Exercise Diminishes Plasma Neurofilament Light Chain and Reroutes the Kynurenine Pathway in **Multiple Sclerosis**

Niklas Joisten, PhD,* Annette Rademacher,* Clemens Warnke, MD, Sebastian Proschinger, Alexander Schenk, PhD, David Walzik, Andre Knoop, PhD, Mario Thevis, PhD, Falk Steffen, Stefan Bittner, MD, Roman Gonzenbach, MD, Jan Kool, PhD, Wilhelm Bloch, MD, Jens Bansi, PhD,† and Philipp Zimmer, PhD, PhD†

Neurol Neuroimmunol Neuroinflamm 2021;8:e982. doi:10.1212/NXI.000000000000982

Abstract

Objective

To examine acute (single-bout) and training effects of high-intensity interval training (HIIT) vs standard exercise therapy (moderate continuous training [MCT]) on plasma neurofilament light chain (pNfL) and kynurenine (KYN) pathway of tryptophan degradation metabolites in persons with multiple sclerosis (pwMS).

Methods

Sixty-nine pwMS (Expanded Disability Status Scale score 3.0–6.0) were randomly assigned to a HIIT or an MCT group. Changes in pNfL and KYN pathway metabolites measured in blood plasma were assessed before, after, and 3 hours after the first training session as well as after the 3-week training intervention.

Results

Acute exercise reduced pNfL and increased the KYN pathway flux toward the neuroprotective kynurenic acid (KA). Changes in pNfL correlated positively with changes in KA and negatively with the quinolinic acid-to-KA ratio. HIIT consistently led to greater effects than MCT. Following the 3-week training intervention, the KYN pathway was activated in HIIT compared with MCT.

Conclusion

Future studies and clinical assessments of pNfL should consider acute exercise as confounding factor for measurement reliability. Moreover, exercise-induced KYN pathway rerouting might mediate neuroprotection, potentially underlying the benefits in rehabilitation for pwMS.

Classification of Evidence

This study provides Class II evidence that acute HIIT diminishes pNfL and increases KA levels, and 3 weeks of HIIT activate the KYN pathway in pwMS.

Trial Registration Information

Clinical trial registration number: NCT03652519.

Go to Neurology.org/NN for full disclosures. Funding information is provided at the end of the article.

Correspondence Dr. Zimmer philipp.zimmer@tu-dortmund.de

MORE ONLINE

Class of Evidence Criteria for rating therapeutic and diagnostic studies NPub.org/coe

^{*}These authors contributed equally to this work.

[†]These authors share senior authorship.

From the Department of "Performance and Health (Sports Medicine)" (N.J., A.S., D.W., P.Z.), Institute of Sport and Sport Science, Technical University Dortmund; Department for Molecular and Cellular Sports Medicine (A.R., S.P., W.B.), Institute of Cardiovascular Research and Sports Medicine, German Sport University Cologne; Department of Neurology (C.W.), Faculty of Medicine and University Hospital Cologne, University of Cologne; Center for Preventive Doping Research/Institute of Biochemistry (A.K., M.T.), German Sport University Cologne; Department of Neurology (F.S., S.B.), Focus Program Translational Neuroscience (FTN), and Immunotherapy (FZI), University Medical Center of the Johannes Gutenberg University, Germany; and Department of Neurology (R.G., J.K., J.B.), Clinics of Valens, Rehabilitation Centre Valens, Switzerland.

The Article Processing Charge was funded by the authors.

This is an open access article distributed under the terms of the Creative Commons Attribution-NonCommercial-NoDerivatives License 4.0 (CC BY-NC-ND), which permits downloading and sharing the work provided it is properly cited. The work cannot be changed in any way or used commercially without permission from the journal.

Glossary

CV = coefficient of variation; **EDSS** = Expanded Disability Status Scale; **HIIT** = high-intensity interval training; **HPLC-MS**/ **MS** = high-performance liquid chromatography–tandem mass spectrometry; **IL-6** = interleukin-6; **KA** = kynurenic acid; **KYN** = kynurenine; **MCT** = moderate continuous training; **pNfL** = plasma neurofilament light chain; **pwMS** = persons with multiple sclerosis; **QA** = quinolinic acid; **TRP** = tryptophan.

Plasma neurofilament light chain (pNfL) has become an integrative biomarker of neurodegeneration in persons with multiple sclerosis (pwMS). Higher levels reflect axonal loss and correlate with acute inflammation and disability progression.¹ The pNfL level is associated with various disease specific clinical and MRI-related outcomes such as acute relapses, T2 lesion load, or future brain atrophy.^{2,3}

The kynurenine (KYN) pathway of tryptophan (TRP) degradation is involved in different neuroinflammatory and neurodegenerative processes.⁴ In pwMS, the KYN pathway is deregulated and has recently attracted extensive attention as therapeutic target.⁵ Persistent overactivations of the KYN pathway are indicated by an elevated KYN/TRP ratio compared with healthy controls.⁶

Recently, 2 studies uncovered a relationship between NfL and the KYN pathway in the context of neurodegeneration. In a cohort of older adults, pNfL levels were positively associated with several KYN pathway metabolites.⁷ In pwMS, NfL levels correlated positively with the neurotoxic KYN pathway metabolite quinolinic acid (QA) in the CSF.⁸

Physical exercise alleviates various symptoms in pwMS, and disease-modifying effects are discussed.⁹ Especially concerning physical fitness and disease-associated biomarkers, high-intensity interval training (HIIT) leads to greater improvements than classical moderate continuous training (MCT).^{10–12} A better understanding of exercise-induced mechanisms would improve our understanding of disease-modifying effects and help to develop targeted exercise recommendations for pwMS.

