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Method parameters' impact on mortality and variability in mouse stroke experiments: a metaanalysis

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Although hundreds of promising substances have been tested in clinical trials, thrombolysis currently remains the only specific pharmacological treatment for ischemic stroke. Poor quality, e.g. low statistical power, in the preclinical studies has been suggested to play an important role in these failures. Therefore, it would be attractive to use animal models optimized to minimize unnecessary mortality and outcome variability, or at least to be able to power studies more exactly by predicting variability and mortality given a certain experimental setup. The possible combinations of methodological parameters are innumerous, and an experimental comparison of them all is therefore not feasible. As an alternative approach, we extracted data from 334 experimental mouse stroke articles and, using a hypothesis-driven meta-analysis, investigated the method parameters' impact on infarct size variability and mortality. The use of Swiss and C57BL6 mice as well as permanent occlusion of the middle cerebral artery rendered the lowest variability of the infarct size while the emboli methods increased variability. The use of Swiss mice increased mortality. Our study offers guidance for researchers striving to optimize mouse stroke models.

Stroke is amongst the most common causes of death and disability worldwide¹. Major advances have been made in the understanding of the pathophysiology of stroke and *in vitro* and animal experiments have suggested numerous substances as promising candidates for treatment of the disease^{2,3}. However, although hundreds of these substances have been tested in clinical trials, thrombolysis is still the only specific pharmacological treatment proven efficacious in acute ischemic stroke². The apparent difficulty of transferring results from experimental studies to the clinical situation ("from bench to bedside") has been referred to as a "translational roadblock"^{2,4} and the possible reasons behind it, particularly lack of methodological quality, has been discussed intensively over the last years^{5,6}. Low statistical power as a results of high outcome variability and mortality in combination with a small group sizes has been suggested to be an important issue⁵ and although this can theoretically be overcome by increasing the group sizes enough, such a solution has several problematic implications. From an ethical point of view, it is recommended to use as few animals as possible according to the "three R principle"⁷ and working with large number of animals is both practically inconvenient (time and space consuming) and costly. Therefore, as a complement, it would be attractive to optimize the animal model by minimizing unnecessary outcome variability and mortality, or at least be able to power studies more exactly by predicting variability and mortality given a certain experimental setup.

Simplified, the standard approach in the majority of the preclinical stroke studies consists of three steps: 1) focal cerebral ischemia is induced in rodents, 2) some kind of treatment is administered and 3) outcome, most often by measuring infarct sizes, is assessed. These basic steps are employed in hundreds of publications each year but unfortunately no consensus exits regarding the ideal setup, and since the variations in methodological

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factors are innumerous, it is very complicated to experimentally evaluate all possible combinations. In an attempt to address this question, we performed a hypothesis-driven meta-analysis in 2013 studying method parameters' impact on mortality and variability in rat stroke experiments⁸. However, since the previous study only used data from rat studies, and since mice are becoming increasingly popular in the preclinical stroke field, we decided to perform a similar analysis on mice. Thus, the objective of the current study was to investigate the effect of methodological variables on infarct size variability and mortality in mouse stroke experiments. Specifically, eight a priori hypotheses concerning factor-outcome relations were formulated:

- 1. Middle cerebral artery occlusion duration affects (A) infarct size variability and (B) mortality.
- 2. Type of focal cerebral ischemia procedure affects (A) infarct size variability and (B) mortality.
- 3. Mouse strain affects (A) infarct size variability and (B) mortality.
- 4. In studies using the intraluminal filament method, the type of occluding filament affects (A) infarct size variability and (B) mortality.

Results

Regression models. The regression model addressing hypotheses 1A, 2A and 3A included 500 control groups while the analysis for hypothesis 4A included 430 (Fig. 1). The r² values were 0.22 and 0.26, meaning that 22% and 26% of the variation in the outcome measures *Infarct size coefficient of variation* were explained by the models, respectively. The two models analyzing impact on *Mortality rate*, one for hypotheses 1B, 2B and 3B and one for hypothesis 4B, included 80 and 73 control groups, respectively. The resulting r² values were 0.72 and 0.78.

Impact of occlusion duration on infarct size variability and mortality (hypotheses 1A and 1B). Regarding the effect of *Occlusion duration* on the outcome *Infarct size coefficient of variation*, only the category *Permanent* turned out to significantly decrease the variability compared to the reference category *Short*

transient (-8.6%, CI: -15.3 to -1.9%; p = 0.012; Fig. 2a). No impact of Occlusion duration on Mortality rate was







Figure 3. Method parameters' impact on mortality rate. Swiss strain was found to significantly increase mortality rate compared to the reference C57BL6. The variables *Occlusion duration, Type of middle cerebral artery occlusion procedure* and *Occluding filament type* were removed in the backward exclusion step of the regression model due to small explanatory value and therefore results of hypotheses 1B, 2B and 4B could not be presented. Bars represent change in *Mortality rate,* measured in absolute percent units. Significant p-values are black, non-significant p-values are grey. N = 80. Error bars represent 0.95 confidence intervals.

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found (those categories were removed in the backward exclusion step of the statistical analysis and therefore not presented in Fig. 3).

Impact of type of focal cerebral ischemia procedure on infarct size variability and mortality (hypotheses 2A and 2B). In the analysis of cerebral ischemia procedures, the *Emboli/clot method* strongly augmented the *Infarct size coefficient of variation* (+25.9, CI: +8.2 to +43.6; p = 0.004; Fig. 2b) in comparison to the reference category *Intraluminal Filament*. *Mortality rate* was not significantly affected by cerebral ischemia procedure (variables removed during the backward exclusion procedure).

Impact of mouse strain on infarct size variability and mortality (hypotheses 3A and 3B). Strain affected both *Infarct size coefficient of variation* and *Mortality rate* significantly. Overall, the majority of the strains seemed to increase the variability compared to the reference category *C57BL6*, with the strongest positive regression coefficient being found for *Mixed C57BL6/129* (+22.8%, CI: +12.5 to 33.1%; p < 0.0001; Fig. 2c) and *129* (+15.9%, CI: +8.3 to 33.1%; p < 0.0001; Fig. 2c). The only strain category that significantly reduced the variability compared to the reference was *Swiss* (-5.7%, CI:-11.2 to -0.3%; p = 0.038; Fig. 2c). Except for the reference,





two strain categories were included in the mortality analysis and only *Swiss* had a significant impact by increasing the *Mortality rate* (+24.2%, CI: +16.2 to +32.2%; p < 0.0001; Fig. 3).

Impact of filament coating type on infarct size variability and mortality (hypotheses 4A and 4B). In the filament subanalyses, including only articles where the intraluminal filament method had been used, none of the coating type categories (*Occluding filament type*) seemed to affect the infarct size variability. Although the categories remained in the final enter model, the regression coefficients were small (Fig. 2d). Regarding *Mortality rate*, coating categories did not make it through the backward exclusion (hence, they were not significant).

Background data. The *Infarct size coefficient of variation* (in the total 500 control groups^{9–257,258–342}) was on average $29.5 \pm 19.2\%$ (range 0.9–135.5%) while *Mortality rate* (calculated from the 80 control groups reporting this) was $14 \pm 12\%$ (range 0–83%). *Number of animals per group* was on average 8.4 ± 3.1 (range 3–26). The reported body weight group means were on average 25.6 ± 4.0 g (range 18–45). The average time from induction of cerebral ischemia until sacrifice and damage evaluation was 65.0 ± 104.5 h (range 1.5–1008 h), with a median of 24 h. Frequencies of the different categories of the categorical variables are presented in Fig. 4.

Discussion

The current study shows that the use of *Swiss* and *C57BL6* mice as well as *Permanent* occlusion of the middle cerebral artery renders the lowest infarct size variability. *Emboli/clot* methods, although represented by few control groups, increased variability. Of the methodological factors investigated, only *Swiss* mice was found to have a significant impact on *Mortality rate* by increasing it compared to the reference strain. Effect sizes were large, with many parameters changing the outcomes more than 10% in absolute terms. In addition to the findings pertaining to the hypotheses, several other interesting observations were made, such as the beneficial effects of *Laser Doppler surveillance* on *Infarct size coefficient of variation* and that *Mortality rate* was higher with *Elderly* mice. However, since this study was designed as a hypothesis-driven meta-analysis, results not related to the factor-outcome relations stated a priori should be interpreted with caution and considered merely hypothesis-generating (nevertheless, all findings are presented in Tables S1, S2, S3 and S4 in the Supplementary for readers with special interest in certain methodological parameters).

As mentioned above, comparing all possible combinations of methodological factors experimentally would be a tedious endeavor. However, there are example of studies that investigated one or a few parameters in order to optimize the ischemia model. The majority of these focused on different mouse strains and they did not specifically present or statically compare effect on outcome variability. However, the coefficients of variation can be calculated from mean infarct size and standard deviation similarly to what was done for the regression model in the current meta-analysis. In line with our findings, 129 mice tended to have smaller infarcts with larger infarct size variation compared to C57BL6³⁴³⁻³⁴⁶, although the extent of difference varied. Not corroborated by the current meta-analysis, two of these studies also included BALB/c in the comparison and found that this strain produced infarcts even bigger than those of C57BL6 but with smaller coefficient of variation^{344,345}. One of the studies presented mortality and concluded that BALB/c had the highest rate, C57BL6 the lowest and 129 was in between the other two strains³⁴⁴. We found an increased mortality with the Swiss strain, but only two other categories were represented in that analysis, C57BL6 and other strains.

A few previous articles describe the effects of different middle cerebral artery occlusion durations but the results are discordant. Similar to what we found, both Tsuchia *et al.*³⁴⁷ and Mao *et al.*³⁴⁸ reported lower coefficients of variation for permanent occlusion compared to transient while in another study, the results were the other way around³⁴³. In a study with occlusion durations corresponding to our categories *Short transient* (up to 60 min) and *Long transient* (>60 min), short transient occlusion was more favorable in terms of infarct size variability. Regarding mortality rate, similar inconsistency was found with one study presenting lower values for transient occlusion³⁴⁷, and one for permanent³⁴⁹.

Proper comparisons between methods for ischemia induction in mice are lacking in the literature. This lack is probably explained by the high cost of introducing a new MCAo method in a laboratory, emphasizing the importance of meta-analyses like the current as an alternative. One study looked at the effect of Poly-L-lysine but, like us, found no effect³⁵⁰. Filament coating length^{351,352} and filament size^{347,353} has been investigated but these parameters were not included in our study due to poor reporting in the included articles.

When comparing the current study with the previous rat meta-analysis (described above), some aspect are worth commenting. Similar to what was described herein, emboli methods were found to render larger coefficient of variation of the infarct size than filament, direct and photothrombosis methods⁸. However, infarcts induced by endothelin (not represented in the current mice analysis) were even more inconsistent. Further, although not included in the main hypotheses of the rat study, permanent ischemia had the lowest variability when comparing different occlusion durations both for rats and mice⁸. The rat and mice studies also differ regarding some parameters. For example, no significant differences were found for mice between types of coatings in the filament subanalysis, whereas silicone decreased variability for rats⁸.

The main problem with high infarct size variability is the resulting lack of statistical power if the sample sizes are not adjusted accordingly, which has been discussed in several reviews^{5,354,355}. Statistical power $(1-\beta)$ is often discussed in relation to negative findings, e.g. to evaluate if a study was adequately designed to detect a treatment effect of a substance and hence if the negative results are to trust or not. However, statistical power is of importance also for studies with positive findings (i.e. when a treatment effect is found)^{5,356}. Low statistical power is associated to the publication bias phenomenon since negative findings are generally less likely to be published, which can distort interpretation of meta-analyses³⁵⁷. To support the claim that statistical power in experimental stroke studies is often low, the average power of the studies included in current meta-analysis can be calculated based on the extracted data: The average group sizes were 8.4 and the average coefficient of variation for infarct sizes 29.5%, which at a significance level of 0.05 gives a power of 59% to detect a 30% difference between groups (calculation based on parametric comparison between two-groups, for more three groups or more and non-parametric methods, the number would be even lower). Ethical boards demanding researchers to minimize number of animals (the three Rs principle⁷) might explain why too small group sizes are often used, but economic as well as practical aspects are also likely to contribute. Lack of adequate statistical training or no available statistician to consult regarding these issues should also be mentioned as an option. So in addition to optimizing the model to produce consistent lesions and minimize mortality, it is important to perform a priori power calculations in order to avoid the abovementioned problems.

The issue of mortality is somewhat related to outcome variability and power calculations in that higher mortality require larger group sizes to attain sufficient power. However, there is another side to the problem as well. Regarding the statistical analysis, it is not uncomplicated to incorporate mortality in the standard parametric methods which might explain why this information in most cases is not even mentioned. A non-parametric approach, with mortality included as worst possible outcome, is an option that has been utilized in our laboratory^{358,359} but either way, the absolute minimum should be to report these data. The risk when omitting mortality rate data can be illustrated by the possible scenario of a toxic substance that seems to decrease infarct sizes compared to a placebo group, only because all mice with large infarcts in the treatment group died. In the current meta-analysis, it might seem surprising that the effects on mortality were generally moderate (e.g. no significant effect of occlusion time). However, mortality data was only available for 80/500 control groups. A low number of observations weakens a regression model with many predictor variables, and this should be considered when conclusions are drawn.

The main strength of the present meta-analysis is the large number of articles included, and that the effects of many methodological factors are investigated together in one single statistical model. However, this approach is relatively novel, warranting a discussion about some aspects of the design:

- The impact of each control group were weighed according to number of animals which might be problematic when analyzing coefficient of variation, since researchers knowing that they have large variability in their model probably compensate by including more animals.
- The effect of publication bias has to be considered, as studies with large coefficients of variation might produce negative results that are more likely to remain unpublished.
- Although many possible confounders were recorded and controlled for, accounting for all details of the included experiments is beyond the reach of even a meta-analytical approach. Impact of different vendors and skill of the surgeon are just a couple of factors that could not be assessed. For mathematical reasons, categories have also, as described in the Methods section, been reduced to larger categories, meaning that differences within categories may be lost.
- 500 control groups are included but only 334 articles, meaning that several articles contributed with more than
 one control group. It is not strictly statistically appropriate to analyze these independently but creating categories for all unique studies would have made the statistical analysis impossible.

In conclusion, the methodological choices are of major importance for consistent results and advantageous animal models. However, although it may be relevant to adjust the experimental setup to minimize infarct size variability and mortality rate, other important components such as similarity to the clinical situation have to be taken into consideration. For this reason, it might be motivated in some studies to use the emboli method or elderly animals even though this might increase the outcome variability and mortality, respectively. In either case, the current study enables a more precise estimation of variability and mortality a priori given a certain experimental setup, thereby facilitating proper power calculations.

Methods

Overview. The basic outline for the study was pre-defined and consisted of the following steps:

- 1. Variables to be studied were chosen.
- 2. Data about chosen variables were extracted from relevant articles.
- 3. Variable categories were refined based on extraction results.
- 4. Statistical analyses were performed on variables left after refinement.

