





Genome Sequences of Three *Microbacterium* Phages Isolated from Flowers

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ABSTRACT Bacteriophages Balsa, Golden, and Lucky3 are cluster EA phages isolated from flowers and infect *Microbacterium foliorum* NRRL B-24224. The genomes of Golden and Lucky3 (subcluster EA1) are closely related, whereas Balsa (subcluster EA4) is a more distant relative.

Numerous bacteriophages infecting actinobacterial hosts have been isolated from environmental samples, such as soil and compost (1–3). The presence of actinobacteria on internal and external structures of flowers suggests that phages of these hosts might also be present (4). We report here the genome sequences of phages Balsa, Golden, and Lucky3, which were isolated from flowers of *Impatiens pallida* (Balsaminaceae), *Solidago canadensis* (Asteraceae), and *Trifolium repens* (Fabaceae), respectively, collected in Pittsburgh, PA, using *Microbacterium foliorum* NRRL B-24224 as a bacterial host. In brief, flowers were washed in peptone-yeast extract-calcium (PYCa) broth, phage growth was enriched by incubation with host bacteria in PYCa broth at 30°C, and plaques were identified following plating on PYCa agar. Phage Balsa forms turbid plaques, whereas Golden and Lucky3 form clear plaques of various sizes.

Phage DNA was isolated from phage lysates using the Wizard DNA extraction kit (Promega), and then sequencing libraries were prepared using an NEB Ultra II FS kit and run on an Illumina MiSeq platform, yielding at least 100,000 single-end 150-base reads for each genome. Reads were assembled using Newbler 2.9, with default settings, and in each case yielded a single phage contig (average coverages for Golden, Lucky3, and Balsa, 1,675-, 3,441-, and 1,992-fold, respectively) which was evaluated with Consed 2.9. No evidence was found for defined genomic termini (5); therefore, the start of the terminase small subunit was chosen as the genome coordinate terminus. Balsa has a 41,862-bp genome with 63.4% G+C content, while Golden and Lucky3 each have 39,640-bp genomes with 64.1% G+C content. The G+C content of the host (68.7%; accession number [NZ_CP031425](https://doi.org/10.1128/MRA.01468-18)) is somewhat higher than that of the three phages (our unpublished data). All three phages belong to cluster EA and share at least 35% average gene content (2). Golden and Lucky3 differ by only three single-base substitutions and were grouped in subcluster EA4. Balsa is similar to a large number of phages grouped in subcluster EA1.

The three genomes were annotated using a previously described genome annotation pipeline (6), together with Glimmer (7), GeneMark (8), BLASTP (9), HHPred (10), and Phamerator (11). Golden and Lucky3 both have 58 predicted protein-coding genes, and Balsa has 62 protein-coding genes. The genome leftmost halves are transcribed rightwards and code for virion structure and assembly, followed by a lysis cassette. With the exception of the rightmost two genes of each genome, the rightmost genome halves are transcribed leftwards and contain genes coding for DNA metabolism functions, including DNA polymerase I, helicase, thymidylate synthase, exonuclease, and a RecA-like recombinase. Although Balsa forms turbid plaques, we did not identify genes

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typically found in temperate phages, such as those for a repressor, integrase, or partitioning functions.

All three phages share common virion structure and assembly genes, including a putative HK97-fold capsid subunit and a tape measure protein gene. The lysis cassettes contain putative holin and endolysin genes, although the Balsa endolysin (gp24), with D-Ala-D-Ala carboxypeptidase and transglycosylase domains, differs from those in Golden and Lucky3 (gp24) that have peptidoglycan binding and amidase domains. Balsa contains several genes near its right end that are absent in subcluster EA4 phages. Only 11 genes found in these phages have homologues in phages outside cluster EA.

Data availability. The GenBank and Sequence Read Archive accession numbers are [MG839030](https://doi.org/10.1093/nar/gkz003) and [SRR7769840](https://doi.org/10.1093/nar/gkz003) for Balsa, [MG925343](https://doi.org/10.1093/nar/gkz003) and [SRR7769839](https://doi.org/10.1093/nar/gkz003) for Golden, and [MG925347](https://doi.org/10.1093/nar/gkz003) and [SRR7769841](https://doi.org/10.1093/nar/gkz003) for Lucky3, respectively.

