matrix remodeling, wound healing, stress, injury, inflammation, and in tumor cells and tumor-associated stromal cells of many cancers (20). Specifically, stimulation of innate immune receptors, e.g., toll-like receptors (TLRs) or cytokine receptors, on immunologically active cells induces expression of the gene encoding uPAR, *PLAUR*, *via* activation of transcription factors, including nuclear factor

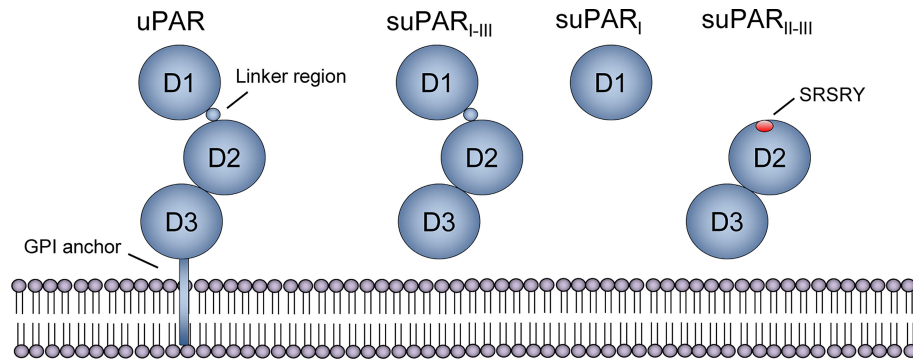


FIGURE 2 | Structure of uPAR and suPAR isoforms. Soluble urokinase plasminogen activator receptor (suPAR) is the soluble form of the membrane-bound receptor uPAR, which is tethered to the membrane by a glycosyl phosphatidylinositol (GPI) anchor. The protein consists of three domains, D1-D3, that are connected with a linker region between D1 and D2D3. Several cleavage sites exist, both in the linker region and the GPI anchor, and proteolytic cleavage generates three suPAR isoforms: full-length suPAR_{I-III}, suPAR_I, and suPAR_{II-III}. Cleavage of uPAR/suPAR in the linker region exposes an SRSRY sequence, which is involved in chemotaxis. Reprinted from Rasmussen, LJH (2018) (19) with permission.

kappa-light-chain-enhancer of activated B cells (NF- κ B) and activator protein 1 (AP1) (21), **Figure 1**. The uPAR mRNA is either degraded or stabilized for translation. After translation, uPAR localizes to the cell surface where it is bound to the cell membrane by the GPI anchor. At the cell surface, uPAR can be cleaved by its ligand uPA or other proteases, as mentioned above, thereby releasing suPAR to the bloodstream or other body fluids.

The specific inflammatory mediators shown to increase the expression of uPAR along with release of suPAR *in vitro* and *in vivo* include lipopolysaccharide (LPS), which increases the mRNA expression of uPAR *in vitro* (39, 40) and stimulates the release of suPAR (41). Injection of LPS in healthy human subjects has also been shown to increase the expression of uPAR on circulating monocytes and increase the blood levels of suPAR (42, 43). IL-8, TNF α , granulocyte-colony stimulating factor (G-CSF), and *N*-formyl-met-leu-phe (fMLP) have been shown to stimulate human peripheral blood mononuclear cells and neutrophils to rapidly increase surface expression of uPAR and induce release of suPAR (44, 45). Similarly, suPAR release from endothelial cells and vascular smooth muscle cells increases in response to IL-1 β *in vitro* (46). In contrast, co-administration of IL-6 abolishes LPS-induced suPAR release (43).

In summary, increased immune activation and stimulation by inflammatory mediators induce the gene expression of uPAR and release of suPAR, *via* major inflammatory transcriptional pathways regulated by NF- κ B and AP1, and increase the blood concentration of suPAR. This suggests that during states of inflammation, the expression of uPAR and suPAR will be increased.

2. uPAR and suPAR Exert Pro-Inflammatory Functions

Cell migration is important for inflammation and immune activation, and uPAR facilitates migration of immune cells through tissues, **Figure 1**. uPAR localizes its ligand uPA to the

cell surface of immune cells, where active uPA cleaves plasminogen to generate the active protease plasmin (20). Plasmin, in turn, activates matrix metalloproteases (47) and cleaves a range of extracellular matrix components, degrades fibrin, and activates the classical complement pathway (generating the anaphylatoxins C3a, C4a, and C5a, and the opsonin C3b) (48). This promotes extracellular matrix degradation, activation of sequestered growth factors (20, 49, 50), cell migration and invasion, fibrinolysis, vasodilation, increased vascular permeability, opsonization, and phagocytosis of foreign pathogens.

Moreover, binding of uPA to uPAR facilitates non-proteolytic functions involved in cell migration. The interaction promotes clustering of uPAR in plasma membrane lipid rafts in the leading edge of migrating cells. It also increases binding of uPAR to the extracellular matrix protein vitronectin and to integrins ($\alpha_3\beta_1$, $\alpha_5\beta_1$, $\alpha_M\beta_2$, $\alpha_V\beta_3$) (20) and their extracellular matrix ligands (e.g., laminins, fibronectin, collagens, vitronectin). This interaction activates intracellular signaling pathways that promote cell adhesion, migration, invasion, proliferation, survival, and immune activity (20), **Figure 1**. These functions are in play during recruitment of monocytes to inflamed tissue, where complexes of uPAR and $\alpha_M\beta_2$ integrin/Mac-1 expressed in leukocytes interact with intracellular Src kinases upon binding to vitronectin or fibrinogen, thereby regulating adhesion and cell migration of mononuclear cells (51). Thus, uPAR, uPA, and β_2 integrin provide the adhesion/degradation interactions between immune cells and endothelial cells or extracellular matrix, required for leukocytes to invade inflamed tissue in response to a chemotactic signal (48). Additional mechanisms by which uPAR regulates inflammatory processes have been suggested (**Figure 1**). These include co-localization of uPAR with cytokeratin-1 (CK1) and globular C1q receptor (gC1qR) on the surface of endothelial cells, which promotes release of the vasodilator bradykinin. Another mechanism is the simultaneous stimulation of uPAR, β_2 integrin, and gC1qR by cleaved high molecular weight kininogen, which induces release of cytokines (IL-1 β , IL-6,

TNF α) and chemokines (IL-8, monocyte chemoattractant protein-1 [MCP-1]) from blood mononuclear cells (52). All these processes contribute to sustaining the inflammatory response and to the cardinal signs of inflammation: swelling (tumor), redness (rubor), heat (calor), pain (dolor), and loss of function.

Although research into the active functions of suPAR has been limited, a number of immunological roles have been suggested. First, full-length suPAR_{I-III} is able to bind vitronectin (53) (**Figure 3**), to form a uPA-suPAR-vitronectin complex, which may allow vitronectin-directed activation of uPA at cellular surfaces or extracellular matrix sites (54). Second, suPAR_{II-III} may directly exert multiple pro-inflammatory functions by exposing an N-terminal SRSRY amino acid sequence (**Figure 2**, **Figure 3**). This SRSRY sequence acts as a chemotactic agent by interacting with the G protein-coupled receptor FPR-like receptor 1 (FPRL1) expressed on immune cells, including monocytes, lymphocytes, and neutrophils (44, 55, 56), and suPAR has also been shown to elicit cancer cell migration *via* this sequence *in vitro* (57). The SRSRY sequence is also involved in chemokine cross-regulation, preventing cell migration mediated by the chemokines MCP-1, CCL5, and fMLP (58). Additionally, the exposed SRSRY sequence stimulates angiogenesis with endothelial sprouting and tube formation, independent of uPA and vascular endothelial growth factor (59). Third, both suPAR and uPAR may impair phagocytic clearance of apoptotic neutrophils and other immune cells (**Figure 3**). This lack of neutrophil efferocytosis might contribute to sustaining the inflammatory response (60).

suPAR has also been ascribed some negative regulatory functions. The cleavage of uPAR into suPAR appears to abrogate uPA-mediated plasminogen activation, integrin-mediated intracellular signaling, and cellular migration (20). Full-length suPAR_{I-III} retains its ability to bind uPA through the D1 domain, and thereby acts as an uPA-scavenger (61), **Figure 3**. suPAR_{II-III} and suPAR_I are not able to bind and activate uPA or vitronectin, and cleavage of uPAR into these suPAR isoforms may

comprise a form of negative regulation of plasminogen activator activity without affecting serum uPA levels (62).

Unlike stimulation with inflammatory cytokines, stimulation of whole blood with high concentrations of suPAR for up to 24 hours has minimal effect on the expression of inflammatory cytokines (IP-10, IL-6, IL-10, and TNF α) (63). This could explain why suPAR is allowed to circulate freely without immediately being cleared, in contrast to most inflammatory biomarkers that often exert strong local-acting effects on immune- and non-immune cells.

Finally, uPAR has been identified as a universal marker of senescent cells, and suPAR release from senescent cells is part of the senescence-associated secretory phenotype (SASP) (64, 65).

In summary, uPAR plays multiple important roles in the inflammatory response, including cell migration, invasion, proliferation, vasodilation, phagocytosis, as well as release of cytokines and chemokines. The full functions of suPAR remain unclear; there are indications that suPAR may mediate chemotaxis of immune cells, promote angiogenesis, and prevent neutrophil efferocytosis.

3. suPAR Is Associated With the Amount of Circulating Immune Cells

Many of the immune system's functions are maintained by circulating immune cells. Inflammation is an essential mechanism of the innate immune system and part of the first line of defense against insults and infections.

suPAR is positively correlated with total white blood cell count (66, 67), and correlation analyses indicate that blood suPAR levels are associated with cells of both the innate and the adaptive immune response, **Appendix I**. Specifically, suPAR has been found to be correlated with innate immune cells including neutrophil count, monocyte count, and eosinophil count (66, 68, 69). For the adaptive immune system, suPAR has been correlated with lymphocyte count (69); however, other studies do not find a correlation between suPAR and lymphocyte

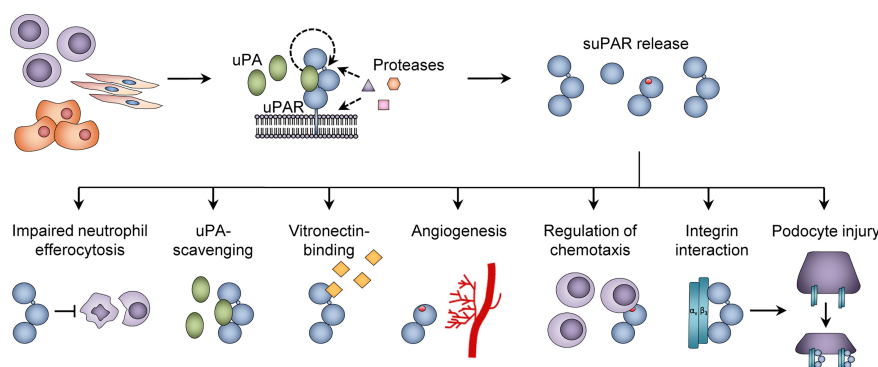


FIGURE 3 | Functions of suPAR. The urokinase plasminogen activator receptor (uPAR) is expressed on the surface of immune cells, endothelial cells, and vascular smooth muscle cells, and proteolytic cleavage in the linker region or glycosyl phosphatidylinositol (GPI) anchor of uPAR generates soluble uPAR (suPAR). Various functions of suPAR have been proposed, including inhibition of neutrophil efferocytosis; binding of urokinase plasminogen activator (uPA) and vitronectin; stimulation of angiogenesis *via* endothelial sprouting and tube formation; promoting chemotaxis; and interactions with β_3 integrin, which is suggested to cause podocyte injury in the glomeruli. Adapted from Rasmussen, LJH (2018) (19) with permission.

count (66, 70, 71), which could suggest that suPAR is mainly associated with innate rather than adaptive immune cells. In line with this, uPAR expression is largely confined to pro-inflammatory monocyte subsets during the inflammatory response of acute liver failure (72) and to monocytes, neutrophils, and macrophages, but not lymphocytes, of patients with cirrhosis (41). LPS stimulation promoted the release of suPAR from monocytes, but not lymphocytes (42, 72).

In mice, suPAR has been found to originate from the expansion of uPAR-expressing bone marrow-derived immature myeloid cells (73). Myeloid expansion occurs under many clinical conditions, and, during inflammation, pro-inflammatory mediators—including cytokines (IL-1, TNF α , interferons [IFNs]), PAMPs, and DAMPs—regulate hematopoiesis and increase the myeloid output of bone marrow cells (74). A sustained overproduction of myeloid cells during SCI could result in increased suPAR levels in various conditions, e.g., chronic infections, autoimmune disease, and chronic inflammatory diseases, such as cardiovascular disease or type 2 diabetes.

In summary, suPAR is associated with the amount of circulating immune cells, mainly neutrophils and monocytes, and has been found to originate from expansion of myeloid lineage cells. This indicates that suPAR is associated with immune activity and could suggest that suPAR, like inflammation, is associated with innate rather than adaptive immune responses.

4. Blood suPAR Levels Correlate With the Levels of Established Inflammatory Biomarkers

The inflammatory response—acute as well as chronic—is mediated by numerous different cell types, inflammatory mediators (e.g., cytokines or chemokines), and their receptors.

Due to the lack of an operational definition of SCI, we are unable to assess suPAR's internal consistency with other measures of SCI. However, suPAR is positively correlated with a multitude of biomarkers of inflammation. Plasma and serum suPAR levels have been found to be positively correlated with traditional markers of inflammation (see **Appendix I**), including CRP (23, 75–77), erythrocyte sedimentation rate (22, 78, 79), fibrinogen (80, 81), procalcitonin (23, 82), white blood cell count (68, 81), neutrophils (66, 68), monocytes (66, 69), and a number of cytokines and chemokines, e.g., IL-1 β (22), IL-6 (83), IL-8 (CXCL8) (68), IL-10 (22, 68), IL-18 (84), MCP-1 (CCL2) (68), and TNF α (23, 24, 68).

But even though suPAR is positively correlated with established markers of inflammation, the correlations with many of these are weak (**Appendix I**). For example, reported correlations between suPAR and CRP range between 0.15–0.30 in population-based studies (69, 75, 76, 85), and between 0.15–0.53 ($P < 0.001$) in clinical studies (71, 86). Similarly, the correlations with IL-1 β (22), IL-10 (22, 68), erythrocyte sedimentation rate (22, 78), and white blood cell count (68, 81) were weak. The correlation of suPAR with other inflammatory markers generally appears to be stronger in patients with severe or exacerbated disease (68, 87, 88), maybe driven by a larger

increase in suPAR levels related to presence of organ damage or dysfunction. When comparing suPAR to CRP, CRP is closer correlated with many of these inflammation markers, including erythrocyte sedimentation rate (89), fibrinogen, and IL-6 (69, 83). This difference suggests that CRP and suPAR reflect different aspects of inflammation, as previously described (90), and are not two measures of the same thing.

In summary, suPAR being positively correlated with established markers of (acute) inflammation, supports the role of suPAR as a marker of inflammation itself, although suPAR's weaker correlation to acute phase proteins and cytokines compared to CRP suggests that suPAR may describe another type of inflammation.

5. suPAR Is Minimally Affected by Acute Changes and Short-Term Influences

Some of the most important pro-inflammatory cytokines have limited value as clinical biomarkers of SCI due to their short half-life, circadian fluctuations (IL-1, IL-12, TNF α , IFN γ), and susceptibility to variations in dietary intake (IL-6, TNF α), physical activity (IL-1 receptor antagonist [IL-1ra], IL-6, IL-10), and sample handling (91, 92). Furthermore, some cytokines, like IL-1 β and TNF α , are even undetectable in healthy individuals with current commercially available assays (93, 94), or otherwise at or near the limits of accurate detection range, creating substantial variability and uncertainty in measured concentrations (95). These factors may obscure the detection and interpretation of clinically relevant changes in the concentrations of these inflammatory markers.

