



Since January 2020 Elsevier has created a COVID-19 resource centre with free information in English and Mandarin on the novel coronavirus COVID-19. The COVID-19 resource centre is hosted on Elsevier Connect, the company's public news and information website.

Elsevier hereby grants permission to make all its COVID-19-related research that is available on the COVID-19 resource centre - including this research content - immediately available in PubMed Central and other publicly funded repositories, such as the WHO COVID database with rights for unrestricted research re-use and analyses in any form or by any means with acknowledgement of the original source. These permissions are granted for free by Elsevier for as long as the COVID-19 resource centre remains active.

- [12] AWAD, J., NAVARRETE, C. & FESTENSTEIN, H., Frequency and associations of HLA-DR and HLA-DQ antigens in a British Caucasoid panel. *Tissue Antigens*, 1987, **29**, 55-59.
- [13] MARCADET, A., MASSART, C., SEMANA, G., FAUCHET, R., SABOURAUD, O., MERIENNE, M., DAUSSET, J. & COHEN, D., Association of class II HLA DQ β -chain DNA restriction fragments with multiple sclerosis. *Immunogenet.*, 1985, **22**, 93-96.
- [14] COHEN, N., BRAUTBAR, C., FONT, M.-P., DAUSSET, J. & COHEN, D., HLA-DR2-associated Dw subtypes correlate with RFLP clusters: most DR2 IDDM patients belong to one of these clusters. *Immunogenet.*, 1986, **23**, 84-89.
- [15] DE MOERLOSE, P., JEANNET, M., MARTINS-DA-SILVA, B., WERNERFAVRE, C., ROHR, J. & GAUTHIER, G., Increased frequency of HLA-DRw2 and DRw3 in multiple sclerosis. *Tissue Antigens*, 1979, **13**, 357-360.
- [16] ZAMVIL, S.S., MITCHELL, D.J., MOORE, A.C., KITAMURA, K., STEINMAN, L. & ROTHBARD, J.B., T-cell epitope of the autoantigen myelin basic protein that induces encephalomyelitis. *Nature (Lond.)*, 1986, **324**, 258-260.
- [17] KONO, D.H., URBAN, J.L., HORVATH, S.J., ANDO, D.G., SAAVEDRA, R.A. & HOOD, L., Two minor determinants of myelin basic protein induce experimental allergic encephalomyelitis in SJL/J mice. *J. exp. Med.*, 1988, **168**, 213-227.
- [18] ZAMVIL, S.S., MITCHELL, D.J., POWELL, M.B., SAKAI, K., ROTHBARD, J.B. & STEINMAN, L., Multiple discrete encephalitogenic epitopes of the autoantigen myelin basic protein include a determinant for I-E class-II-restricted T cells. *J. exp. Med.*, 1988, **168**, 1181-1186.
- [19] HAPP, M.P., WETTSTEIN, P., DIETZSCHOLD, B. & HEBER-KATZ, E., Genetic control of the development of experimental allergic encephalomyelitis in rats. Separation of MHC and non-MHC gene effects. *J. Immunol.*, 1988, **141**, 1489-1494.
- [20] PULLEN, A.M., MARRACK, P. & KAPPLER, J.W., The T-cell repertoire is heavily influenced by tolerance to polymorphic self-antigens. *Nature (Lond.)*, 1988, **335**, 796-801.
- [21] URBAN, J.L., KUMAR, V., KONO, D.H., GOMEZ, C., HORVATH, S.J., CLAYTON, J., ANDO, D.G., SERCARZ, E.E. & HOOD, L., Restricted use of T-cell receptor V genes in murine autoimmune encephalomyelitis raises possibilities for antibody therapy. *Cell*, 1988, **54**, 577-592.

SITES OF ANTIGEN PRESENTATION IN T-CELL MEDIATED DEMYELINATING DISEASES

P.T. Massa

*National Institutes of Health, National Institute of Neurological Diseases and Stroke,
Neuroimmunology Branch, Building 10, Room 5B 16, 9000 Rockville Pike,
Bethesda, MD 20892 (USA)*

MS and animal models for demyelinating diseases.

