



Genome Sequence of *Litorilinea aerophila*, an Icelandic Intertidal Hot Springs Bacterium

Elizabeth G. Maurais,^a Lauren C. Iannazzi,^b ⁽ⁱ⁾Kyle S. MacLea^{a,b,c,d}

^aBiotechnology Undergraduate Program, University of New Hampshire, Manchester, New Hampshire, USA ^bBiology Undergraduate Program, University of New Hampshire, Manchester, New Hampshire, USA ^cGraduate Program in Biotechnology: Industrial and Biomedical Sciences, University of New Hampshire, Manchester, New Hampshire, USA ^dDepartment of Life Sciences, University of New Hampshire, Manchester, New Hampshire, USA

ABSTRACT The hot springs bacterium *Litorilinea aerophila* PRI-4131^T (= ATCC BAA-2444^T) was found in Isafjardardjup, in northwest Iceland. In this paper, we present a draft genome sequence for the type strain, with a total predicted genome length of 6,043,010 bp, 4,608 protein-coding sequences, 54 RNAs, 9 CRISPR arrays, and a G+C content of 64.61%.

The bacterial phylum *Chloroflexi* (1–3) (also *Chlorobacteria* or *Chloroflexota*) is a deepbranching bacterial phylum with significant metabolic diversity, from green non-sulfur photosynthesizers to anaerobic halogen metabolizers to aerobic chemoorganotrophs (4–9). Thermophilic growth is common. The *Chloroflexi* are unusual in that cells stain as Gram negative but possess a single cell wall layer (i.e., they are monoderms) with no evidence of an outer membrane, which is characteristic of most other Gram-negative bacteria (2, 3, 10–12).

Within the *Chloroflexi* class *Caldilineae*, most organisms are anaerobes, but two of these filamentous thermophilic bacteria, including *Litorilinea aerophila*, have shown aerobic growth (4, 9). In this work, we describe a draft genome sequence for the type strain of *Litorilinea aerophila*, PRI-4131 (= DSM 25763 = ATCC BAA-2444). *Litorilinea aerophila* was isolated from an intertidal hot spring (0.6% NaCl) in Iceland (9) but has since been found in other environments, including in the human cervicovaginal microbiota (13), in waste treatment and disposal sites/systems (14–16), in plant cultivation systems (17), and in mines (18).

Lyophilized Litorilinea aerophila ATCC BAA-2444^T was purchased from ATCC (Manassas, VA, USA), resuspended in marine broth 2216 (BD, Franklin Lakes, NJ, USA), and incubated at 50°C for 5 days at 1 atm. It was then subcultured on marine agar (5 days at 50°C), from which a single colony was inoculated into 2 mL marine broth. After growth at 50°C to log phase, genomic DNA (gDNA) was purified using the QIAamp DNA minikit (Qiagen, Valencia, CA, USA). Fragmentation of gDNA and adapter attachment were performed using the KAPA HyperPlus kit v.3.16 (KR1145; Kapa Biosystems, Wilmington, MA, USA). An Illumina HiSeq 2500 instrument (Hubbard Center for Genome Studies, Durham, NH, USA) was used for paired-end 250-bp fragment sequencing. Reads were trimmed using Trimmomatic v.0.38 (settings: paired-end mode with a window size of 4, quality requirement of 15, and minimum read length of 36), and then 1,522,708 trimmed reads were assembled with SPAdes v.3.13.0 (19, 20) with default bacterial assembly parameters. Small contigs (<500 bp) and contigs with low coverage ($<10\times$) were removed. QUAST (21) analysis of this assembly showed 93 contigs (the largest one being 341,248 bp), with an N_{50} value of 180,513 bp. Benchmarking universal single-copy orthologs (BUSCO) v.5.2.2 analysis (with default parameters) showed that the assembly was 95.2% complete (22), and genome coverage of $112 \times$ was calculated. The NCBI Prokaryotic Genome Assembly Pipeline (PGAP) (23) identified and annotated genes in the L. aerophila genome. The assembled genome was 6,043,010 bp in length, and PGAP revealed a total of 4,749 genes, 4,608 protein-coding sequences, 87 pseudogenes, 46 tRNAs, 5 partial or complete copies of the rRNA genes, including 1 complete

Editor Frank J. Stewart, Montana State University

Copyright © 2022 Maurais et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International license.

Address correspondence to Kyle S. MacLea, kyle.maclea@unh.edu.

The authors declare no conflict of interest.

