LAB/IN VITRO RESEARCH

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ysis were performed to determine whether KIAA1199 is a downstream target of miR-486-5p.
Results: We found that KIAA1199 was aberrantly elevated in PTC tissues compared with normal tissues, and upregulation of KIAA1199 was positively correlated with more advanced clinical variables. Additionally, bioinformatic analysis indicated that KIAA1199 was involved in cell migration and invasion. KIAA1199 silencing inhibited the invasive ability of PTC cells by affecting epithelial-mesenchymal transition (EMT) *in vitro* and *in vivo*. Furthermore, miR-486-5p was identified as an upstream microRNA that directly targets the 3'-UTR region of KIAA1199.
Conclusions: The miR-486-5p/KIAA1199/EMT axis might play a critical role in PTC invasion and metastasis and offers a potential therapeutic strategy for PTC.

MeSH Keywords: MicroRNAs • Neoplasm Metastasis • Thyroid Neoplasms

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Background

Thyroid cancer is the most common malignancy in the endocrine system [1]. In recent decades, the incidence of thyroid cancer has been increasing steadily, largely due to advancements in diagnostic techniques for this disease [2]. Approximately 80% of diagnosed thyroid cancers worldwide are papillary thyroid cancer (PTC) [3]. Surgical resection along with radioactive iodine treatment is currently the standard procedure for the management of PTC [4]. However, due to certain aggressive PTC variants, lymph node metastasis or distant metastasis always occurs in PTC patients, who often exhibit poor responses to standard treatments and thus have poor clinical outcomes [5]. Thus, there is an urgent need for a better understanding of the molecular mechanisms behind PTC invasion and metastasis.

KIAA1199 was first identified by Satoko Abe et al. as a gene related to inner-ear function, as mutations in this gene can result in non-syndromic hearing dysfunction [6]. Recent studies have demonstrated that KIAA1199 also plays a critical role in tumor progression, especially invasion and metastasis. Upregulation of KIAA1199 expression was reported to be associated with cancer progression and to predict a poor prognosis in various cancers, including colorectal cancer [7,8], gastric cancer [9,10], breast cancer [11,12], and pancreatic cancer [13]. In gastric cancer, KIAA1199 can activate the Wnt/ β -catenin signalling pathway and then upregulate matrix metalloproteinase (MMP) enzymatic activities, which contribute to epithelial-mesenchymal transition (EMT) [9]. However, the molecular roles of KIAA1199 remain unclear in papillary thyroid cancer.

The aberrant expression of miRNAs has been implicated in the development, progression, and prognosis of cancer [14]. miR-486-5p, widely documented to be a tumor-suppressive microRNA, inhibits proliferation and invasion in many types of cancers [15–17]. However, the clinical and biological role of miR-486-5p in PTC is still unclear.

In this study, we first determined the aberrant mRNA and protein expression of KIAA1199 in PTC. Then, we investigated the clinical significance and molecular role of KIAA1199 using *in vitro* and *in vivo* experiments. Finally, we identified specific sequences in the KIAA1199 3'-UTR that are directly targeted by miR-486-5p in the regulation of PTC invasion and metastasis.

Material and Methods

Data source and bioinformatic analysis

TCGA thyroid cancer level 3 mRNA-seq and miRNA-seq data were acquired from UCSC Xena (*https://tcga.xenahubs.net*),

including 59 paired PTC and adjacent normal tissues along with clinicopathological data.

Normalization and differential expression analysis were performed with the Bioconductor package DESeq2 [18]. A heatmap was plotted using MeV [19]. Gene set enrichment analysis (GSEA) was performed to explore potential pathways with the "hallmark.all.v6.1.symbols.gmt" gene set [20]. The Weighted Gene Coexpression Network Analysis (WGCNA) R package [21] was used to evaluate gene significance (GS) and module membership (MM). The brown module, which was most significantly correlated with KIAA1199 expression, was selected for further study. Genes involved in the blue module were submitted to Metascape for Gene Ontology (GO) enrichment visualization [22].

