

Since January 2020 Elsevier has created a COVID-19 resource centre with free information in English and Mandarin on the novel coronavirus COVID-19. The COVID-19 resource centre is hosted on Elsevier Connect, the company's public news and information website.

Elsevier hereby grants permission to make all its COVID-19-related research that is available on the COVID-19 resource centre - including this research content - immediately available in PubMed Central and other publicly funded repositories, such as the WHO COVID database with rights for unrestricted research re-use and analyses in any form or by any means with acknowledgement of the original source. These permissions are granted for free by Elsevier for as long as the COVID-19 resource centre remains active. Contents lists available at ScienceDirect







journal homepage: www.elsevier.com/locate/mehy

Harnessing adenosine A2A receptors as a strategy for suppressing the lung inflammation and thrombotic complications of COVID-19: Potential of pentoxifylline and dipyridamole

Check for updates

ABSTRACT

Counterproductive lung inflammation and dysregulated thrombosis contribute importantly to the lethality of advanced COVID-19. Adenosine A2A receptors (A2AR), expressed by a wide range of immune cells, as well as endothelial cells and platelets, exert cAMP-mediated anti-inflammatory and anti-thrombotic effects that potentially could be highly protective in this regard. The venerable drug pentoxifylline (PTX) exerts both anti-inflammatory and anti-thrombotic effects that reflect its ability to boost the responsiveness of A2AR to extracellular adenosine. The platelet-stabilizing drug dipyridamole (DIP) blocks intracellular uptake of extracellularly-generated adenosine, thereby up-regulating A2AR signaling in a way that should be functionally complementary to the impact of PTX in that regard. Moreover, DIP has recently been reported to slow the cellular replication of SARS-CoV-2 in clinically feasible concentrations. Both PTX and DIP are reasonably safe, well-tolerated, widely available, and inexpensive drugs. When COVID-19 patients can be treated within several days of symptom onset, using PTX + DIP in conjunction with hydroxychloroquine (HCQ) and an antibiotic – azithromycin (AZM) or doxycycline – might be warranted. HCQ and AZM can suppress SARS-CoV-2 proliferation in vitro and may slow the cell-to-cell spread of the virus; a large case series evaluating this combination in early-stage patients reported an impressively low mortality rate. However, whereas HCQ and AZM can promote QT interval lengthening and may be contraindicated in more advanced COVID-19 entailing cardiac damage, doxycycline has no such effect and exerts a potentially beneficial anti-inflammatory action. In contrast to HCQ, we propose that the combination of PTX + DIP can be used in both early and advanced stages of COVID-19. Concurrent use of certain nutraceuticals – yeast beta-glucan, zinc, vitamin D, spirulina, phase 2 inducers, N-acetylcysteine, glucosamine, quercetin, and magnesium – might also improve therapeutic outcomes in COVID-19.

Versatile anti-inflammatory effects of adenosine A2A receptors

The potential lethality of advanced COVID-19 stems not so much from the direct cytopathic effects of the virus, but from the florid lung inflammation and the endotheliopathy-induced thrombotic complications that it evokes [1-3]. Adenosine A2A receptors (A2AR) exert broad-spectrum anti-inflammatory and anti-thrombotic effects in a range of cells - including neutrophils, macrophages, lymphocytes, platelets, and endothelial cells - that have potential for providing protection in this regard [4-6]. A2AR is a 7-pass G-protein-coupled receptor that stimulates adenylate cyclase activity via $G\alpha_s$ [7]. The intracellular increase in cAMP that this evokes works in multiple complementary ways to suppress oxidant production, cytokine generation, expression of adhesion molecules, trans-endothelial migration of neutrophils, opening of the endothelial barrier, tissue factor generation, and platelet aggregation in A2AR-responsive cells [5]. Downregulation of NF-kappaB activation and JAK-STAT signaling pathways contribute importantly to these effects of cAMP [5]. Importantly, neutrophils, whose activation and transit into lung interstitial tissue and alveolar space is a key mediator of the respiratory distress syndrome associated with COVID-19, are highly responsive to the functionally suppressive effects of A2AR, as are the endothelial cells whose activation attracts and enables transendothelial passage of activated neutrophils [8–11].

These considerations suggest that selective agonists of A2AR may have important potential for blunting the lethality of COVID-19.

As may be expected, such agents have shown protective effects in rodent models of inflammatory lung injury [12–15]. Unfortunately, these agents are not yet clinically available. However, at least two drugs

https://doi.org/10.1016/j.mehy.2020.110051 Received 10 June 2020; Accepted 25 June 2020 0306-9877/ © 2020 Elsevier Ltd. All rights reserved. are currently available – venerable, reasonably safe and well tolerated, and inexpensive – that can function to up-regulate A2AR signaling: pentoxifylline (PTX) and dipyridamole (DIP).

