

## Complete Genome Sequences of Four Isolated Bacteria from an Undergraduate Microbiology Course Using a Hybrid Assembly Approach

**Microbiology**<sup>®</sup>

**Resource Announcements** 

Ciara I. Sanders,<sup>a</sup>\* Christopher J. Ne Ville,<sup>a§</sup> <sup>(D)</sup>Paul M. Orwin<sup>a</sup>

AMERICAN SOCIETY FOR

MICROBIOLOGY

<sup>a</sup>Department of Biology, California State University, San Bernardino, San Bernardino, California, USA

**ABSTRACT** Three Gram-negative bacteria and one Gram-positive bacterium were isolated from environmental samples in an undergraduate microbiology class on the basis of antibiotic resistance. Isolate DNA was purified, sequenced, and assembled using a hybrid approach. Here, we report the genomes of *Acinetobacter johnsonii* CSUSB1, *Aeromonas hydrophila* CSUSB2, *Bacillus velezensis* CSUSB3, and *Comamonas thiooxydans* CSUSB4.

Four isolates were sequenced using the Oxford Nanopore Technologies MinION platform and the Illumina iSeq100 platform, and their genomes were assembled using a hybrid assembly approach. Initial isolations, 16S rRNA gene characterizations, and preliminary MinION sequencing were performed in an undergraduate medical microbiology course.

Acinetobacter johnsonii CSUSB1 was isolated from pond water in Highland, California. Aeromonas hydrophila CSUSB2 and Comamonas thiooxydans CSUSB4 were isolated from a water tank housing Alligator mississippiensis on the California State University, San Bernardino (CSUSB) campus. Pond and alligator water samples were plated on tryptic soy agar (TSA) plates and grown at 30°C for 7 days with five antibiotic disks. Isolates near the disks were restreaked and then retested for resistance. Colonies exhibiting resistance to the antibiotic disks (ampicillin, all isolates; erythromycin, CSUSB2 and CSUSB4; gentamicin and streptomycin, CSUSB2) were streaked for isolation on TSA plates. Single colonies were picked into 100  $\mu$ L of sterile water, boiled, and centrifuged for one minute at maximum speed. The universal 16S rRNA gene primers 27f and 1492r were used to amplify the 16S rRNA gene (1) from the supernatant, and then the gene was sequenced (Retrogen Inc., San Diego, CA) using the universal 16S rRNA gene primer 530f (1). The genera of the isolates were determined using nucleotide BLAST (2), and the best hits were as follows: CSUSB1, GenBank accession number MK184297; CSUSB2, GenBank accession number NR \_118547; CSUSB4, GenBank accession number NR \_029161.

Genomic DNA (gDNA) for sequencing was extracted from a single colony after overnight growth in LB broth. A high-molecular-weight DNA extraction protocol (3) was used for the MinION sequencing, and the Wizard SV gDNA kit (Promega, Madison, WI) was used for the iSeq100 sequencing. Genomic libraries were prepared using the rapid barcoding library preparation kit SQK-RBk004 (Oxford Nanopore Technologies, Oxford, UK) for the MinION sequencing and the Nextera XT library preparation kit (Illumina, San Diego, CA) for the iSeq100 sequencing.

Default parameters were used for all of the following software unless otherwise specified. MinION reads (R9.4.1 flow cell) were demultiplexed with Deepbinner v 0.2.0 (4) and base called with Guppy Basecaller v 2.3.1 using the high-accuracy flip-flop algorithm (5), and adapters were removed with Porechop (Galaxy v 0.2.3) (6). FastQC (Galaxy v 0.72) and Trimmomatic (Galaxy v 0.36.5) were used to identify and clip iSeq100 reads with quality scores of <25 (7). Assembly of the long-read-only data sets

**Editor** J. Cameron Thrash, University of Southern California

**Copyright** © 2022 Sanders et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International license.

Address correspondence to Paul M. Orwin, porwin@pacific.edu.

\*Present address: Ciara I. Sanders, Biology Department, California State University, San Marcos, San Marcos, California, USA. §Present address: Christopher J. Ne Ville,

Molecular and Computational Biology Department, University of Southern California, Los Angeles, California, USA.

◊Present address: Paul M. Orwin, Biological Sciences Department, University of the Pacific, Stockton, California, USA.

The authors declare no conflict of interest.

