

Article

# Radioprotective Effects on Late Third-Instar Bactrocera dorsalis (Diptera: Tephritidae) Larvae in Low-Oxygen Atmospheres



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**Simple Summary:** The oriental fruit fly *Bactrocera dorsalis* Hendel is a highly invasive fruit fly that causes extensive damage to many fruits and vegetables. Irradiation treatment is an economically effective and promising treatment measure. However, treatment efficacy is affected by the presence of low oxygen, i.e., mangoes are treated in modified atmosphere package. In order to investigate the reduced (radioprotective) effects on insects and determine its critical O<sub>2</sub> level, third-instar *B. dorsalis* larvae were irradiated by X-rays at the doses of 8 to 64 Gy with intervals of 8 Gy. The treatments were conducted under ambient air or low-oxygen atomospheres (0%, 2%, 4%, 6%, 8% O<sub>2</sub> and nitrogen). No adult emergence from treatments at 64 Gy in pure nitrogen or 56 Gy under other atmospheres, resulted in significant difference in tolerance. The results from statistical analyses indicate that differences in tolerance to radiation were significant in 0% and 2% O<sub>2</sub> but insignificant in 4%, 6%, and 8% O<sub>2</sub> environments when compared with radiation in ambient air. Therefore, the critical threshold is an O<sub>2</sub> level of  $\geq$ 4% and <6%, but a maximum radiation dose of 14 Gy can compensate for the radioprotective effects when the oriental fruit fly is treated in low-oxygen atmospheres.

Abstract: Ionizing radiation creates free radicals, the effect of which is enhanced by the presence of oxygen; a low oxygen level produces radioprotective effects for insects compared with irradiation in ambient air. Modified (controlled) atmosphere packaging is used for maintaining quality and shelf-life extension; therefore, treatment efficacy may be affected, and there is a need to determine the critical O<sub>2</sub> levels that may cause radioprotective effects. Late third-instar *Bactrocera dorsalis* (Hendel) larvae were irradiated in bags filled with ambient or low-oxygen air (0%, 2%, 4%, 6%, 8% O<sub>2</sub>) and were exposed to radiation doses of 8 to 64 Gy with intervals of 8 Gy. Efficacy was measured by the prevention of adult emergence. Dose–response data on mortality (failure of adult emergence) were analyzed via two-way ANOVA (analysis of variance), ANCOVA (analysis of covariance), and probit regression. The difference in radiotolerance was only significant in 0% O<sub>2</sub> atmospheres through two-way ANOVA; therefore, the 95% confidence limits (CLs) of lethal dose ratios at LD<sub>99</sub> were used to determine significant differences between treatments at different O<sub>2</sub> levels. The differences in radiotolerance were significant in 0% and 2% O<sub>2</sub> but insignificant in 4%, 6%, and 8% O<sub>2</sub> environments when compared with radiation in ambient air. The critical threshold of radioprotective effects for late third-instar *B. dorsalis* larvae is an O<sub>2</sub> level of  $\geq$ 4% and



<6%, but a maximum radiation dose of 14 Gy can compensate for this effect during phytosanitary irradiation treatment.

Keywords: Bactrocera dorsalis; radioprotective effects; critical threshold; low-oxygen radiation

## 1. Introduction

The oriental fruit fly, *Bactrocera dorsalis* Hendel (Diptera: Tephritidae), is a highly invasive species with over 300 known commercial/edible and wild hosts. It is currently found in at least 65 countries, including most of Asia, much of the sub-Saharan countries of Africa, parts of America, Oceania, and Europe, but it is of quarantine significance to many other countries [1–3]. In China, the oriental fruit fly is one of the most destructive quarantine insects of tropical and subtropical fruits and vegetables, causing severe losses to citrus, guava, carambola, and mango [4,5]. Infested commodities are normally required to undergo phytosanitary treatments before export to regulated or quarantine areas [6].

Phytosanitary measures, including fumigation, temperature, and irradiation treatments, are currently used for shipped commodities, and the use of phytosanitary irradiation (PI) treatment has increased in recent years due to its advantages over other treatments; as a result, the global volume of different fresh products was approximately 30,000 tons in 2017 [3,4,7–9]. In order to develop treatment schedules and facilitate the application of PI treatment, many kinds of tephritid fruit fly third-instar larvae have been used for conducting radiotolerance and confirmatory studies, such as the oriental fruit fly, *B. dorsalis* [4,10–12]; the Mediterranean fruit fly, *Ceratitis capitata* Wiedemann [10,12]; the melon fly, *Zeugodacus cucurbitae* Coquillett [10,13]; and *Z. tau* Walker [14]. Currently, the International Plant Protection Convention (IPPC) is discussing draft standards related to irradiation treatment for *B. dorsalis*, *Z. tau*, and the genus *Anastrepha* to formulate international standards, as an annex to International Standard for Phytosanitary Measures (ISPM) No. 28 [15].

Fresh commodities are always allowed to be irradiated with insect-proof packing, so as to protect them from infestation, re-infestation, or contamination afterwards. Modified (controlled) atmosphere packaging (MAP) is widely used to improve the shelf life of fresh fruits by decreasing the  $O_2$  level (below 8%) and/or elevating the  $CO_2$  level (above 10%) [16–18]. Treatment under an  $O_2$  level lower than that in MAP is used as a single or combined phytosanitary treatment measure, and a draft ISPM (Draft ISPM 2014-006: Requirements for the use of modified atmosphere treatments as phytosanitary measures) is under discussion for adoption [15,19,20]. Ionizing radiation creates free radicals, the effect of which is enhanced by the presence of oxygen; thus, high oxygen tension can enhance the effects of radiation. Conversely, a low oxygen level produces radioprotective effects for insects, so radiation in low-oxygen environments is considered to be the most significant factor affecting the efficacy of PI treatment, aside from the radiation dose itself [21,22].

