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New insights on serodiagnosis of trichinellosis during window period: early diagnostic antigens from *Trichinella spiralis* intestinal worms

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Abstract

The clinical diagnosis of trichinellosis is difficult because its clinical manifestations are nonspecific. Detection of anti-*Trichinella* IgG by ELISA using *T. spiralis* muscle larval excretory-secretory (ES) antigens is the most commonly used serological method for diagnosis of trichinellosis, but the main disadvantage is false negativity during the early stage of infection. There is an obvious window period between *Trichinella* infection and antibody positivity. During the intestinal stage of *Trichinella* infection, the ES antigens of intestinal worms (intestinal infective larvae and adults) are exposed to host's immune system at the earliest time and elicit the production of specific anti-*Trichinella* antibodies. Anti-*Trichinella* IgG antibodies in infected mice were detectable by ELISA with ES antigens of intestinal worms as soon as 8–10 days post infection (dpi), but ELISA with muscle larval ES antigens did not permit detection of infected mice before 12 dpi. Therefore, the new early antigens from *T. spiralis* intestinal worms should be screened, identified and characterized for early serodiagnosis of trichinellosis.

Keywords: Trichinellosis, Early serodiagnosis, Intestinal infective larvae, Adult worms

Multilingual abstracts

Please see Additional file 1 for translations of the abstract into the six official working languages of the United Nations.

Background

Trichinellosis is a major food-borne parasitic zoonosis acquired by eating raw or undercooked meat contaminated with the infective larvae of the genus *Trichinella*. Human trichinellosis has been documented in 55 countries in the world [1]. From 1986 to 2009, there were 65 818 cases and 42 deaths reported from 41 countries, and is considered to be a neglected infectious disease, especially in developing countries of east Europe, south America, Asia and Africa [2, 3]. From 2004 to 2009 in China, 15

outbreaks of human trichinellosis affected 1 387 people and caused 4 deaths. Pork is the predominant source of outbreaks of human trichinellosis in China. Out of 15 outbreaks, 12 (85.71%) were caused by eating raw or undercooked pork [4]. Swine trichinellosis is transmitted mostly through garbage (i.e., feeding pigs with raw swill). Pigs infected with Trichinella are predominately from small backyard farms where animals are raised under poor hygienic conditions, and from rural and mountainous areas where they range freely at pasture. Pigs were sometimes slaughtered at home in rural and mountainous areas without veterinary inspection [5, 6]. Recently, outbreaks of trichinellosis also occurred in the mountainous regions of Cambodia, Lao PDR, north Thailand and Vietnam, and most of the patients had consumed traditional raw or undercooked pork dishes at wedding, funeral, or New Year parties [7–10]. Trichinellosis has become an emerging and re-emerging zoonotic disease with health, social, and economic impacts in developing countries [11].

The clinical diagnosis of trichinellosis is difficult because its clinical manifestations are nonspecific. The



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definitive diagnosis of trichinellosis can be made by detecting larvae in a biopsy muscle samples or specific anti-Trichinella antibodies, but muscle biopsy is not sensitive to the light infections and the early stage of infection [12]. Serum specific anti-Trichinella IgE are thought to appear first and are typical of the acute stage of the disease, but IgE antibodies are detected in a few cases only at the onset of the disease because their halflife in serum is relative short [13]. The detection of anti-Trichinella IgG by ELISA using excretory-secretory (ES) antigens of T. spiralis muscle larvae (ML) is the most commonly used method for serodiagnosis of trichinellosis and is recommended by the International Commission on Trichinellosis (ICT), because ML are easily collected from experimentally infected animals and their ES antigens can be prepared by the in vitro cultivation of larvae [14]. However, the main disadvantage of the detection of anti-Trichinella IgG by using ML ES antigens is the occurrence of a high rate of false negative results during the early stage of infection [15], possibly because the majority of ML ES antigens are stagespecific and not recognized by specific antibodies induced by the parasites during the intestinal phase [16]. Previous studies have shown that 100% detection of anti-Trichinella IgG is not possible for at least 1-3 months after initial infection with the parasite [13]. There is an obvious window period of 3-4 weeks between Trichinella infection and specific antibody positivity. The indirect immunofluorescence test (IIF) with frozen sections of infected tissue or isolated whole ML as antigens could detect specific antibodies in sera of experimentally infected mice 1-2 weeks after infection or sera of patients with trichinellosis from day 6 after the onset of illness [17, 18], but cross-reactions with Trichinella antigens were observed in the patients with autoimmune diseases and other helminthiases because IIF is based on cuticle surface antigens of ML [19]. This is particularly important in regions where other human helminthiases (e.g., ascariasis, trichuriasis, clonorchiasis, paragonimiasis, cysticercosis and so on) are common and cross-reactions with these parasites could give could give false positive results [20]. Furthermore, persons reading the fluorescent sections should be aware that only sections with a uniform fluorescence should be considered as positive, whereas those, which show a non-uniform fluorescence along the cuticle, should be considered as false-positive.