Pentoxifylline and dipyridamole work in complementary ways to Up-regulate A2AR signaling

Although PTX is known to have broad anti-inflammatory activity, it is employed primarily in the treatment of intermittent claudication; by lessening neutrophil activation, PTX renders these cells more distensible, so that they can more readily pass through narrow capillaries in affected legs [16,17]. (Upstream stenotic obstructions decrease the transcapillary pressure gradient, rendering the passage of bulky neutrophils through narrow capillaries more difficult in this syndrome.) Although the clinical effects of PTX have usually been ascribed to the ability of this drug to inhibit cAMP phosphodiesterase - thereby boosting cAMP levels - this effect is only significant in vitro at millimolar concentrations that are orders of magnitude higher than the low micromolar concentrations of this drugs achieved clinically [16]. Ironically, however, it does appear that cAMP mediates PTX's clinical effects. Within the last decade, PTX's anti-inflammatory effects have been shown to be contingent on activation of A2AR [18-20]. Whether PTX can act as a direct agonist for A2AR is currently unclear, and some data argue against this [18]. What is clear is that PTX can potentiate the responsiveness of this receptor to adenosine. The latter is produced extracellularly from ATP released into the extracellular space, which is then converted to adenosine by the sequential activity of the CD39 and CD73 ecto-phosphatases expressed on the plasma membranes of A2ARexpressing cells [5,21].

The signaling activity of extracellularly-generated adenosine is terminated by intracellular uptake of the adenosine. The platelet-stabilizing agent DIP is distinguished by its ability to block this re-uptake by platelets [22,23]. Hence, DIP up-regulates the adenosine-mediated activation of platelet A2AR, thereby boosting platelet levels of cAMP, which functions to suppress platelet aggregation. Moreover, DIP blocks adenosine uptake by a range of other A2AR-expressing cell types, including endothelial cells, neutrophils, and monocytes [23–25].

It is evident that PTX and DIP have the potential to work in a complementary fashion to boost A2AR signaling – DIP can be expected to boost the extracellular levels of adenosine whose signaling activity PTX potentiates. Surprisingly, only a handful of studies have evaluated this combination experimentally or clinically – with encouraging results – likely because the mechanism of action of PTX has been clarified only recently [26–28].

Anti-inflammatory and Anti-Thrombotic effects of pentoxifylline

Pre-administration of PTX is protective in rodent models of acute respiratory distress syndrome (ARDS) evoked by lipopolysaccharide (LPS) administration or severe haemorrhage [29-31]. Clinically, it was found to reduce mortality, lower plasma tumor necrosis factor, and achieve clinical and radiological improvements in ARDS associated with cancer [32]. A meta-analysis of clinical studies found that PTX therapy is associated with a decrease in plasma levels of both tumor necrosis factor and C-reactive protein [33]. In chimpanzees, it was shown to blunt LPS-induced activation of coagulation and fibrinolysis [34]. In isolated lungs, PTX pre-treatment reduces the tissue injury induced by neutrophil infusion [35]. In endothelial cells, PTX counteracts the ability of pro-inflammatory cytokines to stimulate expression of adhesion factors and chemokine production [36]. These findings are expectable in light of the known effects of A2AR signaling, and encourage the speculation that PTX could have potential for blunting the exuberant lung inflammation and pro-thrombotic effects of advanced COVID-19. Not surprisingly, the use of PTX for treatment of ARDS associated with SARS infection was suggested in 2003 [37]. (Presumably, this was not studied because the syndrome rapidly disappeared.)

In seeming contradiction, a large multi-center study of lisofylline therapy in ARDS patients failed to show benefit [38]. Lisofylline is the R-isomer of a reductive metabolite of PTX, notable for its protective impact in rodent models of type 1 diabetes [39]. Conceivably, the impact of this agent on A2AR signaling – which has not been reported – is different than that of PTX. Alternatively, this finding may reflect the fact that, for unclear reasons, A2AR agonism is more effective for controlling ARDS when implemented before the syndrome becomes florid. Konrad and colleagues, in interpreting this result, suggest that adenosine levels may be too low in the context of advanced sepsis [19]. If so, the concurrent use of DIP would make logical sense.

Dipyridamole – an Anti-inflammatory agent which can suppress SARS-CoV-2 replication

Most studies with DIP have focused on its platelet-stabilizing effects – which presumably could provide some protection from SARS-CoV-2's pro-thrombotic effects – but experimental studies also show that DIP can act on neutrophils to suppress superoxide production, adhesion to endothelial cells, and, in a mouse model of anti-phospholipid syndrome (a sometime feature of COVID-19), NETosis formation [40–43]. And DIP has been shown to suppress superoxide production and tissue factor expression in monocytes [44].

Of particular pertinence is this new discovery: Chinese researchers have reported that, in clinically relevant concentrations as low as 100 nM, DIP slows the replication of SARS-CoV-2 in Vero E6 cells; this effect may be mediated in part but not entirely by the binding to DIP to the SARS-CoV-2 protease Mpro [45]. In a controlled pilot study, 31 hospitalized COVID-19 patients with respiratory difficulties were treated with either DIP (50 mg 3 times daily) or placebo; of the 14 patients who received DIP, including 8 that were severely ill, all but one recovered, and the remaining patient was in remission at time of the report. Of 12 severely ill patients in the control group, 2 patients died and 2 were in remission [45]. The difference in therapeutic outcome just missed traditional statistical significance (p = 0.06). The response in D-dimer levels was significantly better in the treated than in the control patients.