Here, we (1) investigate the influence of exercise on pNfL in pwMS and (2) shed light on the association between exercise-induced alterations of the KYN pathway and pNfL.

Methods

This study is a secondary analysis of a prospective single-blind randomized controlled trial that was conducted in the inpatient rehabilitation clinic Valens (Switzerland).¹³ The study was approved by the regional ethics committee (EKOS18/96; Project-ID: 2018–01378) and registered at ClinicalTrials.gov (NCT03652519; August 29, 2018). Written informed consent was obtained from all participating patients. Detailed information on eligibility criteria can be depicted elsewhere.¹³ Briefly, the inclusion criteria comprised a definite diagnosis of MS, age >21 years, a relapsing-remitting or secondary progressive MS phenotype, and

an Expanded Disability Status Scale (EDSS) score between 3 and 6. Main exclusion criteria were concomitant diseases, pregnancy, alcohol or drug abuse, and a second participation in this investigation. In total, 69 pwMS with either relapsing-remitting (n = 42) or secondary progressive (n = 27) phenotype and a mean \pm SD EDSS score of 4.51 \pm 1.06 were included. Participant baseline characteristics are shown in table 1. Baseline cognitive performance of participants was assessed using the Brief International Cognitive Assessment in Multiple Sclerosis¹⁴ to verify an association between baseline pNfL levels and a clinical outcome within our sample. The assessment of cognitive performance was conducted within 24 hours after the baseline blood sample collection.

Participants were randomized with concealed allocation in either a HIIT or a MCT (standard therapy) group using physical fitness, EDSS score, age, and fatigue as stratification factors.

Groups and Interventions

Participants of both groups conducted a 3-week training intervention during inpatient rehabilitation with 3 exercise sessions per week. The individual exercise intensity was calculated based on the maximum heart rate achieved in an incremental exercise test until symptom-limited exhaustion before the intervention. Each session in both groups included a warm-up and cool-down period of 3 minutes. Although the participants of the MCT group exercised continuously for 24 minutes at an intensity of 65% of the maximum heart rate, the participants of the HIIT group conducted 5 high-intensity intervals of 1.5 minutes at 95%–100% of their maximum heart rate with 2 minutes of unloaded pedaling in between.

Outcomes and Assessments

To investigate acute (single-bout) and training (3-week intervention) effects, blood samples were collected immediately before (t_0) , immediately after (t_1) , and 3 hours after the first exercise session (t_2) as well as after the training intervention (t_3) . At t_0 and t_3 , blood samples were collected in a resting condition, without any amount of cardiorespiratory arousal due to exercise in the past 24 hours. Blood samples were taken between 8 and 9 AM. Participants were told to refrain from any nutritional intake before blood samples were collected.

After blood samples were drawn from the medial cubital vein in a supine position, samples were rested at room temperature for 10 minutes for clotting. Subsequently, samples were centrifuged at 3,500 rpm for 10 minutes. Plasma samples were aliquoted and stored at -80° C until further analyses. All outcome assessors were blinded to clinical data and intervention group allocation.

Table 1 Participant Baseline Characteristics

	HIIT (n = 35)	MCT (n = 34)	Overall (n = 69)
Sex (F/M)	24/11	19/15	43/26
Age (y)	50.89 (10.31)	49.65 (10.04)	50.28 (10.12)
Height (cm)	170.94 (8.57)	171.79 (7.56)	171.19 (8.15)
Weight (kg)	73.85 (14.60)	74.90 (16.30)	74.37 (15.36)
BMI (kg·m ⁻²)	25.28 (4.09)	25.26 (4.76)	25.27 (4.40)
EDSS score	4.44 (1.06)	4.59 (1.08)	4.51 (1.06)
MS phenotype (RRMS/SPMS)	21/14	21/13	42/27
Time since diagnosis (y)	14.8 (8.26)	12.24 (7.5)	13.54 (7.94)
DMT (yes/no) (%)	78.8/24.2	81.8/18.2	79.1/20.9
Years since last relapse (y)	5.32 (4.59)	4.53 (3.03)	4.95 (3.92)
VO2 _{peak} (mL∙min ^{−1})	1,416.06 (448.71)	1,405.79 (402.17)	1,411.00 (423.31)
VO2 _{peak} (mL·kg ⁻¹ ·min ⁻¹)	19.30 (5.64)	19.09 (5.13)	19.20 (5.36)
Watt _{max} (W)	98.91 (39.82)	96 (32.23)	97.48 (36.04)
Watt _{rel} (W·kg ⁻¹)	1.36 (0.55)	1.31 (0.45)	1.34 (0.50)
SDMT (points)	44.14 (10.30)	42.88 (14.07)	43.53 (12.20)
VLMT (total score. points)	52.43 (8.74)	51.73 (11.71)	52.09 (10.22)
BVMT-R (total score. points)	21.03 (7.33)	19.55 (7.67)	20.31 (7.48)

