



Effects of Hydrated Potato Starch on the Quality of Low-fat Ttoekgalbi (Korean Traditional Patty) Packaged in Modified Atmosphere Conditions during Storage

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ABSTRACT: This study was carried out to investigate the effects of hydrated potato starch on the quality of low-fat ttoekgalbi (Korean traditional patty) packaged in modified atmosphere conditions during storage. The ttoekgalbi was prepared from 53.2% lean beef, 13.9% lean pork, 9.3% pork fat, and 23.6% other ingredients. Two low-fat ttoekgalbi treatments were prepared by substituting pork fat with hydrated potato starch; either by 50% fat replacement (50% FR) or 100% fat replacement (100% FR). Both 50% and 100% FR increased the moisture, crude protein, and decreased fat content, cooking loss, and hardness. For MAP studies, 200 g of ttoekgalbi were placed on the tray and filled with gas composed of 70% O₂: 30% CO₂ (70% O₂-MAP) and 30% CO₂: 70% N₂ (70% N₂-MAP), and were stored at 5°C for 12 d. During the storage time, both 50% and 100% FR showed higher protein deterioration, while no differences were found in CIE a*, CIE L*, lipid oxidation, and bacterial counts in comparison to control. The ttoekgalbi with 70% O₂-MAP was more red, lighter in color, and showed higher TBARS values compared with 70% N₂-MAP. The meat with 70% N₂-MAP showed lower aerobic bacterial counts in control than those with 70% O₂-MAP. The lower anaerobic bacterial counts were observed only in 50% FR and 100% FR packed with 70% N₂-MAP in comparison with 70% O₂-MAP. In conclusion, the fat replacement with hydrated potato starch showed no negative effects on the quality of low fat ttoekgalbi during storage and 70% N₂-MAP was better than 70% O₂-MAP for low-fat ttoekgalbi packaging. (**Key Words:** Low-fat Ttoekgalbi (Korean Traditional Patty), Hydrated Potato Starch, Modified Atmosphere Packaging (MAP))

INTRODUCTION

Ttoekgalbi is a Korean traditional patty, generally formulated with lean beef, lean pork and pork back fat as main ingredients. The use of pork back fat as the main ingredient increases its fat contents. It is well known that the high fat intake is associated with increased risk of some diseases like obesity, cancer, high blood cholesterol, and coronary heart diseases (Khalil, 2000). As the interest to healthy food increased, consumer demands for low-fat meat products also increased.

The major problem in acceptability of low-fat processed meat products is decline in palatability with fat reduction

(Mansour and Khalil, 1997). Water-binding capacity is a critical issue in low-fat meat production (Khalil, 2000), hence it is important to find ingredient that can increase water-holding capacity in low-fat products. It is well known that water-holding capacity is related to cooking loss, and cooking loss itself is an important quality parameter of meat products. Starch is known to have water-binding property and might be used as a fat replacer in meat products (Kim and Lee, 1987; Prabhu and Sebranek, 1997). Starches are glucose polymers derived from corn, potatoes, rice, two polymeric forms, amylose and amylopectin which create three-dimensional gel entrapping water are formed upon hydration (Keeton, 1994). Therefore, incorporation of starch may be beneficial in low-fat, high-added-water processed meat.

Packaging methods are important aspects to maintain the quality of meat products during storage. Modified atmosphere packaging (MAP) has been used by meat industry to extend the shelf-life as well as to keep the quality characteristics of meat products. The main gases used in MAP includes oxygen (O₂), carbon dioxide (CO₂)

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and nitrogen (N₂), and MAP is commonly divided into low oxygen and high oxygen MAP based on the oxygen content. High O₂ MAP preserves the bright red color of meat and increases shelf-life by suppressing the anaerobic microbial growth. High O₂ MAP may contain 25-90% O₂ and 15 to 80% CO₂ (Blakistone, 1999). Low oxygen MAP may contain less than 20% of O₂ and the rest is composed of CO₂ and N₂. Carbon dioxide is well known as an antibacterial gas in MAP, while N₂ is gas filler. Smith et al. (1990) demonstrated that 20 to 60% CO₂ in MAP retarded aerobic spoilage effectively. Low CO₂ concentration (20%) was better than high concentration (60%) in preservation of color and odor of fresh sausages (Martinez et al., 2005). Nitrogen gas is commonly use at 100% in MAP for cooked and cured meat products (Sebranek and Houser, 2006).