Although reduced effects of radiation have been validated by several studies that conducted radiation treatment in <3-5% O<sub>2</sub> atmospheres, most of them performed comparisons between ambient air and pure nitrogen or MAP, whereas the difference in radiation effects was insignificant [23]. In addition, the PI treatment efficacy was not reduced when fruit flies were treated under hypoxia [12]. Therefore, more research is required to determine the critical O<sub>2</sub> levels that may cause radioprotective effects for insects and at which point the efficacy of the recommended doses for commercial PI treatment may decrease when conducting treatments in low-oxygen environments [21–23]. Thus, the objective of this research was to investigate the critical O<sub>2</sub> level (threshold value) that may induce radioprotective effects for late third-instar *B. dorsalis* larvae and to estimate the radiation dose that compensates for this reduced efficacy.

## 2. Materials and Methods

## 2.1. Insect Rearing

The insects used in this research were originally collected in a mango orchard in Chongzuo city, Guangxi Zhuang Autonomous Region, China in July 2018, and then reared in the Laboratory of Phytosanitary Treatment and Equipment, Chinese Academy of Inspection and Quarantine in Beijing, China. Late third-instar *B. dorsalis* larvae that emerged from mango fruits were collected and transferred to plastic boxes containing moist sterile sand for pupation. About 10 days later, the pupae were placed in the rearing cages ( $40 \times 40 \times 50$  cm) for adult emergence; the adults were fed with sterile water, fresh fruit pulp, and a solid mixture of sucrose and hydrolyzed yeast (3:1). Eggs were collected by placing papaya peel domes (emptied of fruit pulp) in the cages, and larvae were reared on artificial diets, the components of which were described by Liu et al. [24]. The rearing room was controlled at a temperature of 24–26 °C with 50–70% relative humidity and a photoperiod of 14:10 h (L/D). The emerged late third-instar larvae (about 8 days old) in the third to sixth generations were used for conducting all the radiation treatments.

#### 2.2. Handling of Larvae

Two-liter gastight air bags, purchased from Dalian Delin Gas Packaging Co., Ltd. Dalian, China, were used for conducting radiation treatments in hypoxia or ambient air. For each treatment, about 100 late third-instar larvae were wrapped in a small net bag and placed into an air bag through its opening, followed by sealing of the bag, exhausting all the air with a one-liter air-tight syringe, and injecting pure nitrogen (<0.001%  $O_2$ ) or controlled atmosphere (CA) (2%, 4%, 6%, or 8%  $O_2$  and nitrogen) (Beijing Green Oxygen Tiangang Technology Development Co., Ltd. Beijing, China) into the bag and holding it for one minute. The procedure was repeated three times to purify the gas in the air bag; the bags containing larvae and a volume of 0.7–0.8 L CA were subjected to irradiation treatment 15 min later. For irradiation treatment in ambient air, the larvae were wrapped and placed in the air bags without any additional procedure.

## 2.3. Radiation Treatments

All the radiation treatments were conducted in an RS-2000 Pro X-ray irradiator (Rad Source Technologies, Inc., Coral Springs, FL, USA) with operating parameters of 220 KV and 17.6 mA. In order to get a good dose uniformity ratio (1.05), the irradiator was equipped with a reflector placed in the bottom of the exposure chamber (width, 17 inch; depth, 15 inch; height, 17 inch) [25]. The bag containing larvae was placed in the exposure chamber to be irradiated at doses of 0 (control) to 64 Gy with intervals of 8 Gy. For each dose treatment, three bags (as replicates) were placed in the exposure chamber and irradiated at the same time. To measure and control the applied radiation doses, a RadCal dosimeter (model 2086, RadCal Corp., Monrovia, CA, USA) with a 10 × 6-0.6 Ion Chamber was placed near the insects. After each radiation treatment run, the cumulative dose was recorded from the dosimeter, and the irradiator was stopped when the cumulative dose was reached. The dose rate monitored in the study was about 5.0 Gy/min.

After treatment, all the bags were opened when the total sealing time reached about one hour; then, the larvae were removed and placed in sterile moist sand for pupation. Three weeks later, the numbers of late third-instar larvae, pupae, and adults were counted.

#### 2.4. Data Analyses

Dose–response data on mortality (failure of adult emergence) were adjusted by using Abbott's formula [26] and then subjected to two-way ANOVA (analysis of variance) to analyze the individual effects of radiation dose and PO<sub>2</sub> (O<sub>2</sub> level: 0%, 2%, 4%, 6%, 8%, and 21%) and the interaction effects of dose  $\times$  PO<sub>2</sub>; means were compared by Tukey's multiple comparison tests [27]. Linear regression after analysis of covariance (ANCOVA) was also used to analyze the mortality data to compare the

radiotolerance of *B. dorsalis* treated in different  $O_2$  atmospheres; all the data less than 100% and the lowest dose causing 100% mortality were used in the analyses [13,27]. All the data derived from treatments below 56 Gy and from the control were subjected to probit analyses using the PoloPlus 2.0 program to estimate the minimum dose for preventing adult emergence at each environmental  $O_2$  level via probit and logit models (using the non-transformed dose) [28]. In addition, to compare the significance of radiotolerance between treatments at different  $O_2$  levels, pairwise comparison tests were performed by calculating the 95% confidence limits (CLs) of the lethal dose ratios at LD<sub>99</sub> (the minimum lethal dose leading to 99% mortality at a specific confidence level, i.e., 95%) [29]. If the 95% CL excludes 1, then the LD<sub>99</sub> values are significantly different [28,30].