Article inclusion. Relevant articles were identified in the Medline database via PubMed using the search string (mcao or "middle cerebral artery occlusion" or "MCA occlusion" or "stroke" or "cerebral ischemia" or "brain ischemia") and (mouse or mice), resulting in over 6,000 hits. The articles were consecutively assessed for inclusion, in order of PubMed identifier, starting with the most recent article January 9th 2012. The inclusion criteria were:

- (I) Article written in English.
- (II) Original research article.
- (III) Experiments performed using living mice.
- (IV) Mice inflicted one single focal cerebral ischemic lesion.
- (V) Infarct sizes measured and results presented.
- (VI) Inclusion of a control group, untreated except for vehicle/placebo treatment.

(VII)Experiment adequately described.

Data extraction. Control group data were extracted from all included articles. If an article described more than one control group, differing in any methodological aspect, these were included separately and analyzed independently. The principle "if it was not described, it was not performed" was adhered to throughout the process. Methodological factors to be extracted were chosen based on our previous rat meta-analysis⁸ and personal experience. See Table 1 for a complete list of all variables that we intended to extract. The goal was to gather as much relevant data as possible in order to build a good statistical model.

To perform a proper power calculation for such a large multiple regression model is a very complex task. Instead, the sample size estimation was based on our previous meta-analysis with a similar design. Furthermore, we performed interim saturation analyses after 400 and 450 included control groups to check when the results had stabilized, i.e. no changes in overall trends occurred. In total, 500 control groups from 334 articles (see Supplementary methods for a complete list of references) were included and 1784 articles were excluded (Fig. 1).

Processing of data. *Category refinement.* To avoid small categories being attributed statistically unsubstantiated explanatory value, categories represented by less than 5 control groups were pooled in an *Other* category for that specific variable. The overall effects on the two outcome variables (*Infarct size variation* and *Mortality*

Factor/outcome measure	Data type	Final categories or unit*	Reference category for regression model
Mouse property factors			
Strain	Category	I. C57BL6	C57BL6
		II. Swiss	
		III. ddY	
		IV. 129	
		V. Mixed C57BL6/129	
		VI. C3H	
		VII. BALB/c	
		VIII. Other strains	
Sex	Category	I. Male	Male
		II. Female	
		III. Ovx female	
		IV. Other sex	
	Category	I. Adult (>2, <12 months)	Adult
		II. Young (0–2 months)	
Age		III. Elderly (>12 months)	
		IV Age not specified	
Weight	Continuous	Grams	NA
Diseases**	Category	I Other diseases	NA
A pesthecia factors	Category	1. Other diseases	14/1
Allestitesia lactors		I. Inhelation anothesia	
		I. IIIIaiauoii ailesuiesia	
		II. Chloral hydrate	
Time of anosthatic	Catagory	III. Ketamine	Inhalation anosthesia
Type of anestnetic	Category	IV. Benzodiazepines and barbiturates	innalation anestnesia
		V. Tribromoethanol	
		VI. Anesthetic not specified	
	Category, Binomial	[No]	[No]
Intubation**		[Yes]	
	Category, Binomial	[No]	[No]
Awakening during occlusion		[Yec]	
		[No]	
Laser Doppler surveillance	Category, Binomial	[No]	[No]
		[les]	[No]
Temperature feedback system	Category, Binomial	[N0]	
Temperature feedback system		[Yes]	
Electroencephalographic surveillance**	Category,	[No]	[No]
	Binomial	[Yes]	
Postoperative antibiotics**	VI. C3HVII. BALB/cVII. Other strainI. MaleI. MaleII. FemaleIII. Ovx femaleIV. Other sexI. Adult (>2, <12 mor	[No]	[No]
•		[Yes]	
Blood pressure monitored	Category, Binomial	[No]	[No]
		[Yes]	
Heart rate monitored	Category, Binomial	[No]	[No]
		[Yes]	
Blood gases/O ₂ saturation analyzed	Category, Binomial	[No]	[No]
		[Yes]	
Plaad hamaglahin analwad	Category, Binomial	[No]	[No]
blood heniogloom analyzed		[Yes]	
Pland alucase analyzed	Category, Binomial	[No]	[No]
Blood glucose analyzed		[Yes]	
Focal ischemia procedure factors			
Type of middle cerebral artery occlusion procedure	Category	I. Intraluminal filament	Intraluminal filament
		II. Direct, mechanical	
		III. Photothrombosis	
		IV. Emboli/clot	
Occlusion duration	Category	I. Short transient (up to	Short transient (up to
	Suitegory	60 minutes)	60 minutes)
Continued			

Factor/outcome measure	Data type	Final categories or unit*	Reference category for regression model
		II. Long transient (>60 minutes)	
		III. Permanent	
	Category	I. Silicone	Silicone
Occluding filament type (only studies using the intraluminal filament method)		II. Poly-L-lysine	
		III. Glue	
		IV. Uncoated	
		V. Other coatings	
Filament coating length (only studies using the intraluminal filament method)**	Continuous	Millimeters	NA
Filament tip diameter**	Continuous	Millimeter	NA
	Category	I. External carotid artery	External carotid artery
Filament insertion (only using the intraluminal filament method)		II. Common carotid artery	
		III. Vessel not specified	
Analysis procedure factors		L	
Time after ischemia for evaluation of damage	Continuous	Hours	NA
Type of staining	Category	I. Triphenyl tetrazolium chloride (TTC)	Triphenyl tetrazolium chloride (TTC)
		II. Acidic/basic stain	
		III. Silver stain	
		IV. Other stains	
Plinding of inforct size measurement procedure	Catagory Pinomial	[No]	[No]
Binding of infarct size measurement procedure	Category, Billonna	[Yes]	
Exclusion based on hemorrhage	Category, Binomial	[No]	[No]
		[Yes]	
Exclusion based on neurological deficit score	Category, Binomial	[No]	[No]
		[Yes]	
Exclusion based on bad clinical condition	Category, Binomial	[No]	[No]
		[Yes]	
Exclusion based on other criteria	Category, Binomial	[No]	[No]
		[Yes]	
Outcome measures			
Infarct size coefficient of variation	Continuous	%	NA
Mortality rate	Continuous	%	NA

Table 1. Extracted factors and outcome measures. *Only categories represented by at least 5 control groups were included in the analysis to avoid statistically inadequate attribution of explanatory value to too small categories. Categories represented by less than 5 control groups were in the analysis included in an Others category. For the same reason, some categories presented here were merged with the Others category for hypotheses 1B, 2B, 3B, 4A and 4B (see Supplementary methods and Tables S2, S3 and S4). Further, some other reductions in number of categories were performed, described in detail in Supplementary methods. **Too few articles reported on this variable; omitted from analysis.

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rate; hypothesis 1A, 1B, 2A, 2B, 3A and 3B) were tested in two independent models and in addition, the filament method subanalysis (hypotheses 4A and 4B) had to be performed separately. Each of the resulting four regression models comprised different numbers of control groups since not all articles reported on mortality and obviously only studies using the intraluminal filament model could be included for the filament subanalysis. Hence, in some cases a category represented by more than 5 control groups in one regression model was reduced to less than 5 groups in another and thus incorporated in the *Others* category, in line with the general category size principle described above. See Supplementary methods for a detailed description of processing of data. Also, in Tables S1, S2, S3 and S4 (Supplementary) the final categories for each regression model are presented.

Excluded variables. The following variables were originally intended to be incorporated into the model, but since none or very few articles reported these data they had to be omitted: *Diseases, Intubation, EEG supervision, Postoperative antibiotics, Filament tip diameter, Filament coating length and Exclusion rate.*

Statistics. As described above, eight main hypotheses were stated a priori:

- 1. Middle cerebral artery occlusion duration affects (A) infarct size variability and (B) mortality.
- 2. Type of focal cerebral ischemia procedure affects (A) infarct size variability and (B) mortality.
- 3. Mouse strain affects (A) infarct size variability and (B) mortality.

4. In studies using the intraluminal filament method, the type of filament affects (A) infarct size variability and (B) mortality.

Since large multiple regression models may suggest a wide range of unexpected associations between variables, a limited set of predefined hypotheses were established to lower the risk of finding falsely significant results due to multiple comparisons (type I errors). Findings not related to these were interpreted with caution and considered merely hypothesis-generating. Due to the risk of type II-errors, corrections for multiple comparisons were not performed.

All categories were dummy-converted before analysis (Table 1). For binomial variables, lack of a specific methodological factor, i.e. [No], was considered the reference category whereas the most common category was chosen as baseline for variables with more than two categories. The data were analyzed using weighted multiple linear regression in two steps. First, a backward exclusion procedure identified factors that contributed significantly to the model and removed the rest. Subsequently, an enter model was performed, in which significant factors identified was manually complemented by lacking dummy variables that were excluded in the previous step (presented in Table S1, S2, S3 and S4). Weighing of cases was performed according to the number of animals in each control group; hence, groups with more animals had larger impact on the statistical model than groups with few animals. Based on the hypotheses, four regression models (one for hypotheses 1A, 2A and 3A; one for hypotheses 1B, 2B and 3B; one for hypothesis 4A and one for hypothesis 4B) were built to test the combined effects of all factors on the two separate outcome measures, Infarct size coefficient of variation or Mortality. In this way, when investigating one of the specific hypotheses, the model controlled for the other predictor variables. The models passed residual checks and multicollinearity tests. All statistical calculations were performed in SPSS (Version 23, IBM Corporation, Armonk, NY, USA). P-values < 0.05 were considered significant. Regarding results from the meta-analysis, 95% confidence interval were provided, otherwise data were presented as mean \pm standard deviation.

References

- 1. Donnan, G. A., Fisher, M., Macleod, M. & Davis, S. M. Stroke. Lancet **371**, 1612–1623, doi: 10.1016/S0140-6736(08)60694-7 (2008).
- Dirnagl, U. & Endres, M. Found in translation: preclinical stroke research predicts human pathophysiology, clinical phenotypes, and therapeutic outcomes. *Stroke* 45, 1510–1518, doi: 10.1161/STROKEAHA.113.004075 (2014).
- 3. O'Collins, V. E. et al. 1,026 experimental treatments in acute stroke. Ann Neurol 59, 467–477, doi: 10.1002/ana.20741 (2006).
- Endres, M. et al. Improving outcome after stroke: overcoming the translational roadblock. Cerebrovasc Dis 25, 268–278, doi: 10.1159/000118039 (2008).
- Dirnagl, U. Bench to bedside: the quest for quality in experimental stroke research. J Cereb Blood Flow Metab 26, 1465–1478, doi: 10.1038/sj.jcbfm.9600298 (2006).
- Macleod, M. R. et al. Evidence for the efficacy of NXY-059 in experimental focal cerebral ischaemia is confounded by study quality. Stroke 39, 2824–2829, doi: 10.1161/STROKEAHA.108.515957 (2008).
- Russel, W. & Burch, R. The principles of humane experimental technique. Universities Federation for Animal Welfare, Wheathamstead, England (1959 (as reprinted 1992)).
- Strom, J. O., Ingberg, E., Theodorsson, A. & Theodorsson, E. Method parameters' impact on mortality and variability in rat stroke experiments: a meta-analysis. BMC Neurosci 14, 41, doi: 10.1186/1471-2202-14-41 (2013).
- Moussaieff, A. et al. Protective effects of incensole acetate on cerebral ischemic injury. Brain Res 1443, 89–97, doi: 10.1016/j. brainres.2012.01.001 (2012).
- Wang, B. et al. Histone deacetylase inhibition activates transcription factor Nrf2 and protects against cerebral ischemic damage. Free Radic Biol Med 52, 928–936, doi: 10.1016/j.freeradbiomed.2011.12.006 (2012).
- Zheng, C. et al. NAD(+) administration decreases ischemic brain damage partially by blocking autophagy in a mouse model of brain ischemia. Neurosci Lett 512, 67–71, doi: 10.1016/j.neulet.2012.01.007 (2012).
- Harada, S. *et al.* Honokiol suppresses the development of post-ischemic glucose intolerance and neuronal damage in mice. J Nat Med 66, 591–599, doi: 10.1007/s11418-011-0623-x (2012).
- 13. Lu, H. et al. Netrin-1 hyperexpression in mouse brain promotes angiogenesis and long-term neurological recovery after transient focal ischemia. Stroke 43, 838–843, doi: 10.1161/STROKEAHA.111.635235 (2012).
- 14. Fujioka, M. *et al.* ADAMTS13 gene deletion enhances plasma high-mobility group box1 elevation and neuroinflammation in brain ischemia-reperfusion injury. *Neurol Sci* 33, 1107–1115, doi: 10.1007/s10072-011-0913-9 (2012).
- Park, J. S. et al. Anti-inflammatory mechanism of compound K in activated microglia and its neuroprotective effect on experimental stroke in mice. J Pharmacol Exp Ther 341, 59–67, doi: 10.1124/jpet.111.189035 (2012).
- Elvington, A. *et al.* Pathogenic natural antibodies propagate cerebral injury following ischemic stroke in mice. *J Immunol* 188, 1460–1468, doi: 10.4049/jimmunol.1102132 (2012).
- Colak, G. & Johnson, G. V. Complete transglutaminase 2 ablation results in reduced stroke volumes and astrocytes that exhibit increased survival in response to ischemia. *Neurobiol Dis* 45, 1042–1050, doi: 10.1016/j.nbd.2011.12.023 (2012).
- Harada, S., Fujita-Hamabe, W. & Tokuyama, S. Ameliorating effect of hypothalamic brain-derived neurotrophic factor against impaired glucose metabolism after cerebral ischemic stress in mice. J Pharmacol Sci 118, 109–116 (2012).
- Liang, J. et al. Participation of MCP-induced protein 1 in lipopolysaccharide preconditioning-induced ischemic stroke tolerance by regulating the expression of proinflammatory cytokines. J Neuroinflammation 8, 182, doi: 10.1186/1742-2094-8-182 (2011).
- Shang, J., Liu, N., Tanaka, N. & Abe, K. Expressions of hypoxic stress sensor proteins after transient cerebral ischemia in mice. J Neurosci Res 90, 648–655, doi: 10.1002/jnr.22776 (2012).
- Hoda, M. N. *et al.* Sex-independent neuroprotection with minocycline after experimental thromboembolic stroke. *Exp Transl Stroke Med* 3, 16, doi: 10.1186/2040-7378-3-16 (2011).
- 22. Hase, Y. *et al.* Cilostazol, a phosphodiesterase inhibitor, prevents no-reflow and hemorrhage in mice with focal cerebral ischemia. *Exp Neurol* **233**, 523–533, doi: 10.1016/j.expneurol.2011.11.038 (2012).
- Kim, G. S. et al. Release of mitochondrial apoptogenic factors and cell death are mediated by CK2 and NADPH oxidase. J Cereb Blood Flow Metab 32, 720–730, doi: 10.1038/jcbfm.2011.176 (2012).
- Venna, V. R., Li, J., Benashski, S. E., Tarabishy, S. & McCullough, L. D. Preconditioning induces sustained neuroprotection by downregulation of adenosine 5'-monophosphate-activated protein kinase. *Neuroscience* 201, 280–287, doi: 10.1016/j. neuroscience.2011.11.014 (2012).
- Masuko, T. et al. Antagonism of NMDA receptors by butanesulfonyl-homospermine guanidine and neuroprotective effects in in vitro and in vivo. Neurosci Lett 506, 251–255, doi: 10.1016/j.neulet.2011.11.017 (2012).