An anti-viral effect of DIP is not unprecedented – cell culture studies have reported that this agent can slow the proliferation of various RNA viruses, and a Russian clinical report some decades ago concluded that DIP administered prophylactically was effective for reducing risk for influenza and upper respiratory infections [45–50].

These considerations suggest that a PTX/DIP regimen might have considerable potential for control of the progression and complications of COVID-19. Provisionally, we recommend dosage schedules for PTX and DIP typically used for their approved indications: PTX 400 mg 3 times daily, and DIP 50 mg 3 times daily.

Another venerable drug which has been suggested for use in COVID-19 management is the anti-parasitic agent ivermectin. Evidence that it can suppress proliferation of SARS-CoV-2 in cell culture is likely of little pertinence, as the IC50 concentration that achieves this – about 2 uM – is vastly higher than the plasma concentrations achievable by doses approved for clinical use [51,52]. Nonetheless, anecdotal claims of its apparent effectiveness in late-stage COVID-19 are encouraging clinical trials with this agent. Largely overlooked is the fact that, in oral doses that are roughly analogous to the standard clinical dose in humans, ivermectin pre-administration can protect mice from a lethal dose of LPS [53,54]. Hence, if ivermectin proves useful in COVID-19, an antiinflammatory mechanism may underlie this benefit.

Concurrent nutraceutical support for antigen-specific immunity

However, it should be acknowledged that A2AR agonism also has potential for suppressing the dendritic cell activity that provides the antigen presentation necessary for developing an antiviral antibody response [4]. The authors have been unavailable to find any studies suggesting that PTX increases infection risk - in marked contrast to the well-known literature on anti-inflammatory corticosteroids - so perhaps this is a relatively minor consideration. Indeed, some studies fail to find an effect of A2AR agonists on dendritic cell antigen presentation [55]. And PTX has actually been suggested as an adjuvant to vaccination, as it boosts memory response to vaccination by increasing survival of activated T cells [56]. Nevertheless, it would seem prudent to complement PTX/DIP therapy with agents such as yeast beta-glucan that specifically boost dendritic cell activity, as a compensatory measure [57–59]. Curiously, beta-glucan administration has been found to be protective in rodent models of sepsis-induced ARDS [60,61]. Supplemental zinc could be another worthwhile adjuvant measure, as it has been found to decrease incidence of infection while lowering systemic markers of inflammation in elderly subjects [62].

Incorporating pentoxifylline/dipyridamole into early-stage protocols

In early-stage ambulatory patients with COVID-19, it would be appropriate to consider using PTX + DIP in conjunction with hydroxychloroquine (HCQ). Currently, this agent is the most commonly used drug for treatment of early-stage COVID-19 [63]. HCQ decreases replication of SARS-CoV-2 in vitro in clinically relevant concentrations [64,65]. Studies examining the molecular biology of SARS-CoV-2 cellto-cell spread have found that endosomal cathepsin L protease activity markedly expedites such spread, presumably because it enables SARS-CoV-2 virions taken up into cellular endosomes to fuse their membranes with that of the endosome, thereby allowing the virion to enter the cytoplasm and begin replication [66]. It is well known that HCQ functions to alkalinize endosomes, and this would be expected to inhibit cathepsin L activity [67,68]. Moreover, HCQ has recently been shown to inhibit activation of NADPH oxidase complexes in endosomes; this would be expected to exert an anti-inflammatory effect that might complement the anti-viral activity of this agent [69]. This model makes evident the desirability of employing HCQ as early during the clinical course of COVID-19 as feasible, as the ability of this agent to slow cell-to-cell spread may be of less utility once the lung epithelium is already widely colonized by the virus. Pharmacokinetic modeling, combined with in vitro data, suggest a regimen for HCQ of 400 mg twice daily for one day, and 200 mg twice daily for a further 4 days; this is predicted to maintain antiviral plasma levels of HCQ for at least 10 days [64].

HCQ therapy often induces modest increases in QT interval, and concurrent administration of azithromycin can amplify this effect. Such increases can increase chances for dangerous torsade de pointes arrhythmias; studies examining ECGs in hospitalized COVID-19 patients treated with this combination reported 2 cases of torsade de pointes in 640 such patients [70–73]. Although it is very rare for these drugs to induce arrhythmias when used for their current indications, SARS-CoV-2 can directly attack the heart, and conceivably this could potentiate the pro-arrhythmic impact of HCQ [74,75]. Hence, monitoring of QT interval appears to be prudent when using HCQ in COVID-19. Fortunately, neither PTX nor DIP have been linked to QT prolongation or torsade de pointes arrhythmias.

Adding an antibiotic such as azithromycin or doxycycline to earlystage treatment of COVID-19 to prevent bacterial super-infection – as many doctors have done when employing HCQ in COVID-19 therapy – would also be a reasonable option [76]. In addition to their antibiotic activities, azithromycin exerts anti-viral effects in vitro against various viruses, and doxycycline has anti-inflammatory properties that likely would be beneficial in SARS-CoV-2-induced cytokine storm [77,78]. However, azithromycin might be inappropriate for late-stage therapy, as it has a greater tendency than HCQ to prolong QT intervals [75]; doxycycline does not have this effect.