Cell culture and transfection

Two papillary thyroid cancer cell lines were purchased from ATCC and cultured in RPMI1640 media (Sigma-Aldrich). Transfection was performed with Lipofectamine 2000 (Invitrogen, USA) according to the manufacturer's protocol. Nonsense RNAi was used as negative control. The efficiency of transfection was measured using Western blot analysis. A miR-486-5p mimic, negative control mimics, and siRNAs targeting KIAA1199 were purchased from Genechem (Shanghai, China). KIAA1199targeted short hairpin RNAs (shRNAs) were designed according to siRNA and nsRNA sequences. Recombinant lentiviral particles were generated, and BCPAP cells were transfected with KIAA1199 or a negative control (sh-KIAA1199 or sh-scramble) for use in the animal study. To steadily overexpress miR-486-5p, recombinant lentiviruses containing the miR-486-5p precursor, as well as the negative control scramble sequences, were purchased from Genechem.

Luciferase reporter assay

A wild-type (WT) 3'-UTR of KIAA1199 cDNA was amplified by PCR and cloned into the Xbal and SacI sites of a pmirGLO dualluciferase miRNA target expression vector. The mutant (Mut) sequence of the KIAA1199 3'-UTR was constructed according to the WT-KIAA1199 3'-UTR region with mutated nucleotides in specific sequences that can predictively bind to miR-486-5p. Vectors (WT-KIAA1199 3'-UTR or Mut-KIAA1199 3'-UTR with miR-486-5p mimic or miR-negative control) were transiently transfected into cells using Lipofectamine 2000 based on the user's manuals. Luciferase activity was detected with the Dual-Luciferase Reporter Assay System after 48 h of transfection.

RNA extraction and qRT-PCR

Total RNA was extracted from cultured cells using TRIzol reagent (Invitrogen, Carlsbad, CA, USA) and qRT-PCR was performed

according to the standard user's manual. KIAA1199 primers used were: forward 5'-CCAGGAATGTTGAATGTCT-3', reverse 5'-ATTGGCTCTTGGTGAATG-3'.

Western blot analysis

Western blot analysis was performed according to standard protocols. In brief, after separation in SDS-PAGE gels, protein was transferred on a PVDF membrane. After incubation with antibodies, membranes were washed in TBS-T. Then, membranes were incubated with secondary antibody. Blots were visualized using the ECL imaging system (Thermo Scientific). Antibodies against β -actin (#4970, CST, MA, USA, 1: 1000), KIAA1199 (NBP2–50336, Novus, USA, 1: 500), Vimentin (#5741, CST, CA, USA, 1: 1000), E-cadherin (sc-71008, Santa Cruz Biotechnology, CA, USA, 1: 1000), and Slug (#9585, CST, MA, USA, 1: 500) were used.

TMA and immunohistochemistry (IHC)

A tissue microarray (TMA) with 58 paired papillary thyroid cancer tissue dots and clinical annotations was used for protein expression validation. Immunohistochemistry (IHC) was performed to measure KIAA1199 protein expression in TMA samples. In brief, tissue sections on the TMA were deparaffinized and rehydrated through graded alcohol solutions. Endogenous peroxidase activity was blocked in 3% H₂O₂. Antigen retrieval was performed with 0.01 M citrate buffer (pH 6.0) and microwave heat treatment. IHC results for each dot were observed independently by 2 experienced pathologists. The staining intensity was classified into 4 grades: 0 points (no staining), 1 point (weak staining), 2 points (intermediate staining), or 3 points (strong staining). The product (percentage of positive regions multiplied by intensity points) was considered the staining score (range from 0 to 300).

Transwell and Matrigel assays

For the Transwell assay, 20 000 transfected PTC cells cultured in serum-free media were seeded in the upper chamber of an insert (8- μ m pore size; Millipore, Billerica, MA, USA). Complete medium with 10% FBS was added to the lower chambers. After culturing for 24 h at 37°C in a 5% CO₂ atmosphere, cells that had migrated through the membrane were fixed and stained, and then, for imaging and quantification by microscopy, cells were counted in at least 3 random fields to obtain an average count. For the Matrigel assay, 20 000 transfected cells were plated into the upper chamber on a Matrigel-coated membrane (BD Biosciences) in serum-free medium. The lower chambers also contained complete medium with 10% FBS. After a 48-h incubation, the cells were harvested under the same conditions as in the Transwell assay described above.