- Denes, A., Ferenczi, S. & Kovacs, K. J. Systemic inflammatory challenges compromise survival after experimental stroke via augmenting brain inflammation, blood- brain barrier damage and brain oedema independently of infarct size. J Neuroinflammation 8, 164, doi: 10.1186/1742-2094-8-164 (2011).
- De Silva, T. M., Brait, V. H., Drummond, G. R., Sobey, C. G. & Miller, A. A. Nox2 oxidase activity accounts for the oxidative stress and vasomotor dysfunction in mouse cerebral arteries following ischemic stroke. *PLoS One* 6, e28393, doi: 10.1371/journal. pone.0028393 (2011).
- Nagai, M. et al. Role of blood cell-associated angiotensin II type 1 receptors in the cerebral microvascular response to ischemic stroke during angiotensin-induced hypertension. Exp Transl Stroke Med 3, 15, doi: 10.1186/2040-7378-3-15 (2011).
- Cipriani, R. et al. CX3CL1 is neuroprotective in permanent focal cerebral ischemia in rodents. J Neurosci 31, 16327–16335, doi: 10.1523/JNEUROSCI.3611-11.2011 (2011).
- Ogle, M. E., Gu, X., Espinera, A. R. & Wei, L. Inhibition of prolyl hydroxylases by dimethyloxaloylglycine after stroke reduces ischemic brain injury and requires hypoxia inducible factor-1alpha. *Neurobiol Dis* 45, 733–742, doi: 10.1016/j.nbd.2011.10.020 (2012).
- Liu, F., Benashski, S. E., Xu, Y., Siegel, M. & McCullough, L. D. Effects of chronic and acute oestrogen replacement therapy in aged animals after experimental stroke. J Neuroendocrinol 24, 319–330, doi: 10.1111/j.1365-2826.2011.02248.x (2012).
- 32. Pleines, I. *et al.* Megakaryocyte-specific RhoA deficiency causes macrothrombocytopenia and defective platelet activation in hemostasis and thrombosis. *Blood* **119**, 1054–1063, doi: 10.1182/blood-2011-08-372193 (2012).
- Strecker, J. K. et al. Monocyte chemoattractant protein-1-deficiency impairs the expression of IL-6, IL-1beta and G-CSF after transient focal ischemia in mice. PLoS One 6, e25863, doi: 10.1371/journal.pone.0025863 (2011).
- Sieber, M. W., Claus, R. A., Witte, O. W. & Frahm, C. Attenuated inflammatory response in aged mice brains following stroke. PLoS One 6, e26288, doi: 10.1371/journal.pone.0026288 (2011).
- Tang, X. N., Zheng, Z., Giffard, R. G. & Yenari, M. A. Significance of marrow-derived nicotinamide adenine dinucleotide phosphate oxidase in experimental ischemic stroke. Ann Neurol 70, 606–615, doi: 10.1002/ana.22476 (2011).
- 36. Li, S. Y. *et al.* Lutein enhances survival and reduces neuronal damage in a mouse model of ischemic stroke. *Neurobiol Dis* 45, 624–632, doi: 10.1016/j.nbd.2011.10.008 (2012).
- Vartanian, K. B. et al. LPS preconditioning redirects TLR signaling following stroke: TRIF-IRF3 plays a seminal role in mediating tolerance to ischemic injury. J Neuroinflammation 8, 140, doi: 10.1186/1742-2094-8-140 (2011).
- Ren, X. et al. Myelin specific cells infiltrate MCAO lesions and exacerbate stroke severity. Metab Brain Dis 27, 7–15, doi: 10.1007/ s11011-011-9267-5 (2012).
- Yung, L. M. et al. Sphingosine kinase 2 mediates cerebral preconditioning and protects the mouse brain against ischemic injury. Stroke 43, 199–204, doi: 10.1161/STROKEAHA.111.626911 (2012).
- Rodrigues, S. F. & Granger, D. N. Cerebral microvascular inflammation in DOCA salt-induced hypertension: role of angiotensin II and mitochondrial superoxide. J Cereb Blood Flow Metab 32, 368–375, doi: 10.1038/jcbfm.2011.139 (2012).
- 41. Zhang, J. Y. et al. Leptin attenuates cerebral ischemia/reperfusion injury partially by CGRP expression. Eur J Pharmacol 671, 61–69, doi: 10.1016/j.ejphar.2011.09.170 (2011).
- Liang, X. et al. Signaling via the prostaglandin E(2) receptor EP4 exerts neuronal and vascular protection in a mouse model of cerebral ischemia. J Clin Invest 121, 4362–4371, doi: 10.1172/JCI46279 (2011).
- Akiyoshi, K. et al. Recombinant T cell receptor ligands improve outcome after experimental cerebral ischemia. Transl Stroke Res 2, 404–410, doi: 10.1007/s12975-011-0085-1 (2011).
- Morancho, A. et al. A new method for focal transient cerebral ischaemia by distal compression of the middle cerebral artery. Neuropathol Appl Neurobiol 38, 617–627, doi: 10.1111/j.1365-2990.2012.01252.x (2012).
- Wang, P., Tian, W. W., Song, J., Guan, Y. F. & Miao, C. Y. Deficiency of NG²⁺ cells contributes to the susceptibility of stroke-prone spontaneously hypertensive rats. CNS Neurosci Ther 17, 327–332, doi: 10.1111/j.1755-5949.2011.00265.x (2011).
- 46. Texel, S. J. et al. Ceruloplasmin deficiency reduces levels of iron and BDNF in the cortex and striatum of young mice and increases their vulnerability to stroke. PLoS One 6, e25077, doi: 10.1371/journal.pone.0025077 (2011).
- Jung, J. E., Kim, G. S. & Chan, P. H. Neuroprotection by interleukin-6 is mediated by signal transducer and activator of transcription 3 and antioxidative signaling in ischemic stroke. *Stroke* 42, 3574–3579, doi: 10.1161/STROKEAHA.111.626648 (2011).
- Chang, C. C. et al. Prodigiosin inhibits gp91(phox) and iNOS expression to protect mice against the oxidative/nitrosative brain injury induced by hypoxia-ischemia. Toxicol Appl Pharmacol 257, 137–147, doi: 10.1016/j.taap.2011.08.027 (2011).
- Balkaya, M. et al. Stress worsens endothelial function and ischemic stroke via glucocorticoids. Stroke 42, 3258–3264, doi: 10.1161/ STROKEAHA.110.607705 (2011).
- Turtzo, L. C., Siegel, C. & McCullough, L. D. X chromosome dosage and the response to cerebral ischemia. J Neurosci 31, 13255–13259, doi: 10.1523/JNEUROSCI.0621-11.2011 (2011).
- De Meyer, S. F., Schwarz, T., Schatzberg, D. & Wagner, D. D. Platelet glycoprotein Ibalpha is an important mediator of ischemic stroke in mice. *Exp Transl Stroke Med* 3, 9, doi: 10.1186/2040-7378-3-9 (2011).
- Kilic, U. *et al.* Evidence that membrane-bound G protein-coupled melatonin receptors MT1 and MT2 are not involved in the neuroprotective effects of melatonin in focal cerebral ischemia. *J Pineal Res* 52, 228–235, doi: 10.1111/j.1600-079X.2011.00932.x (2012).
- 53. Pham, V. *et al.* Insulin-regulated aminopeptidase deficiency provides protection against ischemic stroke in mice. *J Neurotrauma* 29, 1243–1248, doi: 10.1089/neu.2011.1824 (2012).
- Shu, L. et al. Inhibition of neuron-specific CREB dephosphorylation is involved in propofol and ketamine-induced neuroprotection against cerebral ischemic injuries of mice. Neurochem Res 37, 49–58, doi: 10.1007/s11064-011-0582-3 (2012).
- Pfeilschifter, W. et al. Activation of sphingosine kinase 2 is an endogenous protective mechanism in cerebral ischemia. Biochem Biophys Res Commun 413, 212–217, doi: 10.1016/j.bbrc.2011.08.070 (2011).
- Wang, B., Cao, W., Biswal, S. & Dore, S. Carbon monoxide-activated Nrf2 pathway leads to protection against permanent focal cerebral ischemia. *Stroke* 42, 2605–2610, doi: 10.1161/STROKEAHA.110.607101 (2011).
- Nakka, V. P. et al. Increased cerebral protein ISGylation after focal ischemia is neuroprotective. J Cereb Blood Flow Metab 31, 2375–2384, doi: 10.1038/jcbfm.2011.103 (2011).
- Ma, Y., Mehta, S. L., Lu, B. & Li, P. A. Deficiency in the inner mitochondrial membrane peptidase 2-like (Immp21) gene increases ischemic brain damage and impairs mitochondrial function. *Neurobiol Dis* 44, 270–276, doi: 10.1016/j.nbd.2011.06.019 (2011).
- Kim, H. W., Cho, K. J., Lee, S. K. & Kim, G. W. Apoptosis signal-regulating kinase 1 (Ask1) targeted small interfering RNA on ischemic neuronal cell death. Brain Res 1412, 73–78, doi: 10.1016/j.brainres.2011.07.018 (2011).
- 60. Niu, F. N. *et al.* Targeted mutation of Fas ligand gene attenuates brain inflammation in experimental stroke. *Brain Behav Immun* **26**, 61–71, doi: 10.1016/j.bbi.2011.07.235 (2012).
- Shi, Y., Chanana, V., Watters, J. J., Ferrazzano, P. & Sun, D. Role of sodium/hydrogen exchanger isoform 1 in microglial activation and proinflammatory responses in ischemic brains. J Neurochem 119, 124–135, doi: 10.1111/j.1471-4159.2011.07403.x (2011).
- Brede, M. *et al.* alpha(2)-adrenoceptors do not mediate neuroprotection in acute ischemic stroke in mice. J Cereb Blood Flow Metab 31, e1–7, doi: 10.1038/jcbfm.2011.110 (2011).
- 63. Wang, H. W. *et al.* Deciphering the neuroprotective mechanisms of Bu-yang Huan-wu decoction by an integrative neurofunctional and genomic approach in ischemic stroke mice. *J Ethnopharmacol* **138**, 22–33, doi: 10.1016/j.jep.2011.06.033 (2011).