With the increasing concern to high fat intake, it is necessary to develop low-fat meat products as the suitable packaging methods for such products. Moreover, there is little information in the development of low-fat ttoekgalbi using hydrated potato starch as fat replacer as well as the usage of MAP as packaging method. The objective of this study was to evaluate the effect of hydrated potato starch on the quality of low-fat ttoekgalbi packaged in modified atmosphere conditions during storage.

MATERIALS AND METHODS

Sample preparation

Ttoekgalbi was prepared using the ingredients presented in Table 1. The lean pork and lean beef (obtained from local market) were chopped separately through a 3-mm plate using a grinder (DFG 450, Daehan Food Machine Co. Ltd., Korea). The chopped lean pork and lean beef were mixed well with other ingredients using a mixer (RM-20, Mainca, Spain). The low-fat ttoekgalbi was prepared by replacing the pork back fat with hydrated potato starch at 50% (50% FR) and 100% (100% FR). The potato starch was hydrated with water at starch:water = 2:1.5. For packaging studies, 200 g of ttoekgalbi were placed on barrier foam tray (maximum O₂ transmission rate = 0.1 cc/cm² at 23°C, 0% RH; maximum moisture vapor transmission rate = 2.0 g/24 h-254 cm² at 38°C, 100% RH, Cryovac sealed air corp., USA). The trays were sealed with O₂ barrier film (maximum O₂ transmission rate = 0.10 g/24 h-254 m² at 4,

Table 1. Ingredients of ttoekgalbi and low-fat ttoekgalbi formulated with potato starch

Ingredient (%)	Control	Fat replacement	
		50%	100%
Lean of pork	13.90	13.90	13.90
Lean of beef	53.20	53.20	53.20
Mixed spices	17.80	17.80	17.80
Salt	0.30	0.30	0.30
Phosphates	0.20	0.20	0.20
Organic acid salts ¹	0.40	0.40	0.40
Hardtact powder	3.90	3.90	3.90
Corn syrup	1.00	1.00	1.00
Pork fat	9.30	4.65	0.00
Hydrated potato starch ²	0.00	4.65	9.30
Total	100.00	100.00	100.00

¹ Sodium acetate:calcium lactate = 1:1. ² Potato starch:water = 2:1.5.

4°C, 100% RH; Lid 1050, Cryovac sealed air corp., USA). The trays were filled with modified atmosphere with 70% O₂: 30% CO₂ (70% O₂-MAP) or 30% CO₂: 70% N₂ (70% N₂-MAP) using modified atmosphere packaging machine (Hypervac, Korea) and the gas mixture MAP Mix 9001 ME (PBI Dansensor, Denmark). O₂, CO₂, N₂ gases were obtained from Baeklyung Specialty Gas Co., Korea, with purity 99.9%. The details of the experimental design are presented in Table 2.

The ttoekgalbi packs were stored in a refrigerator at 5°C for 12 d. Proximate analysis, sensory evaluation and physical characteristics analysis were performed to investigate the quality of ttoekgalbi at 0, 3, 6, and 12 d storage. The packs were opened in order to evaluate the storage quality of ttoekgalbi.

Proximate analysis

The proximate analysis including moisture, crude protein, crude fat and ash was performed by the Association of Official Analytical Chemists (AOAC) methods (1995).

Sensory evaluation

Panel consisting of 6 trained faculty members and students evaluated the color of raw ttoekgalbi, and the color, flavor, texture and taste of cooked ttoekgalbi. The visual color evaluation was conducted directly from the trays after

Table 2. Experimental design of low-fat ttoekgalbi formulated with hydrated potato starch and packaged in modified atmosphere condition

Meat treatment	Gas composition (O ₂ :CO ₂ :N ₂)	
	70% O ₂ :30% CO ₂ :0% N ₂ (70% O ₂ -MAP)	0% O ₂ :30% CO ₂ :70% N ₂ (70% N ₂ -MAP)
Control ¹	Cont+70% O ₂ -MAP	Cont+70% N ₂ -MAP
50% fat replacement (FR) ²	50% FR+70% O ₂ -MAP	50% FR+70% N ₂ -MAP
100% Fat replacement (FR) ³	100% FR+70% O ₂ -MAP	100% FR+70% N ₂ -MAP

¹ Pork fat = 9.3%, Hydrated potato starch = 0%. ² Pork fat = 4.65%, Hydrated potato starch = 4.65%. ³ Pork fat = 0%, Hydrated potato starch = 9.3%.

gas composition analysis. The panelist scored all parameters with scores ranging from 1 to 9 (9 = extremely like, 7 = like, 5 = moderate like, 3 = dislike, and 1 = extremely dislike).