Received 15 October 2021 Accepted 26 January 2022 Published 10 February 2022

Image: Second S	TABLE 1 G	ienomic data									
Length Coverage Nanopore Nanopore Illumina content predicted best genome match   Solate (b) (x) reads (so) (molti size(s) (bp)) genes (ANI [%])   Solate (b) (x) reads (so) contig size(s) (bp) genes (ANI [%])   CSUSB1 3,655,004 450 17,733 4,674 373,905 41.4 3,562,289,84,108, 3,546 GCA_004337595.1 (96.1)   CSUSB2 4,923,206 272 98,607 41.4 3,562,289,84,108, 3,546 GCA_004337595.1 (96.1)   CSUSB3 4,923,706 272 98,607 41.4 3,562,289,84,108, 3,546 GCA_001395955.1 (97.6)   CSUSB3 4,928,756 491,68,825, 4,538 GCA_0013959595.1 (97.6) 7,470   CSUSB3 4,088,756 499 7,470 7,470 7,470 6GA_012647845.1 (99.17) 6GA_012647845.1 (99.17) 65,254 6GA_00039145.2 (96.62) 7,470 7,470 7,470 7,470 7,411,187,138,008 5				No. of		No. of	gC		No. of	GenBank assembly accession no. for	
CSUSB1 3,655,004 450 17/733 4,674 373,905 41.4 3,562,289,84,108, 3,546 GCA_004337595.1 (96.1)   CSUSB2 4,923,206 272 98,634 5,384 914,196 61.4 4,846,911,68,825, 4,538 GCA_001895965.1 (97.6)   CSUSB2 4,923,206 272 98,634 5,384 914,196 61.4 4,846,911,68,825, 4,538 GCA_001895965.1 (97.6)   CSUSB3 4,088,756 499 30,119 5,729 634,360 46.3 4,088,756 4,041 GCA_012647845.1 (99.17)   CSUSB4 5,642,591 318 272,391 3,217 440,231 61.4 5,441,187,138,008, 5,254 GCA_00093145.2 (96.62)   CSUSB4 5,642,591 318 272,391 3,217 440,231 61.4 5,441,187,138,008, 5,254 GCA_00093145.2 (96.62)	Isolate	Length (bp)	Coverage (×)	Nanopore reads	Nanopore read N <sub>50</sub> (bp)	Illumina reads	content (%)	Contig size(s) (bp)	predicted genes	best genome match (ANI [%])	GenBank accession no.
CSUSB2 4,923,206 272 98,634 5,384 914,196 61.4 4,846,911,68,825, 4,538 GCA_001895965.1 (97.6) 7,470 7,470 CSUSB3 4,088,756 499 30,119 5,729 634,360 46.3 4,088,756 4,041 GCA_012647845.1 (99.17) CSUSB4 5,642,591 318 272,391 3,217 440,231 61.4 5,441,187,138,008, 5,254 GCA_000093145.2 (96.62)	CSUSB1	3,655,004	450	17,733	4,674	373,905	41.4	3,562,289, 84,108,	3,546	GCA_004337595.1 (96.1)	CP083947 to
CSUSB3 4,088,756 499 30,119 5,729 634,360 46.3 4,088,756 4,041 GCA_012647845.1 (99.17) CSUSB4 5,642,591 318 272,391 3,217 440,231 61.4 5,441,187,138,008, 5,254 GCA_00093145.2 (96.62) 57,673,3,429,	CSUSB2	4,923,206	272	98,634	5,384	914,196	61.4	8,607 4,846,911, 68,825, 7 470	4,538	GCA_001895965.1 (97.6)	CP083949 CP083944 to CP083946
CSUSB4 5,642,591 318 272,391 3,217 440,231 61.4 5,441,187,138,008, 5,254 GCA_000093145.2 (96.62) 57,673, 3,429,	<b>CSUSB3</b>	4,088,756	499	30,119	5,729	634,360	46.3	4,088,756	4,041	GCA_012647845.1 (99.17)	CP083943
2,294	CSUSB4	5,642,591	318	272,391	3,217	440,231	61.4	5,441,187, 138,008, 57,673, 3,429, 2,294	5,254	GCA_000093145.2 (96.62)	CP083938 to CP083942

using Unicycler (Galaxy v 0.4.8.0) (6) and subsequent BLAST searches led to the discovery of *Acinetobacter* isolate stock culture contamination with *Bacillus velezensis* CSUSB3. The mixed stock culture was restreaked on TSA plates, and isolated colonies were restreaked for isolation. Single colonies were picked, and gDNA was reisolated. Both isolate gDNA samples were sequenced on the iSEQ100 system, and isolated CSUSB3 gDNA was resequenced on the MinION system (R9.4.1 flow cell). The new CSUSB3 MinION reads were used as known contamination in the Unicycler (Galaxy v 0.4.8.0) long-read alignment parameters. This allowed us to use the data from the original MinION run to assemble the genome of CSUSB1.