# 3. Results

## 3.1. Effects of Radiation Dose and Oxygen Level

The percent mortality of late third-instar *B. dorsalis* larvae treated under low oxygen and ambient air were found to be significantly affected by radiation doses ( $F_{7,143} = 1089.0$ , p < 0.0001), PO<sub>2</sub> ( $F_{5,143} = 4.6$ , p = 0.0026), and dose × PO<sub>2</sub> interactions ( $F_{35,143} = 1.7$ , p = 0.0255) when the dose–response data were analyzed by two-way ANOVA (Table 1). With increasing radiation dose, adult emergence declined across all PO<sub>2</sub> treatment groups, and no insects successfully emerged as adults when treated at 64 Gy in pure nitrogen (<0.001% O<sub>2</sub>) or at 56 Gy in other environments (2%, 4%, 6%, 8%, and 21% O<sub>2</sub>). For treatments in pure nitrogen, the mean mortality at all doses (65.0 ± 31.6%), as well as that at 56 Gy (95.5 ± 2.3%), was significantly lower than those under other treatments, suggesting that the radiation effect is decreased significantly under extreme hypoxia. Meanwhile, other statistical methods were needed to compare the significance in radiotolerance between treatments in ambient air and low-oxygen atmospheres.

O <sub>2</sub> (%)	$\%$ Mortality at the Specified Radiation Dose (Mean $\pm$ SD)									
	0 Gy	8 Gy	16 Gy	24 Gy	32 Gy	40 Gy	48 Gy	56 Gy	64 Gy	
0	6.8 ± 2.1a *	$10.4 \pm 2.6a$	32.3 ± 5.1a	$41.3 \pm 8.6a$	70.5 ± 1.6b	83.2 ± 1.1a	86.5 ± 1.4a	95.5 ± 2.3b	$100.0 \pm 0a$	
2	$8.0 \pm 1.8a$	$12.2 \pm 1.9a$	$36.9 \pm 8.0a$	$43.5 \pm 6.3a$	$78.5 \pm 1.8a$	$85.5 \pm 1.4a$	$87.6 \pm 7.2a$	$100.0 \pm 0a$	$100.0 \pm 0.0$	
4	7.7 ± 3.2a	$13.2 \pm 4.9a$	$31.0 \pm 3.4a$	$52.3 \pm 2.4a$	$78.1 \pm 1.3a$	$86.9 \pm 2.1a$	$91.1 \pm 1.8a$	$100.0 \pm 0a$	$100.0 \pm 0$	
6	$7.0 \pm 0.8a$	$11.9 \pm 4.3a$	$32.1 \pm 1.2a$	$55.5 \pm 4.8a$	$78.5 \pm 3.1a$	$88.5 \pm 3.6a$	$91.4 \pm 0.8a$	$100.0 \pm 0a$	$100.0 \pm 0.0$	
8	$7.9 \pm 0.4a$	$10.9 \pm 2.1a$	$36.0 \pm 6.6a$	$53.8 \pm 4.5a$	76.9 ± 3.8ab	$87.2 \pm 0.3a$	93.5 ± 2.9a	$100.0 \pm 0a$	$100.0 \pm 0$	
21	$7.9 \pm 1.6a$	$12.0 \pm 1.9a$	$31.5 \pm 1.1a$	$54.8 \pm 1.7a$	$77.2 \pm 1.7a$	$87.3 \pm 2.2a$	$92.8 \pm 2.7a$	$100.0 \pm 0a$	$100.0 \pm 0$	

**Table 1.** Radiation effects on the prevention of adult emergence as a result of irradiating late third-instar *Bactrocera dorsalis* larvae with X-rays (0–64 Gy) under ambient air and low-oxygen atmospheres.

\* Within a column, means followed by the same letter are not significantly different at p < 0.05, using the Tukey test.

#### 3.2. Estimating Doses for the Prevention of Adult Emergence

## 3.2.1. Linear Regression

The results obtained from ANCOVA and linear regression (Table 2) showed that all the coefficient of determination ( $R^2$ ) values were larger than 0.95 (maximum of 0.985), which implies that the regression fit the data well. As the interaction effect between dose and PO<sub>2</sub> was significant ( $F_{5,114} = 2.77$ , p = 0.0212), the minimum dose for 100% mortality was predicted by liner regression to compare the relative radiotolerance among treatments in a low-oxygen atmosphere and in ambient air. The estimated dose leading to 100% mortality decreased gradually with the increasing O<sub>2</sub> level from 0% to 4%, and then remained essentially unchanged (Table 2, Figure 1), suggesting that an O<sub>2</sub> level of 4% is likely to be the critical threshold of radioprotective effects for late third-instar *B. dorsalis* larvae. Furthermore, compared with that in ambient air, the minimum radiation doses for 100% mortality increased from 57.6 (in ambient air) to 58.3 (1.2%), 60.3 (4.7%), and 65.8 Gy (14.2%) in 4%, 2%, and 0% O<sub>2</sub> atmospheres, respectively.