- Doeppner, T. R. et al. Enhancement of endogenous neurogenesis in ephrin-B3 deficient mice after transient focal cerebral ischemia. Acta Neuropathol 122, 429–442, doi: 10.1007/s00401-011-0856-5 (2011).
- Iwanami, J. et al. Effect of angiotensin II type 2 receptor deletion in hematopoietic cells on brain ischemia-reperfusion injury. Hypertension 58, 404–409, doi: 10.1161/HYPERTENSIONAHA.111.177873 (2011).
- 66. ElÂli, A. & Hermann, D. M. Liver X receptor activation enhances blood-brain barrier integrity in the ischemic brain and increases the abundance of ATP-binding cassette transporters ABCB1 and ABCC1 on brain capillary cells. *Brain Pathol* 22, 175–187, doi: 10.1111/j.1750-3639.2011.00517.x (2012).
- Ren, X., Akiyoshi, K., Vandenbark, A. A., Hurn, P. D. & Offner, H. Programmed death-1 pathway limits central nervous system inflammation and neurologic deficits in murine experimental stroke. *Stroke* 42, 2578–2583, doi: 10.1161/STROKEAHA.111.613182 (2011).
- Jin, K., Xie, L., Sun, F., Mao, X. & Greenberg, D. A. Corpus callosum and experimental stroke: studies in callosotomized rats and acallosal mice. *Stroke* 42, 2584–2588, doi: 10.1161/STROKEAHA.111.613349 (2011).
- Park, H. A. et al. Natural vitamin E alpha-tocotrienol protects against ischemic stroke by induction of multidrug resistanceassociated protein 1. Stroke 42, 2308–2314, doi: 10.1161/STROKEAHA.110.608547 (2011).
- Lu, C. *et al.* TLR2 ligand induces protection against cerebral ischemia/reperfusion injury via activation of phosphoinositide 3-kinase/Akt signaling. *J Immunol* 187, 1458–1466, doi: 10.4049/jimmunol.1003428 (2011).
- Gibson, C. L., Coomber, B. & Murphy, S. P. Progesterone is neuroprotective following cerebral ischaemia in reproductively ageing female mice. *Brain* 134, 2125–2133, doi: 10.1093/brain/awr132 (2011).
- 72. Liesz, A. *et al.* FTY720 reduces post-ischemic brain lymphocyte influx but does not improve outcome in permanent murine cerebral ischemia. *PLoS One* **6**, e21312, doi: 10.1371/journal.pone.0021312 (2011).
- Ye, R. et al. Ginsenoside Rd attenuates redox imbalance and improves stroke outcome after focal cerebral ischemia in aged mice. Neuropharmacology 61, 815–824, doi: 10.1016/j.neuropharm.2011.05.029 (2011).
- Wang, Z. et al. Fusion of core pathways reveals a horizontal synergistic mechanism underlying combination therapy. Eur J Pharmacol 667, 278-286, doi: 10.1016/j.ejphar.2011.05.046 (2011).
- Ren, X. *et al.* Regulatory B cells limit CNS inflammation and neurologic deficits in murine experimental stroke. *J Neurosci* 31, 8556–8563, doi: 10.1523/JNEUROSCI.1623-11.2011 (2011).
- Stevens, S. L. *et al.* Multiple preconditioning paradigms converge on interferon regulatory factor-dependent signaling to promote tolerance to ischemic brain injury. *J Neurosci* 31, 8456–8463, doi: 10.1523/JNEUROSCI.0821-11.2011 (2011).
- Min, J. et al. Neuroprotective effect of cyanidin-3-O-glucoside anthocyanin in mice with focal cerebral ischemia. Neurosci Lett 500, 157–161, doi: 10.1016/j.neulet.2011.05.048 (2011).
- 78. Su, E. J. *et al.* The thrombomodulin analog Solulin promotes reperfusion and reduces infarct volume in a thrombotic stroke model. *J Thromb Haemost* **9**, 1174–1182, doi: 10.1111/j.1538-7836.2011.04269.x (2011).
- Shin, J. A., Choi, J. H., Choi, Y. H. & Park, E. M. Conserved aquaporin 4 levels associated with reduction of brain edema are mediated by estrogen in the ischemic brain after experimental stroke. *Biochim Biophys Acta* 1812, 1154–1163, doi: 10.1016/j. bbadis.2011.05.004 (2011).
- Yamamoto, H. et al. NDRG4 protein-deficient mice exhibit spatial learning deficits and vulnerabilities to cerebral ischemia. J Biol Chem 286, 26158–26165, doi: 10.1074/jbc.M111.256446 (2011).
- Choe, C. U. et al. CD38 exacerbates focal cytokine production, postischemic inflammation and brain injury after focal cerebral ischemia. PLoS One 6, e19046, doi: 10.1371/journal.pone.0019046 (2011).
- Andrabi, S. A. et al. Iduna protects the brain from glutamate excitotoxicity and stroke by interfering with poly(ADP-ribose) polymer-induced cell death. Nat Med 17, 692–699, doi: 10.1038/nm.2387 (2011).
- Xiong, X. et al. Increased brain injury and worsened neurological outcome in interleukin-4 knockout mice after transient focal cerebral ischemia. Stroke 42, 2026–2032, doi: 10.1161/STROKEAHA.110.593772 (2011).
- Jia, J. et al. Sex differences in neuroprotection provided by inhibition of TRPM2 channels following experimental stroke. J Cereb Blood Flow Metab 31, 2160–2168, doi: 10.1038/jcbfm.2011.77 (2011).
- Tsukuda, K. *et al.* Irbesartan attenuates ischemic brain damage by inhibition of MCP-1/CCR2 signaling pathway beyond AT(1) receptor blockade. *Biochem Biophys Res Commun* 409, 275–279, doi: 10.1016/j.bbrc.2011.04.142 (2011).
- Liu, N., Shang, J., Tian, F., Nishi, H. & Abe, K. In vivo optical imaging for evaluating the efficacy of edaravone after transient cerebral ischemia in mice. Brain Res 1397, 66–75, doi: 10.1016/j.brainres.2011.04.038 (2011).
- Li, L. *et al.* Cerebroside-A provides potent neuroprotection after cerebral ischaemia through reducing glutamate release and Ca(2) (+) influx of NMDA receptors. *Int J Neuropsychopharmacol* 15, 497–507, doi: 10.1017/S1461145711000654 (2012).
- Zhang, Y. *et al.* Dipyrone inhibits neuronal cell death and diminishes hypoxic/ischemic brain injury. *Neurosurgery* 69, 942–956, doi: 10.1227/NEU.0b013e318222afb2 (2011).
- Jin, R. et al. Phosphatidylinositol-3-kinase gamma plays a central role in blood-brain barrier dysfunction in acute experimental stroke. Stroke 42, 2033–2044, doi: 10.1161/STROKEAHA.110.601369 (2011).
- Custodis, F. et al. Heart rate contributes to the vascular effects of chronic mental stress: effects on endothelial function and ischemic brain injury in mice. Stroke 42, 1742–1749, doi: 10.1161/STROKEAHA.110.598607 (2011).
- Oyagi, A. et al. Forebrain specific heparin-binding epidermal growth factor-like growth factor knockout mice show exacerbated ischemia and reperfusion injury. Neuroscience 185, 116–124, doi: 10.1016/j.neuroscience.2011.04.034 (2011).
- Jin, R. C. et al. Glutathione peroxidase-3 deficiency promotes platelet-dependent thrombosis in vivo. Circulation 123, 1963–1973, doi: 10.1161/CIRCULATIONAHA.110.000034 (2011).
- Berny-Lang, M. A. et al. Thrombin mutant W215A/E217A treatment improves neurological outcome and reduces cerebral infarct size in a mouse model of ischemic stroke. Stroke 42, 1736–1741, doi: 10.1161/STROKEAHA.110.603811 (2011).
- Dolga, A. M. et al. KCa2 channels activation prevents [Ca²⁺]i deregulation and reduces neuronal death following glutamate toxicity and cerebral ischemia. Cell Death Dis 2, e147, doi: 10.1038/cddis.2011.30 (2011).
- Hoffmann, U., Lee, J. H., Qin, T., Eikermann-Haerter, K. & Ayata, C. Gabapentin reduces infarct volume but does not suppress peri-infarct depolarizations. J Cereb Blood Flow Metab 31, 1578–1582, doi: 10.1038/jcbfm.2011.50 (2011).
- Chen, J. et al. Circulating endothelial progenitor cells and cellular membrane microparticles in db/db diabetic mouse: possible implications in cerebral ischemic damage. Am J Physiol Endocrinol Metab 301, E62–71, doi: 10.1152/ajpendo.00026.2011 (2011).
- Teramoto, S. *et al.* Exendin-4, a glucagon-like peptide-1 receptor agonist, provides neuroprotection in mice transient focal cerebral ischemia. *J Cereb Blood Flow Metab* 31, 1696–1705, doi: 10.1038/jcbfm.2011.51 (2011).
- Li, N. et al. Age-related differences in experimental stroke: possible involvement of mitochondrial dysfunction and oxidative damage. Rejuvenation Res 14, 261–273, doi: 10.1089/rej.2010.1115 (2011).
- Arumugam, T. V. *et al.* Evidence that gamma-secretase-mediated Notch signaling induces neuronal cell death via the nuclear factor-kappaB-Bcl-2-interacting mediator of cell death pathway in ischemic stroke. *Mol Pharmacol* 80, 23–31, doi: 10.1124/ mol.111.071076 (2011).
- Takamiya, M. et al. Neurological and pathological improvements of cerebral infarction in mice with platinum nanoparticles. J Neurosci Res 89, 1125–1133, doi: 10.1002/jnr.22622 (2011).
- 101. Shen, H. Y. *et al.* Adenosine kinase determines the degree of brain injury after ischemic stroke in mice. *J Cereb Blood Flow Metab* **31**, 1648–1659, doi: 10.1038/jcbfm.2011.30 (2011).

- 102. Kim, J. H. et al. The Traditional Herbal Medicine, Dangkwisoo-San, Prevents Cerebral Ischemic Injury through Nitric Oxide-Dependent Mechanisms. Evid Based Complement Alternat Med 2011, 718302, doi: 10.1155/2011/718302 (2011).
- Zhao, R., Shi, W. Z., Zhang, Y. M., Fang, S. H. & Wei, E. Q. Montelukast, a cysteinyl leukotriene receptor-1 antagonist, attenuates chronic brain injury after focal cerebral ischaemia in mice and rats. *J Pharm Pharmacol* 63, 550–557, doi: 10.1111/j.2042-7158. 2010.01238.x (2011).
- Liu, F. et al. Age-related changes in AMP-activated protein kinase after stroke. Age (Dordr) 34, 157–168, doi: 10.1007/s11357-011-9214-8 (2012).
- Miyazaki, T. *et al.* Distinct effects of tissue-type plasminogen activator and SMTP-7 on cerebrovascular inflammation following thrombolytic reperfusion. *Stroke* 42, 1097–1104, doi: 10.1161/STROKEAHA.110.598359 (2011).
- Zhang, N. et al. Hypoxic preconditioning induced neuroprotection against cerebral ischemic injuries and its cPKCgammamediated molecular mechanism. Neurochem Int 58, 684–692, doi: 10.1016/j.neuint.2011.02.007 (2011).
- 107. Shah, Z. A., Nada, S. E. & Dore, S. Heme oxygenase 1, beneficial role in permanent ischemic stroke and in Gingko biloba (EGb 761) neuroprotection. *Neuroscience* 180, 248–255, doi: 10.1016/j.neuroscience.2011.02.031 (2011).
- Kleinschnitz, C. et al. Glucocorticoid insensitivity at the hypoxic blood-brain barrier can be reversed by inhibition of the proteasome. Stroke 42, 1081–1089, doi: 10.1161/STROKEAHA.110.592238 (2011).
- 109. Qian, Y. R. et al. Neuroprotection by valproic Acid in mouse models of permanent and transient focal cerebral ischemia. Korean J Physiol Pharmacol 14, 435–440, doi: 10.4196/kjpp.2010.14.6.435 (2010).
- Liu, F. *et al.* Sex differences in the response to poly(ADP-ribose) polymerase-1 deletion and caspase inhibition after stroke. *Stroke* 42, 1090–1096, doi: 10.1161/STROKEAHA.110.594861 (2011).
- 111. Chen, H., Kim, G. S., Okami, N., Narasimhan, P. & Chan, P. H. NADPH oxidase is involved in post-ischemic brain inflammation. *Neurobiol Dis* 42, 341–348, doi: 10.1016/j.nbd.2011.01.027 (2011).
- 112. Lu, Q. et al. Betulinic acid protects against cerebral ischemia-reperfusion injury in mice by reducing oxidative and nitrosative stress. *Nitric Oxide* 24, 132–138, doi: 10.1016/j.niox.2011.01.007 (2011).
- Bu, X. *et al.* Proteomic analysis of cPKCbetaII-interacting proteins involved in HPC-induced neuroprotection against cerebral ischemia of mice. *J Neurochem* 117, 346–356, doi: 10.1111/j.1471-4159.2011.07209.x (2011).
- 114. Bahjat, F. R. et al. Proof of concept: pharmacological preconditioning with a Toll-like receptor agonist protects against cerebrovascular injury in a primate model of stroke. J Cereb Blood Flow Metab 31, 1229–1242, doi: 10.1038/jcbfm.2011.6 (2011).
- 115. Wei, Y. *et al.* Fingolimod provides long-term protection in rodent models of cerebral ischemia. *Ann Neurol* **69**, 119–129, doi: 10.1002/ana.22186 (2011).
- Harada, S., Fujita-Hamabe, W. & Tokuyama, S. Effect of orexin-A on post-ischemic glucose intolerance and neuronal damage. J Pharmacol Sci 115, 155–163 (2011).
- 117. Yilmaz, G. *et al.* Selectin-mediated recruitment of bone marrow stromal cells in the postischemic cerebral microvasculature. *Stroke* **42**, 806–811, doi: 10.1161/STROKEAHA.110.597088 (2011).
- Zhao, Y. & Rempe, D. A. Prophylactic neuroprotection against stroke: low-dose, prolonged treatment with deferoxamine or deferasirox establishes prolonged neuroprotection independent of HIF-1 function. J Cereb Blood Flow Metab 31, 1412–1423, doi: 10.1038/jcbfm.2010.230 (2011).
- 119. Bradford, S. T., Stamatovic, S. M., Dondeti, R. S., Keep, R. F. & Andjelkovic, A. V. Nicotine aggravates the brain postischemic inflammatory response. *Am J Physiol Heart Circ Physiol* **300**, H1518–1529, doi: 10.1152/ajpheart.00928.2010 (2011).
- Deplanque, D., Venna, V. R. & Bordet, R. Brain ischemia changes the long term response to antidepressant drugs in mice. *Behav Brain Res* 219, 367–372, doi: 10.1016/j.bbr.2011.01.003 (2011).
- 121. Ikegame, Y. *et al.* Comparison of mesenchymal stem cells from adipose tissue and bone marrow for ischemic stroke therapy. *Cytotherapy* **13**, 675–685, doi: 10.3109/14653249.2010.549122 (2011).
- 122. Qin, L., Kim, E., Ratan, R., Lee, F. S. & Cho, S. Genetic variant of BDNF (Val66Met) polymorphism attenuates stroke-induced angiogenic responses by enhancing anti-angiogenic mediator CD36 expression. J Neurosci 31, 775–783, doi: 10.1523/ JNEUROSCI.4547-10.2011 (2011).
- Valerio, A. et al. Glycogen synthase kinase-3 inhibition reduces ischemic cerebral damage, restores impaired mitochondrial biogenesis and prevents ROS production. J Neurochem 116, 1148–1159, doi: 10.1111/j.1471-4159.2011.07171.x (2011).
- 124. Liao, Y. *et al.* Neuronal Ca²⁺-activated K⁺ channels limit brain infarction and promote survival. *PLoS One* **5**, e15601, doi: 10.1371/journal.pone.0015601 (2010).
- Morrison, H. *et al.* The contribution of mannose binding lectin to reperfusion injury after ischemic stroke. *Curr Neurovasc Res* 8, 52–63 (2011).
- 126. Ferrazzano, P. et al. Inhibiting the Na⁺/H⁺ exchanger reduces reperfusion injury: a small animal MRI study. Front Biosci (Elite Ed.) 3, 81–88 (2011).
- 127. Moyanova, S. G. *et al.* Protective role for type 4 metabotropic glutamate receptors against ischemic brain damage. *J Cereb Blood Flow Metab* **31**, 1107–1118, doi: 10.1038/jcbfm.2010.201 (2011).
- 128. Kraft, P. *et al.* Deficiency of vasodilator-stimulated phosphoprotein (VASP) increases blood-brain-barrier damage and edema formation after ischemic stroke in mice. *PLoS One* **5**, e15106, doi: 10.1371/journal.pone.0015106 (2010).
- 129. Ishiguro, M. *et al.* Phosphodiesterase-III inhibitor prevents hemorrhagic transformation induced by focal cerebral ischemia in mice treated with tPA. *PLoS One* **5**, e15178, doi: 10.1371/journal.pone.0015178 (2010).
- Li, L. & Zuo, Z. Glutamate transporter type 3 knockout reduces brain tolerance to focal brain ischemia in mice. J Cereb Blood Flow Metab 31, 1283–1292, doi: 10.1038/jcbfm.2010.222 (2011).
- Brait, V. H. et al. Chemokine-related gene expression in the brain following ischemic stroke: no role for CXCR2 in outcome. Brain Res 1372, 169–179, doi: 10.1016/j.brainres.2010.11.087 (2011).
- 132. Tureyen, K., Bowen, K., Liang, J., Dempsey, R. J. & Vemuganti, R. Exacerbated brain damage, edema and inflammation in type-2 diabetic mice subjected to focal ischemia. *J Neurochem* **116**, 499–507, doi: 10.1111/j.1471-4159.2010.07127.x (2011).
- Schindowski, K. *et al.* Regulation of GDF-15, a distant TGF-beta superfamily member, in a mouse model of cerebral ischemia. *Cell Tissue Res* 343, 399–409, doi: 10.1007/s00441-010-1090-5 (2011).
- 134. Doeppner, T. R. *et al.* Acute hepatocyte growth factor treatment induces long-term neuroprotection and stroke recovery via mechanisms involving neural precursor cell proliferation and differentiation. *J Cereb Blood Flow Metab* **31**, 1251–1262, doi: 10.1038/jcbfm.2010.211 (2011).
- Bartolini, A., Vigliani, M. C., Magrassi, L., Vercelli, A. & Rossi, F. G-CSF administration to adult mice stimulates the proliferation of microglia but does not modify the outcome of ischemic injury. *Neurobiol Dis* 41, 640–649, doi: 10.1016/j.nbd.2010.11.013 (2011).
- Yoon, J. S. et al. Pregabalin suppresses calcium-mediated proteolysis and improves stroke outcome. Neurobiol Dis 41, 624–629, doi: 10.1016/j.nbd.2010.11.011 (2011).
- 137. Garcia-Yebenes, I. *et al.* A mouse model of hemorrhagic transformation by delayed tissue plasminogen activator administration after *in situ* thromboembolic stroke. *Stroke* **42**, 196–203, doi: 10.1161/STROKEAHA.110.600452 (2011).
- Cai, B. et al. TAT-mediated delivery of neuroglobin protects against focal cerebral ischemia in mice. Exp Neurol 227, 224–231, doi: 10.1016/j.expneurol.2010.11.009 (2011).
- Ren, X., Akiyoshi, K., Vandenbark, A. A., Hurn, P. D. & Offner, H. CD4⁺ FoxP3⁺ regulatory T-cells in cerebral ischemic stroke. *Metab Brain Dis* 26, 87–90, doi: 10.1007/s11011-010-9226-6 (2011).