A total of 12 circular contigs (4 chromosomes and 8 plasmids) were assembled by Unicycler (Galaxy v 0.4.8.0) (8) (Table 1). GToTree (9) was used to identify each isolate's closest relative, and then the two-way average nucleotide identity (ANI) was used to confirm this designation (10) (Table 1).

**Data availability.** The BioProject accession number is PRJNA767399. The GenBank accession numbers are as follows: *Acinetobacter johnsonii* CSUSB1, CP083947 to CP083949; *Aeromonas hydrophila* CSUSB2, CP083944 to CP083946; *Bacillus velezensis* CSUSB3, CP083943; *Comamonas thiooxydans* CSUSB4, CP083938 to CP083942. The BioSample accession numbers are as follows: *Acinetobacter johnsonii* CSUSB1 (TaxID number 40214), SAMN21540781; *Aeromonas hydrophila* CSUSB2 (TaxID number 644), SAMN21540782; *Bacillus velezensis* CSUSB3 (TaxID number 492670), SAMN21540783; *Comamonas thiooxydans* CSUSB4 (TaxID number 363952), SAMN21540784.

## ACKNOWLEDGMENTS

We thank Nicholas Bodman, who cultured CSUSB1 from a local pond, and Tomas Owerkowicz, who allowed us supervised access to his alligators.

## REFERENCES

- Fukuda K, Ogawa M, Taniguchi H, Saito M. 2016. Molecular approaches to studying microbial communities: targeting the 16S ribosomal RNA gene. J UOEH 38:223–232. https://doi.org/10.7888/juoeh.38.223.
- Altschul SF, Gish W, Miller W, Myers EW, Lipman DJ. 1990. Basic local alignment search tool. J Mol Biol 215:403–410. https://doi.org/10.1016/ S0022-2836(05)80360-2.
- Quick J, Loman NJ. 2018. DNA extraction strategies for Nanopore sequencing, p 91–105. *In* Deamer D, Branton D (ed), Nanopore sequencing: an introduction. World Scientific, Singapore.
- Wick RR, Judd LM, Holt KE. 2018. Deepbinner: demultiplexing barcoded Oxford Nanopore reads with deep convolutional neural networks. PLoS Comput Biol 14:e1006583. https://doi.org/10.1371/journal.pcbi.1006583.
- Wick RR, Judd LM, Holt KE. 2019. Performance of neural network basecalling tools for Oxford Nanopore sequencing. Genome Biol 20:129. https:// doi.org/10.1186/s13059-019-1727-y.

- Wick RR, Judd LM, Gorrie CL, Holt KE. 2017. Completing bacterial genome assemblies with multiplex MinION sequencing. Microb Genom 3: e000132. https://doi.org/10.1099/mgen.0.000132.
- 7. Bolger AM, Lohse M, Usadel B. 2014. Trimmomatic: a flexible trimmer for Illumina sequence data. Bioinformatics 30:2114–2120. https://doi.org/10 .1093/bioinformatics/btu170.
- Wick RR, Judd LM, Gorrie CL, Holt KE. 2017. Unicycler: resolving bacterial genome assemblies from short and long sequencing reads. PLoS Comput Biol 13:e1005595. https://doi.org/10.1371/journal.pcbi.1005595.
- Lee MD. 2019. GToTree: a user-friendly workflow for phylogenomics. Bioinformatics 35:4162–4164. https://doi.org/10.1093/bioinformatics/btz188.
- Jain C, Rodriguez-R LM, Phillippy AM, Konstantinidis KT, Aluru S. 2018. High throughput ANI analysis of 90K prokaryotic genomes reveals clear species boundaries. Nat Commun 9:5114. https://doi.org/10.1038/s41467 -018-07641-9.