Abbreviations: BMI = body mass index; BVMT-R = Brief Visuospatial Memory Test–Revised; DMT = disease-modifying treatment; EDSS = Expanded Disability Status Scale; HIIT = high-intensity interval training; MCT = moderate continuous training; RRMS = relapsing-remitting MS; SDMT = Symbol Digit Modalities Test, SPMS = secondary progressive MS; VLMT = verbal learning memory test; VO_{2peak} = peak oxygen uptake; Watt_{max} = peak power output; Watt_{rel} = relative (referring to bodyweight) peak power output. Values are presented as mean (SD).

pNfL Assessment

As described for serum measurements,^{15,16} the pNfL level was determined by means of a SiMoA HD-1 device (Quanterix, Billerica, MA) using NF-Light Advantage Kits (Quanterix) of a single LOT according to the manufacturer's instructions. Although both the samples and the remaining kit components were allowed to come to room temperature, resorufin-β-Dgalactopyranoside was incubated at 33°C for 60 minutes. Samples were then vortexed and centrifuged for 5 minutes at 10,000g. Finally, all samples were measured in duplicates, and the coefficient of variation (CV, as a percentage) was obtained by dividing the SD by the mean value of both replicates multiplied by 100. Samples with a sample CV above 20% (or missing replicate result) were measured a second time. The mean intra-assay CV was 7.3%. Run-to-run variation was monitored by adding duplicates of the same 2 low and high controls, consisting of recombinant human NfL antigen, to each sample run. The mean levels over all runs were 3.0 pg/ mL for the low control and 137.9 pg/mL for the high control with interassay CVs of 4.4% and 1.5%, respectively.

Assessment of KYN Pathway Metabolites

Plasma concentration of TRP, KYN, kynurenic acid (KA), and QA was assessed using high-performance liquid chromatography-tandem mass spectrometry (HPLC-MS/ MS) according to previously published protocols^{17,18} after precipitation of plasma proteins. To provide information on the KYN pathway regulation, the ratios of KYN/TRP, KA/KYN, QA/KYN, and QA/KA were calculated and defined as outcomes. Detailed information on HPLC-MS/MS is provided in supplement 1 (links.lww.com/NXI/A455).

Assessment of Interleukin-6

Plasma levels of interleukin-6 (IL-6) were assessed using ELISA (Human IL-6 Quantikine HS ELISA Kit, R&D Systems). Analysis was conducted according to the manufacturer's instructions. Levels of IL-6 were additionally assessed to consider an inflammatory marker that is relevant as KYN pathway mediator and myokine, which is known to be influenced by physical exercise.

Statistical Analyses

Correlation coefficients were calculated to examine potential baseline associations between pNfL and cognitive performance as well as between IL-6, pNfL, and KYN pathway outcomes. To evaluate acute within- and between-group effects, baselineand IL-6-adjusted analysis of covariance models with repeated measures was performed. Sphericity was checked and adjusted using Greenhouse-Geisser correction if necessary. In case of significant main effects, Bonferroni-corrected post hoc comparisons were applied. Based on the observed acute

Figure 1 Baseline Correlations (Spearman Coefficient) Between Cognitive Performance and pNfL Levels



(A) Correlation between processing speed and pNfL levels. (B) Correlation between verbal learning and pNfL levels. (C) visuospatial memory and pNfL levels. BVMT-R = Brief Visuospatial Memory Test–Revised; pNfL = plasma neurofilament light chain; SDMT = Symbol Digit Modalities Test; VLMT = Verbal Learning and Memory Test.

effects on pNfL and the neuroprotective KYN pathway branch toward KA, we performed delta correlations subsequently. To evaluate training effects, postintervention values were compared between groups in univariate baseline- and IL-6 adjusted analysis of covariance models. Outliers defined as >3 SD were excluded before analyses. Level of significance was set at $p \le 0.05$. Statistical analyses were conducted using SPSS 26 (IBM, Armonk, NY).

Data Availability

Data from this study are available from the authors on reasonable request.

Results

Baseline Associations Between pNfL Levels, Cognitive Performance, and KYN Pathway Outcomes

Baseline correlations indicated negative associations between pNfL and processing speed (Symbol Digit Modalities Test) (r = -0.298; p = 0.015) as well as visuospatial memory (Brief Visuospatial Memory Test–Revised) (r = -0.260, p = 0.035), whereas the association between pNfL and verbal learning (Verbal Learning and Memory Test) did not reach statistical significance (figure 1).

Table 2 Baseline Correlations Between IL-6, pNfL, and KYN Pathway Outcomes



Graphic representation of Spearman's correlation coefficient.