In contrast, suPAR is a very stable protein, which is subject to minimal circadian fluctuation (24, 27, 96, 97), it is readily quantifiable both in healthy (25) and sick individuals (98), and it maintains a steady sample concentration after repeated freezing/thawing cycles (27, 99). Thus, there are no requirements for special collection procedures or need for fasting blood samples. It also has low within-person variability and is stable in individuals with only small changes over time; it had an excellent intraclass correlation coefficient (0.91, 95% CI 0.88–0.93) over 4 months in healthy individuals (100) as well as a high intra-individual correlation in samples taken 5–7 years apart in population-based studies, with $r = 0.55$ for suPAR measured at baseline and 5 years later in the Danish Inter99 Study (101), and $r = 0.58$ for suPAR measured at age 38 and age 45 in the New Zealand Dunedin Study (102), significantly higher than log-transformed CRP levels ($r = 0.48$) and log-transformed IL-6 levels ($r = 0.45$) in the same study (103) (untransformed CRP levels were correlated at $r = 0.26$ and untransformed IL-6 levels at $r = 0.39$, unpublished data).

suPAR and CRP have previously been suggested to reflect different aspects of inflammation, with CRP being a marker of acute infection and metabolic inflammation, and suPAR being a marker of cellular inflammation and subclinical organ damage (90). Corroborating this theory, suPAR has been found to be differently related to cardiometabolic risk factors, for example, it is only weakly correlated with body mass index (BMI) (26, 69, 104), while CRP and BMI were strongly correlated (69).

Furthermore, unlike CRP, suPAR was not correlated with body temperature, week of menstrual cycle, and use of anti-inflammatory medication (69).

Several studies have shown slower, delayed suPAR level increases in response to acute inflammatory stimuli compared to traditional inflammation markers. Knee surgery induced a significant increase in IL-6 and IL-10 between baseline and 1 day after surgery, while suPAR was unchanged the first day after surgery but had increased significantly 4 weeks after surgery (63). Similarly, patients admitted for myocardial infarction had continued rising CRP levels throughout the first 24 hours, while suPAR levels remained stable and unaltered (96), suggesting that the contribution of an acute event to suPAR levels is minimal. Indeed, among acute trauma patients, suPAR measured shortly after trauma was not associated with the severity of the trauma, but was higher in those who later died compared with those who survived (105). Thus, an acute event might not immediately affect the suPAR level substantially, but the basal suPAR level at the time of the event reflects the level of SCI which is associated with the outcome (106). One can therefore speculate that patients with higher SCI—as indicated by elevated suPAR—at the time of traumatic injury or surgery have lower capacity to withstand the immunological challenges and complications caused by the trauma or surgical procedure. This might also be the case for many other conditions, that is, patients with a high basal level of SCI have impaired abilities to manage and tolerate disease.

In summary, the temporal and kinetic stability of suPAR, in addition to the correlations with many of the established inflammatory biomarkers (**Appendix I**), suggest that suPAR reflects a more chronic aspect of inflammation.

6. Like Systemic Chronic Inflammation, suPAR Is Non-Specifically Associated With Multiple Diseases

SCI is not a disease-specific process confined to one single line of pathology. Rather, SCI is associated with diverse diseases affecting different organ systems, such as metabolic syndrome, type 2 diabetes, liver disease, cardiovascular disease, cancers, depression, autoimmune diseases, neurodegenerative diseases, sarcopenia, osteoporosis, and immunosenescence (2).

Likewise, increased suPAR levels are also associated with a wide range of diseases and disorders—non-communicable and infectious diseases alike, **Figure 4** and **Appendix II**. So far suPAR has been shown elevated in cardiovascular disease, including stroke, ischemic heart disease, venous thromboembolism, and incident atrial fibrillation (96, 107, 108); type 1 diabetes and diabetic complications (109, 110); incident and manifest type 2 diabetes (76, 111, 112); different types of cancer (see **Figure 4** and **Appendix II**) (15, 113–130); rheumatic disease, including rheumatoid arthritis (78, 79, 131) and systemic lupus erythematosus (22); asthma and chronic obstructive pulmonary disease (132); acute and chronic pancreatitis (115, 133); chronic liver disease, including non-alcoholic fatty liver disease and

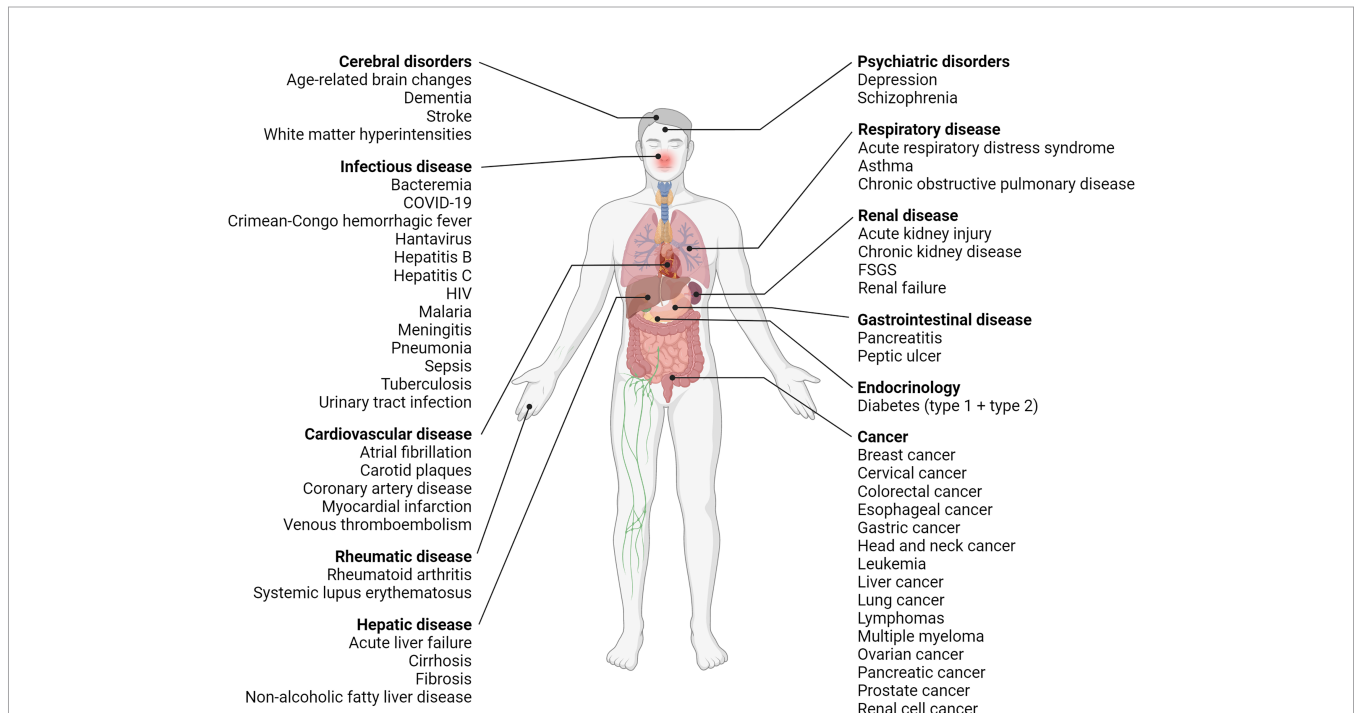


FIGURE 4 | Overview of diseases with elevated suPAR levels. Clinical studies have shown that suPAR levels are elevated and associated with disease severity and prognosis in many diseases, including diseases of the brain, liver, kidneys, and respiratory system, cardiovascular disease, diabetes (type 1 and type 2), cancer as well as infectious, rheumatic, and psychiatric disorders. COVID-19, coronavirus disease 2019; FSGS, focal segmental glomerulosclerosis; HIV, human immunodeficiency virus; suPAR, soluble urokinase plasminogen activator receptor. Created with BioRender.com.

cirrhosis (68, 134, 135); incident acute kidney injury (136, 137) and chronic kidney disease (CKD) (138, 139); and dementia (77). As previously mentioned, suPAR is also elevated in infectious diseases caused by viruses, bacteria, and parasites, e.g., coronavirus disease 2019 (COVID-19) (37, 140), hepatitis B and C (135, 141), human immunodeficiency virus (HIV) (142), bacteremia (143, 144), meningitis (17, 145), urinary tract infection (88), pneumonia (71, 146), sepsis (23, 147), tuberculosis (148), malaria (149, 150), hantavirus (151), and Crimean-Congo hemorrhagic fever (87). suPAR is also elevated in pediatric disorders, including infections and CKD (71, 88, 139). Furthermore, suPAR may be associated with psychiatric disorders, including depression and schizophrenia (25, 152–154).

In summary, suPAR offers little diagnostic value, as it, like SCI, is elevated by many different diseases across multiple organ systems. It does however offer prognostic value.

7. suPAR and Systemic Chronic Inflammation Predict Morbidity and Mortality

SCI is predictive of disease development, progression, and mortality (2). Individual inflammation markers, composite scores including IL-6, CRP, TNF α , albumin, or neutrophil count, and high-dimensional inflammation measures have been found to be associated with morbidity and mortality (9, 155–157).

Elevated suPAR is associated with disease development, progression, severity, and risk of adverse outcomes. Thus, within and across various patient groups (**Figure 4, Appendix II**), high suPAR is associated with more advanced disease, exacerbations, and complications as well as presence of organ damage, comorbidities, and increased risk of adverse events and mortality (77, 158, 159). In critically ill patients, stably elevated or increasing suPAR levels were observed for non-survivors from the time of admission, while that of survivors remained stable or decreased until discharge (82, 146).

A suPAR value may therefore reflect the current health status of a patient, possibly by reporting the level of SCI and organ damage, and could contribute with valuable prognostic information in a clinical setting.

8. suPAR and Systemic Chronic Inflammation Share the Same Risk Factors

A number of risk factors are associated with increased levels of SCI, as recently reviewed by Furman et al. (2). These include higher age, chronic infections, tobacco smoking, physical inactivity, unhealthy diet, obesity, social isolation, psychological stress, and exposure to environmental or industrial toxicants. Moreover, early development and childhood circumstances have been shown to promote SCI in adulthood (2). The same risk factors have been shown to be associated with increased suPAR.

In the general population, suPAR increases with age (25, 76, 101, 102), not only with a person's chronological age, but also with indicators of accelerated aging, such as faster rate of decline across multiple organ systems, older facial age, as well as physical and cognitive decline (102).

Chronic infections caused by viruses and bacteria are associated with elevated suPAR, which has been observed for hepatitis B and C (135, 141), HIV (142), and tuberculosis (148).

With regard to lifestyle, smoking is likely the most devastating cause of poor health, and the inhalation of smoke and toxicants are thought to activate the immune system through DAMPs. Studies in the general population have consistently shown that smokers have increased suPAR levels compared to non-smokers (26, 76, 102, 160), with smokers having around 1 ng/mL higher suPAR (101, 102). The effects of smoking on suPAR appear to be reversible to some degree such that ex-smokers have suPAR levels similar to that of never-smokers (101, 102). In a study of smoking cessation, daily smokers who were randomized to smoking cessation exhibited decreased suPAR levels that were no longer significantly different from that of never-smokers (66). In contrast, smokers and non-smokers did not differ in CRP levels, and smoking cessation had no effect on CRP levels (66).

Individuals with a sedentary lifestyle, unhealthy diet, or morbid obesity also have higher suPAR levels (26, 102, 161). The level of low-density lipoprotein (LDL) cholesterol (a major risk factor of cardiovascular disease due to buildups in the arteries) is positively correlated with suPAR, while high-density lipoprotein (HDL) cholesterol (which helps eliminate LDL cholesterol) is negatively correlated with suPAR (26, 76, 162). Exposure to toxicants like cadmium is also associated with increased suPAR (104).

Experiencing stressful life events—including relationship breakups, job loss, serious illnesses or accidents of self or close relatives, financial problems, being homeless or in jail, being physically or sexually assaulted, death of a friend or family member, and living through disasters—are associated with higher suPAR in midlife (103). In contrast, no associations were observed between stressful life events and CRP or IL-6.

Adult suPAR levels may have origins already in childhood. In two longitudinal birth cohort-studies, we showed that exposure to social and psychological risk factors during childhood—including adverse childhood experiences (such as abuse, neglect, and victimization), low childhood IQ, or poor childhood self-control—was associated with elevated suPAR levels later in life after controlling for adult BMI and smoking, but not with CRP or IL-6 (69, 163). In line with this, findings from previous research studying associations between childhood adversity and adult CRP, IL-6, and TNF α are inconsistent with several studies reporting non-significant associations (164).

In summary, suPAR is elevated in presence of well-established risk factors of SCI, including older chronological age, accelerated biological aging, chronic infections, smoking, physical inactivity, unhealthy diet, obesity, toxicants, and psychosocial stress-exposure during childhood and adulthood.

9. suPAR Is Associated With Risk Factors and Outcomes of Inflammation Above and Beyond Other Inflammatory Biomarkers

A new biomarker of SCI should be strongly and independently associated with outcomes of SCI above and beyond established

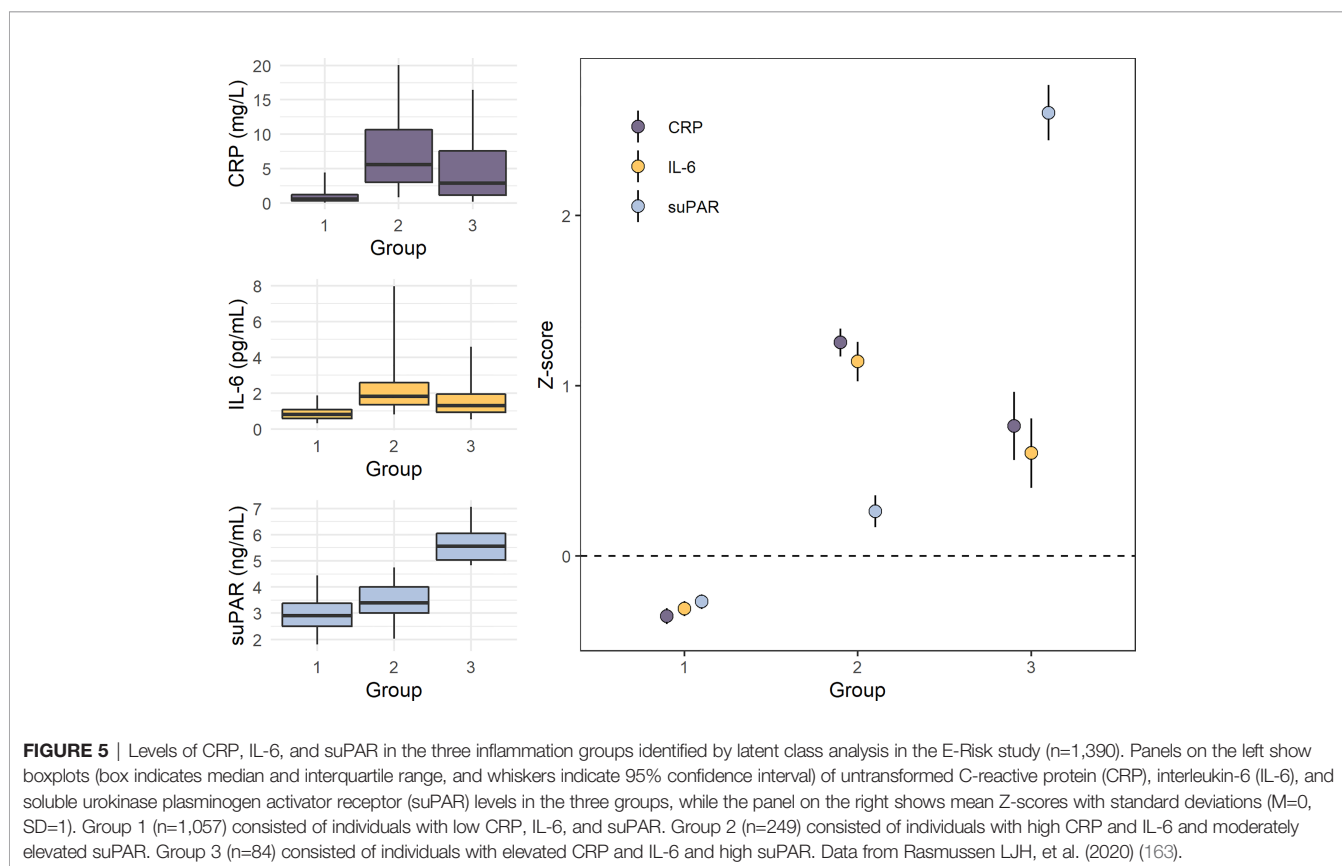
inflammatory biomarkers and other widely available risk scoring systems.

Studies in various settings have shown that suPAR is indeed associated with risk factors as well as outcomes of SCI independently of common inflammation markers.