Thiobarbituric acid reactive substances (TBARS) analysis

The TBARS value was measured according to Sinnhuber and Yu (1977). Briefly, 0.5 g sample was mixed with 3 drops of antioxidant solution (3% BHA-54% propylene glycol-3% BHT-40% Tween20), 3 ml of TBA solution (1% 4,6-Dihydroxy-2-mercaptopyrimidine), and 17 ml of 25% Trichloroacetic acid. The mixture was heated at 100°C for 30 min, and centrifuged at 3,500 rpm for 30 min. An absorbance of supernatant was measured at 532 nm using a spectrophotometer (UV-mini-1240, Shimadzu, Japan). The results were calculated as mg malonaldehyde (MA) per kg sample.

Color measurement

Color changes on the surface of ttoekgalbis were monitored by measuring the CIE lightness (L^*) and redness (a^*) using a color difference meter (CR-400, Konica Minolta Sensing Inc., Japan) and an illuminant C. The color instrument was calibrated using a white plate (Illuminant C: $Y = 93.6$, $x = 0.3134$, and $y = 0.3194$). Color measurements were directly performed on the surface of sample immediately after the packs were opened. The color of ground beef patties surface was measured 30 times in each pack at different locations.

Cooking loss and hardness measurement

Hardness analysis was carried out using a texture analyzer (TA-XT2i version 6.06, Stable Micro Systems Ltd., UK). Thirty two grams of ttoekgalbis were placed in petridish with 60 mm dia.×15 mm depth (SPL Life Science, Korea). The ttoekgalbi was placed in the zipper bag and boiled at 75°C for 45 min. The cooked ttoekgalbi was then cooled at 4°C for 12 h. The water on the cooked ttoekgalbi was removed using Whatman paper No. 1. The weight of cooked ttoekgalbi was measured and the cooking losses were determined. For hardness measurement, the samples were sliced in squares (1 cm×1 cm). The hardness was measured using a 5-mm cylindrical probe. The measurements were performed by three sample replications for every treatment and 10 hardness measurements were performed for every sample. Hardness is expressed as kgf.

pH determination

For pH determination, 10 g sample was added with 100 ml distilled water and then homogenized at 10,000 rpm for 60 s using a homogenizer (PH91, SMT Co. Ltd., Japan). The pH of the homogenized sample was measured using a pH meter (SevenEasy pH, Mettler-Toledo GmbH,

Switzerland). The measurements were performed by three sample replications for every treatment and pH determinations were performed in duplicates for every sample.

Volatile basic nitrogen (VBN) value analysis

The analysis of VBN was measured using the method of Kohsaka (1975). Five gram sample was homogenized with 30 ml of 5% (w/v) TCA using a homogenizer (Ultra-Turrax T25 basic, Ika Werke GmbH and Co., Germany) at 13,500 rpm for 2 min. The homogenate was made up to 50 ml of final volume with 5% (w/v) TCA and filtered through a Whatman No. 1 filter paper. One mL of the filtrate and 1 ml of borate buffer were placed in outer and inner of Conway dish, respectively, and then incubated at 37°C for 100 min. Finally, the inner solution was titrated with 0.01 N HCl.

Microbial analysis

Ten grams of sample was placed in an autoclaved bag (Nasco Co., Ltd., USA) and added with 90 ml of 0.1% (w/v) peptone solution. Each sample was homogenized using a Stomacher (Lab Blender 400, Seward Laboratory, UK) at medium speed for 2 min. Serial 10-fold dilutions were performed by mixing 1 ml of solution with 9 ml of 0.1% (w/v) peptone solution. Plate count agar (Difco, USA) was used for aerobic and anaerobic bacterial counts. Agar plates were prepared according to the manufacturer's instruction. Samples were incubated for 48 h at 35°C. Microbial populations were counted in 30 to 300 colonies and expressed as log CFU per gram of sample.

Statistical analysis

All data were analyzed using SPSS 14.0 for Windows Evaluation Version (2005). The data was analyzed by one way analysis of variance, with treatments (fat replacement and packaging method) as variable. Means of data in every day of storage were compared using Duncan's multiple range tests with examination for significant differences ($p < 0.05$).