Abbreviations: IL-6 = interleukin-6; KA = kynurenic acid; KA/KYN = kynurenic acid-to-kynurenine ratio; KYN = kynurenine; KYN/TRP = kynurenine-to-tryptophan ratio; pNfL = plasma neurofilament light chain; QA = quinolinic acid; QA/KA = quinolinic acid-to-kynurenic acid ratio; QA/KYN = quinolinic acid-to-kynurenine ratio; TRP = tryptophan. * $p \le 0.05$; ** $p \le 0.01$.



Figure 2 Acute Kinetics of pNfL (A), IL-6 (B), and KYN Pathway Outcomes (C-J) Between High-Intensity Interval Training vs Moderate Continuous Training

Data shown as baseline- and IL-6-adjusted outcome kinetics (mean \pm SEM) separated by intervention groups. #Significant between-group effect. IL-6 = interleukin-6; KA = kynurenic acid; KA/KYN = kynurenic acid-to-kynurenine ratio; KYN = kynurenine; KYN/TRP = kynurenine-to-tryptophan ratio; pNfL = plasma neurofilament light chain; QA = quinolinic acid; QA/KA = quinolinic acid-to-kynurenic acid ratio; QA/KYN = quinolinic acid-to-kynurenic acid-to-kynurenic acid ratio; QA/KYN = quinolinic acid-to-kynurenine; tr₁ = immediately postexercise; t₂ = 3 hours postexercise; TRP = tryptophan.

Levels of IL-6 correlated positively with KYN (r = 0.312; p = 0.011), KYN/TRP ratio (r = 0.306; p = 0.013), and QA (r = 0.336; p = 0.006). A negative correlation between pNfL and the KA/KYN ratio (r = -0.271; p = 0.029) was observed, whereas pNfL correlated positively with the QA/KA ratio (r = 0.256; p = 0.040). Baseline correlations between IL-6, pNfL, and KYN pathway outcomes are illustrated in table 2.

Acute Effects of HIIT Versus MCT on pNfL and KYN Pathway Outcomes

Levels of pNfL decreased within MCT from baseline to immediately postexercise (p < 0.001; CI [-0.940 to -0.237]) and subsequently increased to 3 hours postexercise (p = 0.022; CI [0.104–1.711]). pNfL decreased from baseline to immediately postexercise (p < 0.001; CI [-1.555 to -0.830]) and 3 hours postexercise (p = 0.009; CI [-1.961 to -0.219]) within HIIT. A significant interaction effect between the groups was observed at both measurement time points, immediately postexercise (p = 0.005; CI [-1.015 to -0.193) and 3 hours postexercise (p = 0.006; CI [-2.396 to -0.422]), with lower pNfL levels in HIIT compared with MCT.

The acute bout of exercise led to an increase in IL-6 within MCT from baseline to 3 hours postexercise (p < 0.001; 95% CI [0.290–1.008]) and from immediately postexercise to 3 hours postexercise (p = 0.001; CI [0.197–0.964]). Within HIIT, IL-6 increased from baseline to 3 hours postexercise (p = 0.029; CI [0.030–0.748]). No interaction effect between groups was observed.

Levels of TRP decreased within MCT from baseline to 3 hours postexercise (p = 0.006; CI [-1.933 to -00.262]). Within HIIT, TRP decreased from baseline to immediately postexercise (p = 0.048; CI [-1.230 to -0.004]) as well as to 3 hours postexercise (p = 0.019; CI [-1.822 to -0.126]). No interaction effects were observed for TRP. Levels of KYN

Table 3 Delta Correlations Between Acute Changes in pNfL (t₂-t₀) and in KYN Pathway Outcomes Toward Kynurenic Acid



Graphic representation of Spearman's correlation coefficient.

Abbreviations: Δ = delta; IL-6 = interleukin-6; KA = kynurenic acid; KA/KYN = kynurenic acid-to-kynurenine ratio; pNfL = plasma neurofilament light chain; QA/KA = quinolinic acid-to-kynurenic acid ratio; t₀ = baseline; t₁ = immediately postexercise; t₂ = 3 hours postexercise.

p* ≤ 0.05; *p* ≤ 0.01.

decreased from baseline to 3 hours postexercise within MCT (p = 0.031; CI [-42.133 to -1.483]) as well as within HIIT (p = 0.044; CI [-40.425 to -0.401]), but no effects between the groups were observed. Regarding KYN/TRP ratio, QA, and QA/KYN ratio, neither significant withinnor between-group effects were observed. Levels of KA decreased within MCT from baseline to 3 hours postexercise (p = 0.014; CI [-1.600 to -0.143]) as well as from immediately postexercise to 3 hours postexercise (p =0.010; CI [-1.603 to -0.171]). Within HIIT, KA increased from baseline to immediately postexercise (p = 0.001; CI [0.273–1.230]). Between-group differences were observed immediately postexercise (p = 0.009; CI [-0.1.281 to -0.190 and 3 hours hours postexercise (p = 0.024; CI [-1.820 to -0.133), with greater KA levels in HIIT than in MCT. The KA/KYN ratio increased within HIIT from baseline to immediately postexercise (p = 0.002; CI [0.001–0.004]), whereas no other significant effects within or between the groups were detected. The QA/KA ratio increased significantly in MCT from baseline to 3 hours postexercise (p = 0.035; CI [0.074–2.613]) as well as from postexercise to 3 hours postexercise (p = 0.024; CI [0.137–2.590]). Within HIIT, the QA/KA ratio decreased from baseline to immediately postexercise (p = 0.004; CI [-1.881 to -0.285]) and subsequently increased to 3 hours postexercise (p = 0.049; CI [0.002–2.535]). No betweengroup effects for the QA/KA ratio were observed. Acute kinetics of HIIT vs MCT on IL-6, pNfL, and KYN pathway outcomes are shown in figure 2.