Nutraceutical adjuvant measures

Nutraceutical adjuvant measures that support the antigen-specific immune response – such as yeast glucan and zinc – would also appear to be indicated. Nutraceuticals that might be expected to boost the interferon response evoked by SARS-CoV-2 while lessening the contribution of oxidants to lung inflammation have been proposed, including spirulina, phase 2 inducers, N-acetylcysteine [79]. Supplemental glucosamine may likewise up-regulate the type 1 interferon responses to viruses, while exerting anti-inflammatory effects that render it protective in rodent models of sepsis and lung inflammation induced by LPS or cigarette smoke [80–84]. This anti-inflammatory effect might reflect up-regulated activity of the de-ubiquitinase A20, which opposes TRAF6 signaling [85]. Up-regulation of type 1 interferon induction may also play a role in the anti-viral effects of quercetin [86,87].

Theoretical considerations as well as epidemiological findings suggest that good vitamin D status may also be protective [88–90]. In the context of inflammation, lung epithelium and alveolar macrophages can convert circulating 25-hydroxyvitamin D to the active hormone calcitriol; this in turn can boost expression of the antimicrobial protein cathelicidin, which is destructive to many enveloped viruses [91–95]. Intracellular magnesium supports effective function of the A2AR [96]. Lower serum magnesium has been associated with increased thrombotic risk and slowed fibrinolysis [97–101]. Hypomagnesemia predicts poor outcomes in ICU patients, and its correction may improve their prognosis [102,103]. Moreover, magnesium deficiency up-regulates NF-kappaB activation and HMBG1 secretion in LPS-treated macrophages – consistent with other evidence that it may up-regulate inflammation [104,105]. In light of the pro-thrombotic and proinflammatory diathesis associated with COVID-19, magnesium supplementation might be prudent.

Conflict of interest

Dr. DiNicolantonio is Director of Scientific affairs at AIDP.

References

- Helms J, Tacquard C, Severac F, et al. High risk of thrombosis in patients with severe SARS-CoV-2 infection: a multicenter prospective cohort study. Intensive Care Med 2020 May 4.
- [2] Varga Z, Flammer AJ, Steiger P, et al. Endothelial cell infection and endotheliitis in COVID-19. Lancet 2020 April 20.
- [3] Escher R, Breakey N, Lammle B. Severe COVID-19 infection associated with endothelial activation. Thromb Res 2020 April;15(190):62.
- [4] Hasko G, Csoka B, Nemeth ZH, Vizi ES, Pacher P. A(2B) adenosine receptors in immunity and inflammation. Trends Immunol 2009 June;30(6):263–70.
- [5] Milne GR, Palmer TM. Anti-inflammatory and immunosuppressive effects of the A2A adenosine receptor. Sci World J 2011 February;3(11):320–39.
- [6] Guerrero A. A2A adenosine receptor agonists and their potential therapeutic applications. An update. Curr Med Chem 2018;25(30):3597–612.
- [7] Umapathy NS, Fan Z, Zemskov EA, Alieva IB, Black SM, Verin AD. Molecular mechanisms involved in adenosine-induced endothelial cell barrier enhancement. Vasc Pharmacol 2010 May;52(5–6):199–206.
- [8] Fredholm BB, Zhang Y, van dP, I. Adenosine A2A receptors mediate the inhibitory effect of adenosine on formyl-Met-Leu-Phe-stimulated respiratory burst in neutrophil leucocytes. Naunyn Schmiedebergs Arch Pharmacol 1996 August;354(3):262-7.
- [9] Sullivan GW, Linden J, Buster BL, Scheld WM. Neutrophil A2A adenosine receptor inhibits inflammation in a rat model of meningitis: synergy with the type IV phosphodiesterase inhibitor, rolipram. J Infect Dis 1999 November:180(5):1550–60.