- 140. Krajewska, M. et al. Endoplasmic reticulum protein BI-1 modulates unfolded protein response signaling and protects against stroke and traumatic brain injury. Brain Res 1370, 227–237, doi: 10.1016/j.brainres.2010.11.015 (2011).
- Inacio, A. R., Ruscher, K., Leng, L., Bucala, R. & Deierborg, T. Macrophage migration inhibitory factor promotes cell death and aggravates neurologic deficits after experimental stroke. J Cereb Blood Flow Metab 31, 1093–1106, doi: 10.1038/jcbfm.2010.194 (2011).
- 142. Fu, A., Hui, E. K., Lu, J. Z., Boado, R. J. & Pardridge, W. M. Neuroprotection in stroke in the mouse with intravenous erythropoietin-Trojan horse fusion protein. Brain Res 1369, 203–207, doi: 10.1016/j.brainres.2010.10.097 (2011).
- Morrison, H., McKee, D. & Ritter, L. Systemic neutrophil activation in a mouse model of ischemic stroke and reperfusion. *Biol Res Nurs* 13, 154–163, doi: 10.1177/1099800410384500 (2011).
- 144. Macrez, R., Bezin, L., Le Mauff, B., Ali, C. & Vivien, D. Functional occurrence of the interaction of tissue plasminogen activator with the NR1 Subunit of N-methyl-D-aspartate receptors during stroke. *Stroke* 41, 2950–2955, doi: 10.1161/ STROKEAHA.110.592360 (2010).
- Konoeda, F. et al. Therapeutic effect of IL-12/23 and their signaling pathway blockade on brain ischemia model. Biochem Biophys Res Commun 402, 500–506, doi: 10.1016/j.bbrc.2010.10.058 (2010).
- 146. Liu, N. *et al. In vivo* optical imaging of early-stage apoptosis in mouse brain after transient cerebral ischemia. *J Neurosci Res* 88, 3488–3497, doi: 10.1002/jnr.22489 (2010).
- 147. Tanaka, Y., Tanaka, R., Liu, M., Hattori, N. & Urabe, T. Cilostazol attenuates ischemic brain injury and enhances neurogenesis in the subventricular zone of adult mice after transient focal cerebral ischemia. *Neuroscience* **171**, 1367–1376, doi: 10.1016/j. neuroscience.2010.10.008 (2010).
- 148. Kleinschnitz, C. *et al.* Post-stroke inhibition of induced NADPH oxidase type 4 prevents oxidative stress and neurodegeneration. *PLoS Biol* 8, doi: 10.1371/journal.pbio.1000479 (2010).
- 149. Chang, L. et al. Cocaine-and amphetamine-regulated transcript modulates peripheral immunity and protects against brain injury in experimental stroke. Brain Behav Immun 25, 260–269, doi: 10.1016/j.bbi.2010.09.017 (2011).
- Hurtado, O. *et al.* Lack of adrenomedullin, but not complement factor H, results in larger infarct size and more extensive brain damage in a focal ischemia model. *Neuroscience* 171, 885–892, doi: 10.1016/j.neuroscience.2010.09.021 (2010).
- Hyakkoku, K. et al. Toll-like receptor 4 (TLR4), but not TLR3 or TLR9, knock-out mice have neuroprotective effects against focal cerebral ischemia. Neuroscience 171, 258–267, doi: 10.1016/j.neuroscience.2010.08.054 (2010).
- 152. Liu, D. et al. Evidence that OGG1 glycosylase protects neurons against oxidative DNA damage and cell death under ischemic conditions. J Cereb Blood Flow Metab 31, 680-692, doi: 10.1038/jcbfm.2010.147 (2011).
- 153. Marumo, T., Eto, K., Wake, H., Omura, T. & Nabekura, J. The inhibitor of 20-HETE synthesis, TS-011, improves cerebral microcirculatory autoregulation impaired by middle cerebral artery occlusion in mice. *Br J Pharmacol* 161, 1391–1402, doi: 10.1111/j.1476-5381.2010.00973.x (2010).
- Menn, B. et al. Delayed treatment with systemic (S)-roscovitine provides neuroprotection and inhibits in vivo CDK5 activity increase in animal stroke models. PLoS One 5, e12117, doi: 10.1371/journal.pone.0012117 (2010).
- Oida, Y. et al. Post-treatment of a BiP inducer prevents cell death after middle cerebral artery occlusion in mice. Neurosci Lett 484, 43–46, doi: 10.1016/j.neulet.2010.08.015 (2010).
- 156. Xu, X. et al. Synergistic protective effects of humanin and necrostatin-1 on hypoxia and ischemia/reperfusion injury. Brain Res 1355, 189–194, doi: 10.1016/j.brainres.2010.07.080 (2010).
- Qiu, J. et al. High-mobility group box 1 promotes metalloproteinase-9 upregulation through Toll-like receptor 4 after cerebral ischemia. Stroke 41, 2077–2082, doi: 10.1161/STROKEAHA.110.590463 (2010).
- 158. Tai, S. H. *et al.* Melatonin inhibits postischemic matrix metalloproteinase-9 (MMP-9) activation via dual modulation of plasminogen/plasmin system and endogenous MMP inhibitor in mice subjected to transient focal cerebral ischemia. *J Pineal Res* 49, 332–341, doi: 10.1111/j.1600-079X.2010.00797.x (2010).
- 159. Kraft, P., Schwarz, T., Meijers, J. C., Stoll, G. & Kleinschnitz, C. Thrombin-activatable fibrinolysis inhibitor (TAFI) deficient mice are susceptible to intracerebral thrombosis and ischemic stroke. *PLoS One* **5**, e11658, doi: 10.1371/journal.pone.0011658 (2010).
- Shin, J. A. *et al.* Therapeutic effects of resveratrol during acute periods following experimental ischemic stroke. *J Neuroimmunol* 227, 93–100, doi: 10.1016/j.jneuroim.2010.06.017 (2010).
- 161. Ehling, P. *et al.* Two pore domain potassium channels in cerebral ischemia: a focus on K2P9.1 (TASK3, KCNK9). *Exp Transl Stroke Med* **2**, 14, doi: 10.1186/2040-7378-2-14 (2010).
- 162. Ahmad, A. S., Ahmad, M., Maruyama, T., Narumiya, S. & Dore, S. Prostaglandin D2 DP1 receptor is beneficial in ischemic stroke and in acute exicitotoxicity in young and old mice. Age (Dordr) 32, 271–282, doi: 10.1007/s11357-010-9135-y (2010).
- 163. Saleem, S., Shah, Z. A., Maruyama, T., Narumiya, S. & Dore, S. Neuroprotective properties of prostaglandin I2 IP receptor in focal cerebral ischemia. *Neuroscience* 170, 317–323, doi: 10.1016/j.neuroscience.2010.06.060 (2010).
- 164. Li, J., Benashski, S. E., Siegel, C., Liu, F. & McCullough, L. D. Adenosine monophosphate activated protein kinase inhibition is protective in both sexes after experimental stroke. *Neurosci Lett* 482, 62–65, doi: 10.1016/j.neulet.2010.07.007 (2010).
- 165. De Meyer, S. F. et al. Binding of von Willebrand factor to collagen and glycoprotein Ibalpha, but not to glycoprotein IIb/IIIa, contributes to ischemic stroke in mice—brief report. Arterioscler Thromb Vasc Biol 30, 1949–1951, doi: 10.1161/ATVBAHA.110.208918 (2010).
- 166. Ahmad, M. *et al.* The PGE2 EP2 receptor and its selective activation are beneficial against ischemic stroke. *Exp Transl Stroke Med* 2, 12, doi: 10.1186/2040-7378-2-12 (2010).
- 167. Harada, S., Fujita-Hamabe, W. & Tokuyama, S. The importance of regulation of blood glucose levels through activation of peripheral 5'-AMP-activated protein kinase on ischemic neuronal damage. *Brain Res* 1351, 254–263, doi: 10.1016/j. brainres.2010.06.052 (2010).
- Atochin, D. N. *et al.* Soluble guanylate cyclase alpha1beta1 limits stroke size and attenuates neurological injury. *Stroke* 41, 1815–1819, doi: 10.1161/STROKEAHA.109.577635 (2010).
- 169. Fan, Y. Y. et al. Activation of the central histaminergic system is involved in hypoxia-induced stroke tolerance in adult mice. J Cereb Blood Flow Metab 31, 305–314, doi: 10.1038/jcbfm.2010.94 (2011).
- 170. Kahles, T. *et al.* NADPH oxidase Nox1 contributes to ischemic injury in experimental stroke in mice. *Neurobiol Dis* **40**, 185–192, doi: 10.1016/j.nbd.2010.05.023 (2010).
- 171. Zhang, F. et al. Enhanced Delivery of Erythropoietin Across the Blood-Brain Barrier for Neuroprotection against Ischemic Neuronal Injury. Transl Stroke Res 1, 113–121, doi: 10.1007/s12975-010-0019-3 (2010).
- 172. Harada, S. *et al.* Morinda citrifolia fruit juice prevents ischemic neuronal damage through suppression of the development of postischemic glucose intolerance. *J Nat Med* **64**, 468–473, doi: 10.1007/s11418-010-0437-2 (2010).
- 173. Frauenknecht, K. *et al.* Neuroprotective effect of Fn14 deficiency is associated with induction of the granulocyte-colony stimulating factor (G-CSF) pathway in experimental stroke and enhanced by a pathogenic human antiphospholipid antibody. *J Neuroimmunol* 227, 1–9, doi: 10.1016/j.jneuroim.2010.05.043 (2010).
- 174. Manhas, N., Shi, Y., Taunton, J. & Sun, D. p90 activation contributes to cerebral ischemic damage via phosphorylation of Na⁺/H⁺ exchanger isoform 1. J Neurochem 114, 1476–1486, doi: 10.1111/j.1471-4159.2010.06868.x (2010).
- Doeppner, T. R. *et al.* Transplantation of TAT-Bcl-xL-transduced neural precursor cells: long-term neuroprotection after stroke. *Neurobiol Dis* 40, 265–276, doi: 10.1016/j.nbd.2010.05.033 (2010).