O <sub>2</sub> (%)	Slope ± SE	$Intercept \pm SE$	$R^2$	Estimated Dose for 100% Mortality (Gy)
0	$1.454 \pm 0.047$	$-5.640 \pm 1.912$	0.9772	65.8
2	$1.618\pm0.091$	$-7.568 \pm 3.313$	0.9517	60.3
4	$1.658 \pm 0.059$	$-6.668 \pm 2.094$	0.9769	58.3
6	$1.665 \pm 0.061$	$-6.194 \pm 2.198$	0.9748	57.8
8	$1.679\pm0.048$	$-6.505 \pm 1.728$	0.9845	57.5
21	$1.683\pm0.047$	$-6.970 \pm 1.673$	0.9855	57.6

**Table 2.** Linear regressions of data on the prevention of adult emergence from late third-instar

 *Bactrocera dorsalis* larvae irradiated with X-rays under ambient air and low-oxygen atmospheres.



**Figure 1.** The estimated radiation dose for 100% mortality, LD<sub>99</sub>, and LD<sub>99,9968</sub> to prevent adult emergence from late third-instar *Bactrocera dorsalis* larvae irradiated in ambient air and 0%, 2%, 4%, 6%, and 8% O<sub>2</sub> environments.

## 3.2.2. Probit Analyses

Probit analyses were used to analyze the dose-mortality data of late third-instar B. dorsalis larvae. The estimated values of LD<sub>99</sub> and LD<sub>99.9968</sub> (the minimum lethal dose achieving a mortality of 99.9968% at a specific confidence level, i.e., 95%) analyzed by the probit and logit model were very close; thus, only the parameters derived from the probit model, such as slope, intercept, heterogeneity, estimated lethal doses, and their 95% CLs, are listed in Table 3. A small heterogeneity factor and a narrow LD<sub>99</sub> confidence interval clearly showed that the analysis fit the data well. The slopes increased but the estimated values of LD<sub>99</sub> and LD<sub>99,9968</sub> decreased sharply with increasing the O<sub>2</sub> level from 0% to 4% and were almost unchanged until 21% (Table 3, Figure 1). Similarly, the mortality curves obtained from PoloPlus software (Figure 2) can be divided into three groups, in which curve B ( $2\% O_2$ ) is above curve A (0% O<sub>2</sub>) but below the merged curve clusters C (4%, 6%, 8%, and 21% O<sub>2</sub>). Furthermore, the 95% CLs of the lethal dose ratios at  $LD_{99}$  were pairwise compared between treatments at all  $O_2$ levels; the results (Table 3) showed that the radiotolerance of *B. dorsalis* irradiated in the 0% and 2% O<sub>2</sub> environments was significantly greater than that of others, but it was insignificant when irradiated in 4%, 6%, or 8% O<sub>2</sub> or in ambient air. Similar tendencies were obtained by comparing the estimated values of LD<sub>99.9968</sub>, though they were extrapolative values (Table 3, Figure 1). Thus, the sequence of radiotolerance is suggested as follows:  $0\% > 2\% > 4\% \approx 6\% \approx 8\% \approx 21\% O_2$  atmospheres.

Compared with that in ambient air, the  $LD_{99.9968}$  value was increased by 13.9 (17.2%), 8.0 (10.0%), and 1.8 Gy (2.2%) when larvae were irradiated in 0%, 2%, and 4% O<sub>2</sub> atmospheres, respectively. Therefore, the additional radiation dose that might compensate for the radioprotective effects is less than 13.9 Gy (nominally 14 Gy) when the oriental fruit fly is treated in low-oxygen atmospheres.

O <sub>2</sub> (%)	No. Treated	$Slope \pm SE$	Intercept $\pm$ SE	Estimated Lethal E	Hetero-	
02 (78)				LD99	LD <sub>99.9968</sub>	Geneity
0	2155	$0.058 \pm 0.003$	$-1.486 \pm 0.09$	65.6 (61.7–70.5)a	94.5 (87.7–103.0)a	1.61
2	2217	$0.063 \pm 0.003$	$-1.540 \pm 0.099$	61.7 (56.6–68.6)a	88.4 (79.7-100.5)ab	3.13
4	2289	$0.068 \pm 0.003$	$-1.557 \pm 0.096$	57.5 (54.4-61.2)b	82.2 (76.9-88.8)b	1.44
6	2335	$0.069 \pm 0.003$	$-1.553 \pm 0.092$	56.5 (53.3-60.5)b	80.9 (75.3-87.9)b	1.76
8	2142	$0.069 \pm 0.003$	$-1.579 \pm 0.100$	56.2 (53.3-59.8)b	80.4 (75.2-86.7)b	1.32
21	2103	$0.069 \pm 0.003$	$-1.558 \pm 0.100$	56.4 (53.9-59.3)b	80.6 (76.3-85.6)b	0.99

**Table 3.** Probit analysis on the prevention of adult emergence from late third-instar *Bactrocera dorsalis* larvae irradiated with X-rays in ambient air and low-oxygen atmospheres.

\* Within each column, values followed by different letters were significantly different based on lethal dose ratio tests (p < 0.05).



