

# Finished Genome Sequence of the Highly Multidrug-Resistant Human Urine Isolate *Citrobacter freundii* Strain SL151

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***Citrobacter freundii* is a Gram-negative opportunistic pathogen that is increasingly being recognized as a causative agent of hospital-acquired urinary tract infections and an important reservoir of antimicrobial resistance determinants. In this report, we describe the finished genome sequence of *C. freundii* strain SL151, a highly multidrug-resistant human urine isolate.**

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*Citrobacter freundii* is recognized as an emerging opportunistic pathogen and is known to cause a variety of ailments (e.g., urinary tract infections [UTIs], wound infections, gastrointestinal infections, septicemia, meningitis), especially in immunocompromised patients and in hospital settings (1–5). This emergence has coincided with the finding that *C. freundii* is often resistant to multiple classes of antibiotics, suggesting that both clinical and environmental strains may be important reservoirs of antimicrobial resistance determinants (ARDs) (6–10). For example, a recent survey of outpatients in Bo, Sierra Leone, revealed that a surprisingly high number of *C. freundii* were isolated from the urine of individuals with UTI symptoms and that all of these isolates were highly multidrug-resistant (MDR) (11). To determine the underlying genetics responsible for these observed MDR phenotypes, we sequenced the genome of a representative isolate from this collection, *C. freundii* strain SL151, using the Pacific Biosciences RS II sequencing platform (DNA Link USA, Inc., San Diego, CA, USA).

Genomic DNA was extracted using the Gentra Puregene yeast/bacteria Kit (Qiagen, Valencia, CA, USA) and used to prepare a 20-kb insert library that was sequenced using a single-molecule real-time (SMRT) sequencing cell and P6-C4 chemistry. This resulted in 7,148 filtered and preassembled sequence reads with a mean length of 17,444 bp and 23× genome coverage. Assembly and consensus polishing (via SMRT Analysis version 2.3.0 and HGAP.2) yielded one circular chromosome (5,073,255 bp, 51.7% GC) and two circular plasmids (210,673 bp, 54.7% GC, and 154,967 bp, 53.7% GC) with a finished genome size of 5,438,895 bp. Gene prediction and annotation were performed using GeneMarkS+ and the NCBI Prokaryotic Genome Annotation Pipeline, respectively, and identified 5,537 coding sequences, of which 1,127 were predicted to encode hypothetical proteins.

A standard BLAST query of the Comprehensive Antibiotic Resistance Database (CARD) (12) using the SL151 genome sequence resulted in the identification of 97 resistance-associated genes, of which 36 demonstrated ≥95% nucleotide identity to the CARD reference sequences and provided the underlying genotype for every observed resistance phenotype. ARDs found on the chro-

mosome include the intrinsic *C. freundii* AmpC cephalosporinase encoded by the *bla*<sub>CMY-79</sub> gene, a truncated copy of a *qnrB* gene, and a number of multidrug efflux pump-encoding genes. The larger plasmid contained the *aac(3)-III*, *bla*<sub>TEM-1</sub>, and *bla*<sub>CTX-M-15</sub> genes in a Tn2-like structure identical to *Klebsiella pneumoniae* pENVA (13), as well as the *qnrS1*, *sul2*, *catA1*, *strA*, *strB*, and *aadA16* genes. Finally, the smaller plasmid was found to contain an ~11-kb-long, ARD-rich, composite class 1 integron harboring *aac(6′)-Ib-cr*, *arr-3*, *dfrA27*, *aadA16*, and *qnrB6* gene cassettes. This composite element is nearly identical to structures found in *K. pneumoniae* and *K. oxytoca* plasmids (14, 15). Interestingly, a mutated duplication of this region was found adjacent to the fully functional copy. Also detected was an IS26-flanked composite transposon containing the *catA2* and *tetD* genes that is identical to a locus from the *Salmonella* sp. plasmid pTC67. Overall, this genome highlights the mosaic plasmid-mediated accumulation of ARDs in *C. freundii*, particularly those ARDs conferring resistance to aminoglycoside, fluoroquinolone, and sulfonamide compounds.

**Accession number(s).** This whole-genome project has been deposited at DDBJ/EMBL/GenBank under the accession numbers CP016952, CP017058, and CP017059.

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## REFERENCES

- Brenner DJ, Grimont PA, Steigerwalt AG, Fanning GR, Ageron E, Riddle CF. 1993. Classification of citrobacteria by DNA hybridization:

- designation of *Citrobacter farmeri* sp. nov., *Citrobacter youngae* sp. nov., *Citrobacter braakii* sp. nov., *Citrobacter werkmanii* sp. nov., *Citrobacter sedlakii* sp. nov., and three unnamed *Citrobacter* genomospecies. *Int J Syst Bacteriol* 43:645–658. <http://dx.doi.org/10.1099/00207713-43-4-645>.
2. Joaquin A, Khan S, Russel N, al Fayed N. 1991–1992. Neonatal meningitis and bilateral cerebellar abscesses due to *Citrobacter freundii*. *Pediatr Neurosurg* 17:23–24.
  3. Ranjan KP, Ranjan N. 2013. *Citrobacter*: an emerging health care associated urinary pathogen. *Urol Ann* 5:313–314.
  4. Gupta N, Yadav A, Choudhary U, Arora DR. 2003. *Citrobacter* bacteraemia in a tertiary care hospital. *Scand J Infect Dis* 35:765–768. <http://dx.doi.org/10.1080/00365540310016376>.
  5. Samonis G, Karageorgopoulos DE, Kofteridis DP, Matthaiou DK, Sidropoulou V, Maraki S, Falagas ME. 2009. *Citrobacter* infections in a general hospital: characteristics and outcomes. *Eur J Clin Microbiol Infect Dis* 28:61–68. <http://dx.doi.org/10.1007/s10096-008-0598-z>.
  6. Yim G, Kwong W, Davies J, Miao V. 2013. Complex integrons containing *qnrB4-ampC* (*bla*<sub>(DHA-1)</sub>) in plasmids of multidrug-resistant *Citrobacter freundii* from wastewater. *Can J Microbiol* 59:110–116. <http://dx.doi.org/10.1139/cjm-2012-0576>.
  7. Nada T, Baba H, Kawamura K, Ohkura T, Torii K, Ohta M. 2004. A small outbreak of third generation cephem-resistant *Citrobacter freundii* infection on a surgical ward. *Jpn J Infect Dis* 57:181–182.
  8. Pepperell C, Kus JV, Gardam MA, Humar A, Burrows LL. 2002. Low-virulence *Citrobacter* species encode resistance to multiple antimicrobials. *Antimicrob Agents Chemother* 46:3555–3560. <http://dx.doi.org/10.1128/AAC.46.11.3555-3560.2002>.
  9. Feng J, Qiu Y, Yin Z, Chen W, Yang H, Yang W, Wang J, Gao Y, Zhou D. 2015. Coexistence of a novel KPC-2-encoding MDR plasmid and an NDM-1-encoding pNDM-HN380-like plasmid in a clinical isolate of *Citrobacter freundii*. *J Antimicrob Chemother* 70:2987–2991. <http://dx.doi.org/10.1093/jac/dkv232>.
  10. Sheppard AE, Stoesser N, Wilson DJ, Sebra R, Kasarskis A, Anson LW, Giess A, Pankhurst LJ, Vaughan A, Grim CJ, Cox HL, Yeh AJ, Modernising Medical Microbiology (MMM) Informatics Group, Sifri CD, Walker AS, Peto TE, Crook DW, Mathers AJ. 2016. Nested Russian doll-like genetic mobility drives rapid dissemination of the carbapenem resistance gene *bla*<sub>KPC</sub>. *Antimicrob Agents Chemother* 60:3767–3778. <http://dx.doi.org/10.1128/AAC.00464-16>.
  11. Leski TA, Taitt CR, Bangura U, Stockelman MG, Ansumana R, Cooper WH III, Stenger DA, Vora GJ. 2016. High prevalence of multidrug resistant *Enterobacteriaceae* isolated from outpatient urine samples but not the hospital environment in Bo, Sierra Leone. *BMC Infect Dis* 16:167. <http://dx.doi.org/10.1186/s12879-016-1495-1>.
  12. McArthur AG, Waglechner N, Nizam F, Yan A, Azad MA, Baylay AJ, Bhullar K, Canova MJ, De Pascale G, Ejim L, Kalan L, King AM, Koteva K, Morar M, Mulvey MR, O'Brien JS, Pawlowski AC, Piddock LJ, Spanogiannopoulos P, Sutherland AD, Tang I, Taylor PL, Thaker M, Wang W, Yan M, Yu T, Wright GD. 2013. The Comprehensive Antibiotic Resistance Database. *Antimicrob Agents Chemother* 57:3348–3357. <http://dx.doi.org/10.1128/AAC.00419-13>.
  13. Schlüter A, Nordmann P, Bonnin RA, Millemann Y, Eikmeyer FG, Wibberg D, Pühler A, Poirer L. 2014. IncH-type plasmid harboring *bla*<sub>CTX-M-15</sub>, *bla*<sub>DHA-1</sub> and *qnrB4* genes recovered from animal isolates. *Antimicrob Agents Chemother* 58:3768–3773. <http://dx.doi.org/10.1128/AAC.02695-14>.
  14. Cheng C, Sun J, Zheng F, Lu W, Yang Q, Rui Y. 2016. New structures simultaneously harboring class 1 integron and ISCR1-linked resistance genes in multidrug-resistant gram-negative bacteria. *BMC Microbiol* 16:71. <http://dx.doi.org/10.1186/s12866-016-0683-x>.
  15. Ruiz E, Sáenz Y, Zarazaga M, Rocha-Gracia R, Martínez-Martínez L, Arlet G, Torres C. 2012. *Qnr*, *aac(6)-Ib-cr* and *qepA* genes in *Escherichia coli* and *Klebsiella* spp.: genetic environments and plasmid and chromosomal location. *J Antimicrob Chemother* 67:886–897. <http://dx.doi.org/10.1093/jac/dkr548>.