Multiple sclerosis (MS) is a disease of the central nervous system (CNS) involving focal white matter lesions. The characteristic plaque consists of the loss of myelin sheaths and subsequent disappearance of oligodendrocytes, the cells that form and maintain myelin. A com-

mon feature of acute demyelinating lesions in MS is the presence of cellular infiltrates consisting of CD4⁺ and CD8⁺ T lymphocytes and monocytes in varying relative numbers (Traugott *et al.*, 1983). Current opinion is that demyelination in MS is immunologically mediated (McFarlin and McFarland, 1982).

Various experimental animal models of demyelination have provided insight into the possible mechanism of the immune-mediated process. These include experimental autoimmune encephalomyelitis (EAE) and virus-induced encephalomyelitis. In these animal models, T lymphocytes enter the brain and react with antigens presumably localized in white matter. The antigens which are responsible for EAE can be a number of myelin components, including myelin basic protein (MBP) (Paterson, 1976) or myelin proteolipid protein (PLP) (Williams *et al.*, 1982; Yoshimura *et al.*, 1985). Reactivity to these autoantigens may explain the selective destruction of myelin sheaths. In virus-induced demyelinating diseases, the degree of demyelination can often be correlated with the degree of cellular-mediated immune (CMI) response to viral antigens (Clatch *et al.*, 1987; Narayan *et al.*, 1985; Haase *et al.*, 1986), and not necessarily to myelin antigens. This results from either selective infection of oligodendrocytes with direct T-cell cytotoxicity of these cells (Haase *et al.*, 1986), or a "bystander" destruction of myelin. In the latter instance, virus antigen may be associated with cells other than oligodendrocytes, such as astrocytes, microglial cells or endothelial cells, also present in the white matter (Narayan *et al.*, 1985). Viral infections in the CNS can also be associated with auto-sensitization to myelin specific components, thereby complicating the disease process (Watanabe *et al.*, 1983; Johnson *et al.*, 1984; Liebert *et al.*, 1988).

Requirements for antigen presentation in the CNS.

T lymphocytes recognize self or foreign antigens that have been processed and are in direct physical association with either class I or class II major histocompatibility (MHC) molecules on the surface of antigen-presenting cells (APC).

Cells in the CNS are unique in that they do not express detectable levels of MHC molecules on their surfaces

(Schnitzer and Schachner, 1981; Williams *et al.*, 1980; Traugott, 1987). Therefore, APC function within the normal brain is a potential capacity. Inducible expression rather than constitutive expression is uniquely important for APC function within the CNS and rules governing antigen presentation in the periphery are probably different from those in brain. For this reason, tissue-specific mechanisms of MHC antigen expression in the brain are currently being investigated (Massa *et al.*, 1987a, b).

APC in the brain.

The cells that present antigen within the CNS will depend on (1) the self or foreign antigen involved, (2) MHC restriction of T cells reacting to a particular antigen, (3) the availability of the processed antigen to a particular APC, and (4) the capacity of the cells to express class I and/or class II MHC molecules. Expression of various accessory adhesion molecules such as LFA-1, LFA-3 and ICAM-1 may also be crucial (Springer *et al.*, 1987). Whether these molecules are constitutively expressed or are induced on neural cells, as are MHC molecules, is presently unknown.

Current evidence indicates that class I and II antigens are induced on distinct populations of cells in the brain. Both class I and II molecules are inducible on brain macrophages (microglia), astrocytes and endothelial cells. Interferon-gamma is probably the most effective inducer of MHC antigens on these cells (Massa *et al.*, 1987c; McCarron *et al.*, 1986; Wong *et al.*, 1985), however, other factors including viruses are effective as well (Massa *et al.*, 1986a; 1987b; Liu *et al.*, 1987; Suzumura *et al.*, 1986). Astrocytes and endothelial cells have been shown to be effective APC *in vitro* (Fontana *et al.*, 1986; McCarron *et al.*, 1986). Class II antigens are conspicuously lacking on oligodendrocytes (personal observations; Wong *et al.*, 1985) and neurons of laboratory animals exposed to various inducing agents, indicating that

oligodendrocytes cannot function as APC for class-I-restricted T cells. This indicates that the targets of class-II-restricted T cells, especially those that have been shown to mediate demyelination (Fontana *et al.*, 1984; Sun and Wekerle, 1988), are not oligodendrocytes or myelin but other brain APC, at least in rodents. Whether human oligodendrocytes can express class II molecules is unclear (Lisak *et al.*, 1983; Kim *et al.*, 1985).