Correlations of Acute Changes in pNfL and the KYN Pathway Toward KA

Delta correlations (table 3) of acute exercise-induced alterations indicated a negative association between the changes in KA (r = -0.254; p = 0.043) and the KA/KYN ratio (r = -0.264; p = 0.037) from baseline to immediately postexercise and changes in pNfL from baseline to 3 hours postexercise. Moreover, positive correlations between the change in pNfL from baseline to 3 hours postexercise and changes in the QA/ KA ratio (Δt_1 - $t_0 r = 0.412$; p = 0.001 and Δt_2 - $t_0 r = 0.304$; p = 0.016) were revealed.

Training Effects on pNfL Levels and KYN Pathway Outcomes

Concerning training effects, a significant difference between HIIT and MCT was observed following the 3-week intervention period (figure 3). The KYN/TRP ratio was significantly greater in HIIT compared with MCT (p = 0.032). No significant training effects were observed for pNfL, IL-6, TRP, KYN, QA, KA, QA/KYN ratio, KA/KYN ratio, and QA/KA ratio. Detailed analysis of covariance results for both acute and training effects are reported in supplement 2 (links.lww.com/NXI/A456). Raw data shown as mean ± SD are provided in supplement 3 (links. lww.com/NXI/A457).

Classification of Evidence

This secondary investigation was designed to examine acute and training effects of HIIT as the intervention of interest compared with standard exercise therapy on pNfL levels and KYN pathway metabolites in moderately disabled pwMS. In addition, we exploratory investigated a potential association between changes in pNfL levels and changes in the KYN pathway flux toward the neuroprotective KA.

This randomized controlled clinical interventional trial provides class II evidence that acute HIIT diminishes pNfL and increases the metabolic flux of the KYN pathway toward KA. Moreover, this study provides Class II evidence that a 3-week HIIT intervention increases the KYN/TRP ratio as marker of KYN pathway activation compared with standard exercise therapy in pwMS.

Discussion

Here, we showed that acute exercise reduces pNfL levels in pwMS in an intensity-dependent manner. This pNfL reduction is associated with KYN pathway rerouting toward the neuroprotective and immunosuppressive metabolite KA.

С В D A 50 8 25 800 40 20 pNfL (pg/mL) 6 600 FRP (µg/mL) KYN (ng/mL) IL-6 (pg/mL) 30 15 4 400 20 10 2 200 10 5 0 Е F G Н p = 0.0320.08 200 20 0.4 0.06 0.3 150 **KYN/TRP ratio** 15 QA/KYN ratio QA (ng/mL) KA (ng/mL) 0.2 100 10 0.04 50 0.02 0.1 5 C 0 0.00 0.0 I J 0.04 35



Data shown as postexercise intervention values (t₃) separated by intervention groups. Illustrated significance is based on baseline- and IL-6-adjusted ANCOVA results. ANCOVA = analysis of covariance; IL-6 = interleukin-6; KA = kynurenic acid; KA/KYN = kynurenic acid-to-kynurenine ratio; KYN = kynurenine; KYN/TRP = kynurenine-to-tryptophan ratio; pNfL = plasma neurofilament light chain; QA = quinolinic acid; QA/KA = quinolinic acid-to-kynurenic acid ratio; QA/ KYN = quinolinic acid-to-kynurenine ratio; TRP = tryptophan.

HIIT

MCT

Baseline pNfL levels correlated negatively with participants' cognitive performance, underlining the integrative nature and clinical relevance of pNfL as peripheral blood biomarker in pwMS. Baseline pNfL levels were also associated with KYN pathway parameters. Although the KA/KYN ratio correlated negatively, the QA/KA ratio correlated positively with pNfL levels. This observation fits into the literature that attributes KA as neuroprotective and QA as neurotoxic agent of the KYN pathway, respectively.¹⁹ Furthermore, we showed consistent baseline correlations between IL-6 levels and KYN pathway activation as well as between IL-6 and QA. These associations indicate a link between the inflammatory state and KYN pathway activation toward QA within this sample of pwMS.

28

14

7

0

QA/KA ratio 21

The results show that both acute HIIT and MCT reduce pNfL levels, but HIIT induces a greater reduction compared with MCT. Levels of pNfL were still decreased 3 hours after exercise completion in HIIT suggesting a more sustainable effect. Further mechanistic investigations focusing on the neuroprotective effects of single exercise bouts as reflected by

the decrease in pNfL following acute exercise in pwMS are highly warranted to improve our knowledge about how exercise-induced benefits on symptoms and clinical outcomes in neurodegenerative diseases are achieved. Moreover, future clinical trials as well as routine assessments of pNfL should consider acute physical exercise as confounding factor for measurement reliability.