RESULTS AND DISCUSSION

Quality properties of low-fat ttoekgalbi

The quality properties of low-fat ttoekgalbi that includes proximate composition, cooking loss, sensory evaluation and texture (hardness) are presented in Table 3. The replacement of pork fat using potato at 50% and 100% significantly increased ($p < 0.05$) the moisture contents of ttoekgalbi from 62.47% to 66.27% and 68.89%, respectively. This result is in agreement with Choi et al. (2010) who reported higher moisture contents in reduced-fat meat using rice bran fiber. Previous research reported that the fat replacement using pork rind increased the

Table 3. Characteristics and quality properties of low fat ttoekgalbi formulated with hydrated potato starch

Parameter	Fat replacement		
	0% (control)	50% FR	100% FR
Moisture	62.47±1.11 ^c	66.27±1.15 ^b	68.89±0.55 ^a
Crude ash	1.32±0.06 ^a	1.33±0.10 ^a	1.36±0.01 ^a
Crude protein	14.90±0.32 ^c	15.77±0.32 ^b	16.84±0.47 ^a
Crude fat	19.40±1.69 ^a	15.45±0.86 ^b	12.79±0.66 ^c
Cooking loss (%)	7.44±0.90 ^a	2.77±2.81 ^b	0.98±0.80 ^c
Sensory evaluation			
Raw-color	7.33±1.17 ^a	8.40±0.83 ^a	8.53±0.83 ^a
Cooked			
Color	7.88±0.88 ^a	7.56±0.63 ^a	7.50±0.73 ^a
Flavor	8.25±0.86 ^a	8.00±0.82 ^a	8.13±1.14 ^a
Texture	8.25±0.77 ^a	7.81±0.83 ^a	7.31±1.40 ^a
Taste	8.19±0.75 ^a	7.94±0.77 ^a	7.69±1.20 ^a
Hardness (kgf)	15.80±1.71 ^a	14.97±1.76 ^a	14.97±0.62 ^a

^{a-c} Values (Means±SD) with different superscripts in the same row differ significantly ($p < 0.05$), $n = 5$.

moisture contents of sausage (Abiola and Adegbaaju, 2001). The fat replacement using corn starch also increased the moisture contents of beef patties (Khalil, 2000). Higher moisture contents in low-fat ttoekgalbi may be related to the moisture binding ability of potato starch. Berry and Wergin (1993) reported high moisture binding capacity of potato starch and Troutt et al. (1992) reported moisture binding capacity of dietary fibers and starches.

The quality properties of low-fat ttoekgalbi that includes proximate composition, cooking loss, sensory evaluation and texture (hardness) are presented in Table 3. The replacement of pork fat using potato at 50% and 100% significantly increased ($p < 0.05$) the moisture contents of ttoekgalbi from 62.47% to 66.27% and 68.89%, respectively. This result is in agreement with Choi et al. (2010) who reported higher moisture contents in reduced-fat meat using rice brand fiber. Previous research reported that the fat replacement using pork rind increased the moisture contents of sausage (Abiola and Adegbaaju, 2001). The fat replacement using corn starch also increased the moisture contents of beef patties (Khalil, 2000). Higher moisture contents in low-fat ttoekgalbi may be related to the moisture binding ability of potato starch. Berry and Wergin (1993) reported high moisture binding capacity of potato starch and Troutt et al. (1992) reported moisture binding capacity of dietary fibers and starches.

The replacement of pork fat with potato starch at 50% and 100% significantly increased ($p < 0.05$) crude protein and decreased ($p < 0.05$) crude fat content in ttoekgalbi. Protein content in the ttoekgalbi increased from 14.90% to 15.77% and 16.84%, while fat content decreased from 19.40% to 15.45% and 12.79% respectively. The results

showed that the potato starch is effective in lowering fat contents in ttoekgalbi. Abiola and Adegbaaju (2001) reported decrease in fat content and increase in crude protein by replacing pork back fat with increasing level of pork rind in low-fat sausage. In addition, Osburn and Keeton (1994) manufactured low-fat pork sausages with konjac flour gel and concluded that all treated sausages were different than control, exhibiting higher percentage for moisture, protein and lower percentage of fat.

The cooking loss in low-fat ttoekgalbi was lower ($p < 0.05$) compared to control. Our result is in agreement with Park et al. (2005) and Choi et al. (2010), but different with Hughes et al. (1998). The lower cooking loss in low-fat ttoekgalbi may be attributed to higher protein and lower fat contents of low-fat ttoekgalbi. Cooking loss is affected by the amount of fat contents in meat products (Hong et al., 2004) and fat type (Choi et al., 2009) because protein has an ability to trap moisture and reduce moisture loss (Abiola and Adegbaaju, 2001).

