



## **Review The Role of Macrophage in the Pathogenesis of Osteoporosis**

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**Abstract:** Osteoporosis is a systemic disease with progressive bone loss. The bone loss is associated with an imbalance between bone resorption via osteoclasts and bone formation via osteoblasts. Other cells including T cells, B cells, macrophages, and osteocytes are also involved in the pathogenesis of osteoporosis. Different cytokines from activated macrophages can regulate or stimulate the development of osteoclastogenesis-associated bone loss. The fusion of macrophages can form multinucleated osteoclasts and, thus, cause bone resorption via the expression of IL-4 and IL-13. Different cytokines, endocrines, and chemokines are also expressed that may affect the presentation of macrophages in osteoporosis. Macrophages have an effect on bone formation during fracture-associated bone repair. However, activated macrophages may secrete proinflammatory cytokines that induce bone loss by osteoclastogenesis, and are associated with the activation of bone resorption. Targeting activated macrophages at an appropriate stage may help inhibit or slow the progression of bone loss in patients with osteoporosis.

Keywords: osteoporosis; macrophage; cytokine; chemokine; estrogen

### 1. Introduction

Osteoporosis is a systemic skeletal disorder characterized by a generalized increase in bone fragility that results in fractures of the hip, spine, or wrist. There are many risk factors associated with the progression of osteoporosis, including advanced age, low body-mass index, long-term glucocorticoid therapy, history of smoking, family history of hip fracture, excessive alcohol intake and previous fragility fracture. Osteoporosis occurs in all populations with a greater prevalence in postmenopausal women [1]. Women with osteoporosis have poorer musculoskeletal status than women without osteoporosis [2]. Osteoporosis is associated with limitations of daily activities, an increase in the occurrence of falls, and a consequent increased risk of fracture. The etiology of osteoporotic fractures is complex and the incidence of osteoporotic fractures is high among people aged 50 to 54 years [3]. A higher mortality is observed in these patients after osteoporotic fractures occur [4]. Osteoporotic fracture is associated with significant morbidity, mortality, poor quality of life, and increasing health

care costs. Therefore, the pathogenesis of bone loss or fracture is an important issue for the prevention of osteoporosis.

### 2. Osteoblast, Osteoclast and Osteocyte in Bone Formation and Homeostasis

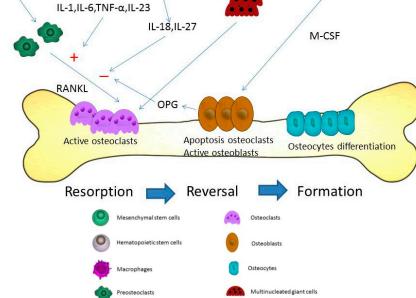
Osteoblasts originate from the mesenchymal stem cells and are important for the progression of bone formation. Co-expression of bone-specific alkaline phosphatase, type I collagen, and non-collagenous matrix proteins is observed in mature osteoblasts. Transforming growth factor- $\beta$  $(TGF-\beta)$  has an important role in bone formation by enhancing osteoblast proliferation [5]. TGF- $\beta$  can also block osteoblast apoptosis and recruit osteoblastic precursors to the bone surface [6–8]. interleukin (IL)-4, IL-10, IL-13, and IL-18 may induce elevation of osteoprotegerin (OPG) and reduction of receptor activator of NF- $\kappa$ B ligand (RANKL) [9]. Final differentiation of osteoblasts can form osteocytes, which become embedded in the mineralized matrix. The processes of bone formation by osteoblasts are enhanced by administration of vitamin D and parathyroid hormone. The Wnt/ $\beta$ -catenin is important for osteoblastic differentiation in skeletal biology and disease [10]. Dickkopf-1 and sclerostin are Wnt inhibitors and can regulate the expression of Wnt/ $\beta$ -catenin in osteoblasts [11]. More than 90% of bone cells are osteocytes, which release chemicals to the bone surface that attract osteoclasts. Osteocytes may secrete sclerostin to limit further bone formation by osteoblasts, and play a major role in matrix mineralization [12]. Bone remodeling involves coupling and regulation of osteoclasts and osteoblasts. Besides these resorptive and formative cells, T-cells, B-cells and macrophages may also influence the immune system and bone loss [13]. The bone remodeling cycle is shown in Figure 1.