Received 16 December 2021 Accepted 7 January 2022 Published 27 January 2022 copy of each, 3 noncoding RNAs, and a G+C content of 64.61%, close to the published value of 64.7% for the species (9). Nine CRISPR arrays were identified, as well as the CRISPR associated genes encoding Cas1 to Cas3 and Cas5e (24). As predicted based on analysis of the *Chloroflexi* (11, 12), *Litorilinea* lacks Gram-negative lipopolysaccharide (LPS) and lipid A metabolic genes such as *lpxC* and also possesses teichoic acid and lipoteichoic acid transport and synthesis genes (24), which are characteristic of monoderms.

Data availability. The *Litorilinea aerophila* ATCC BAA-2444^T whole-genome shotgun sequencing (WGS) project has been deposited in DDBJ/ENA/GenBank under accession number VIGC00000000. The raw Illumina data from BioProject PRJNA551245 were submitted to the NCBI Sequence Read Archive (SRA) under accession number SRX6432641.

ACKNOWLEDGMENTS

K.S.M. acknowledges the contributions of Edna Spurr MacLea (1920 to 2005) to the preliminary stages of this study.

Sequencing and bioinformatic analysis were undertaken at the Hubbard Center for Genome Studies at the University of New Hampshire, supported by New Hampshire-INBRE, with the assistance of Kelley Thomas and Stephen Simpson. This work was a project of the Microbiology Education through Genome Annotation-New Hampshire (MEGA-NH) program. This work was funded by the Department of Life Sciences at the University of New Hampshire, by a Manchester Undergraduate Project Support grant to E.G.M., and by New Hampshire-INBRE through an Institutional Development Award (IDeA) (grant P20GM103506) from the National Institute of General Medical Sciences of the NIH. The funders had no role in study design, data collection and interpretation, or the decision to submit the work for publication.

REFERENCES

- Garrity GM, Holt JG, Castenholz RW, Pierson BK, Keppen OI, Gorlenko VM. 2001. Phylum BVI: *Chloroflexi* phy. nov., p 427–446. *In* Boone DR, Castenholz RW, Garrity GM (ed), Bergey's manual of systematic bacteriology, vol 1. The *Archaea* and the deeply branching and phototrophic *Bacteria*. Springer, New York, NY.
- Cavalier-Smith T. 2006. Rooting the tree of life by transition analyses. Biol Direct 1:19. https://doi.org/10.1186/1745-6150-1-19.
- Cavalier-Smith T, Chao EE-Y. 2020. Multidomain ribosomal protein trees and the planctobacterial origin of neomura (eukaryotes, archaebacteria). Protoplasma 257:621–753. https://doi.org/10.1007/s00709-019-01442-7.
- Sekiguchi Y, Yamada T, Hanada S, Ohashi A, Harada H, Kamagata Y. 2003. Anaerolinea thermophila gen. nov., sp. nov. and Caldilinea aerophila gen. nov., sp. nov., novel filamentous thermophiles that represent a previously uncultured lineage of the domain Bacteria at the subphylum level. Int J Syst Evol Microbiol 53:1843–1851. https://doi.org/10.1099/ijs .0.02699-0.
- Yamada T, Sekiguchi Y, Hanada S, Imachi H, Ohashi A, Harada H, Kamagata Y. 2006. Anaerolinea thermolimosa sp. nov., Levilinea saccharolytica gen. nov., sp. nov. and Leptolinea tardivitalis gen. nov., sp. nov., novel filamentous anaerobes, and description of the new classes Anaerolineae classis nov. and Caldilineae classis nov. in the bacterial phylum Chloroflexi. Int J Syst Evol Microbiol 56: 1331–1340. https://doi.org/10.1099/ijs.0.64169-0.
- Cavalier-Smith T. 2006. Cell evolution and Earth history: stasis and revolution. Philos Trans R Soc Lond B Biol Sci 361:969–1006. https://doi.org/10 .1098/rstb.2006.1842.
- Grégoire P, Bohli M, Cayol J-L, Joseph M, Guasco S, Dubourg K, Cambar J, Michotey V, Bonin P, Fardeau M-L, Ollivier B. 2011. *Caldilinea tarbellica* sp. nov., a filamentous, thermophilic, anaerobic bacterium isolated from a deep hot aquifer in the Aquitaine Basin. Int J Syst Evol Microbiol 61: 1436–1441. https://doi.org/10.1099/ijs.0.025676-0.
- Löffler FE, Yan J, Ritalahti KM, Adrian L, Edwards EA, Konstantinidis KT, Müller JA, Fullerton H, Zinder SH, Spormann AM. 2013. *Dehalococcoides mccartyi* gen. nov., sp. nov., obligately organohalide-respiring anaerobic bacteria relevant to halogen cycling and bioremediation, belong to a novel bacterial class, *Dehalococcoida classis* nov., order *Dehalococcoidales* ord. nov. and family *Dehalococcoidaceae* fam. nov., within the phylum *Chloroflexi*. Int J Syst Evol Microbiol 63: 625–635. https://doi.org/10.1099/ijs.0.034926-0.
- Kale V, Björnsdóttir SH, Friðjónsson ÓH, Pétursdóttir SK, Ómarsdóttir S, Hreggviðsson GÓ. 2013. Litorilinea aerophila gen. nov., sp. nov., an aerobic