There were no negative effects on sensory evaluation by the fat replacement with hydrated potato starch for low fat ttoekgalbi. The scores of taste, flavor, texture, and color of cooked low-fat ttoekgalbi, and color of uncooked low-fat ttoekgalbi were similar to those of control ttoekgalbi. Also, no differences were noticed in hardness of cooked low-fat ttoekgalbi. These are similar results in which no difference was found in flavor score of low fat ground beef formulated with potato starch (Khalil, 2000), and starch (Berry and Wergin, 1993). However, Hughes et al. (1997) and Abiola and Adegbaaju (2001) reported decrease in sensory properties of low-fat sausage formulated with pork rind. The fat replacement in low-fat ground beef decreased the hardness and that might be due to higher moisture content of the low-fat patties (Khalil, 2000).

Quality changes of ttoekgalbi during storage

The pH value: The pH values of low-fat ttoekgalbi formulated with hydrated potato starch and treated with MAP are presented in Table 4. On 0 d of storage, the pH of 50% FR and 100% FR was higher than control. Choi et al. (2010) reported the higher pH value of low-fat meat emulsion system formulated with vegetable oil and rice brand fiber. On 3 d and 6 d, the replacement of pork fat with hydrated potato starch decreased ($p < 0.05$) pH value compared to control, but the different percentage of replacement (50% and 100%) did not affect the pH value. The lower pH value might be related with the lower VBN observed in low-fat ttoekgalbi (Table 4). Ruiz-Capillas and Moral (2001) reported the positive correlation between pH and VBN value of hake stored in ice. At the end of storage, the pH value decreased suddenly. The possible reason of the decreased of pH might be related to the increased of bacterial counts (Table 7). Nassos et al. (1983) observed the

Table 4. Effect of hydrated potato starch on pH and VBN value (% mg sample) of low-fat ttoekgalbi packaged in modified atmosphere conditions during storage

	Storage day	Treatments					
		0% FR		50% FR		100% FR	
		70% O ₂ -MAP	70% N ₂ -MAP	70% O ₂ -MAP	70% N ₂ -MAP	70% O ₂ -MAP	70% N ₂ -MAP
pH	0	6.20±0.02 ^{bc}	6.20±0.02 ^{bc}	6.24±0.01 ^{aC}	6.24±0.01 ^{aA}	6.25±0.01 ^{aA}	6.25±0.01 ^{aB}
	3	6.38±0.03 ^{ab}	6.37±0.02 ^{ab}	6.29±0.01 ^{cBC}	6.32±0.02 ^{ba}	6.29±0.02 ^{cA}	6.29±0.01 ^{cAB}
	6	6.54±0.04 ^{aA}	6.46±0.01 ^{ba}	6.38±0.01 ^{cA}	6.36±0.01 ^{cA}	6.39±0.01 ^{cA}	6.37±0.01 ^{cA}
	9	6.58±0.09 ^{aA}	6.15±0.10 ^{cdC}	6.30±0.07 ^{bb}	6.08±0.05 ^{dA}	6.22±0.12 ^{bcA}	6.31±0.06 ^{baB}
	12	4.90±0.07 ^{aD}	4.70±0.15 ^{aD}	4.77±0.02 ^{aD}	4.68±0.02 ^{ab}	4.74±0.07 ^{ab}	4.72±0.11 ^{aC}
VBN	0	15.45±2.37 ^{aC}	15.45±2.37 ^{aC}	12.03±2.38 ^{bc}	12.02±2.38 ^{bc}	10.28±1.10 ^{bd}	10.28±1.10 ^{bd}
	3	17.75±2.13 ^{abC}	17.56±1.75 ^{ab}	13.54±2.50 ^{bcC}	14.96±2.51 ^{bb}	12.81±1.32 ^{bcC}	11.64±0.88 ^{cdC}
	6	20.22±1.27 ^{ab}	17.92±1.41 ^{bb}	17.22±1.58 ^{bb}	17.11±1.96 ^{baB}	13.26±1.34 ^{cC}	12.26±2.00 ^{cC}
	9	19.01±1.40 ^{ab}	17.35±3.71 ^{ab}	17.22±2.44 ^{ab}	18.55±0.78 ^{aA}	17.15±2.03 ^{ab}	16.44±1.26 ^{ab}
	12	22.15±3.00 ^{aA}	20.74±1.24 ^{aA}	20.71±2.84 ^{aA}	19.43±2.08 ^{aA}	20.76±1.80 ^{aA}	21.89±0.67 ^{aA}

^{a-c} Values (Means±SD) with different superscripts in the same row differ significantly (p<0.05).