T. spiralis circulating antigen (CAg) is the ES antigens produced by live worms and can directly enter the peripheral blood circulation, and CAg appears earlier than antibody in blood circulation [21]. Theoretically, the detection of *T. spiralis* CAg seems an early diagnostic method for trichinellosis. However, because the serum levels of *Trichinella* CAg are usually quite low, the detection rate of CAg in serum samples was usually only 30–50% in the patients with clinical trichinellosis [22]. Thus, detection of *T. spiralis* CAg has not been clinically applied to serodiagnosis of trichinellosis. So far, the detection of anti-*Trichinella* IgG is still the serological test of choice for diagnosis of trichinellosis. Hence, it is necessary to exploit the new source of early diagnostic antigens for detection of anti-*Trichinella* antibodies.

Discussion

After ingestion, T. spiralis ML are released from their nurse-cells in the stomach and activated to intestinal infective larvae (IIL) by intestinal content or bile after 0.9 h post-infection (hpi). The IIL invade host's intestinal epithelium, and undergo four molting to develop to adult worms (AW) in 31 hpi. After mating, the female AW invade intestinal mucosa again and live there for 10-20 days in mice and rats or 4-6 weeks in humans [23]. The IIL and AW are the first two invasive stages in life cycle of *T. spiralis*. During the intestinal stage of *T*. spiralis infection, the ES antigens produced by the IIL and AW result in early exposure to the host's immune system and elicit the production of specific anti-Trichinella antibodies. The ES proteins of intestinal T. spiralis worms might contain the early diagnostic markers of trichinellosis. Therefore, the new early diagnostic antigens should be searched and identified from T. spiralis intestinal worms instead of the conventional ML.

Recent studies have showed that crude antigens of T. spiralis AW were recognized by sera from infected mice or swine 7 days post-infection (dpi) [24]. The recombinant protein from IIL at 6 hpi or pre-adults 20 hpi was recognized by pig antiserum in Western blot as early as 15-20 dpi [25, 26]. Anti-Trichinella IgG in mice infected with 100 T. spiralis larvae were detectable by ELISA with adult worm or IIL ES antigens as soon as 8 or 10 dpi, but ELISA with ML ES antigens did not permit detection of infected mice before 12 dpi. When the sera of patients with trichinellosis at 19 dpi were assayed, the sensitivity (100%, 20/20) of ELISA with adult or IIL ES antigens was evidently higher than 75% (15/20) of ELISA with ML ES antigens. The specificity of ELISA with AW (98.11%, 156/159) and IIL ES (96.86%, 154/159) antigens was also higher than 89.31% (142/159) of ELISA with muscle larval ES antigens. There were no cross-reactions of ELISA with AW and IIL ES antigens with sera of patients with schistosomiasis, clonorchiosis, sparganosis and healthy persons [27, 28]. The sensitivity and specificity of AW and IIL ES antigens were superior to those of muscle larval ES antigens. Therefore, the ES antigens of T. spiralis intestinal worms (IIL and AW) could be considered as a new source of diagnostic antigens and the potential antigens for the early specific serodiagnosis of trichinellosis.

However, there was also low cross-reactivity of the ELISA with the AW and IIL ES antigens with sera of a few of patients with paragonimiosis or cysticercosis. The Trichinella-specific protein bands in the AW and IIL ES antigens should be further identified and validated by Western blotting with a large-scale of sera of the patients with trichinellosis and other helminthiasis. Moreover, Trichinella-specific AW and IIL ES antigens recognized by early infection sera should be characterized by mass spectrometry [29, 30]. Then, the recombinant early antigens of Trichinella should be developed and their sensitivity and specificity were evaluated with a large-scale trial in future. The exploitation of the early diagnostic antigens from T. spiralis invasive stages may provide new insights into early serodiagnosis of other helminthiasis, for example, the early antigens from cercariae and schistosomula of Schistosoma for diagnosis of schistomiasis [31]. Additionally, it will also provide valuable information for further understanding the parasitehost interaction and invasion mechanism [32-34].

