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Carrier Frequency of Spinal Muscular Atrophy in a Large-scale Korean Population

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Spinal muscular atrophy (SMA) is an autosomal recessive disease characterized by progressive proximal muscle weakness and atrophy. Given the recent introduction of gene therapies, knowledge of the SMA carrier frequency in various populations has become important for developing screening programs for this disease. In total, 1,581 anonymous DNA samples from an umbilical cord blood bank were tested for *SMN1* and *SMN2* gene copies using a multiplex ligation-dependent probe amplification assay. Twenty-nine of the 1,581 newborns [1.83%; 95% confidence interval (CI), 1.25–2.66%] were SMA carriers with one copy of *SMN1*, and no homozygous *SMN1* deletion was detected. The carrier frequency in this population was estimated to be 1,834 per 100,000 (95% CI, 1,254– 2,659) or 1 in 55 (95% CI, 1/79-1/38). Our data indicate that SMA carriers are not uncommon in the Korean population and may serve as a reference for designing a population screening program in Korea. Received: August 16, 2019 Revision received: September 29, 2019 Accepted: January 16, 2020

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Spinal muscular atrophy (SMA) is an autosomal recessive neurodegenerative disease characterized by progressive proximal muscle weakness and atrophy resulting from degeneration of the alpha motor neurons of the anterior horn of the spinal cord. SMA is phenotypically heterogeneous and is classified into four clinical subtypes based on the age of onset and motor function achieved [1]. Type I SMA presents marked weakness and developmental motor regression before the age of six months and usually results in death within two years. Type II SMA presents before the age of 18 months and is associated with difficulty in walking independently. Type III SMA presents after the age of 18 months and individuals walk independently with only supportive care and is seldom fatal. Type IV SMA presents in the second or third decade of life, with only minor muscle weak-



ness. The incidence of all types of SMA is estimated to be 1 in 10,000 live births globally [2].

SMA is caused by variants in *SMN1* located on chromosome 5q13.2. Ninety-five percent of SMA patients have homozygous *SMN1* deletions, whereas the remaining 5% have compound heterozygous *SMN1* deletion and point variant. Most patients inherit variants from their parents; only approximately 2% of the affected patients have *de-novo* variants [1]. Although SMA patients lack a functional copy of *SMN1*, the nearly identical gene *SMN2*, located in the centromeric region of the same chromosome, produces a less functional protein through alternative splicing. The number of *SMN2* copies varies within the general population and is inversely associated with disease severity [1].

In December 2016, the US Food and Drug Administration approved nusinersen (SPINRAZA[™], Biogen, Cambridge, MA, USA), the first effective treatment for SMA (www.fda.gov/News-Events/Newsroom/PressAnnouncements/ucm534611.htm). Nusinersen is a modified antisense oligonucleotide that binds to a specific sequence, including exon 7 of SMN2 pre-messenger RNA, and increases the production of full-length SMN proteins. Nusinersen treatment in pre-symptomatic infants meaningfully increased the likelihood of survival and improved motor function [3]. Therefore, it is important to screen for SMA, carrier frequency of SMA, and SMN2 copy number at an early stage. Methods for SMA screening in newborns or adults have been actively studied and analyzed for cost-effectiveness [4, 5]. Although the SMN1 heterozygous deletion carrier frequency reportedly is between 2.00% (95% confidence interval [CI], 0.35–7.74%) and 2.15% (95% CI, 0.94–4.57%) in the Korean population [6, 7], no largescale population studies have been performed. Therefore, we conducted a large-scale study to estimate the SMA carrier frequency in Korean newborns. Using the multiplex ligation-dependent probe amplification (MLPA) assay, we assessed the distribution of SMN1 and SMN2 copy numbers in umbilical cord blood samples from the newborns.

In total, 1,581 anonymous DNA samples obtained from a public umbilical cord blood bank (Seoul Metropolitan Government Cord Blood Bank, Seoul, Korea) were used. This study protocol was exempted from review by the Institutional Review Board (IRB) of Seoul National University Boramae Hospital, Seoul, Korea (07-2019-17) because the samples have been kept as a repository study under the approval of IRB. All samples were collected from mothers who provided informed consent for donation and research use of their cord blood. The MLPA analysis has been conducted since July 2019.