^{A-E} Values (Means±SD) with different superscripts in the same column differ significantly (p<0.05), n = 6.

increased of pH value in ground beef because of the increase of bacterial counts, percentage of gram-positive bacteria and the present of lactic acid-producing bacteria. During the storage time, the different gas compositions in MAP showed no effect on pH value in both control and low-fat ttoekgalbi. This result is similar trend to our earlier study (Muhlisin et al., 2010) that showed no difference in pH value of ground beef packed with 70% O₂-MAP and 70% N₂-MAP, while ground beef added with organic acid and packed with 70% O₂-MAP showed higher pH values than 70% N₂-MAP. In addition, Viana et al. (2005) also reported that MAP did not show strong variation in pH of fresh pork loin.

Volatile basic nitrogen (VBN) value: The protein deterioration (represented by VBN value) of low-fat ttoekgalbi formulated with hydrated potato starch treated with MAP is presented in Table 4. The VBN value of low fat ttoekgalbi was significantly lower (p<0.05) than control on 0 d to 6 d storage. On 3 and 6 d storage, the VBN value was decreased in number as the increasing of percentage of fat replacement. Protein deterioration is associated with the

activity of amino acid decarboxylase of microorganisms (Lin and Lin, 2002). The VBN values increased in all the treatments with the increasing storage time. Arashihar et al. (2004) reported that VBN values increased with the increasing storage time in air packaging, vacuum and MAP of rainbow trout (*Oncorhynchus mykiss*) fillets. MAP did not show any effect on VBN values in all the treatment during the storage time. In control ttoekgalbi, 70% N₂-MAP resulted in lower VBN value but statistically it was not different (p>0.05).

Thiobarbituric acid reactive substance (TBARS) value: The TBARS values in low-fat ttoekgalbi (both in 50% and 100% replacement) were lower compared to the control until 3 d of storage (Table 5). This may be due to the lower fat contents in low fat ttoekgalbi compared to control. The 70% O₂-MAP increased (p<0.05) TBARS value both in control and low-fat ttoekgalbi during the storage compared to 70% N₂-MAP. Similarly, Lund et al. (2007) showed that 100% N₂ MAP significantly lower TBARS value with or without organic acid salts in beef patties. Skibsted et al. (1998) mentioned one of the factors affecting lipid

Table 5. Effect of hydrated potato starch on TBARS value (mg MA/kg sample) of low-fat ttoekgalbi packaged in modified atmosphere conditions during storage

Day	Treatment					
	0% FR		50% FR		100% FR	
	70% O ₂ -MAP	70% N ₂ -MAP	70% O ₂ -MAP	70% N ₂ -MAP	70% O ₂ -MAP	70% N ₂ -MAP
0	0.59±0.10 ^{aC}	0.59±0.10 ^{aC}	0.56±0.10 ^{aC}	0.56±0.10 ^{aB}	0.53±0.11 ^{aC}	0.53±0.11 ^{aB}
3	0.95±0.06 ^{ab}	0.80±0.21 ^{abB}	0.84±0.05 ^{abB}	0.72±0.19 ^{bcA}	0.64±0.20 ^{cC}	0.62±0.14 ^{cB}
6	0.96±0.05 ^{ab}	0.82±0.08 ^{cb}	0.93±0.04 ^{abB}	0.82±0.13 ^{ca}	0.96±0.10 ^{aA}	0.87±0.09 ^{abcA}
9	1.20±0.22 ^{aA}	0.97±0.05 ^{ba}	1.23±0.16 ^{aA}	0.79±0.18 ^{dA}	0.94±0.11 ^{bcA}	0.83±0.06 ^{cdA}
12	0.99±0.20 ^{ab}	0.66±0.08 ^{cdC}	0.82±0.19 ^{bb}	0.52±0.10 ^{dB}	0.78±0.17 ^{bcB}	0.57±0.13 ^{dC}

^{a-c} Values (Means±SD) with different superscripts in the same row differ significantly (p<0.05).

^{A-E} Values (Means±SD) with different superscripts in the same column differ significantly (p<0.05), n = 12.