### 3. Macrophages in the Pathogenesis of Osteoporosis

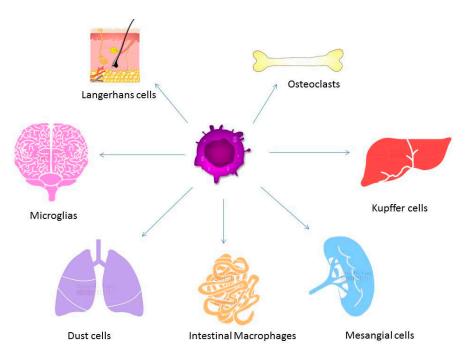
In the process of bone remodeling, different states of resorption, reversal, and formation are found. Activated monocytes or bone marrow macrophage precursors adhere to the bone surface to form multinucleated osteoclasts [14]. The bone remodeling cycle is regulated by local and systemic factors. Osteoclasts and osteoblasts are both important for the pathogenesis and progression of osteoporosis. Osteoclasts induce bone resorption and osteoblasts are associated with bone formation. Normal bone quality involves a neutral balance between resorption and formation. A multinucleated osteoclast is differentiated from the mononuclear osteoclast precursor of hemopoietic stem cell. RANKL and macrophage-colony stimulating factor (M-CSF) can induce the proliferation and activation of osteoclasts via the receptor, RANK [15,16]. Therefore, the balance between resorption and formation determines the progression of osteoporosis. M1 macrophages are associated with exacerbation of inflammation and express proinflammatory cytokines. M2 macrophages are associated with anti-inflammatory reactions through the expression of anti-inflammatory cytokines [17]. When macrophages are exposed the stimulation of RNAKL, macrophages may induce osteolcastogenesis and lead to increased M1/M2 ratio in ovariectomized mice. Estrogen can protect M2 macrophage from RANKL stimulation through estrogen receptor  $\alpha$  and the downstream blockage of NF- $\kappa$ B p65 nuclear translocation [18]. The blocking of estrogen deficiency-mediated M2 macrophage osteoclastogenesis by reducing the M1/M2 ratio may be a potential therapeutic target in treating postmenopausal osteoporosis. Macrophages play a major role in the activation and formation of osteoclasts and are differentiated from monocytes via M-CSF [19]. The different presentations of macrophages in different organs are shown in Figure 2. Besides the differentiation of osteoclasts from the macrophage lineage, macrophage precursors also differentiate into monocytes, macrophages, and dendritic cells. The activation of macrophages may induce the elevation of interferon- $\gamma$  (IFN- $\gamma$ ), IL-1, tumor necrosis factor- $\alpha$  (TNF- $\alpha$ ), complement proteins, and prostaglandins levels [20]. Macrophages are important for the pathogenesis of osteoporosis [19].

M-CSF





**Figure 1.** Microdamages in the bone-remodeling units of cancellous or cortical bone induced by osteoclasts. Osteoclasts may be activated by different cytokines including receptor activator of NF- $\kappa$ B ligand (RANKL), interleukin (IL)-1, IL-6, and macrophage-colony stimulating factor (M-CSF) in the resorption state. After the resorption process, the reversal state progresses. In the reversal state, apoptosis of osteoclasts may be induced to stop the bone resorption. The replacement of osteoblasts is observed at the same time. The activated osteoblasts refill the resorption pits and tunnels on the bone surface. In the formation state, osteoblasts directly adhere to the bone surface and progressively form into osteocytes. The proliferation of osteocytes can use these lacuna-canalicular networks to connect within the bone matrix. The cytokines from macrophages including IL-6, tumor necrosis factor-α (TNF-α), IL-23, IL-18, and IL-27 can induce and inhibit osteoclastogenesis through RANKL in bone remodeling. M-CSF is the most important cytokine in the initial stage of macrophage differentiation from hematopoietic stem cells. Different phases are observed during the formation of the multinucleation. RANKL is the major cytokine that stimulates osteoclasts into mature multinucleated osteoclasts. The cytokines, IL-4 and IL-13, may induce macrophages to form multinucleated giant cells during the course of bone resorption.