In considering viral products as potential target antigens leading to myelin destruction, both animal and human studies indicate T-cell restriction by either class I or II molecules. In this case, the cells that present antigen depend on three factors: 1) cell-specific tropism of the virus, 2) availability of processed virus antigen, and 3) whether the reaction is primarily class-I- or II-restricted. A CMI to the virus that results in demyelination does not necessarily require that the virus is tropic for oligodendrocytes. Examples illustrating this point are Theiler's virus infection of mice (Clatch *et al.*, 1987), JHM corona virus infection of rats (Massa *et al.*, 1986b; Wege *et al.*, 1984) and lentivirus (visna virus) infection of sheep (Narayan *et al.*, 1985). Despite the observation that these viruses have no clear tropism for oligodendrocytes, induction of a CMI to the virus results in widespread demyelination. In the Theiler's virus model, susceptibility appears to be class-I-restricted (Clatch *et al.*, 1987; Rodriguez *et al.*, 1986), indicating that demyelination is related to class-I-restricted killing of APC other than oligodendrocytes, such as astrocytes and/or microglia, the primary targets of this virus. It is interesting that the JHM coronavirus in rats and visna virus in sheep also show strong tropism for these cells (Massa *et al.*, 1986b; Narayan *et al.*, 1982). Demyelination resulting from a CMI reaction to the virus is not always seen; however, LCM or measles infections in mice and rats (Oldstone *et al.*, 1986; Rammohan *et al.*, 1980; Liebert *et al.*, 1988) are exceptions.

Particular cells performing as APC may change with the course of remitting

attacks and chronic progression of the disease. For instance, prior to infiltration of T lymphocytes into prospective regions of demyelination, MHC antigens are absent presumably because of the unavailability of T-cell-derived lymphokines such as IFN- γ . In cases where demyelination is the outcome of viral infection, induction of MHC antigens on brain cells by the virus prior to T-cell infiltration is a possibility related to virus/neural cell interaction (Massa *et al.*, 1986a, 1987b; Suzumura *et al.*, 1986; Liu *et al.*, 1987). The pattern of virus-induced MHC antigen may be distinct from that induced by lymphokines, again restricting the APC function of certain cell types. For instance, neurotropic coronavirus (JHM strain) and measles virus selectively up-regulate class II MHC on rat astrocytes, but not on other cells, including "dedicated" APC such as resident brain macrophages (microglia). Also, depending on the virus, either class II alone (JHM coronavirus) or both class I and II (measles and flaviviruses) may be induced. Species-related differences have also been observed in virus induction of MHC molecules. JHM coronavirus induces class I in both mouse astrocytes and oligodendrocytes, but only MHC class II molecules on rat astrocytes. Following T-cell infiltration and interaction with virus-induced MHC molecules on glial cells, lymphokines are presumably produced in the CNS, leading to a greater diversity of cells expressing MHC molecules, depending on the lymphokines available. IFN- γ for instance, induces class I and II molecules on astrocytes, microglial cells and endothelial cells. However, other T-cell subsets apparently lack the ability to secrete IFN- γ , but secrete IL-4, a potent inducer of class II antigen on macrophages (Crawford *et al.*, 1987) and microglia, but not on astrocytes (personal observation). In addition, lymphotoxin and/or TNF generally amplify the effects of these MHC-inducing agents (Massa *et al.*, 1987b). One notable exception is murine endothelial cells in which MHC expression is apparently down-regulated by TNF (Tanaka, M., personal communica-

tions). Therefore, local secretion of TNF/lymphotoxin in the white matter may be an important factor in a CMI generation of MS lesions.

Studies of tissue from MS patients.