**Figure 2.** The estimated mortality curves derived from probit analyses of the dose–mortality data when late third-instar *Bactrocera dorsalis* larvae were irradiated in ambient air and in 0%, 2%, 4%, 6%, and 8% O<sub>2</sub> environments. (A: 0% O<sub>2</sub>; B: 2% O<sub>2</sub>; C: 4%, 6%, 8%, and 21% O<sub>2</sub>).

## 4. Discussion

Radiotolerance in insects develops with their age and developmental time, so the most developed stage is the most radiotolerant when a common measure of efficacy is used [21]. The late third instar has been defined as the most tolerant stage of *B. dorsalis* larvae, as well as of other tephritid fruit flies, and was selected as the target stage for conducting dose–response tests in our research [4,10-14]. The third-instar larvae were irradiated in a series of O2 levels with the same dose to compare the relative tolerance between treatments, and the prevention of adult emergence was selected as the common efficacy criterions [4,9,11,31,32]. Radiation tolerance in insects is modified by the oxygen level and increases in low-oxygen atmospheres; in particular, insects irradiated in very low oxygen  $(<1\% O_2)$  have repeatedly been shown to have an increased radioprotective response [10,23]. In our study, an increased radiotolerance of *B. dorsalis* in low-oxygen environments (0%, 2%, and 4%) O<sub>2</sub>) was confirmed by comparing mortality rates using two-way ANOVA (Table 1), the estimated minimum doses for 100% mortality by linear regression after ANCOVA (Table 2, Figure 1), and LD<sub>99</sub> by probit analysis (Table 3, Figures 1 and 2). The results are concordant with those from other radiation treatments in low-oxygen atmospheres against the Caribbean fruit fly, Anastrepha suspensa Loew [33]; C. capitata [34]; Z. cucurbitae [13]; Drosophila suzukii Matsumura [35]; the Oriental fruit moth, Grapholita molesta Busck [36]; the European corn borer, Ostrinia nubilalis Hübner [37]; and the cabbage looper moth, Trichoplusia ni Hübner [22].

Compared to those for radiation in ambient air, the minimum dose values for 100% mortality, LD<sub>99</sub>, and LD<sub>99,9968</sub> estimated in pure nitrogen were increased by 8.2 (14.2%), 9.2 (16.2%), and 13.9 Gy (17.2%), respectively (Tables 2 and 3). The estimated LD<sub>99</sub> in ambient air in our research (56.4 (53.9–59.3) Gy)

(Table 3) is very close to that reported by Srimartpirom et al. (58.11 (53.63–64.46) Gy), indicating that the experiments should be replicated and statistically analyzed to ensure that data are verifiable and reproducible [11,38]. The estimated LD<sub>99,9968</sub> values can be used as the minimum dose for PI treatment if they are validated by large-scale confirmatory tests [32,38], and our LD<sub>99,9968</sub> data (80.6 (76.3–85.6) Gy) (Table 3) are very close to the values (84.1 (73.6–99.3) Gy) that were estimated and validated by Zhao et al. [4]; thus, a maximum radiation dose of 14 Gy can compensate for radioprotective effects during the PI treatment of oriental fruit fly. This explains why no radioprotective effects were observed when a radiation dose of 116 Gy (31.9 to 35.4 Gy higher than the LD<sub>99,9968</sub> estimates) was applied to late third-instar *B. dorsalis* larvae under severe hypoxia ( $0.3 \pm 0.02\%$  O<sub>2</sub>, 21.6 ± 0.1% CO<sub>2</sub>) [12].

Commercial PI treatment in MAP or controlled atmospheres is more complicated [39]. The minimum applied dose should be equal to or higher than the maximum dose in the confirmatory tests; therefore, it is much greater than the  $LD_{99.9968}$  estimates, and radioprotective effects are not observed as a result [12,21,31]. Moreover, although the treatment efficacy is reduced by low-oxygen atmospheres, it is increased by the presence of  $CO_2$ , low temperature, and the long duration of modified atmosphere treatment after radiation treatment [11,12,32,38].