- 176. Pfeilschifter, W. et al. Pyrrolidine dithiocarbamate activates p38 MAPK and protects brain endothelial cells from apoptosis: a mechanism for the protective effect in stroke? Neurochem Res 35, 1391–1401, doi: 10.1007/s11064-010-0197-0 (2010).
- 177. Migliati, E. R. *et al.* Na(+)–K(+)–2Cl(–) cotransport inhibitor attenuates cerebral edema following experimental stroke via the perivascular pool of aquaporin-4. *Neurocrit Care* **13**, 123–131, doi: 10.1007/s12028-010-9376-8 (2010).
- 178. Yin, K. J. et al. Peroxisome proliferator-activated receptor delta regulation of miR-15a in ischemia-induced cerebral vascular endothelial injury. J Neurosci **30**, 6398–6408, doi: 10.1523/JNEUROSCI.0780-10.2010 (2010).
- 179. Shah, Z. A. *et al.* The flavanol (–)-epicatechin prevents stroke damage through the Nrf2/HO1 pathway. *J Cereb Blood Flow Metab* **30**, 1951–1961, doi: 10.1038/jcbfm.2010.53 (2010).
- Fan, Y. et al. Endothelial progenitor cell transplantation improves long-term stroke outcome in mice. Ann Neurol 67, 488–497, doi: 10.1002/ana.21919 (2010).
- Xu, Z. et al. Endonuclease G does not play an obligatory role in poly(ADP-ribose) polymerase-dependent cell death after transient focal cerebral ischemia. Am J Physiol Regul Integr Comp Physiol 299, R215–221, doi: 10.1152/ajpregu.00747.2009 (2010).
- Lu, C. et al. Scavenger receptor class-A has a central role in cerebral ischemia-reperfusion injury. J Cereb Blood Flow Metab 30, 1972–1981, doi: 10.1038/jcbfm.2010.59 (2010).
- 183. Vest, R. S., O'Leary, H., Coultrap, S. J., Kindy, M. S. & Bayer, K. U. Effective post-insult neuroprotection by a novel Ca(2+)/ calmodulin-dependent protein kinase II (CaMKII) inhibitor. J Biol Chem 285, 20675–20682, doi: 10.1074/jbc.M109.088617 (2010).
- 184. Haines, B. A., Mehta, S. L., Pratt, S. M., Warden, C. H. & Li, P. A. Deletion of mitochondrial uncoupling protein-2 increases ischemic brain damage after transient focal ischemia by altering gene expression patterns and enhancing inflammatory cytokines. *J Cereb Blood Flow Metab* 30, 1825–1833, doi: 10.1038/jcbfm.2010.52 (2010).
- Liu, F., Akella, P., Benashski, S. E., Xu, Y. & McCullough, L. D. Expression of Na-K-Cl cotransporter and edema formation are age dependent after ischemic stroke. *Exp Neurol* 224, 356–361, doi: 10.1016/j.expneurol.2010.04.010 (2010).
- Kasahara, Y. *et al.* Telmisartan suppresses cerebral injury in a murine model of transient focal ischemia. *Brain Res* 1340, 70–80, doi: 10.1016/j.brainres.2010.03.101 (2010).
- 187. Jin, K., Wang, X., Xie, L., Mao, X. O. & Greenberg, D. A. Transgenic ablation of doublecortin-expressing cells suppresses adult neurogenesis and worsens stroke outcome in mice. *Proc Natl Acad Sci USA* **107**, 7993–7998, doi: 10.1073/pnas.1000154107 (2010).
- Zhu, H. R. *et al.* Icariin protects against brain injury by enhancing SIRT1-dependent PGC-1alpha expression in experimental stroke. *Neuropharmacology* 59, 70–76, doi: 10.1016/j.neuropharm.2010.03.017 (2010).
- 189. Sakata, Y., Zhuang, H., Kwansa, H., Koehler, R. C. & Dore, S. Resveratrol protects against experimental stroke: putative neuroprotective role of heme oxygenase 1. *Exp Neurol* 224, 325–329, doi: 10.1016/j.expneurol.2010.03.032 (2010).
- 190. Abe, T. et al. Key role of CD36 in Toll-like receptor 2 signaling in cerebral ischemia. Stroke 41, 898–904, doi: 10.1161/ STROKEAHA.109.572552 (2010).
- 191. Zechariah, A., ElAli, A. & Hermann, D. M. Combination of tissue-plasminogen activator with erythropoietin induces blood-brain barrier permeability, extracellular matrix disaggregation, and DNA fragmentation after focal cerebral ischemia in mice. *Stroke* 41, 1008–1012, doi: 10.1161/STROKEAHA.109.574418 (2010).
- Zhang, B. *et al.* Estradiol and G1 reduce infarct size and improve immunosuppression after experimental stroke. *J Immunol* 184, 4087–4094, doi: 10.4049/jimmunol.0902339 (2010).
- Kraft, P., Schwarz, T., Pochet, L., Stoll, G. & Kleinschnitz, C. COU254, a specific 3-carboxamide-coumarin inhibitor of coagulation factor XII, does not protect mice from acute ischemic stroke. *Exp Transl Stroke Med* 2, 5, doi: 10.1186/2040-7378-2-5 (2010).
- 194. Li, X. et al. Contributions of poly(ADP-ribose) polymerase-1 and -2 to nuclear translocation of apoptosis-inducing factor and injury from focal cerebral ischemia. J Neurochem 113, 1012–1022, doi: 10.1111/j.1471-4159.2010.06667.x (2010).
- Kleinschnitz, C. et al. Early detrimental T-cell effects in experimental cerebral ischemia are neither related to adaptive immunity nor thrombus formation. Blood 115, 3835–3842, doi: 10.1182/blood-2009-10-249078 (2010).
- Stapels, M. et al. Polycomb group proteins as epigenetic mediators of neuroprotection in ischemic tolerance. Sci Signal 3, ra15, doi: 10.1126/scisignal.2000502 (2010).
- 197. Chen, Z. B. et al. Human urinary kallidinogenase suppresses cerebral inflammation in experimental stroke and downregulates nuclear factor-kappaB. J Cereb Blood Flow Metab 30, 1356–1365, doi: 10.1038/jcbfm.2010.19 (2010).
- Muhammad, S., Aller, M. I., Maser-Gluth, C., Schwaninger, M. & Wisden, W. Expression of the kcnk3 potassium channel gene lessens the injury from cerebral ischemia, most likely by a general influence on blood pressure. *Neuroscience* 167, 758–764, doi: 10.1016/j.neuroscience.2010.02.024 (2010).
- 199. Brait, V. H. et al. Mechanisms contributing to cerebral infarct size after stroke: gender, reperfusion, T lymphocytes, and Nox2derived superoxide. J Cereb Blood Flow Metab **30**, 1306–1317, doi: 10.1038/jcbfm.2010.14 (2010).
- 200. Nagai, N. *et al.* Initial brain lesion size affects the extent of subsequent pathophysiological responses. *Brain Res* **1322**, 109–117, doi: 10.1016/j.brainres.2010.01.077 (2010).
- Jiang, S. X. et al. Neuropilin 1 directly interacts with Fer kinase to mediate semaphorin 3A-induced death of cortical neurons. J Biol Chem 285, 9908–9918, doi: 10.1074/jbc.M109.080689 (2010).
- Ikeda-Matsuo, Y. et al. Microsomal prostaglandin E synthase-1 and cyclooxygenase-2 are both required for ischaemic excitotoxicity. Br J Pharmacol 159, 1174–1186, doi: 10.1111/j.1476-5381.2009.00595.x (2010).
- Mustafa, A. K. *et al.* Serine racemase deletion protects against cerebral ischemia and excitotoxicity. J Neurosci 30, 1413–1416, doi: 10.1523/JNEUROSCI.4297-09.2010 (2010).
- Heurteaux, C. et al. Neuroprotective and neuroproliferative activities of NeuroAid (MLC601, MLC901), a Chinese medicine, in vitro and in vivo. Neuropharmacology 58, 987–1001, doi: 10.1016/j.neuropharm.2010.01.001 (2010).
- 205. Yin, K. J. et al. miR-497 regulates neuronal death in mouse brain after transient focal cerebral ischemia. Neurobiol Dis 38, 17-26, doi: 10.1016/j.nbd.2009.12.021 (2010).
- Elvers, M. et al. Impaired alpha(IIb)beta(3) integrin activation and shear-dependent thrombus formation in mice lacking phospholipase D1. Sci Signal 3, ra1, doi: 10.1126/scisignal.2000551 (2010).
- Shen, Y. et al. Carnosine protects against permanent cerebral ischemia in histidine decarboxylase knockout mice by reducing glutamate excitotoxicity. Free Radic Biol Med 48, 727–735, doi: 10.1016/j.freeradbiomed.2009.12.021 (2010).
- 208. Chi, S. *et al.* Baifuzi reduces transient ischemic brain damage through an interaction with the STREX domain of BKCa channels. *Cell Death Dis* **1**, e13, doi: 10.1038/cddis.2009.10 (2010).
- 209. Nguemeni, C. *et al.* Dietary supplementation of alpha-linolenic acid in an enriched rapeseed oil diet protects from stroke. *Pharmacol Res* **61**, 226–233, doi: 10.1016/j.phrs.2009.12.007 (2010).
- Vagnerova, K. et al. Poly (ADP-ribose) polymerase-1 initiated neuronal cell death pathway—do androgens matter? Neuroscience 166, 476–481, doi: 10.1016/j.neuroscience.2009.12.041 (2010).
- Keum, S. & Marchuk, D. A. A locus mapping to mouse chromosome 7 determines infarct volume in a mouse model of ischemic stroke. *Circ Cardiovasc Genet* 2, 591–598, doi: 10.1161/CIRCGENETICS.109.883231 (2009).
- Strecker, J. K. *et al.* Effects of G-CSF treatment on neutrophil mobilization and neurological outcome after transient focal ischemia. *Exp Neurol* 222, 108–113, doi: 10.1016/j.expneurol.2009.12.012 (2010).
- Samanta, J., Alden, T., Gobeske, K., Kan, L. & Kessler, J. A. Noggin protects against ischemic brain injury in rodents. Stroke 41, 357–362, doi: 10.1161/STROKEAHA.109.565523 (2010).

- Elzer, J. G. et al. Neuronal estrogen receptor-alpha mediates neuroprotection by 17beta-estradiol. J Cereb Blood Flow Metab 30, 935–942, doi: 10.1038/jcbfm.2009.258 (2010).
- Park, J. W. et al. Green tea polyphenol (-)-epigallocatechin gallate reduces matrix metalloproteinase-9 activity following transient focal cerebral ischemia. J Nutr Biochem 21, 1038–1044, doi: 10.1016/j.jnutbio.2009.08.009 (2010).
- Kim, G. S., Jung, J. E., Niizuma, K. & Chan, P. H. CK2 is a novel negative regulator of NADPH oxidase and a neuroprotectant in mice after cerebral ischemia. J Neurosci 29, 14779–14789, doi: 10.1523/JNEUROSCI.4161-09.2009 (2009).
- Felger, J. C. et al. Brain dendritic cells in ischemic stroke: time course, activation state, and origin. Brain Behav Immun 24, 724–737, doi: 10.1016/j.bbi.2009.11.002 (2010).
- Benakis, C., Bonny, C. & Hirt, L. JNK inhibition and inflammation after cerebral ischemia. *Brain Behav Immun* 24, 800–811, doi: 10.1016/j.bbi.2009.11.001 (2010).
- 219. Steele, A. D. et al. Context dependent neuroprotective properties of prion protein (PrP). Prion 3, 240-249 (2009)
- Lathia, J. D. *et al.* Pivotal role for beta-1 integrin in neurovascular remodelling after ischemic stroke. *Exp Neurol* 221, 107–114, doi: 10.1016/j.expneurol.2009.10.007 (2010).
- 221. Leypoldt, F. *et al.* Dimethylarginine dimethylaminohydrolase-1 transgenic mice are not protected from ischemic stroke. *PLoS One* 4, e7337, doi: 10.1371/journal.pone.0007337 (2009).
- 222. Liu, X., Nakayama, S., Amiry-Moghaddam, M., Ottersen, O. P. & Bhardwaj, A. Arginine-vasopressin V1 but not V2 receptor antagonism modulates infarct volume, brain water content, and aquaporin-4 expression following experimental stroke. *Neurocrit Care* 12, 124–131, doi: 10.1007/s12028-009-9277-x (2010).
- 223. Mukerji, S. S., Rainey, R. N., Rhodes, J. L. & Hall, A. K. Delayed activin A administration attenuates tissue death after transient focal cerebral ischemia and is associated with decreased stress-responsive kinase activation. J Neurochem 111, 1138–1148, doi: 10.1111/j.1471-4159.2009.06406.x (2009).
- Karelina, K., Norman, G. J., Zhang, N. & DeVries, A. C. Social contact influences histological and behavioral outcomes following cerebral ischemia. *Exp Neurol* 220, 276–282, doi: 10.1016/j.expneurol.2009.08.022 (2009).
- Czech, B. *et al.* The immunomodulatory sphingosine 1-phosphate analog FTY720 reduces lesion size and improves neurological outcome in a mouse model of cerebral ischemia. *Biochem Biophys Res Commun* 389, 251–256, doi: 10.1016/j.bbrc.2009.08.142 (2009).
- 226. Shi, Q. *et al.* Adenovirus-mediated brain-derived neurotrophic factor expression regulated by hypoxia response element protects brain from injury of transient middle cerebral artery occlusion in mice. *Neurosci Lett* **465**, 220–225, doi: 10.1016/j. neulet.2009.08.049 (2009).
- 227. Gui, L. *et al.* Adenosine A 2A receptor deficiency reduces striatal glutamate outflow and attenuates brain injury induced by transient focal cerebral ischemia in mice. *Brain Res* **1297**, 185–193, doi: 10.1016/j.brainres.2009.08.050 (2009).
- 228. Haraguchi, T. *et al.* Cerebroprotective action of telmisartan by inhibition of macrophages/microglia expressing HMGB1 via a peroxisome proliferator-activated receptor gamma-dependent mechanism. *Neurosci Lett* **464**, 151–155, doi: 10.1016/j. neulet.2009.08.043 (2009).
- Zhao, B. Q. et al. von Willebrand factor-cleaving protease ADAMTS13 reduces ischemic brain injury in experimental stroke. Blood 114, 3329–3334, doi: 10.1182/blood-2009-03-213264 (2009).
- Berthet, C. et al. Neuroprotective role of lactate after cerebral ischemia. J Cereb Blood Flow Metab 29, 1780–1789, doi: 10.1038/ jcbfm.2009.97 (2009).
- Saiki, R. et al. Intense correlation between brain infarction and protein-conjugated acrolein. Stroke 40, 3356–3361, doi: 10.1161/ STROKEAHA.109.553248 (2009).
- 232. Marsh, B. *et al.* Systemic lipopolysaccharide protects the brain from ischemic injury by reprogramming the response of the brain to stroke: a critical role for IRF3. *J Neurosci* **29**, 9839–9849, doi: 10.1523/JNEUROSCI.2496-09.2009 (2009).
- 233. Shichita, T. *et al.* Pivotal role of cerebral interleukin-17-producing gammadeltaT cells in the delayed phase of ischemic brain injury. *Nat Med* **15**, 946–950, doi: 10.1038/nm.1999 (2009).
- Wacker, B. K., Park, T. S. & Gidday, J. M. Hypoxic preconditioning-induced cerebral ischemic tolerance: role of microvascular sphingosine kinase 2. Stroke 40, 3342–3348, doi: 10.1161/STROKEAHA.109.560714 (2009).
- Koumura, A. *et al.* Metallothionein-III knockout mice aggravates the neuronal damage after transient focal cerebral ischemia. *Brain Res* 1292, 148–154, doi: 10.1016/j.brainres.2009.07.050 (2009).
- Choe, C. U. et al. Nitroxyl exacerbates ischemic cerebral injury and oxidative neurotoxicity. J Neurochem 110, 1766–1773, doi: 10.1111/j.1471-4159.2009.06266.x (2009).
- Jackman, K. A., Miller, A. A., Drummond, G. R. & Sobey, C. G. Importance of NOX1 for angiotensin II-induced cerebrovascular superoxide production and cortical infarct volume following ischemic stroke. *Brain Res* 1286, 215–220, doi: 10.1016/j. brainres.2009.06.056 (2009).
- Schilling, M., Strecker, J. K., Ringelstein, E. B., Schabitz, W. R. & Kiefer, R. The role of CC chemokine receptor 2 on microglia activation and blood-borne cell recruitment after transient focal cerebral ischemia in mice. *Brain Res* 1289, 79–84, doi: 10.1016/j. brainres.2009.06.054 (2009).
- 239. Shin, J. A. *et al.* Ischemic preconditioning-induced neuroprotection is associated with differential expression of IL-1beta and IL-1 receptor antagonist in the ischemic cortex. *J Neuroimmunol* **217**, 14–19, doi: 10.1016/j.jneuroim.2009.06.001 (2009).
- Pastor, M. D. *et al.* mTOR/S6 kinase pathway contributes to astrocyte survival during ischemia. *J Biol Chem* 284, 22067–22078, doi: 10.1074/jbc.M109.033100 (2009).
- 241. Kim, H. W., Cho, K. J., Lee, B. I., Kim, H. J. & Kim, G. W. Post-ischemic administration of peptide with apurinic/apyrimidinic endonuclease activity inhibits induction of cell death after focal cerebral ischemia/reperfusion in mice. *Neurosci Lett* 460, 166–169, doi: 10.1016/j.neulet.2009.05.062 (2009).
- Zhang, W. et al. Role of soluble epoxide hydrolase in the sex-specific vascular response to cerebral ischemia. J Cereb Blood Flow Metab 29, 1475–1481, doi: 10.1038/jcbfm.2009.65 (2009).
- 243. Zeynalov, E., Shah, Z. A., Li, R. C. & Dore, S. Heme oxygenase 1 is associated with ischemic preconditioning-induced protection against brain ischemia. *Neurobiol Dis* 35, 264–269, doi: 10.1016/j.nbd.2009.05.010 (2009).
- Harada, S., Fujita, W. H., Shichi, K. & Tokuyama, S. The development of glucose intolerance after focal cerebral ischemia participates in subsequent neuronal damage. *Brain Res* 1279, 174–181, doi: 10.1016/j.brainres.2009.05.014 (2009).
- 245. Subramanian, S. et al. Recombinant T cell receptor ligand treats experimental stroke. Stroke 40, 2539–2545, doi: 10.1161/ STROKEAHA.108.543991 (2009).
- Zhang, Z. et al. Baicalin administration is effective in positive regulation of twenty-four ischemia/reperfusion-related proteins identified by a proteomic study. Neurochem Int 54, 488–496, doi: 10.1016/j.neuint.2009.02.005 (2009).
- 247. Descamps, E. *et al.* Experimental stroke protection induced by 4-hydroxybenzyl alcohol is cancelled by bacitracin. *Neurosci Res* 64, 137–142, doi: 10.1016/j.neures.2009.02.005 (2009).
- Chen, H., Song, Y. S. & Chan, P. H. Inhibition of NADPH oxidase is neuroprotective after ischemia-reperfusion. J Cereb Blood Flow Metab 29, 1262–1272, doi: 10.1038/jcbfm.2009.47 (2009).
- 249. Hua, F. *et al.* Differential roles of TLR2 and TLR4 in acute focal cerebral ischemia/reperfusion injury in mice. *Brain Res* **1262**, 100–108, doi: 10.1016/j.brainres.2009.01.018 (2009).
- 250. Stowe, A. M. *et al.* Neutrophil elastase and neurovascular injury following focal stroke and reperfusion. *Neurobiol Dis* **35**, 82–90, doi: 10.1016/j.nbd.2009.04.006 (2009).

