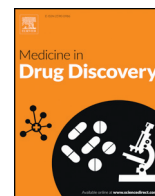




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Short Communication

Novel decoy cellular vaccine strategy utilizing transgenic antigen-expressing cells as immune presenter and adjuvant in vaccine prototype against SARS-CoV-2 virus

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ARTICLE INFO

Article history:

Received 21 March 2020

Received in revised form 22 March 2020

Accepted 22 March 2020

Available online 25 March 2020

Keywords:

COVID-19

SARS-CoV-2

Vaccine

Coronavirus

Antigen

Transgenic cell vaccination

ABSTRACT

A novel approach modifying cells to express viral markers to elicit protective immunity responses (decoy cellular vaccination) in the prevention of COVID-19 disease is currently being explored. Our approach entails utilizing SARS-CoV-2 Spike antigen-expressing, non-replicating cells as carriers and presenters of immunogenic antigens, so called “I-cells”. By using irradiated cells as presenting vehicles of SARS-CoV-2 viral antigens(s) in a cellular context, these presented viral proteins can be recognized by the host immune system, thus, an efficient protective immune response might be elicited. Another advantage of this strategy is that the manufacturing process is scalable and yields uniform cell products allowing for “off-the-shelf” frozen supply availability. To prevent engraftment and proliferation of the cells after administration, the cells will be irradiated post-harvesting abolishing in vivo replication potential. Specifically, immunoreactive Spike-1 proteins from SARS-CoV-2 are expressed on the surface of irradiated target I-cells. Utilizing this innovative strategy, these viral antigen-displaying decoy cells will be developed as a vaccine to protect against COVID-19 disease.

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Enabling the delivery and presentation of pathogen-specific proteins expressed in their native conformation to the host immune system, overcoming the known delivery and conformational inefficiencies associated with nucleic acid and recombinant protein-based approaches is a critical component of successful vaccine development.

The development pathway of I-cells, i.e. utilizing modified “decoy cells” to carry and present target antigens to the host immune system, is depicted in Fig. 1. Engineered Spike-1 protein is expressed on the surface of K562 human myelogenous leukemia cells via introduction of expression constructs into the cellular genome allowing for stable expression of the transgene. Stable-modified K562 clones are selected, profiled for Spike-1 expression as well as overall immunogenic potency, and prepared as GMP master cell bank to be used for large scale manufacturing. Irradiated cells are formulated as vaccine product and administered via intramuscular or subcutaneous injection.

The K562 cell line is HLA-negative and, as such, exquisitely sensitive to NK-mediated killing [1,2]. Expression of SARS-CoV-2-encoded proteins in the context of K562 or other NK-sensitive cell lines provides a means of targeting and activating an innate driver of the host adaptive immune

response against the immunogen and thus, virally infected host cells expressing the antigen. Cancer vaccine treatments utilizing lethally irradiated K562 cells expressing granulocyte-macrophage colony-stimulating factor (GM-CSF) (GM-K562) have been previously demonstrated to be well tolerated in human clinical trials [3,4]. Doses up to 1×10^8 GM-K562 cells have been administered by multiple parenteral routes and in treatment regimens including up to nine dose administrations. GM-CSF expression from GM-K562, in combination with co-delivered autologous tumor antigen have been shown to elicit a robust cellular and humoral anti-tumor immune response in treated patients [5].

This novel virus decoy cellular vaccine is designed to initiate NK cell-driven immune activation with the ultimate goal of inducing protective adaptive host immunity. NK cell-induced I-cell death should result in cytokine release, recruitment of antigen presenting cells (APCs), and maturation of elicited dendritic cells (DCs) at the site of vaccination. Presentation of SARS-CoV-2 Spike protein-derived peptides by recruited APCs will drive adaptive host immune responses against SARS-CoV-2. Interleukin 12 secreted from mature DCs in combination with NK-secreted GM-CSF and interferon- γ (IFN γ) can combine to recruit T lymphocytes and may induce Th1 cell polarization, thereby inducing development of a potent cytotoxic T cell response and clearance of SARS-CoV-2-infected cells. Confirmation of Th1 polarization vs. a mixed Th1/Th2 or predominantly Th2 response