Mouse metastasis model

Animal studies were approved by Jiangsu Province Animal Ethics Committee and were conducted in strict accordance with the Guide for the Care and Use of Laboratory Animals of the National Institutes of Health. Ten male BALB/c nude mice (6 weeks old) were purchased from the Animal Center of the Chinese Academy of Science (Shanghai, China) and maintained in specific pathogen-free (SPF) conditions. Briefly, for the metastasis model, 3×10^6 BCPAP cells stably expressing shRNA-Scramble or shRNA-KIAA1199 were suspended in 100 µl PBS and injected intravenously into the tail vein of mice (5 mice in each group). Six weeks later, the mice were sacrificed and the lungs were harvested. Metastatic nodules on the lung surface were counted and hematoxylin and eosin (HE) staining was performed for histological examination.

Statistical analysis

Data are presented as the mean \pm S.D. Statistical analyses were conducted with SPSS Statistics tool, R software (version 3.5.1), and GraphPad Prism 8 software. We used the *t* test or one-way ANOVA to analyze differences. P<0.05 was considered statistically significant.

Results

Elevation of KIAA1199 is correlated with more advanced clinical variables

To identify potential oncogenes in PTC, we screened wholegenome RNA-seq data from the TCGA. Over 20 000 proteincoding genes were included in the DESeq2 analysis, and the volcano plot in Figure 1A shows significantly upregulated and downregulated genes that met the filtering criteria (llog FC)>2, FDR<0.01). A heatmap based on these differentially expressed genes was plotted to show the detailed expression profile of 59 pairs of PTC and adjacent normal tissues (Figure 1B). KIAA1199 mRNA (read count) was significantly upregulated (p=0.0022) in cancer tissues (Figure 1C) and widely upregulated at a ratio of 42/59 compared with adjacent normal tissues (Figure 1D). Figure 1E shows that KIAA1199 was significantly upregulated in lymph node metastasis (LNM)-positive samples and in more malignant subtypes, but no significant difference was observed in tumor size. IHC analysis of the TMA indicated that KIAA1199 protein level was elevated in papillary thyroid cancer tissues compared to normal tissues (Figure 1F), especially in LNM-positive tissues (Figure 1G).



Figure 1. Upregulation of KIAA1199 is correlated with more advanced clinical variables. (A) The volcano plot showed significantly upregulated and downregulated genes in PTC tissues. (B) The heatmap depicted the detailed expression profile of these genes in 59 pairs of PTC and adjacent normal tissues. (C) KIAA1199 mRNA was significantly upregulated in cancer tissues, and (D) was widely upregulated at a ratio of 42/59 compared with adjacent normal tissues. (E) KIAA1199 was significantly upregulated in lymph node metastasis (LNM)-positive samples and more malignant subtypes. (F) The KIAA1199 protein level was upregulated in PTC tissues, (G) especially in LNM-positive tissues.

WGCNA analysis indicates that KIAA1199 is involved in cell migration and invasion

To construct a gene co-expression network, RNA-seq data from the whole genome of PTC samples were subjected to WGCNA. Genes were assigned to different modules by cluster dendrogram trees, and unassigned genes were categorized into the grey module (Figure 2A). A heatmap of the relationships between clinical traits and gene modules is shown in Figure 2B. We observed that the brown module was most significantly positively correlated with KIAA1199 expression. We then determined if gene significance and module membership exhibited a significant correlation (r=0.46, p=1.7e-16), and the result indicated that genes in the brown module were highly correlated with KIAA1199 (Figure 2C). Next, genes in the brown module were submitted to Metascape for GO enrichment visualization.



Figure 2. WGCNA analysis indicates that KIAA1199 is involved in cell migration and invasion. (A) Cluster dendrogram trees were constructed based on the whole-genome profiling data of TCGA. (B) Heatmap of the relationships between clinical traits and gene modules, and the brown module with highest correlation value (r=0.48, p=8e-20) was chosen for further study. (C) A significant correlation was found between module membership and gene significance of KIAA1199 in the brown module. (D) The major part of the KIAA1199-related network was labelled as "cell adhesion/cell migration/exocytosis/ chemotaxis/ECM organization", which are critical events in cancer invasion and metastasis.

As shown in Figure 2D, the major part of the network was labelled with "cell adhesion/cell migration/exocytosis/chemotaxis/ECM organization", which are critical events in cancer invasion and metastasis.