Investigation of CNS tissue from MS patients may provide evidence of the relevant APC in MS brains. With respect to autoreactivity to myelin components, there have been reports of T-cell sensitization to MBP and PLP in MS, but these studies have not been definitive. Moreover, in early MS plaques undergoing demyelination, oligodendrocytes appear to be spared (Prineas *et al.*, 1984), suggesting no direct insult to these cells in either a primary class I or II CTL attack. Also, astrocyte cytolysis (Sun and Wekerle, 1986) is not apparent in forming MS lesions, arguing against the possibility that CTL attack of these cells plays a role in the demyelinating process. This appears to leave CTL function, either class-I- or II-restricted, as the primary tissue insult, untenable at present. Rather, T cells restricted by either class I or class II MHC molecules may play a more traditional role in this disease compared to experimental models. The expression of class II on CNS cells may primarily mediate a T-helper function in MS. Perhaps, the focus of attention should not be placed on the ability of T cells to lyse various APC in the CNS, but to the secondary effects of lymphokines generated by these T cells, on macrophages and various neural cells, including oligodendrocytes.

The importance of class I and class II MHC-antigen-induction on astrocytes and endothelial cells in this process has been assessed in MS tissue (Traugott *et al.*, 1987). Class-I- and II-positive astrocytes are found primarily at the expanding edge of MS lesions where activated CD4⁺ and CD8⁺ T cells bearing IL-2 receptors are concentrated. The centre of lesions contains a paucity of T cells and numerous myelin-laden macrophages (Traugott *et al.*, 1987). This suggests that astrocytes

and endothelial cells are important in the initial site of antigen presentation.

A hypothesis for lesion formation.

Lesion formation in MS may be initiated by exposure of endothelial cells to lymphokines originating in the periphery, resulting in the induction of MHC molecules and accessory adhesion molecules on these cells. T cells specific for an antigen located in the brain (self or foreign) could then associate with brain endothelial cells, become activated, move into the brain and react with parenchymal APC. T-cell derived IFN- γ , IL-4, and lymphotoxin may function in concert to stimulate microglial cells and astrocytes in plaques. Lymphokine-activated microglial cells and blood-borne monocytes may effect non-specific tissue damage including demyelination, related to their increased oxidation potential, phagocytic capacity and secretion of proteolytic enzymes. This hypothesis is consistent with findings in EAE and some viral models where peripheral activation of T cells has occurred. Alternately, in the case where peripheral lymphokines are not available to induce MHC on endothelial cells, direct induction of class II and/or class I MHC molecules on astrocytes by viruses may also initiate a similar cascade of events leading to non-specific demyelination, primarily by macrophages. This concept that the initial disease process is controlled by class-II-MHC-positive astrocytes is supported by genetic studies showing differing inducibility of class II molecules on these cells from animals susceptible or resistant to demyelinating disease (Massa *et al.*, 1987a, c). It is interesting that endothelial cells from these rat strains show parallel differences (Male and Pryce, 1989). Whether direct T-cell lysis of astrocytes and/or endothelial cells leads to demyelination or whether there exists a more complex mechanism involving neural APC and subsequent macrophage microglia activation is not clear. Variability of brain-specific expression of MHC molecules among in-

dividuals and its possible correlation with disease susceptibility may provide needed information on the relevant site

of antigen presentation in the CNS and on current hypotheses of T-cell-mediated demyelination.

References.