Regarding acute effects on the KYN pathway, we observed a catabolic shift in favor of the end product KA that is in line with previous investigations in other populations.¹⁷ Greater effects of acute exercise on KA levels were observed in HIIT compared with MCT at both postexercise measurement time points. This finding confirms the results of previous studies in both animals and humans showing an elevated KAT expression in skeletal muscle and immune cells.^{17,20} An increase in KA is possibly not only important for neuroprotective properties within the CNS but also concerning its immunomodulatory effects. KA represents a well-described endogenous aryl hydrocarbon receptor (AhR) ligand, which mediates the

0.03

0.02

0.01

0.00

KA/KYN ratio





(A) Interaction of chronic inflammatory conditions in MS, the KYN pathway, and neurodegeneration assessed by pNfL levels. (B) Exercise-induced rerouting of the KYN pathway toward KA leads to reductions in pNfL levels. KA = kynurenic acid; KYN = kynurenine; pNfL = plasma neurofilament light chain; QA = quinolinic acid; TRP = tryptophan.

differentiation of naive CD4⁺ T-helper cells to immunosuppressive regulatory T cells following ligand-dependent activation.²¹ This increased AhR ligand availability in blood plasma provides the foundation for upcoming trials.

Furthermore, the acute exercise-induced changes in KA and in pNfL were associated negatively. We also observed a positive association between changes in pNfL and in the QA/KA ratio. These findings suggest the hypothesis that the short-term increase in KA and consequently the shift of the QA/KA balance leads to neuroprotection as reflected by the reduction in pNfL levels (figure 4).

In contrast to profound acute effects on pNfL and the KYN pathway flux toward KA, solely one significant alteration following the training period was observed. Compared with MCT, an elevated KYN/TRP ratio as parameter of initial KYN pathway activation was detected in HIIT that is in line with results from a previous study.²² A recent EAE mouse model revealed that peripheral IDO boosting for the purpose of KYN pathway activation provokes dendritic cells within lymphatic tissue to suppress the generation of myelin oligodendrocyte glycoprotein (MOG) T cells, which induce neuronal damage.²³ Aside from MOG T cells, evidence indicates that increased IDO activity enhances the differentiation of regulatory T cells,²⁴ suppressing immune overactivation in MS. Although only an elevated KYN/

TRP ratio was observed following the HIIT intervention, this finding further suggests disease-modifying properties of exercise in pwMS and encourages future research to uncover the mentioned neuroimmunologic consequences. However, considering the acute decreases in pNfL levels after HIIT, a reduction following the training intervention remains absent, which is in line with studies investigating the effect of exercise or multimodal rehabilitation on serum NfL levels in other neurologic disorders.^{25,26} Further mechanistic research focusing on the interaction between acute and longer-term training effects on pNfL is needed to understand why pwMS benefit from physical exercise and whether chronic exercise training leads to an alleviation in disease progression.

Although the observed persistent acute decrease in pNfL suggests a chronic reduction following repetitive acute exercise bouts, no training effects on pNfL levels were detected. Nevertheless, 2 major methodological aspects need to be considered. First, the 3-week intervention is very short to assess training effects. A longer intervention period is possibly necessary to reach a chronic reduction in pNfL. However, inpatient rehabilitation usually does not last longer than 4 weeks for pwMS, leading to the question whether this time span is sufficient for conspicuous improvements. Second, we did not include a passive control group due to ethical reasons. In addition, regarding acute effects, we did not compare the

results with a passive resting group, yet it needs to be taken into account that the present results even show superior effects of HIIT compared with the standard exercise therapy. Both groups exercised $3\times$ /week, and only the exercise modalities differed. The importance of higher intensities is underlined by the reported acute effects and supports findings of earlier investigations.^{11,12}

The investigation of the KYN metabolites in peripheral blood provides knowledge about the systemic balance of the KYN pathway and the availability of neuroactive metabolites for the CNS. Nonetheless, the conclusions drawn on neuromodulatory properties remain speculative because KYN pathway metabolites were not assessed in the CSF. However, consistent correlations between CSF and plasma/serum levels of KYN pathway metabolites have been reported,⁶ and, even more importantly, we revealed correlations between the neuroprotective KYN pathway branch toward KA and pNfL levels as marker of neuronal damage.

Acute exercise diminishes pNfL levels in pwMS, indicating a potential alleviation in ongoing neurodegeneration. Clinicians and researchers should consider acute physical exercise as confounding factor for the assessment of pNfL levels. Exercisemediated KYN pathway rerouting toward KA as end product is associated with pNfL reductions and might be responsible for neuroprotective effects. Overall, HIIT leads to greater responses than MCT, emphasizing the importance of higher exercise intensities when exercising with pwMS. Future studies are needed to profoundly address this promising mechanistic link between neuroprotective properties, neuroinflammation, and exercise in pwMS and other neurodegenerative disorders. Finally, the consequences of repetitively occurring exerciseinduced acute effects on pNfL levels and KYN pathway regulation require further research approaches that consider additional (intracellular) biomarkers of physical stress. The KYN/TRP upregulation following 3 weeks of HIIT suggests disease-counterregulatory properties of exercise on immune homeostasis, which remains to be investigated.