Table 6. Effect of hydrated potato starch on instrumental color value of low-fat ttoekgalbi packaged in modified atmosphere conditions during storage

Storage day	Treatments						
	0% FR		50% FR		100% FR		
	70% O ₂ -MAP	70% N ₂ -MAP	70% O ₂ -MAP	70% N ₂ -MAP	70% O ₂ -MAP	70% N ₂ -MAP	
CIE L* ¹	0	53.17±1.89 ^{bb}	53.17±1.89 ^{ba}	54.39±2.32 ^{aa}	54.39±2.32 ^{aa}	54.28±2.10 ^{ab}	54.28±2.01 ^{aa}
	3	53.91±1.93 ^{ab}	53.41±2.17 ^{aa}	53.42±2.16 ^{ab}	53.58±2.16 ^{aa}	53.60±2.32 ^{ab}	53.35±2.01 ^{aa}
	6	53.34±2.91 ^{bb}	53.38±2.39 ^{ba}	54.81±2.64 ^{aa}	53.36±2.25 ^{aa}	54.79±2.11 ^{ab}	53.74±2.09 ^{aa}
	9	54.24±2.91 ^{abc}	53.99±2.31 ^{cd}	54.96±2.35 ^{aa}	53.97±2.59 ^{cd}	54.60±2.22 ^{ab}	53.44±2.86 ^{da}
	12	54.68±2.89 ^{aa}	52.68±2.48 ^{ba}	55.03±2.52 ^{aa}	53.27±2.06 ^{ba}	55.23±2.49 ^{aa}	53.32±2.27 ^{ba}
CIE a* ²	0	10.57±0.93 ^{ba}	10.57±0.93 ^{bc}	11.66±1.10 ^{aa}	11.66±1.10 ^{aa}	10.37±1.65 ^{ba}	10.37±1.65 ^{bb}
	3	7.92±1.02 ^{db}	10.99±1.54 ^{ac}	8.60±1.44 ^{cb}	9.60±1.17 ^{bd}	8.19±1.38 ^{db}	9.92±1.14 ^{bc}
	6	6.86±1.11 ^{ec}	11.62±1.45 ^{ab}	7.68±1.35 ^{dc}	10.40±1.54 ^{bc}	7.79±1.28 ^{dc}	9.78±1.42 ^{cc}
	9	6.95±1.03 ^{dc}	12.50±1.66 ^{aa}	7.28±1.20 ^{cd}	11.21±1.56 ^{bb}	7.42±1.10 ^{cc}	11.32±1.26 ^{ba}
	12	4.45±1.04 ^{dd}	10.98±1.48 ^{ac}	3.40±0.81 ^{ee}	9.50±1.19 ^{cd}	3.71±1.06 ^{ed}	10.35±1.49 ^{bb}

^{a-c} Values (Means±SD) with different superscripts in the same row differ significantly (p<0.05).

^{A-E} Values (Means±SD) with different superscripts in the same column differ significantly (p<0.05), n = 90.

¹ Lightness. ² Redness.

oxidation is oxygen concentration in packaging. These results are also in agreement with those of Cayuela et al. (2004); John et al. (2005); and McMillin (2008) who showed that high oxygen packaging resulted in higher lipid oxidation, compared with vacuum packaging or low oxygen packaging in meat. These results indicated that lipid oxidation of ttoekgalbi could be inhibited by FR with hydrated potato starch and 70% N₂-MAP.

Instrumental color: Fat replacement with hydrated potato starch resulted in lighter (p<0.05) ttoekgalbi compared to control, while different percent of fat replacement (50% and 100%) gave no effect on CIE L* value (Table 6). On 0 d, low-fat ttoekgalbi with 50% fat replacement showed more red (p<0.05) color compared to

control ttoekgalbi and low-fat ttoekgalbi with 100% fat replacement, while no difference was found in redness between control and low-fat ttoekgalbi with 100% fat replacement. Papadima and Bloukas (1999) found that fat level affected the lightness and redness of low-fat traditional Greek sausages, but Garson et al. (2003) noted that no difference were found in CIE L* and CIE a* in low-fat beef patties formulated with carbohydrate-lipid composite. Choi et al. (2010) reported that increasing grape seed oil levels from 0% to 15% as fat replacement significantly increased the lightness of uncooked and cooked batters. Hoffman and Mellet (2003) reported that the patties containing fat were all lighter than that with containing the fat replacer (higher CIE L* values) in low-fat

Table 7. Effect of hydrated potato starch on aerobic and anaerobic bacterial counts (Log CFU/g sample) of low-fat ttoekgalbi packaged in modified atmosphere conditions during storage

