



# Complete Genome Assembly of *Myxococcus xanthus* Strain DZ2 Using Long High-Fidelity (HiFi) Reads Generated with PacBio Technology

 Rikesh Jain,<sup>a,b</sup>  Bianca H. Habermann,<sup>b</sup>  Tãm Mignot<sup>a</sup>

<sup>a</sup>Aix-Marseille Université, CNRS, Institut de Microbiologie de la Méditerranée (UMR 7283), Turing Center for Living Systems, Marseille, France

<sup>b</sup>Aix-Marseille Université, CNRS, Institut de Biologie du Développement de Marseille (UMR 7288), Turing Center for Living Systems, Marseille, France

**ABSTRACT** *Myxococcus xanthus* is a Gram-negative social bacterium belonging to the order *Myxococcales* of the class *Deltaproteobacteria*. It is a facultative social predator found in soils across the globe and is thought to be crucial for the microbial ecosystem. Here, we report a complete high-quality reference genome of the *M. xanthus* strain DZ2.

*Myxococcus xanthus*, a social bacterium, preys on a broad range of microbes, including bacteria as well as fungi (1, 2). Its predatory behavior involves moving or gliding (socially and individually) into prey colonies, killing prey cells that it comes in contact with and consuming them for nutrition (3, 4). The predatory behaviors of *M. xanthus* have been researched for many decades; however, the mechanisms of how it recognizes and kills the diverse preys are not clearly understood yet. Interestingly, different strains of *M. xanthus* show different predation efficiencies against the same prey (1, 2). Therefore, comparing the genomes of these strains might help in identifying key genes, pathways, and molecules involved in prey recognition and killing. It is for this purpose that we sequenced, assembled, and annotated the complete genome sequence of our lab strain, DZ2, which was originally procured from David R. Zusman's lab (5).

DZ2 cells were grown at 32°C in CYE (Casitone yeast extract) medium, and genomic DNA was extracted using a MasterPure DNA purification kit (catalog [cat.] number 85200) (Epicentre/Lucigen, Middleton, WI) following the manufacturer's protocol. The high-fidelity (HiFi) sequencing library was prepared using the SMRTbell express template prep kit (PacBio, CA), and genome sequencing was performed using single-molecule real-time (SMRT) sequencing on the PacBio Sequel II system at the DRESDEN-concept Genome Center, Germany. A total of 98,235 subreads were generated with a mean subread length of 6,019 bp, totaling 591 Mb, which represents approximately 60-fold coverage. Circular consensus reads were called using Code Composer Studio (CCS) version 5 (PacBio) on the raw PacBio subreads, and the resulting consensus reads with a quality value (QV) of >20 or >99% accuracy were used to assemble the genome with HiCanu version 2.1 (6). This resulted in a single circular contig of 9,359,382 bp (approximately 220 kb larger than the DK1622 genome) with a GC content of 68.86%. Genome annotation was performed using the NCBI Prokaryotic Genome Annotation Pipeline (PGAP) version 5.1 (7), which predicted a total of 7,408 protein-coding sequences (CDSs), 12 rRNA genes, 66 tRNA genes, 4 noncoding RNA (ncRNA) genes, 86 pseudogenes, and 4 CRISPR arrays in the DZ2 genome. Default parameters were used for all software unless otherwise noted.

A previous version of the DZ2 genome sequence (8) existed as a draft assembly with 87 contigs. It is around 88 kb shorter than our PacBio complete assembly, suggesting large coverage gaps and possibly missing important loci. It was sequenced with 454 GS-FLX Titanium technology, a second-generation sequencing technology,

**Citation** Jain R, Habermann BH, Mignot T. 2021. Complete genome assembly of *Myxococcus xanthus* strain DZ2 using long high-fidelity (HiFi) reads generated with PacBio technology. *Microbiol Resour Announc* 10:e00530-21. <https://doi.org/10.1128/MRA.00530-21>.

**Editor** Frank J. Stewart, Montana State University

**Copyright** © 2021 Jain et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to Rikesh Jain, [rikeshjain44@gmail.com](mailto:rikeshjain44@gmail.com).