**Figure 2.** Macrophages exist in different tissues including lung, liver, and brain and have different functions. Different forms of macrophages include Kupffer cells in the liver, alveolar macrophages in the lung, osteoclasts in the bone, and microglia in the brain.

### 4. The Cytokines from Macrophages Contribute to the Process of Osteoporosis

Macrophages play a major role in the innate and adaptive immune system. Differentiation of macrophages can be found in various tissues including liver, lung, brain, and bone marrow. Macrophages are divided into M1, which express proinflammatory mediators, and M2 that are involved in anti-inflammatory reactions [21]. In the innate immune system, macrophages can execute phagocytosis and opsonization [22]. Different cell receptors of CD14,  $Fc\gamma$ , and CD25 are found in macrophages. Major histocompatibility complex class II molecules and CD23 are also found. These receptors are important for the progression of phagocytosis. IL-4 and IFN- $\gamma$  can regulate the different functions of macrophages. Macrophages, in turn, may be activated to induce IL-6, TNF- $\alpha$ , IFN- $\gamma$ , complement protein, and prostaglandins in the immune system. The cytokines expressed by macrophages associated with stimulation or inhibition of osteoclastogenesis include IL-6, IL-18, IL-23, IL-27, and TNF- $\alpha$  [23]. IL-6, a proinflammatory cytokine, can activate osteoclastogenesis [24]. During the inflammation state, proinflammatory cytokines including TNF $\alpha$ , IL-1 $\beta$ , and IL-6, may promote the differentiation and activation of osteoclasts [25]. IL-18 secreted via macrophages may regulate the Th1 differentiation and the IFN- $\gamma$  production, and is an inhibitor of the TNF- $\alpha$  mediated osteoclastogenesis [26]. Among osteoporotic women, decreased levels of serum IL-18 binding protein and elevated levels of serum IL-18 are observed [27,28]. IFN- $\gamma$  has a dual role in osteoclasts including the promotion of osteoblast differentiation and inhibition of bone marrow adipocyte formation in different stages [29]. IFN- $\gamma$  can activate macrophages, but macrophages can secrete IL-18 to regulate the IFN- $\gamma$  production. IL-23 has been shown to activate osteoclasts [30]. Adding IL-23 to bone marrow stromal cells led to an increased differentiation towards the osteoblast lineage [31]. In the femur of a rat osteoporosis model, IL-23 is reduced after adequate estrogen therapy for improvement of bone mineral density [32]. IL-27 may suppress the expression of RANKL in Th17 cells and CD4+ T cells [33]. IL-27 also inhibited osteoblast apoptosis through increased Egr-2 expression [34]. TNF- $\alpha$  from macrophages may induce indirect osteoclastic activation through RANKL in bone remodeling [35]. Therefore, inflammatory arthritis, such as rheumatoid arthritis, can induce progressive bone loss when the disease is poorly controlled. The different cytokines of macrophages associated with osteoporosis are shown in Figure 1.

#### 5. The Fusion of Macrophages/Monocytes to Form Multinucleated Cells—Osteoclasts

Macrophages have the ability to fuse and develop into multinucleated cells during an acute infection and inflammation state. The formation of granuloma induced by tuberculosis infection and vasculitis may result in these multinucleated giant cells. In systemic inflammation, macrophages can release reactive oxygen and reactive nitrogen species to induce the formation of multinucleated giant cells. M-CSF is the most important cytokine in the initial stage of macrophages differentiation from hematopoietic stem cells. Different phases are observed during the formation of multinucleated cells. RANKL is the major cytokine responsible for the stimulation of osteoclasts into mature multinucleated giant cells during the course of bone resorption [37–39]. The proliferation and differentiation of macrophages may be stimulated by M-CSF. After the stimulation via M-CSF, RANKL may activate the proliferation of osteoclasts. Fusion-competent osteoclasts may be induced by RANKL. Multinucleated giant cells originate from this fusion of cells, which develops to form multinucleated osteoclasts or giant cells [19]. The fusion of macrophages to form multinucleated osteoclasts is shown in Figure 1.