Storage day	Treatments						
	0% FR		50% FR		100% FR		
	70% O ₂ -MAP	70% N ₂ -MAP	70% O ₂ -MAP	70% N ₂ -MAP	70% O ₂ -MAP	70% N ₂ -MAP	
AE ¹	0	5.37±0.06 ^{ac}	5.37±0.06 ^{ac}	5.35±0.12 ^{ac}	5.35±0.12 ^{ac}	5.31±0.15 ^{ac}	5.31±0.15 ^{ac}
	3	5.39±0.09 ^{ac}	5.41±0.13 ^{ac}	5.38±0.15 ^{ac}	5.48±0.28 ^{ac}	5.53±0.37 ^{ac}	5.47±0.26 ^{ac}
	6	5.38±0.00 ^{ac}	5.47±0.23 ^{ac}	5.43±0.12 ^{ac}	5.33±0.10 ^{ac}	5.59±0.14 ^{ac}	5.64±0.23 ^{ac}
	9	6.60±0.31 ^{ab}	6.62±0.11 ^{ab}	6.95±0.49 ^{ab}	6.24±0.34 ^{bb}	6.84±0.27 ^{ab}	6.64±0.39 ^{ab}
	12	8.52±0.05 ^{aa}	8.46±0.16 ^{aa}	8.54±0.03 ^{aa}	7.90±0.30 ^{ba}	8.51±0.12 ^{aa}	8.41±0.18 ^{aa}
AN ²	0	5.47±0.04 ^{ac}	5.47±0.04 ^{ac}	5.41±0.00 ^{ac}	5.41±0.00 ^{ac}	5.44±0.24 ^{ac}	5.44±0.24 ^{ac}
	3	5.45±0.14 ^{abc}	5.62±0.21 ^{ac}	5.24±0.23 ^{bc}	5.56±0.24 ^{abc}	5.60±0.27 ^{ac}	5.60±0.24 ^{ac}
	6	5.37±0.16 ^{abc}	5.76±0.20 ^{ac}	5.52±0.29 ^{abc}	5.33±0.16 ^{cc}	5.61±0.13 ^{abc}	5.69±0.21 ^{abc}
	9	6.64±0.38 ^{bc}	6.78±0.21 ^{abc}	7.06±0.44 ^{ab}	6.41±0.44 ^{cb}	7.17±0.31 ^{ab}	6.89±0.36 ^{abc}
	12	8.50±0.09 ^{aa}	8.49±0.01 ^{aa}	8.38±0.27 ^{ab}	8.23±0.05 ^{ba}	8.56±0.07 ^{aa}	8.50±0.08 ^{aa}

^{a-c} Values (Means±SD) with different superscripts in the same row differ significantly (p<0.05).

^{A-E} Values (Means±SD) with different superscripts in the same column differ significantly (p<0.05), n = 6.

¹ Aerobic bacteria. ² Anaerobic bacteria.

ostrich but no fixed trend was found in redness.

With MAP treatments, a trend was found in CIE L* in which ttoekgalbi packed with 70% N₂-MAP tends to be darker (lower CIE L*) than ttoekgalbi packed with 70% O₂-MAP from 3 d to 12 d of storage. Our earlier studies (Muhlisin et al., 2010) showed that CIE L* value in 70% N₂-MAP was lower than in that 70% O₂-MAP in Hanwoo ground beef. In contrast, Suman et al. (2010) noticed that difference in gas composition did not affect the CIE L*. In redness (CIE a*), N₂-MAP resulted in more red color compare to 70% O₂-MAP from 3 to 12 d of storage in all treatments. Control ttoekgalbi packed with 70% N₂-MAP showed the highest CIE a* value among the treatments. The treatments with 70% N₂-MAP seemed to be more effective in preserving the red color of ttoekgalbi compared to 70% O₂-MAP.

Microbiology: No differences were found in the bacterial counts of both aerobic and anaerobic bacteria (Table 7). This result implied that fat replacement in low-fat ttoekgalbi had no effect on bacterial counts during storage. Mugerza et al. (2002) reported that fat level had no effects on lactic acid bacterial counts of low fat fermented sausages. Different gas composition in MAP did not affect the aerobic and anaerobic bacterial counts in control ttoekgalbi and low-fat ttoekgalbi with 100% fat replacement. The effects of 70% N₂-MAP was observed on low-fat ttoekgalbi with 50% fat replacement, in which N₂-MAP lowered the aerobic bacterial counts on 9 to 12 d storage, and anaerobic bacterial counts on 6 to 12 d of storage. Similar bacterial counts in 70% O₂-MAP and 70% N₂-MAP may be due to the similar percentage of CO₂ (both contained 30% CO₂). CO₂ is utilized in MAP gas composition because of its antibacterial effects. In addition, Smith et al. (1990) suggested that 20-60% of CO₂ in MAP is required to detain the growth of bacteria.

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