Conclusions

The ES antigens of *T. spiralis* intestinal stages (intestinal infective larvae and adults) are exposed to host's immune system at the earliest time and elicit the production of specific anti-*Trichinella* antibodies, the new antigens from *T. spiralis* intestinal worms should be screened, identified and characterized for early serodiagnosis of trichinellosis. The exploitation of the early diagnostic antigens from the parasite's invasive stages may provide new insights into early serodiagnosis of other helminthiasis.

Additional file

Additional file 1: Multilingual abstracts in the six official working languages of the United Nations. (PDF 698 kb)

Abbreviations

CAg: Circulating antigen; dpi: days post-infection; ES: Excretory–secretory; hpi: hour post-infection; ICT: International Commission on Trichinellosis; IIL: Intestinal infective larvae; ML: Muscle larvae; PDR: People's Democratic Republic.

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Availability of data and materials

The data supporting the results of this paper are included in the paper.

Authors' contributions

WZQ, CYD and CJ designed the study. SYL, LRD, JP and GYY collected and analyzed the data. WZQ, CYD and CJ wrote and revised the paper. All authors read and approved the final manuscript.

Competing interests

The authors declare that they have no competing interests.

Consent for publication

Not applicable.

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References

- 1. Pozio E. World distribution of *Trichinella* spp. infections in animals and humans. Vet Parasitol. 2007;149:3–21.
- Murrell KD, Pozio E. Worldwide occurrence and impact of human trichinellosis, 1986–2009. Emerg Infect Dis. 2011;17:2194–202.
- Bruschi F. Trichinellosis in developing countries: is it neglected? J Infect Dev Ctries. 2012;6:216–22.
- Cui J, Wang ZQ, Xu BL. The epidemiology of human trichinellosis in China during 2004–2009. Acta Trop. 2011;118:1–5.
- Cui J, Wang ZQ. An epidemiological overview of swine trichinellosis in China. Vet J. 2011;190:324–8.
- Cui J, Jiang P, Liu LN, Wang ZQ. Survey of *Trichinella* infection in domestic pigs of northern and eastern Henan, China. Vet Parasitol. 2013;194:133–5.
- Barennes H, Sayasone S, Odermatt P, De Bruyne A, Hongsakhone S, Newton PN, et al. A major trichinellosis outbreak suggesting a high endemicity of *Trichinella* infection in northern Laos. Am J Trop Med Hyg. 2008;78:40–4.
- Burniston S, Okello AL, Khamlome B, Inthavong P, Gilbert J, Blacksell SD, et al. Cultural drivers and health-seeking behaviors that impact on the transmission of pig-associated zoonoses in Lao People's Democratic Republic. Infect Dis Poverty. 2015;4:11.
- Conlan JV, Vongxay K, Khamlome B, Gomez-Morales MA, Pozio E, Blacksell SD, et al. Patterns and risks of *Trichinella* infection in humans and pigs in northern Laos. PLoS Negl Trop Dis. 2014;8:e3034.
- 10. Van De N, Thi Nga V, Dorny P, Vu Trung N, Ngoc Minh P, Trung Dung D, et al. Trichinellosis in Vietnam. Am J Trop Med Hyg. 2015;92:1265–70.
- Onkoba NW, Chimbari MJ, Mukaratirwa S. Malaria endemicity and coinfection with tissue-dwelling parasites in Sub-Saharan Africa: a review. Infect Dis Poverty. 2015;4:35.
- 12. Gottstein B, Pozio E, Nöckler K. Epidemiology, diagnosis, treatment, and control of trichinellosis. Clin Microbiol Rev. 2009;22:127–45.
- 13. Bruschi F, Moretti A, Wassom D, Piergili FD. The use of a synthetic antigen for the serological diagnosis of human trichinellosis. Parasite. 2001;8:S141–3.
- Gamble HR, Pozio E, Bruschi F, Nöckler K, Kapel CM, Gajadhar AA. International Commission on Trichinellosis: recommendations on the use of serological tests for the detection of *Trichinella* infection in animals and man. Parasite. 2004;11:3–13.
- Cui J, Wang L, Sun GG, Liu LN, Zhang SB, Liu RD, et al. Characterization of a *Trichinella spiralis* 31 kDa protein and its potential application for the serodiagnosis of trichinellosis. Acta Trop. 2015;142:57–63.
- Liu JY, Zhang NZ, Li WH, Li L, Yan HB, Qu ZG, et al. Proteomic analysis of differentially expressed proteins in the three developmental stages of *Trichinella spiralis*. Vet Parasitol. 2016;231:32–8.
- Ruitenberg EJ, Ljungström I, Steerenberg PA, Buys J. Application of immunofluorescence and immunoenzyme methods in the serodiagnosis of *Trichinella spiralis* infection. Ann N Y Acad Sci. 1975;254:296–303.
- El-Kadi MA, El-Shazly AM, Abu-Elwafa SA, El-Nahas HA, Abdel-Mageed AA, Soliman M. Serological diagnosis of *Trichinella spiralis* in experimentally infected mice. J Egypt Soc Parasitol. 2005;35:125–36.
- Robert F, Weil B, Kassis N, Dupouy-Camet J. Investigation of immunofluorescence cross-reactions against *Trichinella spiralis* by western blot (immunoblot) analysis. Clin Diagn Lab Immunol. 1996;3:575–7.
- 20. Cui J, Wang Z, Wu F, Jin X, Zhang P, Yang R, Liu J. Diagnosis of trichinosis by indirect fluorescent antibody test with *Trichinella* larva section.