- [10] Iwamoto T, Umemura S, Toya Y, et al. Identification of adenosine A2 receptorcAMP system in human aortic endothelial cells. Biochem Biophys Res Commun 1994 March 15;199(2):905–10.
- [11] Hassanian SM, Dinarvand P, Rezaie AR. Adenosine regulates the proinflammatory signaling function of thrombin in endothelial cells. J Cell Physiol 2014 September;229(9):1292–300.
- [12] Schepp CP, Reutershan J. Bench-to-bedside review: adenosine receptors-promising targets in acute lung injury? Crit Care 2008;12(5):226.
- [13] Folkesson HG, Kuzenko SR, Lipson DA, Matthay MA, Simmons MA. The adenosine 2A receptor agonist GW328267C improves lung function after acute lung injury in rats. Am J Physiol Lung Cell Mol Physiol 2012 August 1;303(3):L259–71.
- [14] Gonzales JN, Gorshkov B, Varn MN, et al. Protective effect of adenosine receptors against lipopolysaccharide-induced acute lung injury. Am J Physiol Lung Cell Mol Physiol 2014 March 15;306(6):L497–507.
- [15] Aggarwal NR, D'Alessio FR, Eto Y, et al. Macrophage A2A adenosinergic receptor modulates oxygen-induced augmentation of murine lung injury. Am J Respir Cell Mol Biol 2013 May;48(5):635–46.
- [16] McCarty MF, O'Keefe JH, DiNicolantonio JJ. Pentoxifylline for vascular health: a brief review of the literature. Open Heart 2016;3(1):e000365.
- [17] Armstrong Jr. M, Needham D, Hatchell DL, Nunn RS. Effect of pentoxifylline on the flow of polymorphonuclear leukocytes through a model capillary. Angiology 1990 April;41(4):253–62.
- [18] Kreth S, Ledderose C, Luchting B, Weis F, Thiel M. Immunomodulatory properties of pentoxifylline are mediated via adenosine-dependent pathways. Shock 2010 July;34(1):10–6.
- [19] Konrad FM, Neudeck G, Vollmer I, Ngamsri KC, Thiel M, Reutershan J. Protective effects of pentoxifylline in pulmonary inflammation are adenosine receptor A2A dependent. FASEB J 2013 September;27(9):3524–35.
- [20] Li H, Tan G, Tong L, et al. Pentoxifylline inhibits pulmonary inflammation induced by infrarenal aorticcross-clamping dependent of adenosine receptor A2A. Am J Transl Res 2016;8(5):2210–21.
- [21] Antonioli L, Pacher P, Vizi ES, Hasko G. CD39 and CD73 in immunity and inflammation. Trends Mol Med 2013 June;19(6):355–67.
- [22] Summers A, Subbaraok Rucinski B, Niewiarowski S. The effect of dipyridamole on adenosine uptake by platelets ex vivo. Thromb Res 1977 November;11(5):611–8.
- [23] Balakumar P, Nyo YH, Renushia R, et al. Classical and pleiotropic actions of dipyridamole: Not enough light to illuminate the dark tunnel? Pharmacol Res 2014 September;87:144–50.
- [24] Cronstein DN, Kramer SB, Weissmann G, Hirschhorn R. Adenosine: a physiological modulator of superoxide anion generation by human neutrophils. J Exp Med 1983 October 1;158(4):1160–77.
- [25] Le V, V, Chen YL, Masson I et al. Inhibition of human monocyte TNF production by adenosine receptor agonists. Life Sci 1993;52(24):1917-24.
- [26] Santos MT, Martinez-Sales V, Valles J, et al. Prostacyclin production by rat aorta "in vitro" is increased by the combined action of dipyridamole plus pentoxifylline. Prostaglandins 1985 January;29(1):113–22.
- [27] Yaya R, Aznar J, Vaya A, et al. Effect of dipyridamole plus pentoxifylline in patients with diffuse cerebrovascular insufficiency. Thromb Haemost 1985 December 17;54(4):896.
- [28] Santos MT, Valles J, Aznar J, Yaya R, Perez-Requejo JL. Effects of dipyridamole,