Elevated suPAR is associated with early-life risk factors and stressful experiences in childhood above and beyond CRP and IL-6 (69, 163). In our Environmental Risk (E-Risk) Study, we showed that children exposed to multiple forms of stress and violence during childhood and adolescence had elevated suPAR levels, but not CRP or IL-6, at age 18, even after adjustment for sex, BMI, and smoking (163). Moreover, participants exposed to cumulative adverse experiences across childhood and adolescence (domestic violence or multiple types of violence in childhood and adolescence) had elevated suPAR, but not CRP or IL-6. This underlines that measuring suPAR can be used to examine the health implications of stressful experiences in childhood beyond the established inflammation markers CRP and IL-6. We found that adverse experiences were prominent in the group of participants with low CRP and low IL-6 but high suPAR—a group of individuals who would have inadvertently been assigned to the low inflammation group if suPAR had not been assayed. Interestingly, we observed the strongest association between stress exposure and inflammation when combining the three biomarkers and thereby utilizing the potentially different inflammatory states they reflect.

The prognostic value of suPAR for various patient outcomes remains significant when controlling for CRP, including incident cancer, readmission, or mortality (77, 89). Of note, in patients with low CRP levels (<10 mg/L) suPAR still remained associated with mortality, further substantiating that suPAR does add prognostic value to CRP (77). In support of this, suPAR predicted all-cause- and cardiovascular disease mortality independent of CRP and IL-6 in a South African population (83).

Another remarkable finding from the E-Risk Study relates to the multidimensionality of inflammation and the utility of suPAR as a biomarker for indexing the chronic dimension of inflammation. We assessed the dimensionality of the three inflammatory biomarkers CRP, IL-6, and suPAR using latent class analysis in this cohort of healthy young adults, and we identified three latent groups of inflammation among participants in the cohort (**Figure 5**). Group 1 consisted of individuals with low CRP, low IL-6, and low suPAR. Group 2 consisted of individuals with high CRP, high IL-6, and moderately elevated suPAR. Lastly, Group 3 consisted of individuals with high suPAR, and moderately elevated CRP and IL-6 (163). We hypothesize that these groups represent three dimensions of systemic inflammation (low, acute, and chronic), identifying individuals with different types and levels of inflammation. Thus, Group 1 would represent low inflammation, Group 2 acute inflammation, and Group 3 SCI, with the key being high CRP/IL-6 as an indication of acute inflammation and high suPAR as an indication of SCI. For these 18-year old participants, members of



Group 3 (high suPAR and moderately elevated CRP and IL-6) had been exposed to more adverse childhood experiences as well as victimization in childhood and adolescence, compared with those in Group 1 and Group 2 (163). These results need to be validated and tested in other and larger cohorts. Recently, a study of 574 adolescents reported similar findings, showing that persistent parent-child separation was more frequently observed among individuals with high suPAR and low CRP or with high suPAR and high CRP, compared to individuals with low suPAR and low or high CRP (165).

Adding suPAR level measurements to existing blood test panels would allow further stratification of inflammation type and level, and add valuable prognostic information above and beyond that of the current inflammatory biomarkers.

10. The suPAR Level Can be Reduced by Anti-Inflammatory Interventions and Treatment of Disease

Anti-inflammatory interventions to lower SCI can aim at targeting risk factors of inflammation, the inflammatory pathway, or diseases that are perpetuating chronic inflammation.

Lifestyle covers multiple risk factors for SCI, and altering lifestyle induces changes in suPAR levels. For example, change in diet or exercise has a positive impact by lowering the suPAR level (102, 166), and as mentioned above, smoking cessation results in a suPAR decrease (66), where former smokers can achieve comparable suPAR levels to that of non-smokers (26, 102). The resultant lowering of suPAR is also associated with lowering the risk of mortality (101), pointing to elevated suPAR as a modifiable risk factor.

Targeting the inflammatory pathway with anti-inflammatory medication has also been shown to lower suPAR levels; for example, suPAR levels were significantly lower after 14 days of corticosteroid treatment for acute exacerbation of COPD (80), after 2-4 weeks of glucocorticoid treatment in pediatric inflammatory bowel disease (167), and after 3-6 months of prednisolone treatment in HIV (168).

Other types of therapy, not aimed at the inflammatory response, have also been associated with reduced suPAR levels in various diseases. These include: long-term treatment with beta-blockers of patients with carotid stenosis (169); lipid-lowering treatment with statins of patients with aortic stenosis (170); surgical tumor resection (120, 123) or chemotherapy in treatment of cancer (15); treatment of acute exacerbation of COPD with bronchodilators, supplemental oxygen, or antibiotics (80, 81); treatment of community-acquired pneumonia with antibiotics (171); highly active antiretroviral therapy in HIV-1-infected patients (172); anti-malaria treatment in pediatric malaria (149); or even nutritional support during hospitalization with a high-protein diet for patients with systemic inflammatory response syndrome (173).

Thus, an increased suPAR level is modifiable and reversible, both by means of lifestyle changes or therapy targeting either disease or inflammatory state. Together, this makes suPAR valuable in assessing the effect of potential interventions aimed at reducing SCI as well as in measuring the risk imposed by an individual's level of SCI.

DISCUSSION

In summary, this body of research provides strong support for the hypothesis that suPAR is a biomarker of SCI. suPAR is upregulated and released to the bloodstream from innate immune cells in response to increased immune activation. It serves inflammatory functions in itself and is positively, although often weakly, correlated with established inflammatory biomarkers, including CRP. suPAR has high stability as a biochemical analyte (temporal stability and low method-specific variance) and is minimally affected by short-term influences, circadian rhythm, and minor acute events. It shares risk factors and outcomes with SCI independently of other inflammatory biomarkers. Finally, suPAR possesses features that are important for a clinical prognostic biomarker (174): it reflects ongoing pathogenic processes with the ability to predict incident (76, 107, 158) or prevalent disease (77, 81, 116, 117, 121, 139), extent or severity of disease (68, 123, 125, 135), and risk of recurrence (96, 175) or fatality (76, 98, 147, 176); the blood concentration of suPAR is significantly altered in response to anti-inflammatory interventions, disease, or remission (80, 81, 149, 172); and it is readily quantifiable both in healthy (25) and sick individuals by means of safe and easy testing. Aside from a potential to index and assess a person's level of SCI, suPAR can be a useful biomarker in the clinic. The stable kinetics of suPAR limits its value in monitoring immediate clinical responses to treatment of acute disease, but suPAR offers clinical value as a prognostic tool for clinical endpoints, due to its strong association with disease severity.

Together, this suggests that suPAR could be the best single marker of SCI, organ damage, and physiologic reserve, contributing added information about the systemic chronic inflammatory state to that of the commonly used (primarily acute) inflammatory markers. suPAR is not specific to any one disease in particular, but is elevated and has a strong prognostic value across many different disorders. As such, suPAR has limited value as a diagnostic tool. The broad associations across diseases suggest that blood suPAR levels reflect a shared feature of disease, which could very well be SCI. The recent studies that link elevated adult suPAR levels with psychosocial stressors in childhood and adolescence (69, 163) as well as physical and cognitive decline and accelerated biological aging (102) further contribute to our understanding of suPAR as a marker of chronic influences and mark the transition from viewing suPAR as a clinical biomarker associated with illness, to being a broader marker of underlying immune activity associated with early development, psychosocial stress, and accelerated aging, before the onset of disease.

Despite the consensus that SCI is health-damaging and constitutes a major risk factor for many diseases, the lack of good stable biomarkers reflecting SCI has so far left this an undiagnosable condition. This greatly limits not only research into chronic inflammation, but also poses a serious problem for treating patients, as emerging disease processes may go unnoticed, leading to development of manifest disease and detrimental health complications. Using suPAR as a measure of SCI has the potential of improving the estimation of a person's

underlying inflammatory burden and provide accurate assessments of interventions aimed at reducing inflammation, creating a valuable window of opportunity for treatment and prevention.

A Marker of Diminished Immunological Capacity?

Based on the research and clinical findings on suPAR in disease reviewed here, the evidence points to suPAR as a quantitative indicator of a person's level of SCI. In other words, the suPAR level mirrors the current level of underlying immune activity and the individual's health state.

As described earlier, the suPAR level measured immediately after an acute trauma is not associated with the severity of trauma while still associated with survival during follow-up (105), suggesting that suPAR is not rapidly released as part of the acute response, which could, in turn, suggest that suPAR does not have an active, functional role in the immune response to acute events. Rather, suPAR may be an indicator of the immunological capacity of a person. That is, the level of SCI, as reflected by the suPAR level, indicates how well a person will tolerate and handle an immune challenge, with individuals with persistently elevated SCI having a lower capacity to withstand and manage injuries, trauma, or disease.

The strong prognostic value of suPAR in infectious diseases, with high suPAR being associated with a higher risk of adverse outcomes, could indicate that the higher the degree of SCI, the less efficient the immune system is at protecting the individual. In patients with COVID-19, an early elevation of suPAR is an indicator of poor prognosis, including increased risk of developing respiratory failure (140), acute kidney injury, and mortality (37). As a result, suPAR has been used to stratify COVID-19 patients in the Emergency Department, where patients with suPAR levels above 6 ng/mL were treated with the IL-1 receptor antagonist anakinra, which significantly reduced time to recovery and lowered mortality compared to the standard of care (177, 178). This raises the question of whether patients with elevated SCI (or suPAR) in general, and not only COVID-19 patients, will benefit from anti-inflammatory treatment through reduced morbidity and mortality.

Immunosenescence is a multifaceted decline in immune effectiveness, resulting in increased susceptibility to infections and age-related inflammatory diseases, diminished vaccine responses, and lower capacity to mediate anti-cancer responses and control tissue homeostasis and repair (179–182). Immunosenescence is characterized by age-related low-grade SCI (*inflammaging*), diminished response to new antigens, and the accumulation of memory T and B cells with a decrease in naïve cells (180). Given that high suPAR is associated with a lower effectiveness of the immune system, as illustrated by the elevated risk of disease progression and adverse events, as well as its associations with inflammation and multiple indicators of accelerated aging and functional decline, suPAR may be a useful biomarker of SCI that can be used to quantify the level of inflammaging and immunosenescence. In support of this, we recently reported elevated suPAR levels in a patient population characterized by accelerated aging and multiple signs of

immunosenescence, including reduced capacity to respond to immune stimulation, defects in NF- κ B signaling, and higher levels of inflammatory biomarkers (CRP, IL-6, IL-18, TNF α , growth differentiation factor 15 [GDF15]) and NLRP3 inflammasome expression compared to age-matched and young healthy controls (183).

Difference From CRP

The most widely used biomarker of inflammation is the acute-phase reactant CRP. CRP and suPAR differ in their respective susceptibility to acute and chronic stressors, temporal specificity (timing of release) and response kinetics (speed, amplitude, and stability of release), and to the type of pathologies that they are most strongly related to.

CRP shares numerous of the 10 characteristics we propose for suPAR. Expression of CRP is induced by IL-6 and other cytokines (IL-1 β , TNF) *via* NF- κ B and other transcription factors as part of the acute-phase response or during inflammatory conditions and infections (184). CRP exerts a functional role in the inflammatory response, through activation of the classical complement pathway, induction of phagocytosis, apoptosis, and release of pro-inflammatory cytokines, as well as chemotaxis and recruitment of leukocytes to areas of inflammation (184, 185). CRP is primarily synthesized by hepatocytes, but it can also be produced by other cell types, including leukocytes, endothelial cells, smooth muscle cells, and adipocytes (185). The blood concentration of CRP correlates with the concentration of various cytokines, including IL-6 and TNF α . CRP is non-specifically associated with multiple diseases, and elevated CRP levels are associated with increased risk of incident disease and mortality (7). Many factors are associated with baseline CRP levels, including age, sex, lifestyle, blood pressure, and in particular, metabolic risk factors such as elevated blood lipids and obesity (185). Lifestyle interventions to reduce cardiovascular risk have been associated with lower CRP levels, and, in case of disease, treatment of the underlying pathology that is causing an acute-phase stimulus can reduce the CRP levels (7).

In contrast to IL-6, TNF α , and other cytokines (186), CRP is also not subject to diurnal variation (187) and does not respond to acute psychological stress challenges (188). CRP increases rapidly in response to acute stimuli, such as severe tissue damage, trauma, or acute infection. Some bacterial infections can dramatically increase CRP levels up to 1,000-fold in the span of 24–72 hours (189). When an inflammatory stimulus is terminated, the CRP level quickly decreases with a half-life of about 19 hours (189). As such, CRP is an excellent biomarker of bacterial infections and the acute inflammatory response. In contrast, the suPAR response to acute stimuli is much slower and the fold-change markedly smaller, e.g., around 1.3-fold increase in suPAR 7 days after diagnosis of ventilator-associated pneumonia and sepsis (146) or cardiac arrest (190). Compared to CRP, suPAR is an inferior diagnostic marker for discriminating between infections of bacterial vs. non-bacterial origins (191). Although suPAR has been found to be significantly elevated in critically ill patients, including patients with sepsis, compared to healthy controls, the ability of blood suPAR levels

to discriminate sepsis from non-sepsis patients was poor compared to that of CRP (23).

Moreover, elevated CRP levels are associated with increased risk of incident disease, such as diabetes and cardiovascular disease, and mortality (192, 193), and CRP has been recommended as an adjunct screening tool for cardiovascular risk prediction in the general population (7). However, there is significant short-term within-person variability in CRP levels in the general population (194, 195), with approximately one-third of persons with elevated CRP levels (≥ 10 mg/L) being reclassified after repeated testing 2.5 weeks later (194). The variation was particularly high at higher CRP values—the cases in which clinicians are most likely to intervene. This variability in CRP means that using a single CRP measure to index SCI may lead to substantial misclassification (194). Moreover, a common approach in research studies that are investigating risk factors of SCI is to systematically remove all observations with CRP > 10 mg/L to exclude participants with acute illness; however, this could introduce bias by also excluding individuals with actual SCI and high CRP (196).

For psychosocial stressors, several different types of adverse childhood experiences, including child maltreatment, bullying, and sexual abuse, have been associated with increased CRP levels. In addition, low socioeconomic status has been shown to be associated with higher CRP (197). However, findings are not consistent; several studies report non-significant associations, and in many cases, associations do not survive control for the confounding effects of BMI or smoking (164, 197). In a meta-analysis investigating associations between socioeconomic status and inflammation, only studies that did not control for BMI or smoking showed significant associations between CRP and socioeconomic status (197). Similarly, CRP and IL-6 did not show consistent associations with adult stressful life events, in contrast to suPAR (103). An explanation could be that these traditional biomarkers of inflammation may mix chronic and acute effects.

The variance in suPAR that can be ascribed to CRP is around 15–30% in general and healthy populations (**Appendix I**). As the two biomarkers appear to identify different classes of people at risk, using suPAR in combination with CRP can provide valuable information about an individual's state of health.

Passive Bystander or Active Disease Agent?

Whether suPAR plays an active role in disease development or is merely a passive bystander that reflects ongoing disease processes remains unresolved. A causal role of suPAR has been described in CKD, primarily in focal segmental glomerulosclerosis (FSGS).

Blood suPAR level is elevated in two thirds of patients with FSGS, and high blood suPAR concentrations induce renal injury in experimental models (175), and infusion of suPAR in uPAR-knockout mice induced proteinuria (175, 198). Moreover, the declining kidney function, which is associated with a high-risk genotype of the gene for apolipoprotein 1 (*APOLI*), is dependent on high plasma suPAR levels (199). This suPAR-induced renal injury is further dependent on suPAR's interaction with β_3

integrin (175), and the suggested pathological mechanism is a synergistic suPAR- and apolipoprotein 1-mediated activation of $\alpha_v\beta_3$ integrin on the podocyte membrane (**Figure 3**), causing renal injury through podocyte foot process effacement, cell detachment, and disruption of the glomerular barrier with resultant proteinuria (199). The pathological suPAR production was caused by expansion of uPAR-expressing immature myeloid cells, which lead to increased suPAR levels and proteinuria in mice (73). Moreover, in uPAR knock-out mice, uPAR expression in transplanted hematopoietic cells was necessary for suPAR production and development of proteinuria (73).

As myeloid expansion occurs under many conditions without necessarily afflicting renal damage, it appears to be the combination of suPAR and high-risk variants of the *APOLI* gene that triggers CKD, and not just the presence of suPAR alone. During inflammation, pro-inflammatory mediators regulate hematopoiesis and increase the myeloid output of bone marrow cells (74). As suPAR is produced from myeloid cells, this chronic overproduction of myeloid cells could be a potential source of increased suPAR, not only in the pathogenesis of CKD, but also in conditions of SCI related to aging or disease, e.g., chronic infections, autoimmune disease, and chronic inflammatory diseases, such as CVD or type 2 diabetes.