Zhongguo Ji Sheng Chong Xue Yu Ji Sheng Chong Bing Za Zhi. 1999;17:28–31 (in Chinese).

- Liu LN, Jing FJ, Cui J, Fu GY, Wang ZQ. Detection of circulating antigen in serum of mice infected with *Trichinella spiralis* by an IgY-IgM mAb sandwich ELISA. Exp Parasitol. 2013;133:150–5.
- Nishiyama T, Araki T, Mizuno N, Wada T, Ide T, Yamaguchi T. Detection of circulating antigens in human trichinellosis. Trans R Soc Trop Med Hyg. 1992;86:292–3.
- Campbell W. Trichinella and trichinosis. New York: Plenum Press; 1983. p. 75–151.
- Yang J, Pan W, Sun X, Zhao X, Yuan G, Sun Q, et al. Immunoproteomic profile of *Trichinella spiralis* adult worm proteins recognized by early infection sera. Parasit Vectors. 2015;8:20.
- Tang B, Liu M, Wang L, Yu S, Shi H, Boireau P, et al. Characterisation of a high-frequency gene encoding a strongly antigenic cystatin-like protein from *Trichinella spiralis* at its early invasion stage. Parasit Vectors. 2015;8:78.
- Zocevic A, Lacour SA, Mace P, Giovani B, Grasset-Chevillot A, Vallee I, et al. Primary characterization and assessment of a *T. spiralis* antigen for the detection of *Trichinella* infection in pigs. Vet Parasitol. 2014;205:558–67.
- Sun GG, Wang ZQ, Jiang P, Liu RD, Wen H, et al. Early serodiagnosis of trichinellosis by ELISA using excretory–secretory antigens of *Trichinella spiralis* adult worms. Parasit Vectors. 2015;8:484.
- Sun GG, Liu RD, Wang ZQ, Jiang P, Wang L, Liu XL, et al. New diagnostic antigens for early trichinellosis: the excretory-secretory antigens of *Trichinella spiralis* intestinal infective larvae. Parasitol Res. 2015;114:4637–44.
- Liu RD, Cui J, Liu XL, Jiang P, Sun GG, Zhang X, et al. Comparative proteomic analysis of surface proteins of *Trichinella spiralis* muscle larvae and intestinal infective larvae. Acta Trop. 2015;150:79–86.
- Liu RD, Jiang P, Wen H, Duan JY, Wang LA, Li JF, et al. Screening and characterization of early diagnostic antigens in excretory–secretory proteins from *Trichinella spiralis* intestinal infective larvae by immunoproteomics. Parasitol Res. 2016;115:615–22.
- Liu M, Ju C, Du XF, Shen HM, Wang JP, Li J, et al. Proteomic analysis on cercariae and schistosomula in reference to potential proteases involved in host invasion of *Schistosoma japonicum* larvae. J Proteome Res. 2015;14:4623–34.
- Liu RD, Qi X, Sun GG, Jiang P, Zhang X, Liu XL, Cui J, Wang ZQ. Proteomic analysis of *Trichinella spiralis* adult worm excretory–secretory proteins recognized by early infection sera. Vet Parasitol. 2016;231:43–6.
- Yang J, Zhu W, Huang J, Wang X, Sun X, Zhan B, Zhu X. Partially protective immunity induced by the 14-3-3 protein from *Trichinella spiralis*. Vet Parasitol. 2016;231:63–8.
- Bermúdez-Cruz RM, Fonseca-Liñán R, Grijalva-Contreras LE, Mendoza-Hernández G, Ortega-Pierres MG. Proteomic analysis and immunodetection of antigens from early developmental stages of *Trichinella spiralis*. Vet Parasitol. 2016;231:22–31.

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