member of the class *Caldilineae*, phylum *Chloroflexi*, isolated from an intertidal hot spring. Int J Syst Evol Microbiol 63:1149–1154. https://doi.org/10.1099/ijs.0 .044115-0.

- Sutcliffe IC. 2010. A phylum level perspective on bacterial cell envelope architecture. Trends Microbiol 18:464–470. https://doi.org/10.1016/j.tim.2010.06.005.
- 11. Sutcliffe IC. 2011. Cell envelope architecture in the *Chloroflexi*: a shifting frontline in a phylogenetic turf war. Environ Microbiol 13:279–282. https://doi.org/10.1111/j.1462-2920.2010.02339.x.
- 12. Errington J. 2013. L-form bacteria, cell walls and the origins of life. Open Biol 3:120143. https://doi.org/10.1098/rsob.120143.
- Huang X, Li C, Li F, Zhao J, Wan X, Wang K. 2018. Cervicovaginal microbiota composition correlates with the acquisition of high-risk human papillomavirus types. Int J Cancer 143:621–634. https://doi.org/10.1002/ijc.31342.
- Wei ZS, He YM, Huang ZS, Xiao XL, Li BL, Ming S, Cheng XL. 2019. Photocatalytic membrane combined with biodegradation for toluene oxidation. Ecotoxicol Environ Saf 184:109618. https://doi.org/10.1016/j.ecoenv .2019.109618.
- Su C, Lin X, Zheng P, Chen Y, Zhao L, Liao Y, Liu J. 2019. Effect of cephalexin after heterogeneous Fenton-like pretreatment on the performance of anaerobic granular sludge and activated sludge. Chemosphere 235: 84–95. https://doi.org/10.1016/j.chemosphere.2019.06.136.
- Lv X, Ma B, Lee K, Ulrich A. 2020. Potential syntrophic associations in anaerobic naphthenic acids biodegrading consortia inferred with microbial interactome networks. J Hazard Mater 397:122678. https://doi.org/10.1016/ j.jhazmat.2020.122678.
- Grunert O, Hernandez-Sanabria E, Buysens S, De Neve S, Van Labeke M-C, Reheul D, Boon N. 2020. In-depth observation on the microbial and fungal community structure of four contrasting tomato cultivation systems in soil based and soilless culture systems. Front Plant Sci 11:520834. https://doi.org/10.3389/fpls.2020.520834.
- Taleski V, Dimkić I, Boev B, Boev I, Živković S, Stanković S. 2020. Bacterial and fungal diversity in the lorandite (TIAsS2) mine 'Allchar' in the Republic of North Macedonia. FEMS Microbiol Ecol 96:fiaa155. https://doi.org/ 10.1093/femsec/fiaa155.
- Bolger AM, Lohse M, Usadel B. 2014. Trimmomatic: a flexible trimmer for Illumina sequence data. Bioinformatics 30:2114–2120. https://doi.org/10 .1093/bioinformatics/btu170.
- 20. Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV,

Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. J Comput Biol 19:455–477. https://doi.org/10.1089/cmb.2012.0021.

- Gurevich A, Saveliev V, Vyahhi N, Tesler G. 2013. QUAST: quality assessment tool for genome assemblies. Bioinformatics 29:1072–1075. https://doi.org/10.1093/bioinformatics/btt086.
- Simão FA, Waterhouse RM, Ioannidis P, Kriventseva EV, Zdobnov EM. 2015. BUSCO: assessing genome assembly and annotation completeness with single-copy orthologs. Bioinformatics 31:3210–3212. https://doi.org/ 10.1093/bioinformatics/btv351.

- Tatusova T, DiCuccio M, Badretdin A, Chetvernin V, Ciufo S, Li W. 2013. Prokaryotic Genome Annotation Pipeline. *In* The NCBI handbook, 2nd ed. National Center for Biotechnology Information, Bethesda, MD. https://www .ncbi.nlm.nih.gov/books/NBK174280.
- Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O. 2008. The RAST Server: Rapid Annotations using Subsystems Technology. BMC Genomics 9:75. https://doi.org/10 .1186/1471-2164-9-75.