If suPAR plays a causal role in CKD, and possibly in the pathogenesis of other diseases, inhibition or removal of suPAR could have a stabilizing effect on disease. Interestingly, renal disease was stabilized or even abrogated when lowering circulating suPAR levels, either through removal of suPAR with plasmapheresis, or by interfering with the suPAR- β_3 integrin interaction using blocking antibodies or small molecule inhibitors (175). It would potentially have major clinical implications, if diseases with elevated suPAR could be treated *via* reduction of suPAR levels, and would suggest that suPAR was not just a passive by-product of uPAR signaling but could have an active role, at least in kidney disease. Experiments are ongoing to further document this causal role of suPAR, with some conflicting evidence (200). Whether increased suPAR levels in other diseases merely reflect the expression and activity of uPAR remain unknown.

As previously mentioned, suPAR is not dramatically upregulated in response to acute events and it is detectable in the blood even during states of normal homeostasis in healthy individuals, in contrast to most active inflammatory mediators. This could be speculated to indicate that suPAR might be less functionally active and therefore allowed to circulate freely without being rapidly cleared from the blood. However, a large buildup of suPAR in the blood over longer time could potentially create toxic concentrations that are inflicting the damage observed in kidney diseases like FSGS. As suPAR is removed from the blood by renal clearance, this mechanism could be particularly exacerbated in patients with poor kidney function.

Research Agenda

While the existing evidence points to suPAR as a potential biomarker of SCI, there are still several questions to answer. In

the following, we describe a research agenda with the purpose of improving the understanding of the link between suPAR and SCI as well as paving the way towards clinical implementation of suPAR.

First, studies should be designed to test the hypothesis that suPAR can actually be used to distinguish acute from chronic systemic inflammation. As previously described, we used latent class analysis in the E-Risk Study of CRP, IL-6, and suPAR, and identified three groups (**Figure 5**). Group 1 consisted of individuals with low CRP, low IL-6, and low suPAR. Group 2 consisted of individuals with high CRP, high IL-6, and moderately elevated suPAR. Group 3 consisted of individuals with high suPAR, and moderately elevated CRP and IL-6 (163). As mentioned, we think these results likely represent a method for determining a person's level and type of inflammation, with Group 1 representing low inflammation, Group 2 acute inflammation, and Group 3 SCI, with suPAR being the indicator that differentiates acute inflammation from SCI. The observation that Group 3 had a higher proportion of traumatic childhood experiences indicates that suPAR levels reflect the long-term health effect of chronic stress that is not sufficiently identified by the established inflammation markers CRP and IL-6. These results need to be tested and validated in other and larger prospective cohort studies, using descriptive statistics to characterize any differences between the three groups and testing associations with both risk factors and long-term outcomes of SCI, such as chronic diseases and early mortality. If Group 3 is in fact characterized by SCI, we would expect to find stronger associations for this group with factors related to SCI. Furthermore, it should be tested if the prognostic information carried by using the composite measure with all three biomarkers can be reasonably approximated using only suPAR. We are currently planning international multi-cohort studies with longitudinal data to replicate and test this model.

Second, mechanistic studies are needed to map the molecular biology behind and the pathways leading to increased suPAR. As research indicates that suPAR is associated with innate immune cells, the association between suPAR and innate immune mechanisms should be further explored to elucidate whether suPAR shares pathways with other known drivers of chronic inflammation, e.g., the inflammasome (201). Testing the hypothesis that high suPAR represents a measure of lower immunological capacity or immunosenescence could be done by comparing individuals with high vs low suPAR in regard to their baseline levels of immune cell subset composition (e.g., ratios of memory:naïve and CD8:CD4 T cells), antibody levels, chronic infection status, as well as ability to elicit an immune response upon stimulation of isolated immune cells as measured by fold-change in cytokine production or immune cell signaling (e.g., STAT and NF- κ B pathways). In addition, uPAR is induced during cellular senescence and released as suPAR as part of the SASP; the role of uPAR and suPAR in senescence of immune cells and the link to immunosenescence should be further explored.

Third, intervention studies aimed at lowering SCI or at preventing outcomes of SCI should use suPAR either as an

effect measure, or to identify the target group for the intervention. Therefore, studies aimed at lowering SCI could test interventions that target risk factors of systemic inflammation and use suPAR as an effect measure to assess whether various lifestyle (e.g., smoking cessation, caloric restriction, physical activity), psychosocial, or clinical interventions have a positive effect on health by lowering the suPAR level. For example, randomized studies of social interventions in high-risk individuals could use suPAR to inform on the effect on health risk. Studies aimed at preventing outcomes of SCI in general populations could use suPAR levels to identify individuals with elevated SCI, either by using suPAR alone (e.g., suPAR >4 ng/mL in general populations), or by using suPAR along with CRP and IL-6 to identify people with inflammation levels consistent with the SCI group identified with latent class analysis (163). Individuals with high suPAR could then be randomized to an intervention or control group, to test whether individuals in the intervention group experienced a positive effect of the intervention on other health-related outcomes. For example, psychosocial interventions that reduce people's psychological distress following trauma or other stressors might be able to reduce inflammation and improve people's health as a result (202), and it should be tested if such interventions can also reduce suPAR levels. Studies aimed at preventing outcomes of SCI in patient populations could randomize patients with high suPAR to interventions that accelerate the diagnostic or treatment procedures, or to novel therapies. Possible interventions for patients with unexpected high suPAR could be referral to a fast-track cancer diagnostics program like the Diagnostic Outpatient Clinics (89), treatment with anti-inflammatory medications like the IL-1 receptor antagonist anakinra (178), or the use of screening with a multiple rule-out CT scanning. This approach was recently shown to be feasible in Emergency Department patients selected based on their prognosis (moderate-to-high risk patients based on the vital sign-based National Early Warning Score) rather than their specific symptoms (203). In that study of 100 patients from the Emergency Department, scanning patients with moderate-to-high risk according to their vital signs led to change in treatment or additional examinations in 37 patients, of which 24 were diagnostically significant, including change in acute treatment in 11 patients and identification of previously unrecognized malignant tumors in 10 patients (203). This intervention might similarly be feasible to test in patients in the Emergency Department presenting without specific symptoms and with high suPAR (>6 ng/mL) for whom the risk of severe disease and mortality is high (77) and the concern regarding radiation exposure is outweighed by the potential benefits of diagnosing a serious illness.

Fourth, establishment of cut-offs and clinical guidelines remains an important task for the successful implementation of suPAR analysis in healthcare settings. These will depend on the context, with certain cut-offs indicating SCI along with risk of incident disease in healthy or general populations, while higher cut-offs could be used to indicate risk of different adverse

outcomes for specific clinical populations. For example, it has been suggested for patients in the Emergency Department that suPAR levels <4 ng/mL indicate that it is safe to discharge the patient (given that the patient does not have other acute indications), whereas levels >6 ng/mL should be considered as an alarming sign of risk for unfavorable outcomes, and levels >12 ng/mL are associated with a high risk of 28-day mortality (204). Direct testing of the benefits of using suPAR above 6 ng/mL for risk stratification is needed. In general populations, broader anti-inflammatory interventions targeting lifestyle behaviors or social risk factors—or even use of mild anti-inflammatory drugs—could be employed for people with elevated suPAR. In patient populations, the nature of an intervention would depend on the underlying diagnosis, made based on other clinical, biochemical, and physiological parameters. Given suPAR's lack of disease specificity, it is not possible to establish one single clinical intervention for patients with high suPAR. A high suPAR level can direct attention to the patient and provide additional characteristics of the underlying health state as well as the extent and severity of disease. Similar recommendations could be made for patients with high suPAR as those made by the Sepsis-3 task force for Intensive Care Unit (ICU) patients with positive Quick Sequential [Sepsis-related] Organ Failure Assessment (SOFA) score, prompting further investigation for organ dysfunction, initiation or escalation of therapy as appropriate, and assessment of need for critical care or increased frequency of monitoring (205).

Fifth, large-scale omics-based approaches could provide further information on suPAR's role in health and disease (206). Thus, integrating suPAR along with proteomic data on inflammatory biomarkers in a systems biology approach to explain SCI and related disease outcomes could identify novel direct and indirect interactions of suPAR with other inflammatory components. With this approach, suPAR emerged as one of the most important markers among 50 plasma proteins in a proteomic panel predicting acute myocardial infarction (207). However, other proteomic-based approaches have had limited ability to correlate suPAR levels and clinical outcomes in contrast to traditional ELISA-based detection methods (208, 209); even for ELISA methods, marked variation has been reported between assays (208, 210). The discrepancy between assay types could relate to different detections of suPAR isoforms, complexes of suPAR with its ligands (e.g., uPA, vitronectin) (210), or general proteomics-related challenges, such as cross-reactivity and non-specific interactions (211). This highlights the importance of understanding assay-related differences for suPAR to ensure robust prognostic capability, as the selection of assay for suPAR measurement could have direct impact on the clinical results obtained.

Implications

Identifying suPAR as a new biomarker of SCI has implications for theory, for methods, for research, and for prevention.

For theory, this hypothesis offers a new way to characterize and define the state of SCI, which has long been acknowledged to be poorly understood (2, 5). Moreover, it contributes with a new understanding of suPAR as a biomarker, with high suPAR levels

indexing presence of SCI along with increased risk of SCI-related outcomes and lower immunological capacity, i.e., the ability to tolerate and deal with incoming challenges to the immune system, such as physiological stress and disease. Additionally, the findings on early life risk factors and elevated adult suPAR provide support for the existing theory that the foundation for SCI in adulthood is laid already in childhood, with a wide variety of early-life risk factors having detrimental effects on life-long health through increased inflammation. Finally, the findings of suPAR being associated with inflammation as well as multiple indicators of accelerated biological aging and functional decline support the existing theories of immunosenescence and inflammation in aging, while also providing a new theory of suPAR being a clinically useful marker of SCI that can be used to quantify the level of inflammaging, and maybe even immunosenescence.

For methods, the measurement of blood suPAR levels comprises a new method to indicate the presence and to quantify and monitor the level of SCI and, with this, to detect and quantify the impact of various risk factors including social and psychological factors on health. Similarly, differentiating individuals in the three inflammation groups described by the E-Risk latent class analysis provides a new way to measure a person's level and type of inflammation (163). Detecting and quantifying SCI further allows for the development of a clinical diagnosis of SCI using elevated suPAR as the main indicator. Being able to diagnose an individual with SCI will enable the identification of individuals at highest risk of developing manifest disease among previously undiagnosed individuals, and provide a more exact assessment of the level of the inflammatory burden of an individual along with any subclinical organ damage and immunological capacity (or lack thereof). This essential information will be missed by relying on markers of acute inflammation. Using suPAR as an adjunct to traditional inflammatory biomarkers provides the most information and allows for the distinction between types of inflammation, such as acute vs. SCI.

For research, suPAR can be used as a quantifiable intermediate outcome between early risk factors and more distal outcomes such as disease development or mortality, as suPAR is associated with early life, social, behavioral, environmental, and pathological risk factors as well as with poor health, accelerated aging, incident and prevalent disease, and mortality. Thus, future intervention studies can use suPAR as an effect measure to assess the effect of social, behavioral, or clinical interventions on health, without having to wait for many years for traditional health outcomes (e.g., disease) to develop, for the impact of the intervention to show. For example, randomized clinical trials of anti-inflammatory or anti-aging interventions intended to slow the course of aging could include suPAR as an outcome measure of SCI. In addition, suPAR could be used to study how stressful experiences become biologically embedded to affect a person's downstream health, or it could be added as a new measure of long-term inflammatory processes to studies that investigate the causes of aging-related illness and the opportunities to improve health throughout the lifespan.

For prevention, since suPAR predicts future disease development, measuring suPAR creates an opportunity for

prevention by targeted interventions among people with the highest risk of adverse health outcomes, as exemplified with the suPAR-guided anakinra treatment in COVID-19 (177, 178). Thus, suPAR analysis in various clinical settings, including assessment of socially vulnerable individuals, could inform imminent serious health risks.

In conclusion, there is cumulating evidence that blood suPAR levels represent a common underlying disease-process shared by many diseases; that is, SCI. We propose that suPAR is a robust measure of SCI with the potential of becoming a gold standard for assessing SCI in research and clinical settings.

AUTHOR CONTRIBUTIONS

LR has written and conceptualized the article. JP and JE-O have contributed to the writing and critically revised it. All authors contributed to the article and approved the submitted version.

FUNDING

LR is supported by an international postdoc fellowship from the Lundbeck Foundation (grant no. R288-2018-380). JP is

supported by an international postdoc fellowship from the Alfred Benzon Foundation (grant no. ABF-2018-91). Preparation of this manuscript was supported, in part, by research grants from the National Institute on Aging (AG032282 and AG049789), National Institute of Child Health and Human Development (HD077482), and Medical Research Council (MR/P005918/1, G1002190). The funders had no role in the conception, writing, or preparation of this manuscript, nor in the decision to publish.

ACKNOWLEDGMENTS

The authors thank Dr. Avshalom Caspi for supervision and revision of the paper.

SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: <https://www.frontiersin.org/articles/10.3389/fimmu.2021.780641/full#supplementary-material>

REFERENCES

- World Health Organization. (2014). *Global Status Report on Noncommunicable Diseases 2014*. World Health Organization. <https://apps.who.int/iris/handle/10665/148114>
- Furman D, Campisi J, Verdin E, Carrera-bastos P, Targ S, Franceschi C, et al. Chronic Inflammation in the Etiology of Disease Across the Life Span. *Nat Med* (2019) 25:1822–32. doi: 10.1038/s41591-019-0675-0
- Yuan N, Chen Y, Xia Y, Dai J, Liu C. Inflammation-Related Biomarkers in Major Psychiatric Disorders: A Cross-Disorder Assessment of Reproducibility and Specificity in 43 Meta-Analyses. *Transl Psychiatry* (2019) 9:233. doi: 10.1038/s41398-019-0570-y
- Miller AH. Beyond Depression: The Expanding Role of Inflammation in Psychiatric Disorders. *World Psychiatry* (2020) 19:108–9. doi: 10.1002/wps.20723
- Medzhitov R. Origin and Physiological Roles of Inflammation. *Nature* (2008) 454:428–35. doi: 10.1038/nature07201
- Pearson TA, Mensah GA, Alexander RW, Anderson JL, Cannon RO, Criqui M, et al. Markers of Inflammation and Cardiovascular Disease: Application to Clinical and Public Health Practice: A Statement for Healthcare Professionals From the Centers for Disease Control and Prevention and the American Heart Association. *Circulation* (2003) 107:499–511. doi: 10.1161/01.CIR.0000052939.59093.45
- Ridker PM. Clinical Application of C-Reactive Protein for Cardiovascular Disease Detection and Prevention. *Circulation* (2003) 107:363–9. doi: 10.1161/01.CIR.0000053730.47739.3C
- Alpert A, Pickman Y, Leipold M, Rosenberg-Hasson Y, Ji X, Gaujoux R, et al. A Clinically Meaningful Metric of Immune Age Derived From High-Dimensional Longitudinal Monitoring. *Nat Med* (2019) 25:487–95. doi: 10.1038/s41591-019-0381-y
- Sayed N, Huang Y, Nguyen K, Krejciova-Rajaniemi Z, Grawe AP, Gao T, et al. An Inflammatory Aging Clock (Iage) Based on Deep Learning Tracks Multimorbidity, Immunosenescence, Frailty and Cardiovascular Aging. *Nat Aging* (2021) 1:598–615. doi: 10.1038/s43587-021-00082-y
- Zeier M, Reiser J. suPAR and Chronic Kidney Disease—A Podocyte Story. *Pflugers Arch* (2017) 469:1017–20. doi: 10.1007/s00424-017-2026-7
- Ni W, Han Y, Zhao J, Cui J, Wang K, Wang R, et al. Serum Soluble Urokinase-Type Plasminogen Activator Receptor as a Biological Marker of Bacterial Infection in Adults: A Systematic Review and Meta-Analysis. *Sci Rep* (2016) 6:39481. doi: 10.1038/srep39481
- Hodges GW, Bang CN, Wachtell K, Eugen-Olsen J, Jeppesen JL. suPAR: A New Biomarker for Cardiovascular Disease? *Can J Cardiol* (2015) 31:1293–302. doi: 10.1016/j.cjca.2015.03.023
- Desmedt S, Desmedt V, Delanghe JR, Speeckaert R, Speeckaert MM. The Intriguing Role of Soluble Urokinase Receptor in Inflammatory Diseases. *Crit Rev Clin Lab Sci* (2017) 54:117–33. doi: 10.1080/10408363.2016.1269310
- Marsland AL. suPAR: A Newer Biomarker of Systemic Chronic Inflammation. *Brain Behav Immun* (2021) 98:263–4. doi: 10.1016/j.bbi.2021.08.224
- Mustjoki S, Sidenius N, Sier CF, Blasi F, Elonen E, Alitalo R, et al. Soluble Urokinase Receptor Levels Correlate With Number of Circulating Tumor Cells in Acute Myeloid Leukemia and Decrease Rapidly During Chemotherapy. *Cancer Res* (2000) 60:7126–32.
- Kjellman A, Akre O, Gustafsson O, Høyer-Hansen G, Lilja H, Norming U, et al. Soluble Urokinase Plasminogen Activator Receptor as a Prognostic Marker in Men Participating in Prostate Cancer Screening. *J Intern Med* (2011) 269:299–305. doi: 10.1111/j.1365-2796.2010.02284.x
- Tzanakaki G, Paparoupa M, Kyprianou M, Barbouni A, Eugen-Olsen J, Kourea-Kremastinou J. Elevated Soluble Urokinase Receptor Values in CSF, Age and Bacterial Meningitis Infection Are Independent and Additive Risk Factors of Fatal Outcome. *Eur J Clin Microbiol Infect Dis* (2012) 31:1157–62. doi: 10.1007/s10096-011-1423-7
- Gustafsson A, Ajeti V, Ljunggren L. Detection of suPAR in the Saliva of Healthy Young Adults: Comparison With Plasma Levels. *Biomark Insights* (2011) 6:119–25. doi: 10.4137/BMI.S8326
- Rasmussen LJH. Clinical Prognostication With the Inflammatory Biomarker suPAR. PhD Thesis. University of Copenhagen. (2018).
- Smith HW, Marshall CJ. Regulation of Cell Signalling by uPAR. *Nat Rev Mol Cell Biol* (2010) 11:23–36. doi: 10.1038/nrm2821
- Thunø M, Macho B, Eugen-Olsen J. suPAR: The Molecular Crystal Ball. *Dis Markers* (2009) 27:157–72. doi: 10.3233/DMA-2009-0657
- Enocsson H, Wetterö J, Skogh T, Sjöwall C. Soluble Urokinase Plasminogen Activator Receptor Levels Reflect Organ Damage in Systemic Lupus Erythematosus. *Transl Res* (2013) 162:287–96. doi: 10.1016/j.trsl.2013.07.003
- Koch A, Voigt S, Kruschinski C, Sanson E, Dücker H, Horn A, et al. Circulating Soluble Urokinase Plasminogen Activator Receptor Is Stably