In order to define the critical threshold of radioprotective effects, radiation treatments in a series of oxygen levels should be carried out and the dose-response data subjected to statistical analysis to determine the statistically significant differences in radiotolerance [22,23]. The probit model is commonly used to analyze dose-response data [38,40]; however, the values of the slope obtained in this research (Table 3) are unequal, which means that the regression lines are unparallel, whereas they were parallel when Srimartpirom et al. treated B. dorsalis in MAP [11]. Therefore, the 95% CLs of the lethal dose ratios at LD<sub>99</sub> (ratio test) were used for comparing the relative radiotolerance [28,39]. As a result, oriental fruit fly irradiated in 0% and 2% O<sub>2</sub> showed significantly higher tolerance than that irradiated in ambient air and other low-oxygen atmospheres (Table 3, Figures 1 and 2). There were no differences among 6% and 8%  $O_2$  and ambient air; however, visible but insignificant tolerance differences were present between the treatments in 4% and 6% O<sub>2</sub> (Tables 2 and 3; Figures 1 and 2). To analyze the trends more intuitively, like how Chao et al. [23] predicted the critical  $O_2$  level for the radiation treatment of *T. ni*, the estimated values (100% mortality,  $LD_{99}$ , and  $LD_{99,9968}$ ) at all  $O_2$  levels in our testing were plotted in Figure 1. All three curves remained flat through the treatment at 21%, 8%, and 6%  $O_2$ but turned up slowly for treatments at 6-4% O2 and then steepened rapidly; therefore, the critical threshold of radioprotective effects is an O<sub>2</sub> level of  $\geq$ 4% and <6% when late third-instar *B. dorsalis* larvae are irradiated in low-oxygen atmospheres. Since there is a small difference in LD<sub>99</sub> values of 1.1 Gy (1.95% of the radiation doses) between the treatments in 4% and 6%  $O_2$  (Table 3), more precise research is required to reduce the range of critical values, for example, by conducting radiation testing in 4.5% or 5%  $O_2$  atmospheres. Similarly, radioprotective effects have been observed in other fruit insects irradiated at  $\langle 3-5\% \rangle O_2$  levels, such as A. suspensa [33], C. capitata [34], Z. cucurbitae [13], D. suzukii [35], G. molesta [36], O. nubilalis [37], and T. ni [22,23]. Furthermore, all these results support the APHIS' (Animal and Plant Health Inspection Service, United States Department of Agriculture) changes to administrative requirements, wherein the minimum  $O_2$  level was reduced from 18% to 10% for conducting PI treatments [22,41]. However, the IPPC prohibits the use of PI treatment under low-oxygen atmospheres except for the treatment of G. molesta [20]; therefore, more insect species in different orders or families still need to be irradiated under low-oxygen environments to determine their critical threshold of radioprotective effects or the extra radiation dose that can compensate for the reduced efficacy. After that, a generic threshold of radioprotective effects can be established to facilitate the application of PI treatment of commodities with MAP or in low-oxygen environments.

Probit analysis has been widely used to analyze dose–response data for a number of fruit fly studies, and the ratios test is often used to determine the relative toxicity among a number of chemicals to determine the relative susceptibility of populations to pesticide resistance [4,11,14,29,42–44]. In addition, confidence interval (CI) overlap, relative median potency, and one-way ANOVA (on LD<sub>95</sub>) have been used to determine the significance of radiotolerance differences [11,23,30]. The 95% CLs of the lethal

dose ratios at a specific efficacy level, which are calculated automatically using computer software, are more general than the alternative statistics: Relative potency, which assumes that the regression lines are parallel; the CI overlap test, which should be used only when no alternative test exists; and one-way ANOVA, which requires at least three replicates of the LD<sub>X</sub> (for example, four replicates were conducted in radiation treatment of the cabbage looper moth, resulting in significant radioprotective effects determined in <0.1% and 2.5% O<sub>2</sub> atmospheres) [11,23,40]. The ratio test showed that the LD<sub>99</sub> value in 2% O<sub>2</sub> atmospheres is significantly larger than those in air and in 4%, 6%, and 8% atmospheres (Table 3), but all the 95% CIs overlapped, suggesting that the ratio test is more sensitive than the CI overlap test, which provided a lower observed type I error rate of 0.004 to 0.005 [30]. Since probit-9 (LD<sub>99,9968</sub> at 95% CL), which is well known for use as the criterion for the PI treatment of insects, is an extrapolated value, the LD<sub>99</sub> that is calculated from the dose–response curve of tests in the most tolerant life stage is used as the minimum treatment level [38]; therefore, the 95% CLs of the lethal dose ratios at LD<sub>99</sub> are recommend for comparing the significance of tolerance in phytosanitary treatments.

## 5. Conclusions

In this study, radioprotective effects were demonstrated and their critical threshold was determined by irradiating late third-instar *B. dorsalis* larvae with 220 KV X-rays in ambient air and low-oxygen environments. The ratio test indicated that the differences in radiotolerance were significant in 0% and 2% O<sub>2</sub> but insignificant in 4%, 6%, and 8% O<sub>2</sub> environments when compared with treatment in ambient air; the critical threshold of radioprotective effects is, therefore, an O<sub>2</sub> level of  $\geq$ 4% and <6%, but a maximum radiation dose of 14 Gy can compensate for this effect during PI treatment. We recommend that the 95% CLs of the lethal dose ratios at LD<sub>99</sub> be used for comparing the significance of tolerance in phytosanitary treatments, and a generic threshold of radioprotective effects should be established by testing a variety of insects.

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## References

- Crop Protection Compendium. *Bactrocera dorsalis* (Oriental fruit fly). Available online: https://www.cabi.org/ ISC/datasheet/17685 (accessed on 24 January 2020).
- 2. Nugnes, F.; Russo, E.; Viggiani, G.; Bernardo, U. First record of an invasive fruit fly belonging to *Bactrocera dorsalis* complex (Diptera: Tephritidae) in Europe. *Insects* **2018**, *9*, 182. [CrossRef]
- 3. Fang, Y.; Kang, F.F.; Zhan, G.P.; Ma, C.; Li, Y.G.; Wang, L.; Wei, Y.D.; Gao, X.W.; Li, Z.H.; Wang, Y.J. The effects of a cold disinfestation on *Bactrocera dorsalis* survival and navel orange quality. *Insects* **2019**, *10*, 452. [CrossRef]
- Zhao, J.P.; Ma, J.; Wu, M.T.; Jiao, X.G.; Wang, Z.G.; Liang, F.; Zhan, G.P. Gamma radiation as a phytosanitary treatment against larvae and pupae of *Bactrocera dorsalis* (Diptera: Tephritidae) in guava fruits. *Food Control* 2017, 72, 360–366. [CrossRef]
- 5. Liu, H.; Zhang, D.J.; Xu, Y.J.; Wang, L.; Cheng, D.F.; Qi, Y.X.; Zeng, L.; Lu, Y.Y. Invasion, expansion, and control of *Bactrocera dorsalis* (Hendel) in China. *J. Integr. Agric.* **2019**, *18*, 771–787. [CrossRef]
- 6. Hallman, G.J. Process control in phytosanitary irradiation of fresh fruits and vegetables as a model for other phytosanitary treatment processes. *Food Control* **2017**, *72*, 372–377. [CrossRef]