- CLATCH, R.J., MELVOLD, R.W., DAL CANTO, M.C., MILLER, S.D. & LIPTON, H.L. (1987), The Theiler's murine encephalomyelitis virus (TMEV) model for multiple sclerosis shows a strong influence of the murine equivalents of HLA-A, B, and C. *J. Neuroimmunol.*, **15**, 121-135.
- CRAWFORD, R.M., FINBLOOM, D.S., O'HARA, J., PAUL, W.E. & MELTZER, M.S. (1987), B-cell stimulatory factor-1 (interleukin 4) activates macrophages for increased tumoricidal activity and expression of Ia antigens. *J. Immunol.*, **139**, 135-141.
- FONTANA, A., ERB, P., PIRCHER, H., ZINKERNAGEL, R., WEBER, E. & FIERZ, W. (1986), Astrocytes as antigen-presenting cells. Part II: unlike H-2K-dependent cytotoxic T cells, H-2 Ia-restricted T cells are only stimulated in the presence of interferon-gamma. *J. Neuroimmunol.*, **12**, 15-28.
- FONTANA, A., FIERZ, W. & WEKERLE, H. (1984), Astrocytes present myelin basic protein to encephalitogenic T-cell lines. *Nature (Lond.)*, **307**, 273-276.
- HAASE, A.T. (1986), Pathogenesis of lentivirus infections. *Nature (Lond.)*, **322**, 130-136.
- JOHNSON, R.T., GRIFFIN, D.E., HIRSCH, R.L., WOLINSKY, J.S., ROEDENBECK, S., LINDO DE SORIANO, I. & VAISBERG, A. (1984), Measles encephalomyelitis: clinical and immunological studies. *New Engl. J. Med.*, **310**, 137-141.
- KIM, S.U., MORETTO, G. & SHIN, D.H. (1985), Expression of Ia antigens on the surface of human oligodendrocytes and astrocytes in culture. *J. Neuroimmunol.*, **10**, 141-149.
- LIEBERT, U.G., LININGTON, C. & TER MEULEN, V. (1988), Induction of autoimmune reactions to myelin basic protein in measles virus encephalitis in Lewis rats. *J. Neuroimmunol.*, **17**, 103-118.
- LISAK, R., HIRAYAMA, M., KUCHMY, D., ROSENZEIG, A., KIM, S.U., PLEASURE, D. & SILBERBERG, D. (1983), Cultured human and rat oligodendrocytes and rat Schwann cells do not have immune response gene-associated antigen (Ia) on their surface. *Brain Res.*, **289**, 285-292.
- LIU, Y., KING, N., KESSON, A., BLANDEN, R.V. & MULLBACHER, A. (1987), Flavivirus infection modulates cell surface MHC antigen expression on astrocytes. *J. Neuroimmunol.*, **16**, 109a.
- MALE, D. & PRYCE, G. (1989), Induction of Ia molecules on brain endothelium is related to susceptibility to experimental allergic encephalomyelitis. *J. Neuroimmunol.*, **21**, 87-90.
- MASSA, P.T., BRINKMANN, R. & TER MEULEN, V. (1987a), Inducibility of Ia antigen on astrocytes by murine coronavirus JHM is rat-strain dependent. *J. exp. Med.*, **166**, 259-264.
- MASSA, P.T., DORRIES, R. & TER MEULEN, V. (1986a), Viral particles induce Ia antigen expression on astrocytes. *Nature (Lond.)*, **320**, 543-546.
- MASSA, P.T., SCHIMPL, A., WECKER, E. & TER MEULEN, V. (1987b), Tumor necrosis factor amplifies measles-virus-mediated Ia induction on astrocytes. *Proc. nat. Acad. Sci. (Wash.)*, **84**, 7242-7245.
- MASSA, P.T., TER MEULEN, V. & FONTANA, A. (1987c), Hyperinducibility of Ia antigen on astrocytes correlates with strain-specific susceptibility to experimental autoimmune encephalomyelitis. *Proc. nat. Acad. Sci. (Wash.)*, **84**, 4219-4223.
- MASSA, P.T., WEGE, H. & TER MEULEN, V. (1986b), Analysis of murine hepatitis virus (JHM strain) tropism toward Lewis rat glial cells *in vitro*: type I astrocytes and brain macrophages (microglia) as primary glial cell targets. *Lab. Invest.*, **55**, 318-327.
- MCCARRON, R.M., SPATZ, M., KEMPSKI, O., HOGAN, R.N., MEUHL, L. & MCFARLIN, D.E. (1986), Interaction between myelin-basic-protein-sensitized T lymphocytes and murine cerebral vascular endothelial cells. *J. Immunol.*, **137**, 3428-3435.
- MCFARLIN, D.E. & MCFARLAND, H.F. (1982), Multiple sclerosis. *New Engl. J. Med.*, **307**, 1183-1188 and 1246-1251.
- NARAYAN, O., WOLINSKI, J.S., CLEMENTS, J.E., STRANDBERG, J.D., GRIFFIN, D.E. & CORK, L.C. (1982), Slow virus replication: the role of macrophages in the persistence and expression of visna virus of sheep and goats. *J. gen. Virol.*, **59**, 345-356.