**Received** 27 May 2021

**Accepted** 18 June 2021

**Published** 15 July 2021

which generates much shorter reads (average read length, 450 bp) than the third-generation sequencing platform PacBio Sequel II (average read length, 10,000 bp). Highly accurate (99.8%) long high-fidelity PacBio reads enabled the resolving of large repetitive and low-complexity genomic regions, thus closing all the gaps, a task which remains challenging with short-read sequencing (9).

Uninterrupted and accurate genome sequence information is crucial for many downstream genomic analyses, such as biosynthetic gene cluster mining and gene synteny comparison, as well as for understanding genome plasticity, the patterns of transposon distribution, and gene expression regulation.

**Data availability.** The raw sequencing data can be found at the NCBI SRA database under the accession number [PRJNA701418](https://www.ncbi.nlm.nih.gov/sra/PRJNA701418). The complete genome sequence has been deposited in GenBank under accession number [CP070500](https://www.ncbi.nlm.nih.gov/genbank/CP070500).

## ACKNOWLEDGMENTS

We thank Sylke Winkler and Thomas Brown for the useful suggestions and data analysis.

This work was supported by a research grant from the European Research Council (ERC JAWS 885145) to T.M. and a postdoctoral fellowship from the Turing Center for Living Systems (CENTURI) to R.J.

## REFERENCES

- Morgan AD, MacLean RC, Hillesland KL, Velicer GJ. 2010. Comparative analysis of *Myxococcus* predation on soil bacteria. *Appl Environ Microbiol* 76:6920–6927. <https://doi.org/10.1128/AEM.00414-10>.
- Livingstone PG, Morphey RM, Whitworth DE. 2017. Myxobacteria are able to prey broadly upon clinically-relevant pathogens, exhibiting a prey range which cannot be explained by phylogeny. *Front Microbiol* 8:1593. <https://doi.org/10.3389/fmicb.2017.01593>.
- Muñoz-Dorado J, Marcos-Torres FJ, García-Bravo E, Moraleda-Muñoz A, Pérez J. 2016. Myxobacteria: moving, killing, feeding, and surviving together. *Front Microbiol* 7:781. <https://doi.org/10.3389/fmicb.2016.00781>.
- Seef S, Herrou J, De Boissier P, My L, Brasseur G, Robert D, Jain R, Mercier R, Cascales E, Habermann B, Mignot T. 2020. A Tad-like apparatus is required for contact-dependent prey killing in predatory social bacteria. *bioRxiv* <https://doi.org/10.1101/2021.02.25.432843>.
- Blackhart BD, Zusman DR. 1985. “Frizzy” genes of *Myxococcus xanthus* are involved in control of frequency of reversal of gliding motility. *Proc Natl Acad Sci U S A* 82:8767–8770. <https://doi.org/10.1073/pnas.82.24.8767>.
- Nurk S, Walenz BP, Rhie A, Vollger MR, Logsdon GA, Grothe R, Miga KH, Eichler EE, Phillippy AM, Koren S. 2020. HiCanu: accurate assembly of segmental duplications, satellites, and allelic variants from high-fidelity long reads. *Genome Res* 30:1291–1305. <https://doi.org/10.1101/gr.263566.120>.
- Tatusova T, DiCuccio M, Badretdin A, Chetvernin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI Prokaryotic Genome Annotation Pipeline. *Nucleic Acids Res* 44:6614–6624. <https://doi.org/10.1093/nar/gkw569>.
- Müller S, Willett JW, Bahr SM, Darnell CL, Hummels KR, Dong CK, Vlamakis HC, Kirby JR. 2013. Draft genome sequence of *Myxococcus xanthus* wild-type strain DZ2, a model organism for predation and development. *Genome Announc* 1:e00217-13. <https://doi.org/10.1128/genomeA.00217-13>.
- Logsdon GA, Vollger MR, Eichler EE. 2020. Long-read human genome sequencing and its applications. *Nat Rev Genet* 21:597–614. <https://doi.org/10.1038/s41576-020-0236-x>.