pentoxifylline or dipyridamole plus pentoxifylline on platelet reactivity in patients with ischemic cerebrovascular insufficiency. Thromb Res 1993 November 1:72(3):219–29.

- [29] Barroso-Aranda J, Schmid-Schonbein GW. Pentoxifylline pretreatment decreases the pool of circulating activated neutrophils, in-vivo adhesion to endothelium, and improves survival from hemorrhagic shock. Biorheology 1990;27(3–4):401–18.
- [30] Barroso-Aranda J, Schmid-Schonbein GW. Pentoxifylline pretreatment decreases neutrophil activation during endotoxic shock and improves survival. *Pentoxifylline* and analogues: Effects on leukocyte function.Basel. Skarger 1990:97–103.
- [31] Deree J, Martins J, de CT et al. Pentoxifylline attenuates lung injury and modulates transcription factor activity in hemorrhagic shock. J Surg Res 2007 November:143(1):99-108.
- [32] Ardizzoia A, Lissoni P, Tancini G, et al. Respiratory distress syndrome in patients with advanced cancer treated with pentoxifylline: a randomized study. Support Care Cancer 1993 November;1(6):331–3.
- [33] Brie D, Sahebkar A, Penson PE, et al. Effects of pentoxifylline on inflammatory markers and blood pressure: a systematic review and meta-analysis of randomized controlled trials. J Hypertens 2016 December;34(12):2318–29.
- [34] Levi M, ten CH, Bauer KA et al. Inhibition of endotoxin-induced activation of coagulation and fibrinolysis by pentoxifylline or by a monoclonal anti-tissue factor antibody in chimpanzees. J Clin Invest 1994 January;93(1):114-20.
- [35] McDonald RJ. Pentoxifylline reduces injury to isolated lungs perfused with human neutrophils. Am Rev Respir Dis 1991 December;144(6):1347–50.
- [36] Krakauer T. Pentoxifylline inhibits ICAM-1 expression and chemokine production induced by proinflammatory cytokines in human pulmonary epithelial cells. Immunopharmacology 2000 March;46(3):253–61.
- [37] Bermejo Martin JF, Jimenez JL, Munoz-Fernandez A. Pentoxifylline and severe acute respiratory syndrome (SARS): a drug to be considered. Med Sci Monit 2003 June;9(6):SR29-SR34.
- [38] Randomized, placebo-controlled trial of lisofylline for early treatment of acute lung injury and acute respiratory distress syndrome. Crit Care Med 2002 January;30(1):1-6.
- [39] Yang Z, Chen M, Nadler JL. Lisofylline: a potential lead for the treatment of diabetes. Biochem Pharmacol 2005 January 1;69(1):1–5.
- [40] Suzuki S, Sugai K, Sato H, Sakatume M, Arakawa M. Inhibition of active oxygen generation by dipyridamole in human polymorphonuclear leukocytes. Eur J Pharmacol 1992 December 1;227(4):395–401.
- [41] Chello M, Mastroroberto P, Malta E, Cirillo F, Celi V. Inhibition by dipyridamole of neutrophil adhesion to vascular endothelium during coronary bypass surgery. Ann Thorac Surg 1999 May;67(5):1277–82.
- [42] Hallevi H, Hazan-Halevy I, Paran E. Modification of neutrophil adhesion to human endothelial cell line in acute ischemic stroke by dipyridamole and candesartan. Eur J Neurol 2007 September;14(9):1002–7.
- [43] Ali RA, Gandhi AA, Meng H, et al. Adenosine receptor agonism protects against NETosis and thrombosis in antiphospholipid syndrome. Nat Commun 2019 April 23;10(1):1916.
- [44] Brozna JP, Horan M, Carson SD. Dipyridamole inhibits O2- release and expression of tissue factor activity by peripheral blood monocytes stimulated with lipopolysaccharide. Thromb Res 1990 October 15;60(2):141–56.
- [45] Liu X, Li Z, Liu S, et al. Potential therapeutic effects of dipyridamole in the severely ill patients with COVID-19. Acta Pharm Sin B 2020;April:20.
- [46] Korbecki M, Bankowski A, Filczak K, Klimek A. Dipyridamole as an inhibitor of vaccinia virus replication. Mol Gen Mikrobiol Virusol 1985 January;1:29–32.
- [47] Hay KA, Gaydos A, Tenser RB. Inhibition of herpes simplex virus reactivation by dipyridamole in a mouse model. J Med Virol 1996 October;50(2):198–203.
- [48] Tenser RB, Gaydos A, Hay KA. Inhibition of herpes simplex virus reactivation by dipyridamole. Antimicrob Agents Chemother 2001 December;45(12):3657–9.
- [50] Kuzmov K, Galabov AS, Radeva K, Kozhukharova M, Milanov K. Epidemiological trial of the prophylactic effectiveness of the interferon inducer dipyridamole with respect to influenza and acute respiratory diseases. Zh Mikrobiol Epidemiol Immunobiol 1985 June;6:26–30.
- [51] Caly L, Druce JD, Catton MG, Jans DA, Wagstaff KM. The FDA-approved drug ivermectin inhibits the replication of SARS-CoV-2 in vitro. Antiviral Res 2020 April;3(178):104787.
- [52] Schmith VD, Zhou JJ, Lohmer LR. The approved dose of ivermectin alone is not the ideal dose for the treatment of COVID-19. Clin Pharmacol Ther 2020 May 7.
- [53] Zhang X, Song Y, Ci X, et al. Ivermectin inhibits LPS-induced production of inflammatory cytokines and improves LPS-induced survival in mice. Inflamm Res 2008 November;57(11):524–9.
- [54] DiNicolantonio JJ, Barroso-Aranda J, McCarty MF. Ivermectin may be a clinically useful anti-inflammatory agent for late-stage COVID-19. submitted for publication 2020.
- [55] Li L, Huang L, Ye H, et al. Dendritic cells tolerized with adenosine A(2)AR agonist attenuate acute kidney injury. J Clin Invest 2012 November;122(11):3931–42.
- [56] Suresh R, Vig M, Bhatia S, et al. Pentoxifylline functions as an adjuvant in vivo to enhance T cell immune responses by inhibiting activation-induced death. J Immunol 2002 October 15;169(8):4262–72.
- [57] Vetvicka V, Vannucci L, Sima P, Richter J. Beta glucan: supplement or drug? From laboratory to clinical trials. Molecules 2019;24(7).
- [58] Talbott SM, Talbott JA. Baker's yeast beta-glucan supplement reduces upper respiratory symptoms and improves mood state in stressed women. J Am Coll Nutr 2012 August;31(4):295–300.
- [59] Wang M, Zhang L, Yang R, et al. Improvement of immune responses to influenza vaccine (H5N1) by sulfated yeast beta-glucan. Int J Biol Macromol 2016

December;93(Pt A):203-7.