- OLDSTONE, M.B.A., BLOUNT, P., SOUTHERN, P.J. & LAMPERT, P.W. (1986), Cytoimmunotherapy for persistent virus infection reveals a unique clearance pattern from the central nervous system. *Nature (Lond.)*, **321**, 239-243.
- PATERSON, P.Y. (1976), Experimental autoimmune (allergic) encephalomyelitis, in "Textbook of Immunopathology" (P.A. Miescher and H.J. Mueller-Eberhard) (pp. 179-213). Grune and Stratton, New York.
- PRINEAS, J.W., KWON, E.E., CHO, E.-S. & SHARER, L.R. (1984), In "Multiple Sclerosis: experimental and clinical aspects" (L.C. Scheinberg and C.S. Raine) (pp. 11-33). Annals of the New York Academy of Sciences, New York.
- RAMMOHAN, K.W., MCFARLIN, D.E. & MCFARLAND, H.F. (1980), Chronic measles encephalitis in mice. *J. infect. Dis.*, **142**, 608-613.
- RODRIGUEZ, M., LEIBOWITZ, J. & DAVID, C.S. (1986), Susceptibility to Theiler's-virus-induced demyelination. Mapping of the gene within the H-2D region. *J. exp. Med.*, **163**, 620-631.
- SCHNITZER, J. & SCHACHNER, M. (1981), Expression of Thy-1, H-2 and NS-4 cell surface antigens and tetanus toxin receptors in early postnatal and adult mouse cerebellum. *J. Neuroimmunol.*, **1**, 429-456.
- SPRINGER, T.A., DUSTIN, M.L., KISHIMOTO, T.K. & MARLIN, S.D. (1987), The lymphocyte function-associated LFA-1, CD-2, and LFA-3 molecules: cell adhesion receptors of the immune system. *Ann. Rev. Immunol.*, **5**, 223-252.
- SUN, D. & WEKERLE, H. (1986), Ia-restricted encephalitogenic T lymphocytes mediating EAE lyse autoantigen-presenting astrocytes. *Nature (Lond.)*, **320**, 70-72.
- SUZUMURA, A., LAVI, E., WEISS, S.R. & SILBERBERG, D.H. (1986), Coronavirus infection induces H-2 antigen expression on oligodendrocytes and astrocytes. *Science*, **232**, 991-993.
- TRAUGOTT, U., REINHERZ, E.L. & RAINE, C.S. (1983), Multiple sclerosis: distribution of T cells, T-cell subsets and Ia-positive macrophages in lesions of different ages. *J. Neuroimmunol.*, **4**, 201-221.
- TRAUGOTT, U. (1987), Multiple sclerosis: relevance of class I and class II MHC-expressing cells to lesion development. *J. Neuroimmunol.*, **16**, 283-302.
- WATANABE, R., WEGE, H. & TER MEULEN, V. (1983), Adoptive transfer of EAE-like lesions from rats with coronavirus-induced demyelinating encephalomyelitis. *Nature (Lond.)*, **305**, 150-153.
- WEGE, H., WATANABE, R. & TER MEULEN, V. (1984), Relapsing subacute demyelinating encephalomyelitis in rats during the course of coronavirus JHM infection. *J. Neuroimmunol.*, **6**, 325-336.
- WILLIAMS, K.A., HART, D.N.J., FABRE, J.W. & MORRIS, P.J. (1980), Distribution and quantitation of HLA-ABC and DR (Ia) antigens on human kidney and other tissues. *Transplantation*, **29**, 274-279.
- WILLIAMS, R.M., LEES, M.B., CAMBI, F. & MACKLIN, W.B. (1982), Chronic experimental allergic encephalomyelitis induced in rabbits with bovine white matter proteolipid apoprotein. *J. Neuropath. Exp. Neurol.*, **41**, 508-521.
- WONG, G.H.W., BARTLETT, P.F., CLARK-LEWIS, I., MCKIMM-BRESHKIN, J.L. & SCHRADER, J.W. (1985), Interferon-gamma induces the expression of H-2 and Ia antigens on brain cells. *J. Neuroimmunol.*, **7**, 255-278.
- YOSHIMURA, T., KUNISHIHA, T., SAKAI, K., ENDOH, M., NAMIKOWA, T. & TABIRA, T. (1985), Chronic experimental allergic encephalomyelitis in guinea pigs induced by proteolipid protein. *J. Neurol. Sci.*, **69**, 47-58.