Acknowledgment

The authors thank Daniel Caminada and Francesca Ferrara from Labor Risch for support with blood sample preparation and storage. Furthermore, the authors thank Ramona Sylvester for supervising exercise training sessions.

Study Funding

German MS Society; Swiss MS Society (SMSG-12D-2018); Grenzen überschreiten (GRENZ-JB/18-007); Blumenau-Léonie Hartmann-Stiftung; Stiftung Ergotherapie (ergo-ZH-28-2018).

Disclosure

The authors declare that they have no conflict of interest. C. Warnke: institutional support: Biogen, Novartis, Roche, Sanofi, and Alexion. Go to Neurology.org/NN for full disclosures.

Publication History

Received by Neurology: Neuroimmunology & Neuroinflammation October 21, 2020. Accepted in final form January 21, 2021.

Appendix Authors

		a . u . u
Name	Location	Contribution
Niklas Joisten, PhD	Department of "Performance and Health (Sports Medicine)", Institute of Sport and Sport Science, Technical University Dortmund, Germany	Drafting/revision of the manuscript for content, including medical writing for content; major role in the acquisition of data; and analysis or interpretation of data
Annette Rademacher	Department for Molecular and Cellular Sports Medicine, Institute of Cardiovascular Research and Sports Medicine, German Sport University Cologne	Drafting/revision of the manuscript for content, including medical writing for content; major role in the acquisition of data; and analysis or interpretation of data
Clemens Warnke, MD	Department of Neurology, Faculty of Medicine and University Hospital Cologne, University of Cologne, Germany	Drafting/revision of the manuscript for content, including medical writing for content, and study concept or design
Sebastian Proschinger	Department for Molecular and Cellular Sports Medicine, Institute of Cardiovascular Research and Sports Medicine, German Sport University Cologne, Germany	Drafting/revision of the manuscript for content, including medical writing for content, and major role in the acquisition of data
Alexander Schenk, PhD	Department of "Performance and Health (Sports Medicine)", Institute of Sport and Sport Science, Technical University Dortmund, Germany	Drafting/revision of the manuscript for content, including medical writing for content, and major role in the acquisition of data
David Walzik	Department of "Performance and Health (Sports Medicine)", Institute of Sport and Sport Science, Technical University Dortmund	Drafting/revision of the manuscript for content, including medical writing for content, and major role in the acquisition of data
Andre Knoop, PhD	Center for Preventive Doping Research/Institute of Biochemistry, German Sport University Cologne, Germany	Analysis or interpretation of data; additional contributions: designed and conducted outcome measurement
Mario Thevis, PhD	Center for Preventive Doping Research/ Institute of Biochemistry, German Sport University Cologne	Analysis or interpretation of data; additional contributions: designed and conducted outcome measurement
Falk Steffen	Department of Neurology, Focus Program Translational Neuroscience (FTN), and Immunotherapy (FZI), University Medical Center of the Johannes Gutenberg University, Germany	Analysis or interpretation of data; additional contributions: designed and conducted outcome measurement

Continued

Appendix (continued)

Name	Location	Contribution
Stefan Bittner, MD	Department of Neurology, Focus Program Translational Neuroscience (FTN), and Immunotherapy (FZI), University Medical Center of the Johannes Gutenberg University, Germany	Analysis or interpretation of data; additional contributions: designed and conducted outcome measurement
Roman Gonzenbach, MD	Department of Neurology, Clinics of Valens, Rehabilitation Centre Valens, Switzerland	Study concept or design
Jan Kool, PhD	Department of Neurology, Clinics of Valens, Rehabilitation Centre Valens, Switzerland	Study concept or design
Wilhelm Bloch, MD	Department for Molecular and Cellular Sports Medicine, Institute of Cardiovascular Research and Sports Medicine, German Sport University Cologne, Germany	Study concept or design
Jens Bansi, PhD	Department of Neurology, Clinics of Valens, Rehabilitation Centre Valens, Switzerland	Major role in the acquisition of data; study concept or design; and additional contributions: supervised manuscript drafting and obtained funding
Philipp Zimmer, PhD, PhD	Department of "Performance and Health (Sports Medicine)", Institute of Sport and Sport Science, Technical University Dortmund, Germany	Study concept or design; analysis or interpretation of data; and additional contributions: supervised manuscript drafting and obtained funding

References

- Siller N, Kuhle J, Muthuraman M, et al. Serum neurofilament light chain is a biomarker of acute and chronic neuronal damage in early multiple sclerosis. Mult Scler 2019;25:678–686.
- Kuhle J, Kropshofer H, Haering DA, et al. Blood neurofilament light chain as a biomarker of MS disease activity and treatment response. Neurology 2019;92: e1007–e1015.
- Kuhle J, Plavina T, Barro C, et al. Neurofilament light levels are associated with longterm outcomes in multiple sclerosis. Mult Scler J 2019;26:1691–1699.
- Lovelace MD, Varney B, Sundaram G, et al. Recent evidence for an expanded role of the kynurenine pathway of tryptophan metabolism in neurological diseases. Neuropharmacology 2017;112:373–388.
- Fakan B, Szalardy L, Vecsei L. Exploiting the therapeutic potential of endogenous immunomodulatory systems in multiple sclerosis-special focus on the peroxisome

proliferator-activated receptors (PPARs) and the kynurenines. Int J Mol Sci 2019;20: 426.