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REFERENCES

1. Pope WH, Bowman CA, Russell DA, Jacobs-Sera D, Asai DJ, Cresawn SG, Jacobs WR, Hendrix RW, Lawrence JG, Hatfull GF, Science Education Alliance Phage Hunters Advancing Genomics and Evolutionary Science, Phage Hunters Integrating Research and Education, Mycobacterial Genetics Course. 2015. Whole genome comparison of a large collection of mycobacteriophages reveals a continuum of phage genetic diversity. *Elife* 4:e06416. <https://doi.org/10.7554/eLife.06416>.
2. Pope WH, Mavrich TN, Garlena RA, Guerrero-Bustamante CA, Jacobs-Sera D, Montgomery MT, Russell DA, Warner MH, Science Education Alliance-Phage Hunters Advancing G, Evolutionary S, Hatfull GF. 2017. Bacteriophages of *Gordonia* spp. display a spectrum of diversity and genetic relationships. *mBio* 8:e01069-17. <https://doi.org/10.1128/mBio.01069-17>.
3. Klyczek KK, Bonilla JA, Jacobs-Sera D, Adair TL, Afram P, Allen KG, Archambault ML, Aziz RM, Bagnasco FG, Ball SL, Barrett NA, Benjamin RC, Blasi CJ, Borst K, Braun MA, Broomell H, Brown CB, Brynell ZS, Bue AB, Burke SO, Casazza W, Cautela JA, Chen K, Chimalakonda NS, Chudoff D, Connor JA, Cross TS, Curtis KN, Dahlke JA, Deaton BM, Degroote SJ, DeNigris DM, DeRuff KC, Dolan M, Dunbar D, Egan MS, Evans DR, Fahnstock AK, Farooq A, Finn G, Fratus CR, Gaffney BL, Garlena RA, Garrigan KE, Gibbon BC, Goedde MA, Guerrero Bustamante CA, Harrison M, Hartwell MC, Heckman EL, et al. 2017. Tales of diversity: genomic and morphological characteristics of forty-six *Arthrobacter* phages. *PLoS One* 12:e0180517. <https://doi.org/10.1371/journal.pone.0180517>.
4. Turner TR, James EK, Poole PS. 2013. The plant microbiome. *Genome Biol* 14:209. <https://doi.org/10.1186/gb-2013-14-6-209>.
5. Russell DA. 2018. Sequencing, assembling, and finishing complete bacteriophage genomes. *Methods Mol Biol* 1681:109–125. https://doi.org/10.1007/978-1-4939-7343-9_9.
6. Pope WH, Jacobs-Sera D. 2018. Annotation of bacteriophage genome sequences using DNA Master: an overview. *Methods Mol Biol* 1681: 217–229. https://doi.org/10.1007/978-1-4939-7343-9_16.
7. Delcher AL, Harmon D, Kasif S, White O, Salzberg SL. 1999. Improved microbial gene identification with Glimmer. *Nucleic Acids Res* 27: 4636–4641. <https://doi.org/10.1093/nar/27.23.4636>.
8. Borodovsky M, McIninch J. 1993. Recognition of genes in DNA sequence with ambiguities. *Biosystems* 30:161–171. [https://doi.org/10.1016/0303-2647\(93\)90068-N](https://doi.org/10.1016/0303-2647(93)90068-N).
9. Altschul SF, Gish W, Miller W, Myers EW, Lipman DJ. 1990. Basic local alignment search tool. *J Mol Biol* 215:403–410. [https://doi.org/10.1016/S0022-2836\(05\)80360-2](https://doi.org/10.1016/S0022-2836(05)80360-2).
10. Zimmermann L, Stephens A, Nam SZ, Rau D, Kubler J, Lozajic M, Gabler F, Soding J, Lupas AN, Alva V. 2018. A completely reimplemented MPI bioinformatics toolkit with a new HHpred server at its core. *J Mol Biol* 430:2237–2243. <https://doi.org/10.1016/j.jmb.2017.12.007>.
11. Russell DA, Hatfull GF. 2017. PhagesDB: the actinobacteriophage database. *Bioinformatics* 33:784–786. <https://doi.org/10.1093/bioinformatics/btw711>.