- Hallman, G.J. Phytosanitary applications of irradiation. *Compr. Rev. Food Sci. Food Saf.* 2011, 10, 143–151. [CrossRef]
- 8. Hallman, G.J.; Blackburn, C.M. Phytosanitary Irradiation. Foods 2016, 5, 8. [CrossRef]
- 9. International Irradiation Association, Thailand Institute of Nuclear Technology, and the Joint FAO/IAEA Programme. The 8th Annual Chapman University Phytosanitary Forum. June, 2018. Bangkok, Thailand. Available online: https://iiaglobal.com/news/8th-chapman-annual-forum-phytosanitary/ (accessed on 24 January 2019).
- 10. Follett, P.A.; Armstrong, J.W. Revised irradiation doses to control melon fly, Mediterranean fruit fly, and oriental fruit fly (Diptera: Tephritidae) and a generic dose for tephritid fruit flies. *J. Econ. Entomol.* **2004**, *97*, 1254–1262. [CrossRef]
- Srimartpirom, M.; Burikam, I.; Limohpasmanee, W.; Kongratarporn, T.; Thannarin, T.; Bunsiri, A.; Follett, P.A. Low-Dose Irradiation with modified atmosphere packaging for mango against the oriental fruit fly (Diptera: Tephritidae). *J. Econ. Entomol.* 2018, *111*, 135–140. [CrossRef]
- 12. Dias, V.S.; Hallman, G.J.; Martínez-Barrera, O.Y.; Hurtado, N.V.; Cardoso, A.A.S.; Parker, A.G.; Caravantes, L.A.; Rivera, C.; Araújo, A.S.; Maxwell, F.; et al. Modified Atmosphere Does Not Reduce the Efficacy of Phytosanitary Irradiation Doses Recommended for Tephritid Fruit Flies. *Insects* **2020**, *11*. [CrossRef]
- Follett, P.A.; Wall, M.M.; Bailey, W. Influence of modified atmosphere packaging on radiation tolerance in the phytosanitary pest melon fly (Diptera: Tephritidae). *J. Econ. Entomol.* 2013, 106, 2020–2026. [CrossRef] [PubMed]
- 14. Zhan, G.P.; Ren, L.L.; Shao, Y.; Wang, Q.L.; Yu, D.J.; Wang, Y.J.; Li, T.X. Gamma irradiation as a phytosanitary treatment of *Bactrocera tau* (Diptera: Tephritidae) in pumpkin fruits. *J. Econ. Entomol.* **2015**, *108*, 88–94.
- 15. IPPC (International Plant Protection Convention). List of Topics for IPPC Standards. Available online: https://www.ippc.int/en/core-activities/standards-setting/list-topics-ippc-standards/list) (accessed on 19 February 2020).
- 16. Wall, M. Quality of postharvest horticultural crops after irradiation treatment. *Stewart Postharvest Rev.* **2015**, *4*, 1–7. [CrossRef]
- 17. Follett, P.A.; Wall, M.M. Phytosanitary irradiation for export of fresh produce: Commercial adoption in Hawaii and current issues. *J. Radioanal. Nucl. Chem.* **2013**, *296*, 517–522. [CrossRef]
- 18. Lacroix, M.; Follett, P.A. Combination irradiation treatments for food safety and phytosanitary uses. *Stewart Postharvest Rev.* **2015**, *11*, 1–10.
- 19. Neven, L.G.; Hansen, L.D. Effects of temperature and controlled atmospheres on codling moth metabolism. *Ann. Entomol. Soc. Am.* **2010**, *103*, 418–423. [CrossRef]
- 20. IPPC (International Plant Protection Convention) ISPM 28. *Annex 1 PT 11: Irradiation Treatment for Grapholita Molesta;* FAO: Rome, Italy, 2010.
- 21. Hallman, G.J.; Levang-Brilz, N.M.; Zettler, J.L.; Winborne, I.C. Factors affecting ionizing radiation phytosanitary treatments, and implications for research and generic treatments. *J. Econ. Entomol.* **2010**, *103*, 1950–1963. [CrossRef]
- 22. Condon, C.H.; White, S.; Meagher, R.L.; Jeffers, L.A.; Bailey, W.D.; Hahn, D.A. Effects of low-oxygen environments on the radiation tolerance of the cabbage looper moth (Lepidoptera: Noctuidae). *J. Econ. Entomol.* **2017**, *110*, 80–86. [CrossRef]
- Chen, C.; Condon, C.H.; Boardman, L.; Meagher, R.L.; Jeffers, L.A.; Beam, A.; Bailey, W.D.; Hahn, D.A. Critical PO<sub>2</sub> as a diagnostic biomarker for the effects of low-oxygen modified and controlled atmospheres on phytosanitary irradiation treatments in the cabbage looper *Trichoplusia ni* (Hübner). *Pest Manag. Sci.* 2020, 76, 76. [CrossRef]
- 24. Liu, B.; Li, B.S.; Zhan, G.P.; Zha, T.; Wang, Y.J.; Ma, C. Forced hot-air treatment against *Bactrocera papayae* (Diptera: Tephritidae) in papaya. *Appl. Entomol. Zool.* **2017**, *52*, 531–541. [CrossRef]
- Gueorguiev, G. Irradiation System and Method Using X-Ray and Gamma-Ray Reflector. U.S. Patent 6,389,099, 14 May 2002.
- 26. Abbott, W.S. A method for computing the effectiveness of an insecticide. *J. Econ. Entomol.* **1925**, *18*, 265–267. [CrossRef]