The MLPA assay was conducted using SALSA MLPA probe-

mix P460-A1 SMA kits (MRC-Holland, Amsterdam, Netherlands), according to the manufacturer's instructions (https://www.mlpa.com/WebForms/WebFormDBData.aspx?FileOID =_ImdLw-Ti-aZY.). The PCR products were analyzed using an ABI PRISM 3100 ABI Genetic Analyzer (Applied Biosystems, Foster City, CA, USA). The data were analyzed using GeneMarker software, version 2.6 (Softgenetics, State College, PA, USA). Gene frequencies were calculated with VassarStats (http://vassarstats.net/), with 95% CI. The Hardy-Weinberg equilibrium was assessed by a professional web-based program (https://wpcalc.com/en/equilibrium-hardy-weinberg/).

The distribution of *SMN1* and *SMN2* copy numbers in the population is shown in Fig. 1. Of the 1,581 newborns, 29 (1.83%; 95% CI, 1.25–2.66%) were SMA carriers. Therefore, the carrier frequency in this population was estimated to be 1,834 per 100,000 (95% CI, 1,254–2,659) or 1 in 55 (95% CI, 1/79–1/38). Homozygous *SMN1* deletion was not detected. One newborn was suspected as an *SMN1* silent (2+0) carrier with two *SMN1* copies on one chromosome 5 and deletion on the other [8]. We also assessed *SMN2* copy numbers in this study population: 63.1% (N=998) had two copies of *SMN2*, 29.8% (N=471) had 0 or one copy, and 7.1% (N=112) had three or more copies. Among the SMA carriers, 24.1% (N=7) had 0 or one copy of *SMN2*, 58.6% (N=17) had two copies, and 17.2% (N=5) had three or more copies.

In this study, the SMN1 heterozygous deletion carrier frequency

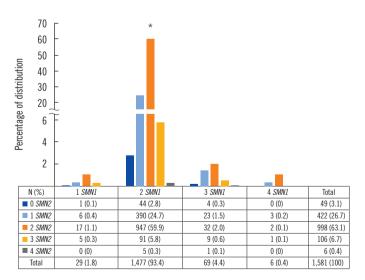


Fig. 1. Distribution of *SMN1* and *SMN2* copy numbers. Each bar represents the percentage of *SMN1* and *SMN2* copy numbers in the 1,581 umbilical cord blood samples of newborns included in this study. *One newborn was suspected as a *SMN1* silent (2+0) carrier.

Total samples (N)	Carrier frequency % (95% CI)	Method	Reference
569	1.58 (0.77–3.09)	Real-time PCR	Chan, <i>et al.</i> 2004 [10]
1,712	2.39 (1.75–3.27)	DHPLC	Sheng-Yuan, <i>et al.</i> 2010 [11]
1,741	2.58 (1.91–3.47)	Real-time PCR	Qu, <i>et al.</i> 2013 [12]
4,719	1.90 (1.54–2.35)	DHPLC	Gong, <i>et al.</i> 2013 [13]
4,931	1.24 (0.96–1.60)	DHPLC and multiplex PCR	Tan, <i>et al.</i> 2018 [14]
310	1.94 (0.79–4.37)	MLPA	Huang, <i>et al.</i> 2007 [15]
163	2.50 (0.79–6.56)	Multiplex PCR and CE	Chen, <i>et al.</i> 2011 [16]
107,611	2.10 (2.03–2.20)	DHPLC and multiplex PCR	Su, <i>et al.</i> 2011 [17]
453	3.97 (2.44–6.33)	HRMA, DHPLC, and MLPA	Wang, et al. 2015 [18]
326	2.15 (0.94–4.57)	Real-time PCR	Lee, <i>et al.</i> 2004 [7]
100	2.00 (0.35–7.74)	MLPA	Yoon, <i>et al.</i> 2010 [6]
1,581	1.83 (1.25–2.66)	MLPA	This study
	569 1,712 1,741 4,719 4,931 310 163 107,611 453 326 100	569 1.58 (0.77–3.09) 1,712 2.39 (1.75–3.27) 1,741 2.58 (1.91–3.47) 4,719 1.90 (1.54–2.35) 4,931 1.24 (0.96–1.60) 310 1.94 (0.79–4.37) 163 2.50 (0.79–6.56) 107,611 2.10 (2.03–2.20) 453 3.97 (2.44–6.33) 326 2.15 (0.94–4.57) 100 2.00 (0.35–7.74)	569 1.58 (0.77–3.09) Real-time PCR 1,712 2.39 (1.75–3.27) DHPLC 1,741 2.58 (1.91–3.47) Real-time PCR 4,719 1.90 (1.54–2.35) DHPLC 4,931 1.24 (0.96–1.60) DHPLC and multiplex PCR 310 1.94 (0.79–4.37) MLPA 163 2.50 (0.79–6.56) Multiplex PCR and CE 107,611 2.10 (2.03–2.20) DHPLC and multiplex PCR 453 3.97 (2.44–6.33) HRMA, DHPLC, and MLPA 326 2.15 (0.94–4.57) Real-time PCR 100 2.00 (0.35–7.74) MLPA