KIAA1199 promotes PTC invasion by influencing EMT

Two pairs of siRNAs were designed to inhibit KIAA1199 expression level in PTC cell lines (Figure 3A). Transwell and Matrigel assays indicated that silencing of KIAA1199 greatly impaired the migratory and invasive abilities of TPC-1 and BCPAP cells in response to siRNA-treatment compared with the group treated with siRNA-scramble (si-scr) (Figure 3B, 3C). To further validate the effect of KIAA1199 silencing on metastatic potential *in vivo*, we established a mouse metastasis model, as described in the Methods section above. We found that the incidence of lung metastasis was decreased in mice injected with BCPAP-shKIAA1199 cells compared with those injected with BCPAP-shScramble. Histological examination confirmed that silencing of KIAA1199 decreased the size and number of lung metastatic nodules (Figure 3D).

PTC samples in the TCGA dataset with KIAA1199 expression <Q1 and >Q3 according to a quartile method were selected, and GSEA was performed based on RNA-seq data. Among all the outputs, EMT ranked first, with an enrichment score of 0.64 (Figure 3E). Thus, it is plausible to hypothesize that KIAA1199 promotes PTC progression via EMT. Then, the protein levels



Figure 3. KIAA1199 promotes PTC invasion by influencing EMT. (A) Two pairs of siRNAs were designed to knock down the KIAA1199 expression level in PTC cell lines. (B, C) Transwell and Matrigel assays revealed that silencing of KIAA1199 greatly reduces the migratory and invasive abilities of TPC-1 and BCPAP cells in response to siRNA-treatment compared with the cells treated with siRNA-scramble. (D) Representative images showing that the incidence of lung metastasis was decreased in mice injected with BCPAP-shKIAA1199 cells compared with those injected with BCPAP-shScramble (p=0.0022). (E) GSEA showed that EMT ranked first in the KIAA1199-related predictive outputs. (F) Knock down of KIAA1199 upregulated E-cadherin but downregulated Slug and Vimentin in TPC-1 and BCPAP cells.



Figure 4. KIAA1199 is a direct target of miR-486-5p. (A) miR-486-5p was identified with a high predictive score in the intersection of 3 databases (TargetScan, miRDB, and PicTar). (B) TCGA data showed that miR-486 was significantly downregulated in cancer tissues compared to adjacent nontumoral tissues. (C) 2378-2385 of the KIAA1199 3'-UTR was a predicted sequence of miR-485-5p. (D) Luciferase reporter assays determined that miR-486-5p regulates KIAA1199 expression by directly binding to the predicted site in the 3'-UTR of KIAA1199. (E, F) qRT-PCR and Western blot analysis revealed the inhibitory effect of miR-486-5p on KIAA1199 expression in both cell lines. (G, H) A Matrigel assay showed that the invasive capacity of TPC-1 and BCPAP cells transfected with miR-486-5p was greatly impaired compared with that of the negative control group.

of EMT markers such as E-cadherin, Slug, and Vimentin were measured by Western blot. We observed that knockdown of KIAA1199 upregulated E-cadherin but downregulated Slug and Vimentin in TPC-1 and BCPAP cells (Figure 3F). These results indicated that KIAA1199 might exert its metastatic function by influencing EMT in PTC.

KIAA1199 is a direct target of miR-486-5p

To identify potential microRNAs targeting KIAA1199, we analyzed the predictive results from the intersection of 3 predictive databases – TargetScan, miRDB, and PicTar. Among the overlapping candidates, miR-486-5p attracted our attention due to its high predictive score (Figure 4A). Then, we investigated

the expression level of miR-486-5p in PTC, and observed in the TCGA miRNA-seq dataset that miR-486 expression was significantly downregulated in papillary thyroid cancer tissues compared to normal tissues (Figure 4B). As shown in Figure 4C, 2378-2385 of the KIAA1199 3'-UTR region was a predictive target sequence of miR-485-5p. Luciferase reporter assays were performed to validate whether miR-486-5p could bind to the predicted site described above. The data showed that miR-486-5p significantly inhibited luciferase activity in both TPC-1 and BCPAP cells with a reporter plasmid carrying the WT-KIAA1199 3'-UTR sequence. In contrast, we did not observe significant inhibition in cells with the reporter plasmid carrying a Mut-KIAA1199 3'-UTR (Figure 4D). Next, we measured the inhibitory effect of miR-486-5p on KIAA1199 expression in 2 PTC cell lines. As shown in Figure 4E and 4F, miR-486-5p overexpression significantly inhibited KIAA1199 mRNA and protein expression compared with the negative control group. Then, we investigated the invasion-promoting role of miR-486-5p in vitro. Matrigel assays indicated that miR-486-5p transfection markedly impaired the invasive ability of TPC-1 and BCPAP cells compared with the negative control group (Figure 4G, 4H).

