

Draft Genome Sequence of the Yeast Vanrija humicola (Formerly Cryptococcus humicola) Strain UJ1, a Producer of D-Aspartate Oxidase

gen@meAnnouncements™

Daiki Imanishi,^a Katsumasa Abe,^a Yoshio Kera,^a Shouji Takahashi^a

AMERICAN SOCIETY FOR MICROBIOLOGY

^aDepartment of Bioengineering, Nagaoka University of Technology, Nagaoka, Niigata, Japan

ABSTRACT Vanrija humicola (Cryptococcus humicola) strain UJ1 is a basidiomycetous yeast that produces p-aspartate oxidase, which is highly specific to p-aspartate. Here, we report the 22.6-Mb draft genome sequence of V. humicola strain UJ1, which comprises 22.6 Mb in 46 scaffolds, with an overall G+C content of 62.82%, comprising 46 scaffolds with an N_{50} of 1.34 Mb.

Vanrija humicola strain UJ1 (=JCM 9575), formerly known as *Cryptococcus humicola* strain UJ1, is a basidiomycetous yeast that utilizes D-aspartate as a sole source of carbon, nitrogen, or both, which is caused by a flavin enzyme, D-aspartate oxidase (DDO) (1, 2). DDO of *V. humicola* UJ1 is produced only in the presence of D-aspartate in culture media and has higher catalytic activity and specificity toward D-aspartate than DDOs from other origins (1, 3), which makes it useful for D-aspartate identification and quantification. The yeast draft genome sequence provides information on the D-aspartate-specific induction mechanism and the physiological significance of DDO in the yeast.

The genome sequence of *V. humicola* strain UJ1 was generated using an Illumina HiSeq 2500 platform. The sequencing generated 44,746,782 paired ends that were used for *de novo* assembly with Velvet version 1.2.10 (4) and Platanus version 1.2.4 (5) software. This assembly represents a total of 46 scaffolds with total and average lengths of 22,628,423 and 491,922 bp, respectively. The maximum and minimum scaffold lengths were 3,532,612 and 151 bp, respectively. The scaffold N_{50} and N_{90} values were 1,340,400 and 602,907 bp, respectively. The overall G+C content was determined to be 62.82%. Gene prediction using AUGUSTUS (6) trained with the parameters of the species *Cryptococcus neoformans* resulted in 8,919 genes and 37,033 exons. Additionally, 320 tRNAs were predicted using tRNAscan-SE (7). An automatic annotation of predicted open reading frames was carried out using Blast2GO Basic (8).

Accession number(s). This whole-genome shotgun sequencing project has been deposited in DDBJ/EMBL/GenBank under the accession numbers BFAH01000001 to BFAH01000046 (scaffolds 1 to 19 and 21 to 47, respectively). This paper describes the first version of the genome.

ACKNOWLEDGMENTS

The genome sequencing was supported by the Hokkaido System Science Co., Ltd. (Sapporo, Hokkaido, Japan).

We thank Enago for the English language review.

Received 18 January 2018 Accepted 29 January 2018 Published 15 March 2018

Citation Imanishi D, Abe K, Kera Y, Takahashi S. 2018. Draft genome sequence of the yeast *Vanrija humicola* (formerly *Cryptococcus humicola*) strain UJ1, a producer of D-aspartate oxidase. Genome Announc 6:e00068-18. https://doi.org/10.1128/genomeA.00068-18.

Copyright © 2018 Imanishi et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International license.

Address correspondence to Shouji Takahashi, shoutaka@vos.nagaokaut.ac.jp.

REFERENCES

- Yamada R, Ujiie H, Kera Y, Nakase T, Kitagawa K, Imasaka T, Arimoto K, Takahashi M, Matsumura Y. 1996. Purification and properties of D-aspartate oxidase from *Cryptococcus humicolus* UJ1. Biochim Biophys Acta 1294: 153–158. https://doi.org/10.1016/0167-4838(96)00012-X.
- Takahashi S, Kakuichi T, Fujii K, Kera Y, Yamada RH. 2005. Physiological role of D-aspartate oxidase in the assimilation and detoxification of D-aspartate in the yeast *Cryptococcus humicola*. Yeast 22:1203–1212. https://doi.org/10.1002/yea.1303.
- Takahashi S, Takahashi T, Kera Y, Matsunaga R, Shibuya H, Yamada RH. 2004. Cloning and expression in *Escherichia coli* of the p-aspartate oxidase gene from the yeast *Cryptococcus humicola* and characterization of the recombinant enzyme. J Biochem 135:533–540. https://doi.org/10.1093/jb/ mvh068.
- Zerbino DR, Birney E. 2008. Velvet: algorithms for *de novo* short read assembly using de Bruijn graphs. Genome Res 18:821–829. https://doi .org/10.1101/gr.074492.107.
- Kajitani R, Toshimoto K, Noguchi H, Toyoda A, Ogura Y, Okuno M, Yabana M, Harada M, Nagayasu E, Maruyama H, Kohara Y, Fujiyama A, Hayashi T, Itoh T. 2014. Efficient *de novo* assembly of highly heterozygous genomes from whole-genome shotgun short reads. Genome Res 24:1384–1395. https://doi.org/10.1101/gr.170720.113.
- Hoff KJ, Stanke M. 2013. WebAUGUSTUS—a Web service for training AUGUSTUS and predicting genes in eukaryotes. Nucleic Acids Res 41: W123–W128. https://doi.org/10.1093/nar/gkt418.
- Lowe TM, Eddy SR. 1997. tRNAscan-SE: a program for improved detection of transfer RNA genes in genomic sequence. Nucleic Acids Res 25: 955–964.
- Conesa A, Götz S, Garcia-Gomez JM, Terol J, Talon M, Robles M. 2005. Blast2GO: a universal tool for annotation, visualization and analysis in functional genomics research. Bioinformatics 21:3674–3676. https://doi .org/10.1093/bioinformatics/bti610.