- 27. DPS (Data Processing System). *User's Guide*; Version 13.5; Hangzhou RuiFeng Information Technology Co., Lt.: Hangzhou, China, 2010; Available online: http://www.chinadps.net/files/dps2nd.pdf (accessed on 25 January 2020).
- 28. Robertson, J.L.; Preisler, K.H.; Russell, R.M. A user's guide to probit or logit analysis. In *PoloPlus Package Version 2.0*; LeOra Software: Berkeley, CA, USA, 2007.
- Myers, S.W.; Cancio-Martinez, E.; Hallman, G.Y.; Fontenot, E.A.; Vreysen, M.J.B. Relative tolerance of six *Bactrocera* (Diptera: Tephritidae) to phytosanitary cold treatment. *J. Econ. Entomol.* 2016, 109, 2341–2347. [CrossRef] [PubMed]
- 30. Wheeler, M.W.; Robert, M.; Park, R.M.; Bailer, J. Comparing median lethal concentration values using confidence interval overlap or ratio tests. *Environ. Toxicol. Chem.* **2006**, *25*, 1441–1444. [CrossRef] [PubMed]
- 31. IPPC (International Plant Protection Convention), ISPM 18. *Guidelines for the Use of Irradiation as a Phytosanitary Measure;* FAO: Rome, Italy, 2003.
- 32. IPPC (International Plant Protection Convention), ISPM 28. *Phytosanitary Treatments for Regulated Pests*; FAO: Rome, Italy, 2007.
- 33. López-Martínez, G.; Hahn, D.A. Short-term anoxic conditioning hormesis boosts antioxidant defenses, lowers oxidative damage following irradiation and enhances male sexual performance in the Caribbean fruit fly, *Anastrepha suspensa*. J. Exp. Biol. **2012**, 215, 2150–2161. [CrossRef]
- Nestel, D.; Nemny-Lavy, E.; Islam, A.; Wornoayporn, V.; Cáceres, C. Effects of pre-irradiation conditioning of medfly pupae (Diptera: Tephritidae): Hypoxia and quality of sterile males. *Fla. Entomol.* 2007, *90*, 80–87. [CrossRef]
- 35. Follett, P.A.; Swedman, A.; Mackey, B. Effect of low oxygen conditions created by modified atmosphere packaging on radiation tolerance in *Drosophila suzukii* (Diptera: Drosophilidae) in sweet cherries. *J. Econ. Entomol.* **2018**, *111*, 141–145. [CrossRef]
- 36. Hallman, G.J. Ionizing irradiation quarantine treatment against oriental fruit moth (Lepidoptera: Tortricidae) in ambient and hypoxic atmospheres. *J. Econ. Entomol.* **2004**, *97*, 824–827. [CrossRef]
- 37. Hallman, G.J.; Hellmich, R.L. Ionizing radiation as a phytosanitary treatment against European corn borer (Lepidoptera: Crambidae) in ambient, low oxygen, and cold conditions. *J. Econ. Entomol.* **2009**, *102*, 64–68. [CrossRef]
- 38. NAPPO (North American Plant Protection Organization), RSPM 34. *Development of Phytosanitary Treatment Protocols for Regulated Arthropod Pests of Fresh Fruits or Vegetables;* NAPPO: Ottawa, ON, Canada, 2011.
- Follett, P.A.; Neven, L.G. Phytosanitary irradiation: Does modified atmosphere packaging or controlled atmosphere storage creating a low oxygen environment threaten treatment efficacy? *Radiat. Phys. Chem.* 2020, 173. [CrossRef]
- 40. Robertson, J.R.; Ryssekk, R.M.; Preisler, H.K.; Savin, N.E. *Bioassays with Arthropods*, 2nd ed.; CRC Press: Boca Raton, FL, USA, 2007; pp. 26–32.
- 41. USDA Treatment Manual. Available online: https://www.aphis.usda.gov/import\_export/plants/manuals/ ports/downloads/treatment.pdf (accessed on 1 May 2020).
- 42. Bustos, M.E.; Enkerlin, W.; Reyes, J.; Tolrdo, J. Irradiation of Mangoes as a postharvest quarantine treatment for fruit flies. *J. Econ. Entomol.* **2004**, *97*, 286–292. [CrossRef]
- 43. Hallman, G.J.; Thomas, D.B. Ionizing radiation as a phytosanitary treatment against fruit flies (Diptera: Tephritidae): Efficacy in naturally versus artificially infested fruit. *J. Econ. Entomol.* **2010**, *103*, 1129–1134. [CrossRef] [PubMed]
- 44. Lei, C.F.; Sun, X.L. Comparing lethal dose ratios using probit regression with arbitrary slopes. *BMC Pharmacol. Toxicol.* **2018**, *19*, 61. [CrossRef] [PubMed]



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