- Saleem, S., Ahmad, A. S., Maruyama, T., Narumiya, S. & Dore, S. PGF(2alpha) FP receptor contributes to brain damage following transient focal brain ischemia. *Neurotox Res* 15, 62–70, doi: 10.1007/s12640-009-9007-3 (2009).
- 252. Zeynalov, E. & Dore, S. Low doses of carbon monoxide protect against experimental focal brain ischemia. *Neurotox Res* 15, 133–137, doi: 10.1007/s12640-009-9014-4 (2009).
- Doeppner, T. R. et al. TAT-Hsp70-mediated neuroprotection and increased survival of neuronal precursor cells after focal cerebral ischemia in mice. J Cereb Blood Flow Metab 29, 1187–1196, doi: 10.1038/jcbfm.2009.44 (2009).
- 254. Nonaka, Y. *et al.* Cilostazol protects against hemorrhagic transformation in mice transient focal cerebral ischemia-induced brain damage. *Neurosci Lett* **452**, 156–161, doi: 10.1016/j.neulet.2009.01.039 (2009).
- Krishnamurthy, R. G. *et al.* Asiatic acid, a pentacyclic triterpene from Centella asiatica, is neuroprotective in a mouse model of focal cerebral ischemia. J Neurosci Res 87, 2541–2550, doi: 10.1002/jnr.22071 (2009).
- Schilling, M., Strecker, J. K., Schabitz, W. R., Ringelstein, E. B. & Kiefer, R. Effects of monocyte chemoattractant protein 1 on bloodborne cell recruitment after transient focal cerebral ischemia in mice. *Neuroscience* 161, 806–812, doi: 10.1016/j. neuroscience.2009.04.025 (2009).
- 257. Kim, H. W., Cho, K. J., Park, S. C., Kim, H. J. & Kim, G. W. The adenoviral vector-mediated increase in apurinic/apyrimidinic endonuclease inhibits the induction of neuronal cell death after transient ischemic stroke in mice. *Brain Res* 1274, 1–10, doi: 10.1016/j.brainres.2009.04.006 (2009).
- Nonaka, Y. *et al.* Combination treatment with normobaric hyperoxia and cilostazol protects mice against focal cerebral ischemiainduced neuronal damage better than each treatment alone. *J Pharmacol Exp Ther* 330, 13–22, doi: 10.1124/jpet.109.151548 (2009).
- Zhang, M. et al. CB2 receptor activation attenuates microcirculatory dysfunction during cerebral ischemic/reperfusion injury. Microvasc Res 78, 86–94, doi: 10.1016/j.mvr.2009.03.005 (2009).
- Sevimli, S. *et al.* Endogenous brain protection by granulocyte-colony stimulating factor after ischemic stroke. *Exp Neurol* 217, 328–335, doi: 10.1016/j.expneurol.2009.03.018 (2009).
- Zhou, F. et al. Attenuation of neuronal degeneration in thioredoxin-1 overexpressing mice after mild focal ischemia. Brain Res 1272, 62–70, doi: 10.1016/j.brainres.2009.03.023 (2009).
- Karelina, K. *et al.* Social isolation alters neuroinflammatory response to stroke. *Proc Natl Acad Sci USA* 106, 5895–5900, doi: 10.1073/pnas.0810737106 (2009).
- Wang, X. et al. Methazolamide and melatonin inhibit mitochondrial cytochrome C release and are neuroprotective in experimental models of ischemic injury. Stroke 40, 1877–1885, doi: 10.1161/STROKEAHA.108.540765 (2009).
- Wang, C. Y. et al. Obesity increases vascular senescence and susceptibility to ischemic injury through chronic activation of Akt and mTOR. Sci Signal 2, ra11, doi: 10.1126/scisignal.2000143 (2009).
- 265. Liu, D., Gharavi, R., Pitta, M., Gleichmann, M. & Mattson, M. P. Nicotinamide prevents NAD⁺ depletion and protects neurons against excitotoxicity and cerebral ischemia: NAD⁺ consumption by SIRT1 may endanger energetically compromised neurons. *Neuromolecular Med* 11, 28–42, doi: 10.1007/s12017-009-8058-1 (2009).
- Li, R. C. et al. Heme-hemopexin complex attenuates neuronal cell death and stroke damage. J Cereb Blood Flow Metab 29, 953–964, doi: 10.1038/jcbfm.2009.19 (2009).
- Yuan, M. *et al.* Sex differences in the response to activation of the poly (ADP-ribose) polymerase pathway after experimental stroke. *Exp Neurol* 217, 210–218, doi: 10.1016/j.expneurol.2009.02.012 (2009).
- 268. Liu, F. et al. Sex differences in caspase activation after stroke. Stroke 40, 1842–1848, doi: 10.1161/STROKEAHA.108.538686 (2009).
- Saleem, S., Shah, Z. A., Urade, Y. & Dore, S. Lipocalin-prostaglandin D synthase is a critical beneficial factor in transient and permanent focal cerebral ischemia. *Neuroscience* 160, 248–254, doi: 10.1016/j.neuroscience.2009.02.039 (2009).
- Harada, S. *et al.* Preventive effect of Morinda citrifolia fruit juice on neuronal damage induced by focal ischemia. *Biol Pharm Bull* 32, 405–409 (2009).
- 271. Jin, G. *et al.* CD47 gene knockout protects against transient focal cerebral ischemia in mice. *Exp Neurol* **217**, 165–170, doi: 10.1016/j.expneurol.2009.02.004 (2009).
- Liu, F., Yuan, R., Benashski, S. E. & McCullough, L. D. Changes in experimental stroke outcome across the life span. J Cereb Blood Flow Metab 29, 792–802, doi: 10.1038/jcbfm.2009.5 (2009).
- Saleem, S., Kim, Y. T., Maruyama, T., Narumiya, S. & Dore, S. Reduced acute brain injury in PGE2 EP3 receptor-deficient mice after cerebral ischemia. J Neuroimmunol 208, 87–93, doi: 10.1016/j.jneuroim.2009.01.015 (2009).
- Pradillo, J. M. *et al.* Toll-like receptor 4 is involved in neuroprotection afforded by ischemic preconditioning. *J Neurochem* 109, 287–294, doi: 10.1111/j.1471-4159.2009.05972.x (2009).
- Lambertsen, K. L. et al. Microglia protect neurons against ischemia by synthesis of tumor necrosis factor. J Neurosci 29, 1319–1330, doi: 10.1523/JNEUROSCI.5505-08.2009 (2009).
- Li, J. & McCullough, L. D. Sex differences in minocycline-induced neuroprotection after experimental stroke. J Cereb Blood Flow Metab 29, 670–674, doi: 10.1038/jcbfm.2009.3 (2009).
- Kleinschnitz, C. et al. Deficiency of von Willebrand factor protects mice from ischemic stroke. Blood 113, 3600–3603, doi: 10.1182/ blood-2008-09-180695 (2009).
- Jackman, K. A. *et al.* Reduction of cerebral infarct volume by apocynin requires pretreatment and is absent in Nox2-deficient mice. *Br J Pharmacol* 156, 680–688, doi: 10.1111/j.1476-5381.2008.00073.x (2009).
- 279. Liesz, A. *et al.* Regulatory T cells are key cerebroprotective immunomodulators in acute experimental stroke. *Nat Med* **15**, 192–199, doi: 10.1038/nm.1927 (2009).
- Doeppner, T. R. et al. TAT-Bcl-x(L) improves survival of neuronal precursor cells in the lesioned striatum after focal cerebral ischemia. Neurobiol Dis 34, 87–94, doi: 10.1016/j.nbd.2008.12.013 (2009).
- Liu, F., Schafer, D. P. & McCullough, L. D. TTC, fluoro-Jade B and NeuN staining confirm evolving phases of infarction induced by middle cerebral artery occlusion. *J Neurosci Methods* 179, 1–8, doi: 10.1016/j.jneumeth.2008.12.028 (2009).
- Hyakkoku, K. et al. Thalidomide protects against ischemic neuronal damage induced by focal cerebral ischemia in mice. Neuroscience 159, 760-769, doi: 10.1016/j.neuroscience.2008.12.043 (2009).
- Kuroki, K. et al. Effects of transient focal cerebral ischemia in mice deficient in puma. Neurosci Lett 451, 237–240, doi: 10.1016/j. neulet.2009.01.019 (2009).
- 284. Zhang, C., An, J., Strickland, D. K. & Yepes, M. The low-density lipoprotein receptor-related protein 1 mediates tissue-type plasminogen activator-induced microglial activation in the ischemic brain. Am J Pathol 174, 586–594, doi: 10.2353/ ajpath.2009.080661 (2009).
- Ji, S. *et al.* Acute neuroprotection by pioglitazone after mild brain ischemia without effect on long-term outcome. *Exp Neurol* 216, 321–328, doi: 10.1016/j.expneurol.2008.12.007 (2009).
- Lee, S. R., Kim, H. Y., Hong, J. S., Baek, W. K. & Park, J. W. PPARgamma agonist pioglitazone reduces matrix metalloproteinase-9 activity and neuronal damage after focal cerebral ischemia. *Biochem Biophys Res Commun* 380, 17–21, doi: 10.1016/j. bbrc.2008.12.181 (2009).
- 287. Lee, E. J., Hung, Y. C., Chen, H. Y., Wu, T. S. & Chen, T. Y. Delayed treatment with carboxy-PTIO permits a 4-h therapeutic window of opportunity and prevents against ischemia-induced energy depletion following permanent focal cerebral ischemia in mice. *Neurochem Res* 34, 1157–1166, doi: 10.1007/s11064-008-9892-5 (2009).