- Elevated During the First Week of Treatment in the Intensive Care Unit and Predicts Mortality in Critically Ill Patients. *Crit Care* (2011) 15:R63. doi: 10.1186/cc10037
24. Andersen O, Eugen-Olsen J, Kofoed K, Iversen J, Haugaard SB. Soluble Urokinase Plasminogen Activator Receptor Is a Marker of Dysmetabolism in HIV-Infected Patients Receiving Highly Active Antiretroviral Therapy. *J Med Virol* (2008) 80:209–16. doi: 10.1002/jmv.21114
 25. Haastруп E, Grau K, Eugen-Olsen J, Thorball C, Kessing LV, Ullum H. Soluble Urokinase Plasminogen Activator Receptor as a Marker for Use of Antidepressants. *PLoS One* (2014) 9:e110555. doi: 10.1371/journal.pone.0110555
 26. Haupt TH, Kallemosе T, Ladelund S, Rasmussen LJH, Thorball CW, Andersen O, et al. Risk Factors Associated With Serum Levels of the Inflammatory Biomarker Soluble Urokinase Plasminogen Activator Receptor in a General Population. *Biomark Insights* (2014) 9:91–100. doi: 10.4137/BMI.S19876
 27. Chew-Harris J, Appleby S, Richards AM, Troughton RW, Pemberton CJ. Analytical, Biochemical and Clearance Considerations of Soluble Urokinase Plasminogen Activator Receptor (suPAR) in Healthy Individuals. *Clin Biochem* (2019) 69:36–44. doi: 10.1016/j.clinbiochem.2019.05.010
 28. Wlazел RN, Szwabe K, Guligowska A, Kostka T. Soluble Urokinase Plasminogen Activator Receptor Level in Individuals of Advanced Age. *Sci Rep* (2020) 10:15462. doi: 10.1038/s41598-020-72377-w
 29. Stephens RW, Pedersen AN, Nielsen HJ, Hamers MJ, Høyer-Hansen G, Rønne E, et al. ELISA Determination of Soluble Urokinase Receptor in Blood From Healthy Donors and Cancer Patients. *Clin Chem* (1997) 43:1868–76. doi: 10.1093/clinchem/43.10.1868
 30. Dowsett J, Ferkingstad E, Rasmussen LJH, Thørner LW, Magnússon MK, Sugden K, et al. Eleven Genomic Loci Affect Plasma Levels of Chronic Inflammation Marker Soluble Urokinase-Type Plasminogen Activator Receptor. *Commun Biol* (2021) 4:655. doi: 10.1038/s42003-021-02144-8
 31. Manetti M, Allanore Y, Revillod L, Fatini C, Guiducci S, Cuomo G, et al. A Genetic Variation Located in the Promoter Region of the UPAR (CD87) Gene Is Associated With the Vascular Complications of Systemic Sclerosis. *Arthritis Rheum* (2011) 63:247–56. doi: 10.1002/art.30101
 32. Zandifar A, Soleimani S, Iraj N, Haghdoost F, Tajaddini M, Javanmard SH. Association Between Promoter Region of the uPAR (Rs344781) Gene Polymorphism in Genetic Susceptibility to Migraine Without Aura in Three Iranian Hospitals. *Clin Neurol Neurosurg* (2014) 120:45–8. doi: 10.1016/j.clineuro.2014.02.003
 33. Weng C-J, Tsai C-M, Chen Y-C, Hsieh Y-H, Lin C-W, Liu Y-F, et al. Evaluation of the Association of Urokinase Plasminogen Activator System Gene Polymorphisms With Susceptibility and Pathological Development of Hepatocellular Carcinoma. *Ann Surg Oncol* (2010) 17:3394–401. doi: 10.1245/s10434-010-1124-y
 34. Andraweera PH, Dekker GA, Thompson SD, Nowak RC, Jayasekara RW, Dissanayake VHW, et al. Polymorphisms in the Fibrinolytic Pathway Genes and the Risk of Recurrent Spontaneous Abortion. *Reprod BioMed Online* (2014) 29:745–51. doi: 10.1016/j.rbmo.2014.08.014
 35. Drechsler C, Hayek SS, Wei C, Sever S, Genser B, Krane V, et al. Soluble Urokinase Plasminogen Activator Receptor and Outcomes in Patients With Diabetes on Hemodialysis. *Clin J Am Soc Nephrol* (2017) 12:1265–73. doi: 10.2215/CJN.10881016
 36. Wlazел RN, Szadkowska I, Bartnicki P, Rośniak-Bak K, Rysz J. Clinical and Prognostic Usefulness of Soluble Urokinase Plasminogen Activator Receptor in Hemodialysis Patients. *Int Urol Nephrol* (2018) 50:339–45. doi: 10.1007/s11255-017-1778-5
 37. Azam TU, Shadid HR, Blakely P, O'Hayer P, Berlin H, Pan M, et al. Soluble Urokinase Receptor (SuPAR) in COVID-19-Related AKI. *J Am Soc Nephrol* (2020) 31:2725–35. doi: 10.1681/asn.2020060829
 38. Wei C, Li J, Adair BD, Zhu K, Cai J, Merchant M, et al. uPAR Isoform 2 Forms a Dimer and Induces Severe Kidney Disease in Mice. *J Clin Invest* (2019) 129:1946–59. doi: 10.1172/jci124793
 39. Bhandary YP, Velusamy T, Shetty P, Shetty RS, Idell S, Cines DB, et al. Post-Transcriptional Regulation of Urokinase-Type Plasminogen Activator Receptor Expression in Lipopolysaccharide-Induced Acute Lung Injury. *Am J Respir Crit Care Med* (2009) 179:288–98. doi: 10.1164/rccm.200712-1787OC
 40. Matsumoto H, Ueshima S, Fukao H, Mitsui Y, Matsuo O. Effects of Lipopolysaccharide on the Expression of Fibrinolytic Factors in an Established Cell Line From Human Endothelial Cells. *Life Sci* (1996) 59:85–96. doi: 10.1016/0024-3205(96)00265-2
 41. Zimmermann HW, Reuken PA, Koch A, Bartneck M, Adams DH, Trautwein C, et al. Soluble Urokinase Plasminogen Activator Receptor Is Compartmentally Regulated in Decompensated Cirrhosis and Indicates Immune Activation and Short-Term Mortality. *J Intern Med* (2013) 274:86–100. doi: 10.1111/joim.12054
 42. Dekkers PE, ten Hove T, te Velde AA, van Deventer SJ, van der Poll T. Upregulation of Monocyte Urokinase Plasminogen Activator Receptor During Human Endotoxemia. *Infect Immun* (2000) 68:2156–60. doi: 10.1128/IAI.68.4.2156-2160.2000
 43. Ostrowski SR, Plomgaard P, Fischer CP, Steensberg AS, Møller K, Høyer-Hansen G, et al. Interleukin-6 Infusion During Human Endotoxaemia Inhibits *In Vitro* Release of the Urokinase Receptor From Peripheral Blood Mononuclear Cells. *Scand J Immunol* (2005) 61:197–206. doi: 10.1111/j.0300-9475.2005.01547.x
 44. Pliyev BK. Activated Human Neutrophils Rapidly Release the Chemotactically Active D2D3 Form of the Urokinase-Type Plasminogen Activator Receptor (uPAR/CD87). *Mol Cell Biochem* (2009) 321:111–22. doi: 10.1007/s11010-008-9925-z
 45. Selleri C, Montuori N, Ricci P, Visconte V, Carriero MV, Sidenius N, et al. Involvement of the Urokinase-Type Plasminogen Activator Receptor in Hematopoietic Stem Cell Mobilization. *Blood* (2005) 105:2198–205. doi: 10.1182/blood-2004-06-2424
 46. Chavakis T, Willuweit AK, Lupu F, Preissner KT, Kanse SM. Release of Soluble Urokinase Receptor From Vascular Cells. *Thromb Haemost* (2001) 86:686–93. doi: 10.1055/s-0037-1616105
 47. Carmeliet P, Moons L, Lijnen R, Baes M, Lemaître V, Tipping P, et al. Urokinase-Generated Plasmin Activates Matrix Metalloproteinases During Aneurysm Formation. *Nat Genet* (1997) 17:439–44. doi: 10.1038/ng1297-439
 48. Del Rosso M, Fibbi G, Pucci M, Margheri F, Serrati S. The Plasminogen Activation System in Inflammation. *Front Biosci* (2008) 13:4667–86. doi: 10.2741/3032
 49. Houck K, Leung D, Rowland A, Winer J, Ferrara N. Dual Regulation of Vascular Endothelial Growth Factor Bioavailability by Genetic and Proteolytic Mechanisms. *J Biol Chem* (1992) 267:26031–7. doi: 10.1016/S0021-9258(18)35712-0
 50. Lyons RM, Gentry LE, Purchio AF, Moses HL. Mechanism of Activation of Latent Recombinant Transforming Growth Factor Beta 1 by Plasmin. *J Cell Biol* (1990) 110:1361–7. doi: 10.1083/jcb.110.4.1361
 51. Bohuslav J, Horejsi V, Hansmann C, Stöckl J, Weidle UH, Majdic O, et al. Urokinase Plasminogen Activator Receptor, Beta 2-Integrins, and Src-Kinases Within a Single Receptor Complex of Human Monocytes. *J Exp Med* (1995) 181:1381–90. doi: 10.1084/jem.181.4.1381
 52. Khan MM, Bradford HN, Isordia-Salas I, Liu Y, Wu Y, Espinola RG, et al. High-Molecular-Weight Kininogen Fragments Stimulate the Secretion of Cytokines and Chemokines Through uPAR, Mac-1, and Gc1qr in Monocytes. *Arterioscler Thromb Vasc Biol* (2006) 26:2260–6. doi: 10.1161/01.ATV.0000240290.70852.c0
 53. Sidenius N, Andolfo A, Fesce R, Blasi F. Urokinase Regulates Vitronectin Binding by Controlling Urokinase Receptor Oligomerization. *J Biol Chem* (2002) 277:27982–90. doi: 10.1074/jbc.M111736200
 54. Chavakis T, Kanse SM, Yutzy B, Lijnen HR, Preissner KT. Vitronectin Concentrates Proteolytic Activity on the Cell Surface and Extracellular Matrix by Trapping Soluble Urokinase Receptor-Urokinase Complexes. *Blood* (1998) 91:2305–12. doi: 10.1182/blood.V91.7.2305
 55. Resnati M, Pallavicini I, Wang JM, Oppenheim J, Serhan CN, Romano M, et al. The Fibrinolytic Receptor for Urokinase Activates the G Protein-Coupled Chemotactic Receptor FPRL1/LXA4R. *Proc Natl Acad Sci USA* (2002) 99:1359–64. doi: 10.1073/pnas.022652999
 56. Resnati M, Guttinger M, Valcamonica S, Sidenius N, Blasi F, Fazioli F. Proteolytic Cleavage of the Urokinase Receptor Substitutes for the Agonist-Induced Chemotactic Effect. *EMBO J* (1996) 15:1572–82. doi: 10.1002/j.1460-2075.1996.tb00502.x
 57. Bifulco K, Longanesi-Cattani I, Masucci MT, De Chiara A, Fazioli F, Di Carluccio G, et al. Involvement of the Soluble Urokinase Receptor in