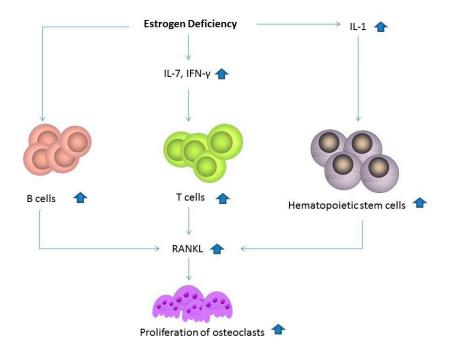
### 6. Different Cytokines for the Pathogenesis of Osteoporosis

RANKL can be expressed by different cells including T cells, B cells, bone-marrow stromal cells, and bone-forming osteoblasts. Mice with depletion of RANKL gene show severe osteopetrosis and lack mature circulating osteoclasts [40]. The differentiation of osteoclasts may be inhibited by the decoy receptor OPG, which is produced by osteoblasts [41]. Proinflammatory cytokines including IL-1 and TNF- $\alpha$  can stimulate osteoclastogenesis in vitro [42]. Other osteoclastogenic cytokines include IL-6, IL-8, IL-15, IL-17, and IFN- $\gamma$  [9,43]. High dosage of IFN- $\gamma$  may promote the differentiation of osteoclasts, and the effect of bone loss is enhanced in situations of estrogen deficiency [44,45]. The immune response in osteoclastogenesis via IFN- $\gamma$  include activation of RANKL/RANK pathway and promotion of fused mononucleated osteoclasts [29]. In patients with rheumatoid arthritis (RA), activated T cells can directly trigger osteoclastogenesis through RANKL/RANK/OPG pathway [46,47]. Therefore, juxta-articular osteopenia of both hands and osteoporotic fracture are usually found during the disease course of RA. The role of T cells in regulating osteoclastogenesis is associated with the formation of osteoclasts. B cells may participate in osteoclastogenesis by expression of RANKL for osteoclast differentiation and serve as osteoclast progenitors [48]. Osteoclast-associated receptor may be expressed by macrophages or monocytes in order to modulate the innate and adaptive immune response [49].

### 7. Estrogen Deficiency Induced the Expression of Different Cytokines in Osteoporosis

Estrogen can directly inhibit osteoclastic bone resorption by inducing apoptosis of osteoclasts [50]. Estrogen may induce osteoblast differentiation in bone formation by binding the estrogen receptor through the upregulation of PACE4 expression [51], and it also has an anabolic effect on the function of osteoblasts [52]. Estrogen serves different biological functions in the regulation of osteogenic differentiation with involvement of the Wnt/ $\beta$ -catenin signaling pathway [53]. Estrogen loss may also influence the immune system through upregulation of T and B cells [54]. Higher expression of circulating IL-1, IL-7, and IFN- $\gamma$  are found in patients with estrogen withdrawal [55,56]. Estrogen deficiency can stimulate T-cell activation and production of pro-osteoclastogenic cytokines. The levels of follicle-stimulating hormone (FSH) are increased during the development of estrogen deficiency. FSH receptors are present on osteoclasts, osteoclast precursors, and mesenchymal stem cells, and promote osteoclast differentiation, activity, and survival [57]. The net effect of estrogen deficiency on the bone is an increased activation of bone remodeling and osteoclasts. The bone loss induced by estrogen deficiency has a complex mechanism with predominant involvement of the immune system rather than a direct action of estrogen on bone cells [56]. The possible mechanism underlying the

association of estrogen and bone loss is shown in Figure 3. Therefore, estrogen deficiency is associated with bone loss by influencing activity and formation of osteoclasts or proliferation of osteoblasts.



**Figure 3.** Estrogen loss may also influence the immune system by the upregulation of T and B cells. Higher expression of circulating IL-1, IL-7, and IFN- $\gamma$  is found in patients with estrogen withdrawal. Estrogen deficiency can stimulate T-cell activation and production of pro-osteoclastogenic cytokines.