- Hassid, B. G. et al. Neuronal RAGE expression modulates severity of injury following transient focal cerebral ischemia. J Clin Neurosci 16, 302–306, doi: 10.1016/j.jocn.2007.12.011 (2009).
- Stetler, R. A. *et al.* Hsp27 protects against ischemic brain injury via attenuation of a novel stress-response cascade upstream of mitochondrial cell death signaling. *J Neurosci* 28, 13038–13055, doi: 10.1523/JNEUROSCI.4407-08.2008 (2008).
- Valerio, A. *et al.* Leptin is induced in the ischemic cerebral cortex and exerts neuroprotection through NF-kappaB/c-Rel-dependent transcription. *Stroke* 40, 610–617, doi: 10.1161/STROKEAHA.108.528588 (2009).
- Muhammad, S. et al. The HMGB1 receptor RAGE mediates ischemic brain damage. J Neurosci 28, 12023–12031, doi: 10.1523/ JNEUROSCI.2435-08.2008 (2008).
- 292. Dogrukol-Ak, D. *et al.* Isolation of peptide transport system-6 from brain endothelial cells: therapeutic effects with antisense inhibition in Alzheimer and stroke models. *J Cereb Blood Flow Metab* **29**, 411–422, doi: 10.1038/jcbfm.2008.131 (2009).
- 293. Austinat, M. *et al.* Blockade of bradykinin receptor B1 but not bradykinin receptor B2 provides protection from cerebral infarction and brain edema. *Stroke* **40**, 285–293, doi: 10.1161/STROKEAHA.108.526673 (2009).
- 294. Mito, T. *et al.* Decreased damage from transient focal cerebral ischemia by transfusion of zero-link hemoglobin polymers in mouse. *Stroke* **40**, 278–284, doi: 10.1161/STROKEAHA.108.526731 (2009).
- 295. Hirt, L. *et al.* Protective role of early aquaporin 4 induction against postischemic edema formation. *J Cereb Blood Flow Metab* **29**, 423–433, doi: 10.1038/jcbfm.2008.133 (2009).
- 296. Jia, J. et al. Estrogen inhibits Fas-mediated apoptosis in experimental stroke. Exp Neurol 215, 48-52, doi: 10.1016/j. expneurol.2008.09.015 (2009).
- 297. Gurer, G., Gursoy-Ozdemir, Y., Erdemli, E., Can, A. & Dalkara, T. Astrocytes are more resistant to focal cerebral ischemia than neurons and die by a delayed necrosis. *Brain Pathol* **19**, 630–641, doi: 10.1111/j.1750-3639.2008.00226.x (2009).
- Meuth, S. G. *et al.* The neuroprotective impact of the leak potassium channel TASK1 on stroke development in mice. *Neurobiol Dis* 33, 1–11, doi: 10.1016/j.nbd.2008.09.006 (2009).
- 299. Saleem, S., Zhuang, H., Biswal, S., Christen, Y. & Dore, S. Ginkgo biloba extract neuroprotective action is dependent on heme oxygenase 1 in ischemic reperfusion brain injury. *Stroke* **39**, 3389–3396, doi: 10.1161/STROKEAHA.108.523480 (2008).
- Dong, K., Xu, W., Yang, J., Qiao, H. & Wu, L. Neuroprotective effects of Tanshinone IIA on permanent focal cerebral ischemia in mice. *Phytother Res* 23, 608–613, doi: 10.1002/ptr.2615 (2009).
- Zhou, X. Q., Zeng, X. N., Kong, H. & Sun, X. L. Neuroprotective effects of berberine on stroke models *in vitro* and *in vivo*. Neurosci Lett 447, 31–36, doi: 10.1016/j.neulet.2008.09.064 (2008).
- 302. Koumura, A. et al. A novel calpain inhibitor, ((1S)-1((((1S)-1-benzyl-3-cyclopropylamino-2,3-di-oxopropyl)amino)carbonyl)-3-met hylbutyl) carbamic acid 5-methoxy-3-oxapentyl ester, protects neuronal cells from cerebral ischemia-induced damage in mice. Neuroscience 157, 309–318, doi: 10.1016/j.neuroscience.2008.09.007 (2008).
- Mastroiacovo, F. et al. Induction of the Wnt antagonist, Dickkopf-1, contributes to the development of neuronal death in models of brain focal ischemia. J Cereb Blood Flow Metab 29, 264–276, doi: 10.1038/jcbfm.2008.111 (2009).
- Sato, Y., Meller, R., Yang, T., Taki, W. & Simon, R. P. Stereo-selective neuroprotection against stroke with vitamin A derivatives. Brain Res 1241, 188–192, doi: 10.1016/j.brainres.2008.09.020 (2008).
- Oyamada, N. et al. Transplantation of vascular cells derived from human embryonic stem cells contributes to vascular regeneration after stroke in mice. J Transl Med 6, 54, doi: 10.1186/1479-5876-6-54 (2008).
- 306. Kilic, E. et al. ABCC1: a gateway for pharmacological compounds to the ischaemic brain. Brain 131, 2679–2689, doi: 10.1093/ brain/awn222 (2008).
- Morales, J. R. et al. Activation of liver X receptors promotes neuroprotection and reduces brain inflammation in experimental stroke. Circulation 118, 1450–1459, doi: 10.1161/CIRCULATIONAHA.108.782300 (2008).
- Belayev, L. et al. LAU-0901, a novel platelet-activating factor antagonist, is highly neuroprotective in cerebral ischemia. Exp Neurol 214, 253–258, doi: 10.1016/j.expneurol.2008.08.009 (2008).
- 309. Yu, J., Zhu, H., Ko, D. & Kindy, M. S. Motoneuronotrophic factor analog GM6 reduces infarct volume and behavioral deficits following transient ischemia in the mouse. *Brain Res* 1238, 143–153, doi: 10.1016/j.brainres.2008.08.053 (2008).
- Li, W. L. et al. Chronic fluoxetine treatment improves ischemia-induced spatial cognitive deficits through increasing hippocampal neurogenesis after stroke. J Neurosci Res 87, 112–122, doi: 10.1002/jnr.21829 (2009).
- Wong, C. H., Bozinovski, S., Hertzog, P. J., Hickey, M. J. & Crack, P. J. Absence of glutathione peroxidase-1 exacerbates cerebral ischemia-reperfusion injury by reducing post-ischemic microvascular perfusion. J Neurochem 107, 241–252, doi: 10.1111/j.1471-4159.2008.05605.x (2008).
- Zeynalov, E. *et al.* The perivascular pool of aquaporin-4 mediates the effect of osmotherapy in postischemic cerebral edema. *Crit Care Med* 36, 2634–2640, doi: 10.1097/CCM.0b013e3181847853 (2008).
- Mayanagi, K., Katakam, P. V., Gaspar, T., Domoki, F. & Busija, D. W. Acute treatment with rosuvastatin protects insulin resistant (C57BL/6J ob/ob) mice against transient cerebral ischemia. J Cereb Blood Flow Metab 28, 1927–1935, doi: 10.1038/jcbfm.2008.81 (2008).
- 314. Wang, Y. et al. Gene inactivation of Na⁺/H⁺ exchanger isoform 1 attenuates apoptosis and mitochondrial damage following transient focal cerebral ischemia. Eur J Neurosci 28, 51–61, doi: 10.1111/j.1460-9568.2008.06304.x (2008).
- Nakamura, J. et al. Targeted disruption of Hsp110/105 gene protects against ischemic stress. Stroke 39, 2853–2859, doi: 10.1161/ STROKEAHA.107.506188 (2008).
- Schwarting, S. et al. Hematopoietic stem cells reduce postischemic inflammation and ameliorate ischemic brain injury. Stroke 39, 2867–2875, doi: 10.1161/STROKEAHA.108.513978 (2008).
- 317. Li, J., Lu, Z., Li, W. L., Yu, S. P. & Wei, L. Cell death and proliferation in NF-kappaB p50 knockout mouse after cerebral ischemia. Brain Res 1230, 281–289, doi: 10.1016/j.brainres.2008.06.130 (2008).
- Song, Y. S. et al. The role of Akt signaling in oxidative stress mediates NF-kappaB activation in mild transient focal cerebral ischemia. J Cereb Blood Flow Metab 28, 1917–1926, doi: 10.1038/jcbfm.2008.80 (2008).
- Iwanami, J. et al. Deletion of angiotensin II type 2 receptor attenuates protective effects of bone marrow stromal cell treatment on ischemia-reperfusion brain injury in mice. Stroke 39, 2554–2559, doi: 10.1161/STROKEAHA.107.513275 (2008).
- Moisse, K., Welch, I., Hill, T., Volkening, K. & Strong, M. J. Transient middle cerebral artery occlusion induces microglial priming in the lumbar spinal cord: a novel model of neuroinflammation. J Neuroinflammation 5, 29, doi: 10.1186/1742-2094-5-29 (2008).
- Shen, H., Luo, Y., Kuo, C. C. & Wang, Y. BMP7 reduces synergistic injury induced by methamphetamine and ischemia in mouse brain. *Neurosci Lett* 442, 15–18, doi: 10.1016/j.neulet.2008.06.052 (2008).
- 322. Xu, X. *et al.* Neuroprotective effect of humanin on cerebral ischemia/reperfusion injury is mediated by a PI3K/Akt pathway. *Brain Res* **1227**, 12–18, doi: 10.1016/j.brainres.2008.06.018 (2008).
- Hua, F. et al. Preconditioning with a TLR2 specific ligand increases resistance to cerebral ischemia/reperfusion injury. J Neuroimmunol 199, 75-82, doi: 10.1016/j.jneuroim.2008.05.009 (2008).
- 324. Nonaka, Y., Shimazawa, M., Yoshimura, S., Iwama, T. & Hara, H. Combination effects of normobaric hyperoxia and edaravone on focal cerebral ischemia-induced neuronal damage in mice. *Neurosci Lett* **441**, 224–228, doi: 10.1016/j.neulet.2008.06.033 (2008).
- Denes, A., Ferenczi, S., Halasz, J., Kornyei, Z. & Kovacs, K. J. Role of CX3CR1 (fractalkine receptor) in brain damage and inflammation induced by focal cerebral ischemia in mouse. *J Cereb Blood Flow Metab* 28, 1707–1721, doi: 10.1038/jcbfm.2008.64 (2008).

- 326. Dong, W. et al. Resveratrol attenuates ischemic brain damage in the delayed phase after stroke and induces messenger RNA and protein express for angiogenic factors. J Vasc Surg 48, 709–714, doi: 10.1016/j.jvs.2008.04.007 (2008).
- 327. Fujimoto, M. et al. Tissue inhibitor of metalloproteinases protect blood-brain barrier disruption in focal cerebral ischemia. J Cereb Blood Flow Metab 28, 1674–1685, doi: 10.1038/jcbfm.2008.59 (2008).
- Varga-Szabo, D. *et al.* The calcium sensor STIM1 is an essential mediator of arterial thrombosis and ischemic brain infarction. J Exp Med 205, 1583–1591, doi: 10.1084/jem.20080302 (2008).
- Min, J. et al. Differential neuroprotective effects of carnosine, anserine, and N-acetyl carnosine against permanent focal ischemia. J Neurosci Res 86, 2984–2991, doi: 10.1002/jnr.21744 (2008).
- Tang, X. N., Cairns, B., Cairns, N. & Yenari, M. A. Apocynin improves outcome in experimental stroke with a narrow dose range. *Neuroscience* 154, 556–562, doi: 10.1016/j.neuroscience.2008.03.090 (2008).
- Kilic, U., Kilic, E., Matter, C. M., Bassetti, C. L. & Hermann, D. M. TLR-4 deficiency protects against focal cerebral ischemia and axotomy-induced neurodegeneration. *Neurobiol Dis* 31, 33–40, doi: 10.1016/j.nbd.2008.03.002 (2008).
- 332. Li, J. et al. Misoprostol, an anti-ulcer agent and PGE2 receptor agonist, protects against cerebral ischemia. Neurosci Lett 438, 210–215, doi: 10.1016/j.neulet.2008.04.054 (2008).
- Haddad, M. *et al.* Reduction of hemorrhagic transformation by PJ34, a poly(ADP-ribose)polymerase inhibitor, after permanent focal cerebral ischemia in mice. *Eur J Pharmacol* 588, 52–57, doi: 10.1016/j.ejphar.2008.04.013 (2008).
- Mori, T. et al. Overexpression of human S100B exacerbates brain damage and periinfarct gliosis after permanent focal ischemia. Stroke 39, 2114–2121, doi: 10.1161/STROKEAHA.107.503821 (2008).
- Kim, E. et al. CD36/fatty acid translocase, an inflammatory mediator, is involved in hyperlipidemia-induced exacerbation in ischemic brain injury. J Neurosci 28, 4661–4670, doi: 10.1523/JNEUROSCI.0982-08.2008 (2008).
- 336. Li, J. M. et al. Temporary pretreatment with the angiotensin II type 1 receptor blocker, valsartan, prevents ischemic brain damage through an increase in capillary density. Stroke 39, 2029–2036, doi: 10.1161/STROKEAHA.107.503458 (2008).
- Oyamada, N. *et al.* The role of mineralocorticoid receptor expression in brain remodeling after cerebral ischemia. *Endocrinology* 149, 3764–3777, doi: 10.1210/en.2007-1770 (2008).
- Weise, J. et al. Overexpression of cellular prion protein alters postischemic Erk1/2 phosphorylation but not Akt phosphorylation and protects against focal cerebral ischemia. Restor Neurol Neurosci 26, 57–64 (2008).
- Wang, X. et al. Effects of neuroglobin overexpression on acute brain injury and long-term outcomes after focal cerebral ischemia. Stroke 39, 1869–1874, doi: 10.1161/STROKEAHA.107.506022 (2008).
- Polavarapu, R., An, J., Zhang, C. & Yepes, M. Regulated intramembrane proteolysis of the low-density lipoprotein receptor-related protein mediates ischemic cell death. Am J Pathol 172, 1355–1362, doi: 10.2353/ajpath.2008.070975 (2008).
- 341. Kim, H. H. *et al.* Additive effects of statin and dipyridamole on cerebral blood flow and stroke protection. *J Cereb Blood Flow Metab* 28, 1285–1293, doi: 10.1038/jcbfm.2008.24 (2008).
- Zhang, W. et al. Soluble epoxide hydrolase gene deletion is protective against experimental cerebral ischemia. Stroke 39, 2073–2078, doi: 10.1161/STROKEAHA.107.508325 (2008).
- Connolly, E. S., Jr., Winfree, C. J., Stern, D. M., Solomon, R. A. & Pinsky, D. J. Procedural and strain-related variables significantly affect outcome in a murine model of focal cerebral ischemia. *Neurosurgery* 38, 523–531, discussion 532 (1996).
- Schulte-Herbruggen, O., Klehmet, J., Quarcoo, D., Meisel, C. & Meisel, A. Mouse strains differ in their susceptibility to poststroke infections. *Neuroimmunomodulation* 13, 13–18, doi: 10.1159/000092109 (2006).
- 345. Majid, A. *et al.* Differences in vulnerability to permanent focal cerebral ischemia among 3 common mouse strains. *Stroke* **31**, 2707–2714 (2000).
- Maeda, K., Hata, R. & Hossmann, K. A. Regional metabolic disturbances and cerebrovascular anatomy after permanent middle cerebral artery occlusion in C57black/6 and SV129 mice. *Neurobiol Dis* 6, 101–108, doi: 10.1006/nbdi.1998.0235 (1999).
- 347. Tsuchiya, D., Hong, S., Kayama, T., Panter, S. S. & Weinstein, P. R. Effect of suture size and carotid clip application upon blood flow and infarct volume after permanent and temporary middle cerebral artery occlusion in mice. *Brain Res* 970, 131–139 (2003).
- Mao, Y., Yang, G. Y., Zhou, L. F., Stern, J. D. & Betz, A. L. Focal cerebral ischemia in the mouse: description of a model and effects of permanent and temporary occlusion. *Brain Res Mol Brain Res* 63, 366–370 (1999).
- 349. Barber, P. A. et al. Early T1- and T2-weighted MRI signatures of transient and permanent middle cerebral artery occlusion in a murine stroke model studied at 9.4T. Neurosci Lett 388, 54–59, doi: 10.1016/j.neulet.2005.06.067 (2005).
- Huang, J., Kim, L. J., Poisik, A., Pinsky, D. J. & Connolly, E. S., Jr. Does poly-L-lysine coating of the middle cerebral artery occlusion suture improve infarct consistency in a murine model? J Stroke Cerebrovasc Dis 7, 296–301 (1998).
- Yuan, F. et al. Optimizing suture middle cerebral artery occlusion model in C57BL/6 mice circumvents posterior communicating artery dysplasia. J Neurotrauma 29, 1499–1505, doi: 10.1089/neu.2011.2105 (2012).
- 352. Akamatsu, Y., Shimizu, H., Saito, A., Fujimura, M. & Tominaga, T. Consistent focal cerebral ischemia without posterior cerebral artery occlusion and its real-time monitoring in an intraluminal suture model in mice. J Neurosurg 116, 657–664, doi: 10.3171/2011.11.JNS111167 (2012).
- Tureyen, K., Vemuganti, R., Sailor, K. A. & Dempsey, R. J. Ideal suture diameter is critical for consistent middle cerebral artery occlusion in mice. *Neurosurgery* 56, 196–200, discussion 196-200 (2005).
- Macleod, M. R. *et al.* Good laboratory practice: preventing introduction of bias at the bench. *Stroke* 40, e50–52, doi: 10.1161/ STROKEAHA.108.525386 (2009).
- 355. Schlattmann, P. & Dirnagl, U. Statistics in experimental cerebrovascular research-comparison of two groups with a continuous outcome variable. J Cereb Blood Flow Metab 30, 474–479, doi: 10.1038/jcbfm.2009.266 (2010).
- 356. Sterne, J. A. & Davey Smith, G. Sifting the evidence-what's wrong with significance tests? BMJ 322, 226–231 (2001).
- 357. Korevaar, D. A., Hooft, L. & ter Riet, G. Systematic reviews and meta-analyses of preclinical studies: publication bias in laboratory animal experiments. *Lab Anim* 45, 225–230, doi: 10.1258/la.2011.010121 (2011).
- 358. Strom, J. O., Ingberg, E., Theodorsson, E. & Theodorsson, A. Effects of high and low 17beta-estradiol doses on focal cerebral ischemia: negative results. *Sci Rep* **3**, 3111, doi: 10.1038/srep03111 (2013).
- 359. Strom, J. O., Strid, T. & Hammarstrom, S. Disruption of the alox5ap gene ameliorates focal ischemic stroke: possible consequence of impaired leukotriene biosynthesis. *BMC Neurosci* 13, 146, doi: 10.1186/1471-2202-13-146 (2012).

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Author Contributions

E.I. contributed to designing the study, extracted data, performed the statistical analyses and drafted the manuscript. H.D. contributed to designing the study, extracted data and revised the manuscript. E.T. and A.T. contributed to designing the study and revising the manuscript. J.O.S. contributed to designing the study, analyzing the results and revising the manuscript.

Additional Information

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