- [60] Babayigit H, Kucuk C, Sozuer E, Yazici C, Kose K, Akgun H. Protective effect of beta-glucan on lung injury after cecal ligation and puncture in rats. Intensive Care Med 2005 June;31(6):865–70.
- [61] Bedirli A, Kerem M, Pasaoglu H, et al. Beta-glucan attenuates inflammatory cytokine release and prevents acute lung injury in an experimental model of sepsis. Shock 2007 April;27(4):397–401.
- [62] Prasad AS, Beck FW, Bao B, et al. Zinc supplementation decreases incidence of infections in the elderly: effect of zinc on generation of cytokines and oxidative stress. Am J Clin Nutr 2007 March;85(3):837–44.
- [63] Meo SA, Klonoff DC, Akram J. Efficacy of chloroquine and hydroxychloroquine in the treatment of COVID-19. Eur Rev Med Pharmacol Sci 2020 April:24(8):4539–47.
- [64] Yao X, Ye F, Zhang M, et al. In vitro antiviral activity and projection of optimized dosing design of hydroxychloroquine for the treatment of severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2). Clin Infect Dis 2020 March 9.
- [65] Liu J, Cao R, Xu M, et al. Hydroxychloroquine, a less toxic derivative of chloroquine, is effective in inhibiting SARS-CoV-2 infection in vitro. Cell Discov 2020;6:16.
- [66] Ou X, Liu Y, Lei X, et al. Characterization of spike glycoprotein of SARS-CoV-2 on virus entry and its immune cross-reactivity with SARS-CoV. Nat Commun 2020 March 27;11(1):1620.
- [67] Fox RI. Mechanism of action of hydroxychloroquine as an antirheumatic drug. Semin Arthritis Rheum 1993 October;23(2 Suppl 1):82–91.
- [68] Dufour E, Dive V, Toma F. Delineation of chicken cathepsin L secondary structure; relationship between pH dependence activity and helix content. BBA 1988 June 29;955(1):58–64.
- [69] Muller-Calleja N, Manukyan D, Canisius A, Strand D, Lackner KJ. Hydroxychloroquine inhibits proinflammatory signalling pathways by targeting endosomal NADPH oxidase. Ann Rheum Dis 2017 May;76(5):891–7.
- [70] Saleh M, Gabriels J, Chang D, et al. The effect of chloroquine, hydroxychloroquine and azithromycin on the corrected QT interval in patients with SARS-CoV-2 infection. Circ Arrhythm Electrophysiol 2020;29.
- [71] Mercuro NJ, Yen CF, Shim DJ et al. Risk of QT Interval Prolongation Associated With Use of Hydroxychloroquine With or Without Concomitant Azithromycin Among Hospitalized Patients Testing Positive for Coronavirus Disease 2019 (COVID-19). JAMA Cardiol 2020 May 1.
- [72] Chorin E, Wadhwani L, Magnani S, et al. QT interval prolongation and torsade de pointes in patients with COVID-19 treated with hydroxychloroquine/azithromycin. Heart Rhythm 2020 May 11.
- [73] Ramireddy A, Chugh H, Reinier K, et al. Experience with hydroxychloroquine and azithromycin in the COVID-19 pandemic: implications for QT interval monitoring. J Am Heart Assoc 2020 May:28:e017144.
- [74] Shi S, Qin M, Shen B et al. Association of Cardiac Injury With Mortality in Hospitalized Patients With COVID-19 in Wuhan, China. JAMA Cardiol 2020 March 25.
- [75] Naksuk N, Lazar S, Peeraphatdit TB. Cardiac safety of off-label COVID-19 drug therapy: a review and proposed monitoring protocol. Eur Heart J Acute Cardiovasc Care 2020 May;6.
- [76] Risch HA. Early outpatient treatment of symptomatic, high-risk covid-19 patients that should be ramped-up immediately as key to the pandemic crisis. Am J Epidemiol 2020 May 27.
- [77] Damle B, Vourvahis M, Wang E, Leaney J, Corrigan B. Clinical pharmacology perspectives on the antiviral activity of azithromycin and use in COVID-19. Clin Pharmacol Ther 2020 April 17.
- [78] Conforti C, Giuffrida R, Zalaudek I, Di MN. Doxycycline, a widely used antibiotic in dermatology with a possible anti-inflammatory action against IL-6 in COVID-19 outbreak. Dermatol Ther 2020 April;20:e13437.
- [79] McCarty MF, DiNicolantonio JJ. Nutraceuticals have potential for boosting the type 1 interferon response to RNA viruses including influenza and coronavirus. Prog Cardiovasc Dis 2020 February 12.
- [80] Song N, Qi Q, Cao R, et al. MAVS O-GlcNAcylation is essential for host antiviral immunity against lethal RNA viruses. Cell Rep 2019 August 27;28(9):2386–96.
- [81] Silva JF, Olivon VC, Mestriner FLAC, et al. Acute increase in O-GlcNAc improves survival in mice with LPS-induced systemic inflammatory response syndrome. Front Physiol 2019;10:1614.
- [82] Hwang JS, Kim KH, Park J, et al. Glucosamine improves survival in a mouse model of sepsis and attenuates sepsis-induced lung injury and inflammation. J Biol Chem 2019 January 11;294(2):608–22.
- [83] Wu YL, Lin AH, Chen CH, et al. Glucosamine attenuates cigarette smoke-induced lung inflammation by inhibiting ROS-sensitive inflammatory signaling. Free Radic Biol Med 2014 April;69:208–18.
- [84] Chuang KH, Peng YC, Chien HY, Lu ML, Du HI, Wu YL. Attenuation of LPS-induced lung inflammation by glucosamine in rats. Am J Respir Cell Mol Biol 2013 December;49(6):1110–9.
- [85] Yao D, Xu L, Xu O, et al. O-Linked beta-N-Acetylglucosamine Modification of A20 Enhances the Inhibition of NF-kappaB (Nuclear Factor-kappaB) Activation and Elicits Vascular Protection After Acute Endoluminal Arterial Injury. Arterioscler Thromb Vasc Biol 2018 June;38(6):1309–20.
- [86] DiNicolantonio JJ, McCarty MF. Targeting Casein kinase 2 with quercetin or enzymatically modified isoquercitrin as a strategy for boosting the type 1 interferon response to viruses and promoting cardiovascular health. Med Hypotheses 2020 May;4(142):109800.
- [87] Peng D, Chen L, Sun Y, et al. Melanoma suppression by quercein is correlated with RIG-I and type I interferon signaling. Biomed Pharmacother 2020 May;125:109984.