Abbreviations: CI, confidence interval; CE, capillary electrophoresis; DHPLC, denaturing high-performance liquid chromatography; HRMA, high-resolution DNA melting analysis; MLPA, multiplex ligation-dependent probe amplification.

was 1.83–1.90% (including silent carriers), which is similar to or lower than the frequencies reported in the previous Korean studies [6, 7]; however, in these studies, the CI was wide because of small sample numbers. Given the narrower CI in the current study, the SMA carrier frequency found here might be more accurate. Similar to findings in a previous study, the carrier frequency in our study was higher than that reported for other ethnicities, except Caucasians [9]. However, it was similar to or lower than that reported for the East Asian population (Table 1) [10–18]. The incidence of SMA caused by *SMN1* homozygous deletion calculated using the Hardy–Weinberg equation was 1/11,815 in Koreans.

According to data from the Korean Statistical Information Service (http://kosis.kr/; accessed on 17 May 2019), in 2017, the total population of South Korea was 51.42 million, with 357,771 births. Based on the carrier frequency in this study, the number of carriers was estimated to be 0.94 million in total, and 6,547 in newborns per year. Thus, the estimated incidence rate of SMA due to *SMN1* deletion in Korea based on Hardy–Weinberg equilibrium is 30 cases per year.

With the development of drugs for the treatment of SMA, the need for newborn or carrier screening for the early diagnosis of SMA has become important. The US Secretary of Health and Human Services' Advisory Committee on Heritable Disorders in Newborns and Children recommends the addition of SMA screening to default newborn screening programs (https://www.hrsa. gov/advisory-committees/heritable-disorders/rusp/index.html), and the American College of Medical Genetics and the American College of Obstetricians and Gynecologists recommend routine SMA carrier screening [19, 20]. Cost-effectiveness analyses of SMA screening have been reported. Little, *et al.* [4] reported that although universal prenatal screening for SMA is not costeffective, screening of high-risk populations, such as those with a family history, may be. Zhang, *et al.* [5] reported that while SMA-only preconception carrier screening is not cost-effective, a combination screen for other conditions, such as cystic fibrosis and fragile X syndrome may prove to be and may prevent up to 30 SMA patients per million (all adults between the ages of 18 and 25 in the Australian health care system). Since the approval of nusinersen, there has been a lack of cost-effective analyses for newborn or carrier screening of SMA. Therefore, large-scale prospective newborn or carrier screening studies are needed for predicting the efficacy of nusinersen treatment.

Our study had some limitations. First, while 5% of SMA patients are known to have *SMN1* missense variant, the MLPA assay is not a suitable method for detecting missense variants. Second, we experienced difficulties in identifying silent carrier phenotypes (e.g., *SMN1* '2+0' genotype) due to limitations of the MLPA method. Therefore, the SMA carrier frequency might be underestimated. This study also had various advantages. This is the first large-scale population study on the SMA carrier frequency in Koreans conducted using samples from a public umbilical cord blood bank. Further, because the CI used herein was narrower than that reported in previous studies [6, 7], the SMA carrier frequency and incidence predicted here might be more accurate. Finally, we evaluated the distribution of *SMN2*



copy number in the general Korean population, which is important for predicting both the prognosis of patients with SMA and the efficacy of nusinersen treatment.

Our data indicate that carriers of *SMN1* heterozygous deletion are not uncommon in the general Korean population. The carrier frequency was similar to or lower than that in the East Asian population and higher than that in other ethnicities. To our knowledge, this is the largest cohort study of *SMN1* and *SMN2* copy numbers in the Korean population to date, and our data are expected to serve as a reference for further investigation of SMA in the Korean population.

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AUTHOR CONTRIBUTIONS

CSK and SS conceived and designed the study. EYR and JHY performed umbilical cord blood data acquisition. Multiplex ligation-dependent probe amplification was performed by SAY and analyzed by JEP. JEP wrote the paper. All authors read and approved the final manuscript.

CONFLICTS OF INTEREST

No potential conflicts of interest relevant to this article were reported.

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