- Chondrosarcoma Cell Mobilization. *Sarcoma* (2011) 2011:842842. doi: 10.1155/2011/842842
58. Furlan F, Orlando S, Laudanna C, Resnati M, Basso V, Blasi F, et al. The Soluble D2D3(88-274) Fragment of the Urokinase Receptor Inhibits Monocyte Chemotaxis and Integrin-Dependent Cell Adhesion. *J Cell Sci* (2004) 117:2909–16. doi: 10.1242/jcs.01149
 59. Bifulco K, Longanesi-Cattani I, Gala M, Di Carluccio G, Masucci MT, Pavone V, et al. The Soluble Form of Urokinase Receptor Promotes Angiogenesis Through Its Ser88-Arg-Ser-Arg-Tyr92 Chemotactic Sequence. *J Thromb Haemost* (2010) 8:2789–99. doi: 10.1111/j.1538-7836.2010.04075.x
 60. Park Y-J, Liu G, Tsuruta Y, Lorne E, Abraham E. Participation of the Urokinase Receptor in Neutrophil Efferocytosis. *Blood* (2009) 114:860–70. doi: 10.1182/blood-2008-12-193524
 61. Masucci MT, Pedersen N, Blasi F. A Soluble, Ligand Binding Mutant of the Human Urokinase Plasminogen Activator Receptor. *J Biol Chem* (1991) 266:8655–8. doi: 10.1016/S0021-9258(18)31492-3
 62. Hoyer-Hansen G, Pessara U, Holm A, Pass J, Weidle U, Danø K, et al. Urokinase-Catalyzed Cleavage of the Urokinase Receptor Requires an Intact Glycolipid Anchor. *Biochem J* (2001) 358:673–9. doi: 10.1042/0264-6021:3580673
 63. Langkilde A, Jakobsen TL, Bandholm TQ, Eugen-Olsen J, Blauenfeldt T, Petersen J, et al. Inflammation and Post-Operative Recovery in Patients Undergoing Total Knee Arthroplasty—Secondary Analysis of a Randomized Controlled Trial. *Osteoarthritis Cartilage* (2017) 25:1265–73. doi: 10.1016/j.joca.2017.03.008
 64. Coppé J-P, Desprez P-Y, Krtolica A, Campisi J. The Senescence-Associated Secretory Phenotype: The Dark Side of Tumor Suppression. *Annu Rev Pathol* (2010) 5:99–118. doi: 10.1146/annurev-pathol-121808-102144
 65. Amor C, Feucht J, Leibold J, Ho YJ, Zhu C, Alonso-Curbelo D, et al. Senolytic CAR T Cells Reverse Senescence-Associated Pathologies. *Nature* (2020) 583:127–32. doi: 10.1038/s41586-020-2403-9
 66. Eugen-Olsen J, Ladelund S, Sørensen LT. Plasma suPAR Is Lowered by Smoking Cessation: A Randomized Controlled Study. *Eur J Clin Invest* (2016) 46:305–11. doi: 10.1111/eci.12593
 67. Haupt TH, Petersen J, Ellekilde G, Klausen HH, Thorball CW, Eugen-Olsen J, et al. Plasma suPAR Levels Are Associated With Mortality, Admission Time, and Charlson Comorbidity Index in the Acutely Admitted Medical Patient: A Prospective Observational Study. *Crit Care* (2012) 16:R130. doi: 10.1186/cc11434
 68. Zimmermann HW, Koch A, Seidler S, Trautwein C, Tacke F. Circulating Soluble Urokinase Plasminogen Activator Is Elevated in Patients With Chronic Liver Disease, Discriminates Stage and Aetiology of Cirrhosis and Predicts Prognosis. *Liver Int* (2012) 32:500–9. doi: 10.1111/j.1478-3231.2011.02665.x
 69. Rasmussen LJH, Moffitt TE, Eugen-Olsen J, Belsky DW, Danese A, Harrington HL, et al. Cumulative Childhood Risk Is Associated With a New Measure of Chronic Inflammation in Adulthood. *J Child Psychol Psychiatry* (2019) 60:199–208. doi: 10.1111/jcpp.12928
 70. Ostrowski SR, Ullum H, Goka BQ, Hoyer-Hansen G, Obeng-Adjei G, Pedersen BK, et al. Plasma Concentrations of Soluble Urokinase-Type Plasminogen Activator Receptor Are Increased in Patients With Malaria and Are Associated With a Poor Clinical or a Fatal Outcome. *J Infect Dis* (2005) 191:1331–41. doi: 10.1086/428854
 71. Wrotek A, Jackowska T, Pawlik K. Soluble Urokinase Plasminogen Activator Receptor: An Indicator of Pneumonia Severity in Children. *Adv Exp Med Biol* (2015) 835:1–7. doi: 10.1007/5584_2014_40
 72. Koch A, Zimmermann HW, Gassler N, Jochum C, Weiskirchen R, Bruensing J, et al. Clinical Relevance and Cellular Source of Elevated Soluble Urokinase Plasminogen Activator Receptor (suPAR) in Acute Liver Failure. *Liver Int* (2014) 34:1330–9. doi: 10.1111/liv.12512
 73. Hahm E, Wei C, Fernandez I, Li J, Tardi NJ, Tracy M, et al. Bone Marrow-Derived Immature Myeloid Cells Are a Main Source of Circulating suPAR Contributing to Proteinuric Kidney Disease. *Nat Med* (2017) 23:100–6. doi: 10.1038/nm.4242
 74. Pietras EM. Inflammation: A Key Regulator of Hematopoietic Stem Cell Fate in Health and Disease. *Blood* (2017) 130:1693–8. doi: 10.1182/blood-2017-06-780882
 75. Botha S, Fourie CMT, Schutte R, Kruger A, Schutte AE. Associations of suPAR With Lifestyle and Cardiometabolic Risk Factors. *Eur J Clin Invest* (2014) 44:619–26. doi: 10.1111/eci.12278
 76. Eugen-Olsen J, Andersen O, Linneberg A, Ladelund S, Hansen TW, Langkilde A, et al. Circulating Soluble Urokinase Plasminogen Activator Receptor Predicts Cancer, Cardiovascular Disease, Diabetes and Mortality in the General Population. *J Intern Med* (2010) 268:296–308. doi: 10.1111/j.1365-2796.2010.02252.x
 77. Rasmussen LJH, Ladelund S, Haupt TH, Ellekilde G, Poulsen JH, Iversen K, et al. Soluble Urokinase Plasminogen Activator Receptor (suPAR) in Acute Care: A Strong Marker of Disease Presence and Severity, Readmission and Mortality. A Retrospective Cohort Study. *Emerg Med J* (2016) 33:769–75. doi: 10.1136/emmermed-2015-205444
 78. Toldi G, Bekő G, Kádár G, Mácsai E, Kovács L, Vásárhelyi B, et al. Soluble Urokinase Plasminogen Activator Receptor (suPAR) in the Assessment of Inflammatory Activity of Rheumatoid Arthritis Patients in Remission. *Clin Chem Lab Med* (2013) 51:327–32. doi: 10.1515/cclm-2012-0221
 79. Slot O, Brünnner N, Loch H, Oxholm P, Stephens RW. Soluble Urokinase Plasminogen Activator Receptor in Plasma of Patients With Inflammatory Rheumatic Disorders: Increased Concentrations in Rheumatoid Arthritis. *Ann Rheum Dis* (1999) 58:488–92. doi: 10.1136/ard.58.8.488
 80. AboEl-Magd GH, Mabrouk MM. Soluble Urokinase-Type Plasminogen Activator Receptor as a Measure of Treatment Response in Acute Exacerbation of COPD. *J Bras Pneumol* (2018) 44:36–41. doi: 10.1590/S1806-37562017000000151
 81. Gumus A, Altintas N, Cinarca H, Kirbas A, Hazirolu M, Karatas M, et al. Soluble Urokinase-Type Plasminogen Activator Receptor Is a Novel Biomarker Predicting Acute Exacerbation in COPD. *Int J Chron Obstruct Pulmon Dis* (2015) 10:357–65. doi: 10.2147/COPD.S77654
 82. Zeng M, Chang M, Zheng H, Li B, Chen Y, He W, et al. Clinical Value of Soluble Urokinase-Type Plasminogen Activator Receptor in the Diagnosis, Prognosis, and Therapeutic Guidance of Sepsis. *Am J Emerg Med* (2016) 34:375–80. doi: 10.1016/j.ajem.2015.11.004
 83. Botha S, Fourie CM, Schutte R, Eugen-Olsen J, Pretorius R, Schutte AE. Soluble Urokinase Plasminogen Activator Receptor as a Prognostic Marker of All-Cause and Cardiovascular Mortality in a Black Population. *Int J Cardiol* (2015) 184:631–6. doi: 10.1016/j.ijcard.2015.03.041
 84. Almroth G, Lönn J, Uhlin F, Brudin L, Andersson B, Hahn-Zoric M. Sclerostin, TNF-Alpha and Interleukin-18 Correlate and Are Together With Klotho Related to Other Growth Factors and Cytokines in Haemodialysis Patients. *Scand J Immunol* (2016) 83:58–63. doi: 10.1111/sji.12392
 85. Diederichsen MZ, Diederichsen SZ, Mickley H, Steffensen FH, Lambrechtsen J, Sand NPR, et al. Prognostic Value of suPAR and Hs-CRP on Cardiovascular Disease. *Atherosclerosis* (2018) 271:245–51. doi: 10.1016/j.atherosclerosis.2018.01.029
 86. Böcskei RM, Benczúr B, Losonczy G, Illyés M, Cziráki A, Müller V, et al. Soluble Urokinase-Type Plasminogen Activator Receptor and Arterial Stiffness in Patients With COPD. *Lung* (2019) 197:189–97. doi: 10.1007/s00408-019-00211-w
 87. Yilmaz G, Mentese A, Kaya S, Uzun A, Karahan SC, Koksali I. The Diagnostic and Prognostic Significance of Soluble Urokinase Plasminogen Activator Receptor in Crimean-Congo Hemorrhagic Fever. *J Clin Virol* (2011) 50:209–11. doi: 10.1016/j.jcv.2010.11.014
 88. Wittenhagen P, Andersen JB, Hansen A, Lindholm L, Rønne F, Theil J, et al. Plasma Soluble Urokinase Plasminogen Activator Receptor in Children With Urinary Tract Infection. *Biomark Insights* (2011) 6:79–82. doi: 10.4137/BMI.S6876
 89. Rasmussen LJH, Schultz M, Gaardsting A, Ladelund S, Garred P, Iversen K, et al. Inflammatory Biomarkers and Cancer: CRP and suPAR as Markers of Incident Cancer in Patients With Serious Nonspecific Symptoms and Signs of Cancer. *Int J Cancer* (2017) 141:191–9. doi: 10.1002/ijc.30732
 90. Lyngbæk S, Sehested T, Marott JL, Hansen TW, Olsen MH, Andersen O, et al. CRP and suPAR Are Differently Related to Anthropometry and Subclinical Organ Damage. *Int J Cardiol* (2013) 167:781–5. doi: 10.1016/j.ijcard.2012.03.040
 91. Zhou X, Fragala MS, McElhaney JE, Kuchel GA. Conceptual and Methodological Issues Relevant to Cytokine and Inflammatory Marker

- Measurements in Clinical Research. *Curr Opin Clin Nutr Metab Care* (2010) 13:541–7. doi: 10.1097/MCO.0b013e32833cf3bc
92. Petrovsky N, McNair P, Harrison LC. Diurnal Rhythms of Pro-Inflammatory Cytokines: Regulation by Plasma Cortisol and Therapeutic Implications. *Cytokine* (1998) 10:307–12. doi: 10.1006/cyto.1997.0289
 93. Kleiner G, Marcuzzi A, Zanin V, Monasta L, Zauli G. Cytokine Levels in the Serum of Healthy Subjects. *Mediators Inflammation* (2013) 2013:434010. doi: 10.1155/2013/434010
 94. Zelová H, Hošek J. TNF- α Signalling and Inflammation: Interactions Between Old Acquaintances. *Inflammation Res* (2013) 62:641–51. doi: 10.1007/s00011-013-0633-0
 95. Aziz N. Measurement of Circulating Cytokines and Immune-Activation Markers by Multiplex Technology in the Clinical Setting: What Are We Really Measuring? *For Immunopathol Dis Therap* (2015) 6:19–22. doi: 10.1615/ForumImmunDisTher.2015014162
 96. Lyngbæk S, Marott JL, Møller DV, Christiansen M, Iversen KK, Clemmensen PM, et al. Usefulness of Soluble Urokinase Plasminogen Activator Receptor to Predict Repeat Myocardial Infarction and Mortality in Patients With ST-Segment Elevation Myocardial Infarction Undergoing Primary Percutaneous Intervention. *Am J Cardiol* (2012) 110:1756–63. doi: 10.1016/j.amjcard.2012.08.008
 97. Sier CF, Sidenius N, Mariani A, Aletti G, Agape V, Ferrari A, et al. Presence of Urokinase-Type Plasminogen Activator Receptor in Urine of Cancer Patients and Its Possible Clinical Relevance. *Lab Invest* (1999) 79:717–22.
 98. Rasmussen LJH, Ladelund S, Haupt TH, Ellekilde GE, Eugen-Olsen J, Andersen O. Combining National Early Warning Score With Soluble Urokinase Plasminogen Activator Receptor (suPAR) Improves Risk Prediction in Acute Medical Patients: A Registry-Based Cohort Study. *Crit Care Med* (2018) 46:1961–8. doi: 10.1097/CCM.0000000000003441
 99. Kofoed K, Schneider UV, Scheel T, Andersen O, Eugen-Olsen J. Development and Validation of a Multiplex Add-on Assay for Sepsis Biomarkers Using xMAP Technology. *Clin Chem* (2006) 52:1284–93. doi: 10.1373/clinchem.2006.067595
 100. Schenk M, Eichelmann F, Schulze MB, Rudovich N, Pfeiffer AF, di Giuseppe R, et al. Reproducibility of Novel Immune-Inflammatory Biomarkers Over 4 Months: An Analysis With Repeated Measures Design. *Biomark Med* (2019) 13:639–48. doi: 10.2217/bmm-2018-0351
 101. Haupt TH, Rasmussen LJH, Kallemose T, Ladelund S, Andersen O, Pisinger C, et al. Healthy Lifestyles Reduce suPAR and Mortality in a Danish General Population Study. *Immun Ageing* (2019) 16:1. doi: 10.1186/s12979-018-0141-8
 102. Rasmussen LJH, Caspi A, Ambler A, Danese A, Elliott M, Eugen-Olsen J, et al. Association Between Elevated suPAR, a New Biomarker of Inflammation, and Accelerated Aging. *J Gerontol A Biol Sci Med Sci* (2021) 76:318–27. doi: 10.1093/gerona/glaa178
 103. Bourassa KJ, Rasmussen LJH, Danese A, Eugen-Olsen J, Harrington H, Houts R, et al. Linking Stressful Life Events and Chronic Inflammation Using suPAR (Soluble Urokinase Plasminogen Activator Receptor). *Brain Behav Immun* (2021) 97:79–88. doi: 10.1016/j.bbi.2021.06.018
 104. Fagerberg B, Borné Y, Barregard L, Sallsten G, Forsgard N, Hedblad B, et al. Cadmium Exposure Is Associated With Soluble Urokinase Plasminogen Activator Receptor, A Circulating Marker of Inflammation and Future Cardiovascular Disease. *Environ Res* (2017) 152:185–91. doi: 10.1016/j.envres.2016.10.019
 105. Timmermans K, Vaneker M, Scheffer GJ, Maassen P, Janssen S, Kox M, et al. Soluble Urokinase-Type Plasminogen Activator Levels Are Related to Plasma Cytokine Levels But Have Low Predictive Value for Mortality in Trauma Patients. *J Crit Care* (2015) 30:476–80. doi: 10.1016/j.jccr.2015.01.006
 106. Loosen SH, Tacke F, Pütke N, Binneboesel M, Wiltberger G, Alizai PH, et al. High Baseline Soluble Urokinase Plasminogen Activator Receptor (suPAR) Serum Levels Indicate Adverse Outcome After Resection of Pancreatic Adenocarcinoma. *Carcinogenesis* (2019) 40:947–55. doi: 10.1093/carcin/bgz033
 107. Persson M, Östling G, Smith G, Hamrefors V, Melander O, Hedblad B, et al. Soluble Urokinase Plasminogen Activator Receptor: A Risk Factor for Carotid Plaque, Stroke, and Coronary Artery Disease. *Stroke* (2014) 45:18–23. doi: 10.1161/STROKEAHA.113.003305
 108. Westin O, Rasmussen LJH, Andersen O, Buch E, Eugen-Olsen J, Friberg J. Soluble Urokinase Plasminogen Activator Receptor (suPAR) as a Predictor of Incident Atrial Fibrillation. *J Atr Fibrillation* (2018) 10:1801. doi: 10.4022/jafib.1801
 109. Theilade S, Lyngbæk S, Hansen TW, Eugen-Olsen J, Fenger M, Rossing P, et al. Soluble Urokinase Plasminogen Activator Receptor Levels Are Elevated and Associated With Complications in Patients With Type 1 Diabetes. *J Intern Med* (2015) 277:362–71. doi: 10.1111/joim.12269
 110. Curovic VR, Theilade S, Winther SA, Tofte N, Eugen-Olsen J, Persson F, et al. Soluble Urokinase Plasminogen Activator Receptor Predicts Cardiovascular Events, Kidney Function Decline, and Mortality in Patients With Type 1 Diabetes. *Diabetes Care* (2018) 42:1112–9. doi: 10.2337/dc18-1427
 111. Heraclides A, Jensen TM, Rasmussen SS, Eugen-Olsen J, Haugeard SB, Borch-Johnsen K, et al. The Pro-Inflammatory Biomarker Soluble Urokinase Plasminogen Activator Receptor (suPAR) Is Associated With Incident Type 2 Diabetes Among Overweight But Not Obese Individuals With Impaired Glucose Regulation: Effect Modification by Smoking and Body Weight. *Diabetologia* (2013) 56:1542–6. doi: 10.1007/s00125-013-2914-0
 112. Guthoff M, Wagner R, Randrianarisoa E, Hatzigelaki E, Peter A, Häring H-U, et al. Soluble Urokinase Receptor (suPAR) Predicts Microalbuminuria in Patients at Risk for Type 2 Diabetes Mellitus. *Sci Rep* (2017) 7:40627. doi: 10.1038/srep40627
 113. Mustjoki S, Alitalo R, Stephens RW, Vaheri A. Blast Cell-Surface and Plasma Soluble Urokinase Receptor in Acute Leukemia Patients: Relationship to Classification and Response to Therapy. *Thromb Haemost* (1999) 81:705–10. doi: 10.1055/s-0037-1614558
 114. Wach S, Al-Janabi O, Weigelt K, Fischer K, Greither T, Marcou M, et al. The Combined Serum Levels of miR-375 and Urokinase Plasminogen Activator Receptor Are Suggested as Diagnostic and Prognostic Biomarkers in Prostate Cancer. *Int J Cancer* (2015) 137:1406–16. doi: 10.1002/ijc.29505
 115. Sorio C, Mafficini A, Furlan F, Barbi S, Bonora A, Brocco G, et al. Elevated Urinary Levels of Urokinase-Type Plasminogen Activator Receptor (uPAR) in Pancreatic Ductal Adenocarcinoma Identify a Clinically High-Risk Group. *BMC Cancer* (2011) 11:448. doi: 10.1186/1471-2407-11-448
 116. Cobos E, Jumper C, Lox C. Pretreatment Determination of the Serum Urokinase Plasminogen Activator and Its Soluble Receptor in Advanced Small-Cell Lung Cancer or Non-Small-Cell Lung Cancer. *Clin Appl Thromb* (2003) 9:241–6. doi: 10.1177/107602960300900309
 117. Miyake H, Hara I, Yamanaka K, Gohji K, Arakawa S, Kamidono S. Elevation of Serum Levels of Urokinase-Type Plasminogen Activator and Its Receptor Is Associated With Disease Progression and Prognosis in Patients With Prostate Cancer. *Prostate* (1999) 39:123–9. doi: 10.1002/(SICI)1097-0045(199905051)39:2<123::AID-PROS7>3.0.CO;2-2
 118. Rigolin GM, Tieghi A, Ciccone M, Bragotti LZ, Cavazzini F, Della Porta M, et al. Soluble Urokinase-Type Plasminogen Activator Receptor (suPAR) as an Independent Factor Predicting Worse Prognosis and Extra-Bone Marrow Involvement in Multiple Myeloma Patients. *Br J Haematol* (2003) 120:953–9. doi: 10.1046/j.1365-2141.2003.04176.x
 119. Braybrooke JP, O'Byrne KJ, Propper DJ, Blann A, Saunders M, Dobbs N, et al. A Phase II Study of Razoxane, an Antiangiogenic Topoisomerase II Inhibitor, in Renal Cell Cancer With Assessment of Potential Surrogate Markers of Angiogenesis. *Clin Cancer Res* (2000) 6:4697–704.
 120. Schmidt M, Hoppe F. Increased Levels of Urokinase Receptor in Plasma of Head and Neck Squamous Cell Carcinoma Patients. *Acta Otolaryngol* (1999) 119:949–53. doi: 10.1080/00016489950180342
 121. Riisbro R, Christensen IJ, Piironen T, Greenall M, Larsen B, Stephens RW, et al. Prognostic Significance of Soluble Urokinase Plasminogen Activator Receptor in Serum and Cytosol of Tumor Tissue From Patients With Primary Breast Cancer. *Clin Cancer Res* (2002) 8:1132–41.
 122. Loosen SH, Gorgulho J, Jördens MS, Schulze-Hagen M, Beier F, Vucur M, et al. Serum Levels of Soluble Urokinase Plasminogen Activator Receptor Predict Tumor Response and Outcome to Immune Checkpoint Inhibitor Therapy. *Front Oncol* (2021) 11:646883. doi: 10.3389/fonc.2021.646883
 123. Jing J, Zheng S, Han C, Du L, Guo Y, Wang P. Evaluating the Value of uPAR of Serum and Tissue on Patients With Cervical Cancer. *J Clin Lab Anal* (2012) 26:16–21. doi: 10.1002/jcla.20499
 124. Riisbro R, Stephens RW, Brünner N, Christensen IJ, Nielsen HJ, Heilmann L, et al. Soluble Urokinase Plasminogen Activator Receptor in Preoperatively Obtained Plasma From Patients With Gynecological Cancer or Benign