- [89] Rhodes JM, Subramanian S, Laird E, Kenny RA. Editorial: low population mortality from COVID-19 in countries south of latitude 35 degrees North supports vitamin D as a factor determining severity. Aliment Pharmacol Ther 2020 April 20.
- [90] Marik PE, Kory P, Varon J. Does vitamin D status impact mortality from SARS-CoV-2 infection? Med Drug Discov 2020 April;29:100041.
- [91] Telcian AG, Zdrenghea MT, Edwards MR, et al. Vitamin D increases the antiviral activity of bronchial epithelial cells in vitro. Antiviral Res 2017 January;137:93–101.
- [92] Pryke AM, Duggan C, White CP, Posen S, Mason RS. Tumor necrosis factor-alpha induces vitamin D-1-hydroxylase activity in normal human alveolar macrophages. J Cell Physiol 1990 March;142(3):652–6.
- [93] Tripathi S, Tecle T, Verma A, Crouch E, White M, Hartshorn KL. The human cathelicidin LL-37 inhibits influenza A viruses through a mechanism distinct from that of surfactant protein D or defensins. J Gen Virol 2013 January;94(Pt 1):40–9.
- [94] Brice DC, Diamond G. Antiviral activities of human host defense peptides. Curr Med Chem 2020;27(9):1420–43.
- [95] Beard JA, Bearden A, Striker R. Vitamin D and the anti-viral state. J Clin Virol 2011 March;50(3):194–200.
- [96] Ye L, Neale C, Sljoka A, et al. Mechanistic insights into allosteric regulation of the A2A adenosine G protein-coupled receptor by physiological cations. Nat Commun 2018 April 10;9(1):1372.
- [97] Sobczak AIS, Phoenix FA, Pitt SJ, Ajjan RA, Stewart AJ. Reduced plasma magnesium levels in type-1 diabetes associate with prothrombotic changes in fibrin clotting and fibrinolysis. Thromb Haemost 2020 February;120(2):243–52.
- [98] Gromova OA, Torshin IY, Kobalava ZD, et al. Deficit of magnesium and states of

hypercoagulation: intellectual analysis of data obtained from a sample of patients aged 18–50 years from medical and preventive facilities in Russia. Kardiologiia 2018 November 18;58(4):22–35.

- [99] Cicek G, Acikgoz SK, Yayla C, Kundi H, Ileri M. Magnesium as a predictor of acute stent thrombosis in patients with ST-segment elevation myocardial infarction who underwent primary angioplasty. Coron Artery Dis 2016 January;27(1):47–51.
- [100] Sheu JR, Hsiao G, Shen MY, Lee YM, Yen MH. Antithrombotic effects of magnesium sulfate in in vivo experiments. Int J Hematol 2003 May;77(4):414–9.
- [101] Shechter M, Merz CN, Rude RK, et al. Low intracellular magnesium levels promote platelet-dependent thrombosis in patients with coronary artery disease. Am Heart J 2000 August;140(2):212–8.
- [102] Jiang P, Lv Q, Lai T, Xu F. Does hypomagnesemia impact on the outcome of patients admitted to the intensive care unit? A systematic review and meta-analysis. Shock 2017 March;47(3):288–95.
- [103] Charles BS, Menon I, Girish TS, Cherian AM. Hypomagnesemia in the. J Assoc Physicians India 2016 November;64(11):15–9.
- [104] Liu Z, Chang Y, Zhang J, et al. Magnesium deficiency promotes secretion of highmobility group box 1 protein from lipopolysaccharide-activated macrophages in vitro. J Surg Res 2013 April;180(2):310–6.
- [105] Nielsen FH. Magnesium deficiency and increased inflammation: current perspectives. J Inflamm Res 2018;11:25–34.

James J. DiNicolantonio^{a,*}, Jorge Barroso-Aranda^b ^a Mid America Heart Institute, Kansas City, MO, USA ^b Clinica Libre de Adicciones. Tijuana, B.C., Mexico E-mail address: jjdinicol@gmail.com (J.J. DiNicolantonio).

^{*} Corresponding author.