Discussion

Activation of invasion and metastasis is a critical hallmark of cancer, but the underlying mechanism remains unclear [23]. Although most PTC patients have a favorable prognosis, accumulating evidence has shown that regional invasion and lymph node or distant metastasis increase the risk of recurrence and a worse survival [24–26]. Therefore, a better understanding of tumor invasion and metastasis is essential to identify reliable biomarkers and potential therapeutic targets for PTC.

In our study, we revealed that KIAA1199 mRNA and protein expression is aberrantly and frequently elevated in PTC tissues compared with normal tissues. Furthermore, KIAA1199 expression is positively correlated with more advanced clinicopathological characteristics. A bioinformatic analysis indicated that KIAA1199 is involved in cell migration and invasion, and *in vitro* and *in vivo* experiments showed that KIAA1199 enhances PTC invasion and metastasis by influencing EMT.

KIAA1199 was first identified as a novel large, long cDNA in the Human Unidentified Gene Encoded (HUGE) protein database [27]. Mutations in KIAA1199 were reported to be associated with loss of function of non-syndromic hearing [6]. Subsequently, some researchers reported that KIAA1199 is a novel hyaluronan-binding protein (HYBID) involved in hyaluronan depolymerization [28]. Recently, it was reported that tumor hypoxia-induced aberrant overexpression of KIAA1199 enhances tumor invasive capacity; thus, this protein is also known as cell migration-inducing protein (CEMIP) [29]. Some recently published studies have revealed the clinical and biological roles of KIAA1199 in cancer progression. KIAA1199 expression is aberrantly upregulated in gastric cancer [9], colon cancer [7,8], breast cancer [11,12], and pancreatic cancer [13]. In cervical cancer, KIAA1199 upregulation can be induced by human papillomavirus infection, and its expression is also aberrantly upregulated in pre-cancerous lesions [30]. Furthermore, KIAA1199 was reported to act as a predictor of poor clinical outcomes in lung cancer [31], pancreatic cancer [13], and colon cancer [7].

The malignant progression of PTC is considered to be a comprehensive event that includes a gene expression network and alterations in the tumor microenvironment, in which microRNAs play critical roles [32]. From the intersection of predictive results from 3 databases, we identified miR-486-5p as a promising upstream microRNA, which specifically targets KIAA1199 with the highest predictive score. Previously, Ma et al. reported that miR-486-5p inhibits cell growth of PTC by targeting fibrillin-1 [33]. We validated the miR-486-5p expression profile in PTC and revealed its downregulation in PTC tissues compared with adjacent normal tissues. Subsequently, by directly targeting the 3'-UTR region of KIAA1199, we demonstrated its invasion-suppressive role in PTC. In previous studies, KIAA1199 was reported to interact with many microRNAs such as miR-29c [34], miR-216a [35] and miR-600 [36], and we present here the first evidence that KIAA1199 enhances PTC invasion and metastasis by promoting EMT and that KIAA1199 is regulated by the tumor-suppressor miR-486-5p, which provides more clues to the regulation network of KIAA1199.

Conclusions

In summary, our study demonstrates the biological function of the miR-486-5p/KIAA1199/EMT axis, which provides strong evidence for the invasive ability of KIAA1199 in PTC. KIAA1199 might serve as a potential therapeutic target in PTC, and further studies are required to investigate the plausibility and potential pathways involved in KIAA1199-mediated cell invasion and metastasis.

Data availability

Normalized expression of KIAA1199 and miR-486-5p in PTC tissues and corresponding adjacent normal tissues in the TCGA database can be downloaded from the following website: *https://xenabrowser.net/*. The original data from the PTC TMA can be downloaded from the following website: *http://www.superchip.com.cn/*.

Conflicts of interest

None.

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