- Gynecological Diseases. *Gynecol Oncol* (2001) 82:523–31. doi: 10.1006/gyno.2001.6324
125. Lomholt AF, Høyer-Hansen G, Nielsen HJ, Christensen IJ. Intact and Cleaved Forms of the Urokinase Receptor Enhance Discrimination of Cancer From Non-Malignant Conditions in Patients Presenting With Symptoms Related to Colorectal Cancer. *Br J Cancer* (2009) 101:992–7. doi: 10.1038/sj.bjc.6605228
 126. Usnarska-Zubkiewicz L, Strutyńska-Karpińska M, Zubkiewicz-Kucharska A, Zarebski P, Grabowski K. Soluble Urokinase-Type Plasminogen Activator Receptor and Ferritin Concentration in Patients With Advanced Alimentary Tract Carcinoma. Relationship to Localization, Surgical Treatment and the Stage of the Disease - Preliminary Report. *Adv Clin Exp Med* (2014) 23:959–67. doi: 10.17219/acem/30817
 127. Fidan E, Mentese A, Ozdemir F, Deger O, Kavgaci H, Caner Karahan S, et al. Diagnostic and Prognostic Significance of CA IX and suPAR in Gastric Cancer. *Med Oncol* (2013) 30:540. doi: 10.1007/s12032-013-0540-9
 128. Chounta A, Ellinas C, Tzanetakou V, Pliarhopoulou F, Mplani V, Oikonomou A, et al. Serum Soluble Urokinase Plasminogen Activator Receptor as a Screening Test for the Early Diagnosis of Hepatocellular Carcinoma. *Liver Int* (2015) 35:601–7. doi: 10.1111/liv.12705
 129. Rubio-Jurado B, Tello-González A, Bustamante-Chávez L, de la Peña A, Riebeling-Navarro C, Nava-Zavala AH. Circulating Levels of Urokinase-Type Plasminogen Activator Receptor and D-Dimer in Patients With Hematological Malignancies. *Clin Lymphoma Myeloma Leuk* (2015) 15:621–6. doi: 10.1016/j.clml.2015.07.632
 130. Henic E, Borgfeldt C, Christensen IJ, Casslén B, Høyer-Hansen G. Cleaved Forms of the Urokinase Plasminogen Activator Receptor in Plasma Have Diagnostic Potential and Predict Postoperative Survival in Patients With Ovarian Cancer. *Clin Cancer Res* (2008) 14:5785–93. doi: 10.1158/1078-0432.CCR-08-0096
 131. Enocsson H, Lukic T, Ziegelasch M, Kastbom A. Serum Levels of the Soluble Urokinase Plasminogen Activator Receptor (suPAR) Correlates With Disease Activity in Early Rheumatoid Arthritis and Reflects Joint Damage Over Time. *Transl Res* (2021) 232:142–9. doi: 10.1016/j.trsl.2021.02.007
 132. Portelli MA, Siedlinski M, Stewart CE, Postma DS, Nieuwenhuis MA, Vonk JM, et al. Genome-Wide Protein QTL Mapping Identifies Human Plasma Kallikrein as a Post-Translational Regulator of Serum uPAR Levels. *FASEB J* (2014) 28:923–34. doi: 10.1096/fj.13-240879
 133. Long D, Wang Y, Wang H, Wu X, Yu L. Correlation of Serum and Ascitic Fluid Soluble Form Urokinase Plasminogen Activator Receptor Levels With Patient Complications, Disease Severity, Inflammatory Markers, and Prognosis in Patients With Severe Acute Pancreatitis. *Pancreas* (2019) 48:335–42. doi: 10.1097/MPA.0000000000001247
 134. Wiese S, Mortensen C, Götze JP, Christensen E, Andersen O, Bendtsen F, et al. Cardiac and Proinflammatory Markers Predict Prognosis in Cirrhosis. *Liver Int* (2014) 34:e19–30. doi: 10.1111/liv.12428
 135. Sjöwall C, Martinsson K, Cardell K, Ekstedt M, Kechagias S. Soluble Urokinase Plasminogen Activator Receptor Levels Are Associated With Severity of Fibrosis in Nonalcoholic Fatty Liver Disease. *Transl Res* (2015) 165:658–66. doi: 10.1016/j.trsl.2014.09.007
 136. Hayek SS, Leaf DE, Samman Tahhan A, Raad M, Sharma S, Waikar SS, et al. Soluble Urokinase Receptor and Acute Kidney Injury. *N Engl J Med* (2020) 382:416–26. doi: 10.1056/NEJMoa1911481
 137. Iversen E, Houliand MB, Kallemose T, Rasmussen L JH, Hornum M, Feldt-Rasmussen B, et al. Elevated suPAR Is an Independent Risk Marker for Incident Kidney Disease in Acute Medical Patients. *Front Cell Dev Biol* (2020) 8:339. doi: 10.3389/fcell.2020.00339
 138. Meijers B, Poesen R, Claes K, Dietrich R, Bammens B, Sprangers B, et al. Soluble Urokinase Receptor Is a Biomarker of Cardiovascular Disease in Chronic Kidney Disease. *Kidney Int* (2015) 87:210–6. doi: 10.1038/ki.2014.197
 139. Schaefer F, Trachtman H, Wühl E, Kirchner M, Hayek SS, Anarat A, et al. Association of Serum Soluble Urokinase Receptor Levels With Progression of Kidney Disease in Children. *JAMA Pediatr* (2017) 171:e172914. doi: 10.1001/jamapediatrics.2017.2914
 140. Rovina N, Akinosoglou K, Eugen-Olsen J, Hayek S, Reiser J, Giamarellos-Bourboulis EJ. Soluble Urokinase Plasminogen Activator Receptor (suPAR) as an Early Predictor of Severe Respiratory Failure in Patients With COVID-19 Pneumonia. *Crit Care* (2020) 24:187. doi: 10.1186/s13054-020-02897-4
 141. Sevgi DY, Bayraktar B, Gündüz A, Özgüven BY, Togay A, Bulut E, et al. Serum Soluble Urokinase-Type Plasminogen Activator Receptor and Interferon- γ -Induced Protein 10 Levels Correlate With Significant Fibrosis in Chronic Hepatitis B. *Wien Klin Wochenschr* (2016) 128:28–33. doi: 10.1007/s00508-015-0886-4
 142. Sidenius N, Sier CF, Ullum H, Pedersen BK, Cozzi-Lepri A, Blasi F, et al. Serum Level of Soluble Urokinase-Type Plasminogen Activator Receptor Is a Strong and Independent Predictor of Survival in Human Immunodeficiency Virus Infection. *Blood* (2000) 96:4091–5. doi: 10.1182/blood.V96.13.4091
 143. Hoenigl M, Raggam RB, Wagner J, Valentini T, Leitner E, Seeber K, et al. Diagnostic Accuracy of Soluble Urokinase Plasminogen Activator Receptor (suPAR) for Prediction of Bacteremia in Patients With Systemic Inflammatory Response Syndrome. *Clin Biochem* (2013) 46:225–9. doi: 10.1016/j.clinbiochem.2012.11.004
 144. Wittenhagen P, Kronborg G, Weis N, Nielsen H, Obel N, Pedersen SS, et al. The Plasma Level of Soluble Urokinase Receptor Is Elevated in Patients With Streptococcus Pneumoniae Bacteraemia and Predicts Mortality. *Clin Microbiol Infect* (2004) 10:409–15. doi: 10.1111/j.1469-0691.2004.00850.x
 145. Østergaard C, Benfield T, Lundgren JD, Eugen-Olsen J. Soluble Urokinase Receptor Is Elevated in Cerebrospinal Fluid From Patients With Purulent Meningitis and Is Associated With Fatal Outcome. *Scand J Infect Dis* (2004) 36:14–9. doi: 10.1080/00365540310017366
 146. Savva A, Raftogiannis M, Baziaka F, Routsis C, Antonopoulou A, Koutoukas P, et al. Soluble Urokinase Plasminogen Activator Receptor (suPAR) for Assessment of Disease Severity in Ventilator-Associated Pneumonia and Sepsis. *J Infect* (2011) 63:344–50. doi: 10.1016/j.jinf.2011.07.016
 147. Donadello K, Scolletta S, Taccone FS, Covajes C, Santonocito C, Cortes DO, et al. Soluble Urokinase-Type Plasminogen Activator Receptor as a Prognostic Biomarker in Critically Ill Patients. *J Crit Care* (2014) 29:144–9. doi: 10.1016/j.jcrc.2013.08.005
 148. Rabna P, Andersen A, Wejse C, Oliveira I, Gomes VF, Haaland MB, et al. Utility of the Plasma Level of suPAR in Monitoring Risk of Mortality During TB Treatment. *PLoS One* (2012) 7:e43933. doi: 10.1371/journal.pone.0043933
 149. Perch M, Kofoed P, Fischer TK, Cò F, Rombo L, Aaby P, et al. Serum Levels of Soluble Urokinase Plasminogen Activator Receptor Is Associated With Parasitemia in Children With Acute Plasmodium Falciparum Malaria Infection. *Parasite Immunol* (2004) 26:207–11. doi: 10.1111/j.0141-9838.2004.00695.x
 150. Plewes K, Royakkers AA, Hanson J, Hasan MMU, Alam S, Ghose A, et al. Correlation of Biomarkers for Parasite Burden and Immune Activation With Acute Kidney Injury in Severe Falciparum Malaria. *Malar J* (2014) 13:91. doi: 10.1186/1475-2875-13-91
 151. Outinen TK, Tervo L, Mäkelä S, Huttunen R, Mäenpää N, Huhtala H, et al. Plasma Levels of Soluble Urokinase-Type Plasminogen Activator Receptor Associate With the Clinical Severity of Acute Puumala Hantavirus Infection. *PLoS One* (2013) 8:e71335. doi: 10.1371/journal.pone.0071335
 152. Ventorp F, Gustafsson A, Träskman-Bendz L, Westrin Å, Ljunggren L. Increased Soluble Urokinase-Type Plasminogen Activator Receptor (suPAR) Levels in Plasma of Suicide Attempters. *PLoS One* (2015) 10:e0140052. doi: 10.1371/journal.pone.0140052
 153. Nielsen J, Røge R, Pristed SG, Viuff AG, Ullum H, Thøner LW, et al. Soluble Urokinase-Type Plasminogen Activator Receptor Levels in Patients With Schizophrenia. *Schizophr Bull* (2015) 41:764–71. doi: 10.1093/schbul/sbu118
 154. Bigseth TT, Fredriksen M, Egeland J, Andersen E, Andreassen OA, Bang-Kittelsen G, et al. Elevated Levels of Soluble Urokinase Plasminogen Activator Receptor as a Low-Grade Inflammation Marker in Schizophrenia: A Case-Control Study. *Schizophr Res* (2021) 228:190–2. doi: 10.1016/j.schres.2020.11.051
 155. Giovannini S, Onder G, Liperoti R, Russo A, Carter C, Capoluongo E, et al. Interleukin-6, C-Reactive Protein, and Tumor Necrosis Factor-Alpha as Predictors of Mortality in Frail, Community-Living Elderly Individuals. *J Am Geriatr Soc* (2011) 59:1679–85. doi: 10.1111/j.1532-5415.2011.03570.x
 156. Proctor MJ, McMillan DC, Horgan PG, Fletcher CD, Talwar D, Morrison DS. Systemic Inflammation Predicts All-Cause Mortality: A Glasgow Inflammation Outcome Study. *PLoS One* (2015) 10:e0116206. doi: 10.1371/journal.pone.0116206
 157. Emerging Risk Factors Collaboration Kaptoge S, Di Angelantonio E, Lowe G, Pepys MB, Thompson SG, et al. C-Reactive Protein Concentration and Risk