- Lim CK, Bilgin A, Lovejoy DB, et al. Kynurenine pathway metabolomics predicts and provides mechanistic insight into multiple sclerosis progression. Sci Rep 2017;7: 41473.
- Chatterjee P, Zetterberg H, Goozee K, et al. Plasma neurofilament light chain and amyloid-beta are associated with the kynurenine pathway metabolites in preclinical Alzheimer's disease. J Neuroinflammation 2019;16:186.
- Rajda C, Galla Z, Polyak H, et al. Cerebrospinal fluid neurofilament light chain is associated with kynurenine pathway metabolite changes in multiple sclerosis. Int J Mol Sci 2020;21:2665.
- Dalgas U, Langeskov-Christensen M, Stenager E, Riemenschneider M, Hvid LG. Exercise as medicine in multiple sclerosis-time for a paradigm shift: preventive, symptomatic, and disease-modifying aspects and perspectives. Curr Neurol Neurosci Rep 2019;19:88.
- Campbell E, Coulter EH, Paul L. High intensity interval training for people with multiple sclerosis: a systematic review. Mult Scler Relat Disord 2018;24:55–63.
- Joisten N, Proschinger S, Rademacher A, et al. High-intensity interval training reduces neutrophil-to-lymphocyte ratio in persons with multiple sclerosis during inpatient rehabilitation. Mult Scler 2020. doi: 10.1177/1352458520951382.
- Zimmer P, Bloch W, Schenk A, et al. High-intensity interval exercise improves cognitive performance and reduces matrix metalloproteinases-2 serum levels in persons with multiple sclerosis: a randomized controlled trial. Mult Scler 2018;24:1635–1644.
- Joisten N, Rademacher A, Bloch W, et al. Influence of different rehabilitative aerobic exercise programs on (anti-) inflammatory immune signalling, cognitive and functional capacity in persons with MS—study protocol of a randomized controlled trial. BMC Neurol 2019;19:37.
- Langdon DW, Amato MP, Boringa J, et al. Recommendations for a Brief International Cognitive Assessment for Multiple Sclerosis (BICAMS). Mult Scler 2012;18: 891–898.
- Engel S, Friedrich M, Muthuraman M, et al. Intrathecal B-cell accumulation and axonal damage distinguish MRI-based benign from aggressive onset in MS. Neurol Neuroimmunol Neuroinflamm 2019;6:e595.
- Bittner S, Steffen F, Uphaus T, et al. Clinical implications of serum neurofilament in newly diagnosed MS patients: a longitudinal multicentre cohort study. EBioMedicine 2020;56:102807.
- Joisten N, Kummerhoff F, Koliamitra C, et al. Exercise and the Kynurenine pathway: current state of knowledge and results from a randomized cross-over study comparing acute effects of endurance and resistance training. Exerc Immunol Rev 2020;26: 24–42.
- Henykova E, Vranova HP, Amakorova P, et al. Stable isotope dilution ultra-high performance liquid chromatography-tandem mass spectrometry quantitative profiling of tryptophan-related neuroactive substances in human serum and cerebrospinal fluid. J Chromatogr A 2016;1437:145–157.
- Schwarcz R, Stone TW. The kynurenine pathway and the brain: challenges, controversies and promises. Neuropharmacology 2017;112:237–247.
- Agudelo LZ, Femenia T, Orhan F, et al. Skeletal muscle PGC-1alpha1 modulates kynurenine metabolism and mediates resilience to stress-induced depression. Cell 2014;159:33–45.
- DiNatale BC, Murray IA, Schroeder JC, et al. Kynurenic acid is a potent endogenous aryl hydrocarbon receptor ligand that synergistically induces interleukin-6 in the presence of inflammatory signaling. Toxicol Sci 2010;115:89–97.
- Bansi J, Koliamitra C, Bloch W, et al. Persons with secondary progressive and relapsing remitting multiple sclerosis reveal different responses of tryptophan metabolism to acute endurance exercise and training. J Neuroimmunol 2018;314:101–105.
- Lemos H, Mohamed E, Ou R, et al. Co-treatments to boost IDO activity and inhibit production of downstream catabolites induce durable suppression of experimental autoimmune encephalomyelitis. Front Immunol 2020;11:1256.
- Baban B, Chandler PR, Sharma MD, et al. IDO activates regulatory T cells and blocks their conversion into Th17-like T cells. J Immunol 2009;183:2475–2483.
- Jensen CS, Portelius E, Hogh P, et al. Effect of physical exercise on markers of neuronal dysfunction in cerebrospinal fluid in patients with Alzheimer's disease. Alzheimers Dement (N Y) 2017;3:284–290.
- Bartlett DM, Govus A, Rankin T, et al. The effects of multidisciplinary rehabilitation on neuroimaging, biological, cognitive and motor outcomes in individuals with premanifest Huntington's disease. J Neurol Sci 2020;416:117022.