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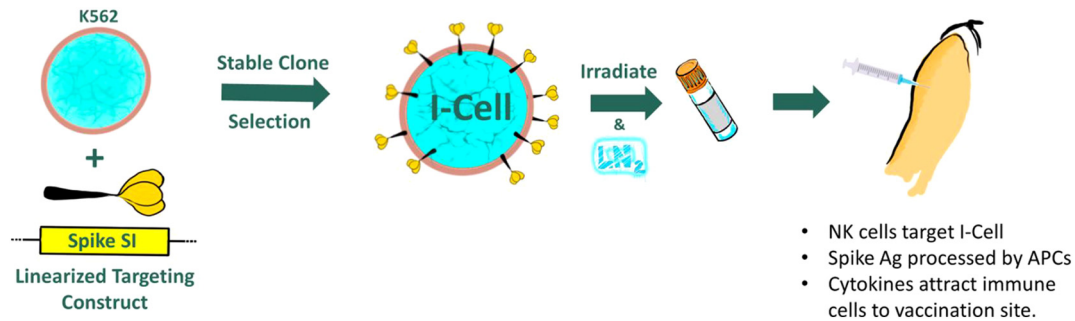


Fig. 1. Targeted expression of SARS-CoV-2 Spike immunogen and resulting I-cell vaccine candidate.

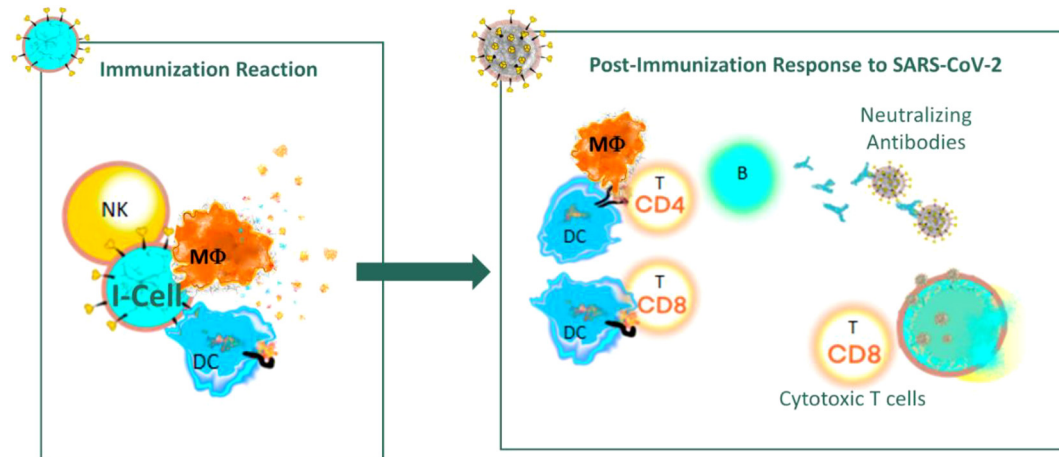


Fig. 2. Elicitation of humoral and cellular immune responses following virus decoy cell vaccination.

is currently underway in preclinical immunization models. Previous efforts to develop respiratory virus vaccines to protect against Respiratory Syncytial Virus (RSV) and SARS-related disease have demonstrated the potential clinical benefits of eliciting a Th1 adaptive immune response over the disease-exacerbating effects of a Th2 polarized response [3,4]. Immunization studies in mice with four candidate SARS vaccines (VLP, whole virus, and an rDNA-produced Spike protein) led to pulmonary immunopathology upon challenge with SARS virus, an effect that was signified by Th2 polarization in mice immunized with each candidate vaccine [4]. The decoy cell vaccine can drive the host cellular immune response toward Th1, generating both potent cytotoxic T cell immunity against the major determinant of SARS-CoV-2 cellular entry and pathogenesis (Fig. 2).

Utilizing pathogen-derived cellular immunogens (virus-encoded antigens) as initiators of NK-mediated immune responses might provide an exciting method to produce novel vaccine candidates. It is important to highlight the flexibility of the decoy cell vaccination strategy as additional cell vaccines can be generated in an expeditious manner once new antigens of interest have been identified. Thus, successful demonstration of the efficacy of this current anti-SARS-CoV-2 approach in clinical trials would pave the way for use of the decoy cell platform to address a large number of unmet medical needs in the clinic as we seek to protect populations from viral, parasitic, fungal and bacterial diseases.

Funding source

Sorrento Therapeutics is funding the discovery, development and clinical supply manufacturing of the vaccine platform mentioned in this communication.

CRediT authorship contribution statement

Henry Ji: Conceptualization. **Ying Yan:** Conceptualization. **Beibei Ding:** Conceptualization. **Wenzhong Guo:** Conceptualization. **Mark Brunswick:** Writing - review & editing. **Andreas Niethammer:** Original writing. **Williams SooHoo:** Visualization. **Robin Smith:** Writing - review & editing. **Alexis Nahama:** Writing - review & editing. **Yanliang Zhang:** Conceptualization.

Declaration of competing interest

The authors are employed (or Directors) of Sorrento Therapeutics, Inc. The current discovery strategy is being implemented for vaccine development under the I-Cell™ label and within Sorrento's cGMP manufacturing facilities.

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