- of Coronary Heart Disease, Stroke, and Mortality: An Individual Participant Meta-Analysis. *Lancet* (2010) 375:132–40. doi: 10.1016/S0140-6736(09)61717-7
158. Hayek SS, Sever S, Ko YA, Trachtman H, Awad M, Wadhvani S, et al. Soluble Urokinase Receptor and Chronic Kidney Disease. *N Engl J Med* (2015) 373:1916–25. doi: 10.1056/NEJMoal506362
 159. Enocsson H, Wirestam L, Dahle C, Padyukov L, Jönsen A, Urowitz MB, et al. Soluble Urokinase Plasminogen Activator Receptor (suPAR) Levels Predict Damage Accrual in Patients With Recent-Onset Systemic Lupus Erythematosus. *J Autoimmun* (2020) 106:102340. doi: 10.1016/j.jaut.2019.102340
 160. Persson M, Engström G, Björkbacka H, Hedblad B. Soluble Urokinase Plasminogen Activator Receptor in Plasma Is Associated With Incidence of CVD. Results From the Malmö Diet and Cancer Study. *Atherosclerosis* (2012) 220:502–5. doi: 10.1016/j.atherosclerosis.2011.10.039
 161. Törnkvist PBS, Haupt TH, Rasmussen LJH, Ladelund S, Toft U, Pisinger C, et al. Soluble Urokinase Plasminogen Activator Receptor Is Linearly Associated With Dietary Quality and Predicts Mortality. *Br J Nutr* (2019) 121:699–708. doi: 10.1017/S0007114518003720
 162. Borné Y, Persson M, Melander O, Smith JG, Engström G. Increased Plasma Level of Soluble Urokinase Plasminogen Activator Receptor Is Associated With Incidence of Heart Failure But Not Atrial Fibrillation. *Eur J Heart Fail* (2014) 16:377–83. doi: 10.1002/ehf.49
 163. Rasmussen LJH, Moffitt TE, Arseneault L, Danese A, Eugen-Olsen J, Fisher HL, et al. Association of Adverse Experiences and Exposure to Violence in Childhood and Adolescence With Inflammatory Burden in Young People. *JAMA Pediatr* (2020) 174:38–47. doi: 10.1001/jamapediatrics.2019.3875
 164. Baumeister D, Akhtar R, Ciufolini S, Pariante CM, Mondelli V. Childhood Trauma and Adulthood Inflammation: A Meta-Analysis of Peripheral C-Reactive Protein, Interleukin-6 and Tumour Necrosis Factor- α . *Mol Psychiatry* (2016) 21:642–9. doi: 10.1038/mp.2015.67
 165. Liu W, Zhang A, He H, Zhao X, Tao F, Sun Y. Inflammatory Burden in Adolescents With Prolonged Parent-Child Separation. *Brain Behav Immun* (2021) 98:257–62. doi: 10.1016/j.bbi.2021.08.227
 166. Valsdottir TD, Henriksen C, Odden N, Nellemann B, Jeppesen PB, Hisdal J, et al. Effect of a Low-Carbohydrate High-Fat Diet and a Single Bout of Exercise on Glucose Tolerance, Lipid Profile and Endothelial Function in Normal Weight Young Healthy Females. *Front Physiol* (2019) 10:1499. doi: 10.3389/fphys.2019.01499
 167. Kolho K-L, Valtonen E, Rintamäki H, Savilahti E. Soluble Urokinase Plasminogen Activator Receptor suPAR as a Marker for Inflammation in Pediatric Inflammatory Bowel Disease. *Scand J Gastroenterol* (2012) 47:951–5. doi: 10.3109/00365521.2012.699549
 168. Kasang C, Kalluvya S, Majinge C, Kongo G, Mlewa M, Massawe I, et al. Effects of Prednisolone on Disease Progression in Antiretroviral-Untreated HIV Infection: A 2-Year Randomized, Double-Blind Placebo-Controlled Clinical Trial. *PLoS One* (2016) 11:e0146678. doi: 10.1371/journal.pone.0146678
 169. Ascitto G, Edsfieldt A, Dias NV, Nilsson J, Prehn C, Adamski J, et al. Treatment With Beta-Blockers Is Associated With Lower Levels of Lp-PLA2 and suPAR in Carotid Plaques. *Cardiovasc Pathol* (2013) 22:438–43. doi: 10.1016/j.carpath.2013.04.005
 170. Hodges GW, Bang CN, Forman JL, Olsen MH, Boman K, Ray S, et al. Effect of Simvastatin and Ezetimibe on suPAR Levels and Outcomes. *Atherosclerosis* (2018) 272:129–36. doi: 10.1016/j.atherosclerosis.2018.03.030
 171. Tsai P-K, Tsao S-M, Yang W-E, Yeh C-B, Wang H-L, Yang S-F. Plasma Soluble Urokinase-Type Plasminogen Activator Receptor Level as a Predictor of the Severity of Community-Acquired Pneumonia. *Int J Environ Res Public Health* (2019) 16:1035. doi: 10.3390/ijerph16061035
 172. Ostrowski SR, Katzenstein TL, Piironen T, Gerstoft J, Pedersen BK, Ullum H. Soluble Urokinase Receptor Levels in Plasma During 5 Years of Highly Active Antiretroviral Therapy in HIV-1-Infected Patients. *J Acquir Immunodefic Syndr* (2004) 35:337–42. doi: 10.1097/00126334-200404010-00002
 173. Tavenier J, Haupt TH, Andersen AL, Buhl SF, Langkilde A, Andersen JR, et al. A High-Protein Diet During Hospitalization Is Associated With an Accelerated Decrease in Soluble Urokinase Plasminogen Activator Receptor Levels in Acutely Ill Elderly Medical Patients With SIRS. *Nutr Res* (2017) 41:56–64. doi: 10.1016/j.nutres.2017.04.006
 174. Biomarkers Definitions Working Group. Biomarkers and Surrogate Endpoints: Preferred Definitions and Conceptual Framework. *Clin Pharmacol Ther* (2001) 69:89–95. doi: 10.1067/mcp.2001.113989
 175. Wei C, El Hindi S, Li J, Fornoni A, Goes N, Sageshima J, et al. Circulating Urokinase Receptor as a Cause of Focal Segmental Glomerulosclerosis. *Nat Med* (2011) 17:952–60. doi: 10.1038/nm.2411
 176. Khater WS, Salah-Eldeen NN, Khater MS, Saleh AN. Role of suPAR and Lactic Acid in Diagnosing Sepsis and Predicting Mortality in Elderly Patients. *Eur J Microbiol Immunol* (2016) 6:178–85. doi: 10.1556/1886.2016.00011
 177. Kyriazopoulou E, Panagopoulos P, Metallidis S, Dalekos GN, Poulakou G, Gatselis N, et al. An Open Label Trial of Anakinra to Prevent Respiratory Failure in COVID-19. *Elife* (2021) 10:e66125. doi: 10.7554/eLife.66125
 178. Kyriazopoulou E, Poulakou G, Milonias H, Metallidis S, Adamis G, Tsiakos K, et al. Early Treatment of COVID-19 With Anakinra Guided by Soluble Urokinase Plasminogen Receptor Plasma Levels: A Double-Blind, Randomized Controlled Phase 3 Trial. *Nat Med* (2021) 27:1752–60. doi: 10.1038/s41591-021-01499-z
 179. Goronzy JJ, Weyand CM. Understanding Immunosenescence to Improve Responses to Vaccines. *Nat Immunol* (2013) 14:428–36. doi: 10.1038/ni.2588
 180. Aiello A, Farzaneh F, Candore G, Caruso C, Davinelli S, Gambino CM, et al. Immunosenescence and Its Hallmarks: How to Oppose Aging Strategically? A Review of Potential Options for Therapeutic Intervention. *Front Immunol* (2019) 10:2247. doi: 10.3389/fimmu.2019.02247
 181. Pera A, Campos C, López N, Hassouneh F, Alonso C, Tarazona R, et al. Immunosenescence: Implications for Response to Infection and Vaccination in Older People. *Maturitas* (2015) 82:50–5. doi: 10.1016/j.maturitas.2015.05.004
 182. Cunha LL, Perazzio SF, Azzi J, Cravedi P, Riella LV. Remodeling of the Immune Response With Aging: Immunosenescence and Its Potential Impact on COVID-19 Immune Response. *Front Immunol* (2020) 11:1748. doi: 10.3389/fimmu.2020.01748
 183. Tavenier J, Rasmussen LJH, Houliand MB, Andersen AL, Panum I, Andersen O, et al. Alterations of Monocyte NF- κ B p65/RelA Signaling in a Cohort of Older Medical Patients, Age-Matched Controls, and Healthy Young Adults. *Immun Ageing* (2020) 17:25. doi: 10.1186/s12979-020-00197-7
 184. Rhodes B, Fürnrohr BG, Vyse TJ. C-Reactive Protein in Rheumatology: Biology and Genetics. *Nat Rev Rheumatol* (2011) 7:282–9. doi: 10.1038/nrrheum.2011.37
 185. Sproston NR, Ashworth JJ. Role of C-Reactive Protein at Sites of Inflammation and Infection. *Front Immunol* (2018) 9:754. doi: 10.3389/fimmu.2018.00754
 186. Nilsson G, Lekander M, Åkerstedt T, Axelsson J, Ingre M. Diurnal Variation of Circulating Interleukin-6 in Humans: A Meta-Analysis. *PLoS One* (2016) 11:e0165799. doi: 10.1371/journal.pone.0165799
 187. Meier-Ewert HK, Ridker PM, Rifai N, Price N, Dinges DF, Mullington JM. Absence of Diurnal Variation of C-Reactive Protein Concentrations in Healthy Human Subjects. *Clin Chem* (2001) 47:426–30. doi: 10.1093/clinchem/47.3.426
 188. Marsland AL, Walsh C, Lockwood K, John-Henderson NA. The Effects of Acute Psychological Stress on Circulating and Stimulated Inflammatory Markers: A Systematic Review and Meta-Analysis. *Brain Behav Immun* (2017) 64:208–19. doi: 10.1016/j.bbi.2017.01.011
 189. Pepys MB, Hirschfield GM. *C-Reactive Protein: Crit Update J Clin Invest* (2003) 111:1805–12. doi: 10.1172/JCI18921
 190. Rana OR, Schröder JW, Koch A, Tacke F, Koos R, Schwinger RHG, et al. Soluble Urokinase Plasminogen Activator Receptor (suPAR): Its Relation to Neurological Outcome in Patients With Survived Cardiac Arrest. *IJC Metab Endocr* (2016) 12:8–13. doi: 10.1016/j.ijcme.2016.05.008
 191. Kofoed K, Andersen O, Kronborg G, Tvede M, Petersen J, Eugen-Olsen J, et al. Use of Plasma C-Reactive Protein, Procalcitonin, Neutrophils, Macrophage Migration Inhibitory Factor, Soluble Urokinase-Type Plasminogen Activator Receptor, and Soluble Triggering Receptor Expressed on Myeloid Cells-1 in Combination to Diagnose Infections. *Crit Care* (2007) 11:R38. doi: 10.1186/cc5723
 192. Parrinello CM, Lutsey PL, Ballantyne CM, Folsom AR, Pankow JS, Selvin E. Six-Year Change in High-Sensitivity C-Reactive Protein and Risk of

- Diabetes, Cardiovascular Disease, and Mortality. *Am Heart J* (2015) 170:380–9. doi: 10.1016/j.ahj.2015.04.017
193. Ridker PM, Rifai N, Rose L, Buring JE, Cook NR. Comparison of C-Reactive Protein and Low-Density Lipoprotein Cholesterol Levels in the Prediction of First Cardiovascular Events. *N Engl J Med* (2002) 347:1557–65. doi: 10.1056/NEJMoa021993
194. Bower JK, Lazo M, Juraschek SP, Selvin E. Within-Person Variability in High-Sensitivity C-Reactive Protein. *Arch Intern Med* (2012) 172:1519–21. doi: 10.1001/archinternmed.2012.3712
195. Bogaty P, Dagenais GR, Joseph L, Boyer L, Leblanc A, Bélisle P, et al. Time Variability of C-Reactive Protein: Implications for Clinical Risk Stratification. *PLoS One* (2013) 8:e60759. doi: 10.1371/journal.pone.0060759
196. Mac Giollabhui N, Ellman LM, Coe CL, Byrne ML, Abramson LY, Alloy LB. To Exclude or Not to Exclude: Considerations and Recommendations for C-Reactive Protein Values Higher Than 10 Mg/L. *Brain Behav Immun* (2020) 87:898–900. doi: 10.1016/j.bbi.2020.01.023
197. Muscatell KA, Brosso SN, Humphreys KL. Socioeconomic Status and Inflammation: A Meta-Analysis. *Mol Psychiatry* (2020) 25:2189–99. doi: 10.1038/s41380-018-0259-2
198. Alfano M, Cinque P, Giusti G, Proietti S, Nebuloni M, Danese S, et al. Full-Length Soluble Urokinase Plasminogen Activator Receptor Down-Modulates Nephron Expression in Podocytes. *Sci Rep* (2015) 5:13647. doi: 10.1038/srep13647
199. Hayek SS, Koh KH, Grams ME, Wei C, Ko Y-A, Li J, et al. A Tripartite Complex of suPAR, APOL1 Risk Variants and $\alpha\text{v}\beta\text{3}$ Integrin on Podocytes Mediates Chronic Kidney Disease. *Nat Med* (2017) 23:945–53. doi: 10.1038/nm.4362
200. Spinale JM, Mariani LH, Kapoor S, Zhang J, Weyant R, Song PX, et al. A Reassessment of Soluble Urokinase-Type Plasminogen Activator Receptor in Glomerular Disease. *Kidney Int* (2015) 87:564–74. doi: 10.1038/ki.2014.346
201. Caseley EA, Poulter JA, Rodrigues F. Immunome Project Consortium for Autoinflammatory Disorders (ImmunAID), Mcdermott MF. Inflammation Inhibition Under Physiological and Pharmacological Conditions. *Genes Immun* (2020) 21:211–23. doi: 10.1038/s41435-020-0104-x
202. Shields GS, Spahr CM, Slavich GM. Psychosocial Interventions and Immune System Function: A Systematic Review and Meta-Analysis of Randomized Clinical Trials. *JAMA Psychiatry* (2020) 77:1031–43. doi: 10.1001/jamapsychiatry.2020.0431
203. Pries-Heje MM, Hasselbalch RB, Raaschou H, Rezanavaz-Gheshlagh B, Heebøll H, Rehman S, et al. Utility of Multiple Rule Out CT Screening of High-Risk Atraumatic Patients in an Emergency Department — A Feasibility Study. *Emerg Radiol* (2018) 25:357–65. doi: 10.1007/s10140-018-1584-0
204. Velissaris D, Dimopoulos G, Parissis J, Alexiou Z, Antonakos N. Prognostic Role of Soluble Urokinase Plasminogen Activator Receptor at the Emergency Department: A Position Paper by the Hellenic Sepsis Study Group. *Infect Dis Ther* (2020) 9:407–16. doi: 10.1007/s40121-020-00301-w
205. Singer M, Deutschman CS, Seymour CW, Shankar-Hari M, Annane D, Bauer M, et al. The Third International Consensus Definitions for Sepsis and Septic Shock (Sepsis-3). *JAMA* (2016) 315:801–10. doi: 10.1001/jama.2016.0287
206. Joshi A, Rienks M, Theofilatos K, Mayr M. Systems Biology in Cardiovascular Disease: A Multiomics Approach. *Nat Rev Cardiol* (2021) 18:313–30. doi: 10.1038/s41569-020-00477-1
207. Hoogeveen RM, Pereira JPB, Nurmohamed NS, Zampoleri V, Bom MJ, Baragetti A, et al. Improved Cardiovascular Risk Prediction Using Targeted Plasma Proteomics in Primary Prevention. *Eur Heart J* (2020) 41:3998–4007. doi: 10.1093/eurheartj/ehaa648
208. Hayek SS, Raffield LM, Gao Y, Engstrom G, Quyyumi AA, Reiner AP, et al. Assay-Related Differences in SuPAR Levels: Implications for Measurement and Data Interpretation. *medRxiv* (2021). doi: 10.1101/2021.06.23.21259148 PREPRINT.
209. Abraham AG, Xu Y, Roem JL, Greenberg JH, Weidemann DK, Sabbisetti VS, et al. Variability in CKD Biomarker Studies: Soluble Urokinase Plasminogen Activator Receptor (suPAR) and Kidney Disease Progression in the Chronic Kidney Disease in Children (CKiD) Study. *Kidney Med* (2021) 3:712–21.e1. doi: 10.1016/j.xkme.2021.04.007
210. Winnicki W, Sunder-Plassmann G, Sengölge G, Handisurya A, Herkner H, Kornauth C, et al. Diagnostic and Prognostic Value of Soluble Urokinase-Type Plasminogen Activator Receptor (suPAR) in Focal Segmental Glomerulosclerosis and Impact of Detection Method. *Sci Rep* (2019) 9:13783. doi: 10.1038/s41598-019-50405-8
211. Joshi A, Mayr M. In Aptamers They Trust: The Caveats of the SOMAscan Biomarker Discovery Platform From SomaLogic. *Circulation* (2018) 138:2482–5. doi: 10.1161/CIRCULATIONAHA.118.036823

Conflict of Interest: JE-O is a named inventor on patents on suPAR as a prognostic biomarker. The patents are owned by Copenhagen University Hospital Amager and Hvidovre, Denmark, and is licensed to ViroGates A/S. JE-O is a co-founder, shareholder, and CSO of ViroGates A/S.

The remaining authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's Note: All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Copyright © 2021 Rasmussen, Petersen and Eugen-Olsen. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.