# 8. The Activation and Differentiation of Macrophages to Osteoclasts in the Development of Osteoporosis

The differentiations of osteoclasts are both from hematopoietic precursor cells and macrophage lineage [58]. Osteoclastogenesis from macrophages is activated by M-CSF and RANKL, and the blockage of RANKL signaling pathway may prevent the progression of osteoporosis in mice models [59,60]. The bone loss in ovariectomized mice is also associated with osteoclast differentiation of bone marrow-derived macrophages [61]. The expression of TNF receptor associated factor (TRAF) 6 and TRAF3 are both important in the differentiation of early osteoclasts in osteoclast's precursors and macrophages. The level of TRAF3 protein decreases in bone and bone marrow with aging [62]. TRAF3 has been revealed to be a powerful negative regulator in B cells [63]. Proliferation of B cells can induce the expression of RANKL. Therefore, TRAF3 may be a target for the prevention of immune related bone loss.

M1 macrophages can induce exacerbation of inflammation and are associated with the development of osteoporosis. Bisphosphonates are used for the treatment of osteoporosis, and associated osteonecrosis of the jaw is an unusual complication. The related osteonecrosis may be due to an abnormal activation of M1 macrophages [64]. The differentiation of osteoclast from macrophage can be activated by M-CSF and RANKL, and RANKL signaling pathway activates the major regulator of NFATc1 [65]. Down-regulating NFATc1 may inhibit RANKL-mediated osteoclastogenesis [66].

### 9. The Effect of Macrophages in Osteoblasts

Osteoblasts are the major cells responsible for bone formation and originated from mesenchymal stem cells. The osteoblastogenesis of mature osteoblasts is controlled by different transcription factors. Mature osteoblasts may differentiate to osteocytes and lining cells. In the cycle of bone remodeling, macrophages have a major role in the induction of osteoclastogenic differentiation during the state of resorption. The differentiation of osteoclasts serves as a regulatory step in the formation of osteoblasts

and osteocytes (Figure 1). However, young macrophage cells present during the rejuvenation process and are associated with bone repair in mice [67]. Adequate activation of osteoblasts may be found during the progression of bone repair. Activated macrophages are the most likely candidates to promote bone formation and have also been implicated in tissue repair processes [17]. Osteoblasts are important for the progression of bone formation under adequate activation and proliferation. IL-4 and IL-13 can recruit and activate osteoblasts for the progression of bone resorption and healing [68,69]. IL-4 and IL-13 may inhibit osteoclast differentiation and bone resorption via activation of the receptors on osteoblasts and osteoclasts that affect the RANKL/RANK/OPG system [70]. In osteoblasts, IL-4 and IL-13 can suppress the production of prostaglandin through the induction of IL-1. TNF- $\alpha$  has the effect of stimulating osteoblast chemotaxis in vitro [71]. However, osteoblasts are regulated by different cytokines during the phase of bone resorption. IL-6 may inhibit osteoblast differentiation with disrupting the balance of healthy bone turnover [72]. TNF- $\alpha$  may suppress osteoblast differentiation in RA patients [73]. Inflammatory cytokines can activate osteoclasts and promote bone resorption. At the same time, inhibition or attenuation of the osteoblasts is also observed. Therefore, adequate activation and regulation of osteoblasts are important for the pathogenic presentation of bone remodeling. The balance between bone formation and resorption is regulated by different cytokines.

### 10. The Activation and Regulation of Chemokines in Macrophages-Associated Osteoporosis

Chemokines are important for the migration of circulating hematopoietic cells into different tissues. CXCL-8 and CCL-20 are elevated in inflammatory arthritis, and they may enhance osteoblast-mediated osteoclastogenesis through the production of IL-6 [74]. The active expression of CCL-6 is observed during the differentiation of osteoclasts, and can progress to bone loss in vitro [75]. CCL-4 can inhibit migration of osteoclast precursor cells from the bone marrow into the bone surfaces, and cannot affect the differentiation of osteoclasts [76]. Elevation of CXCL-10 is found during osteoclast differentiation, and CXCL-10-neutralizing antibodies can reduce the effect of osteoclastogenesis [77,78]. Higher expression of CX3CR-1 is found on osteoclast precursor cells, and CX3CL-1 from osteoblasts can bind the receptor of CX3CR-1 and induce migration and adhesion of osteoclasts to the bone marrow [79]. During activation of osteoclasts, expression and activation of CCL-3 is observed in vitro [75]. Therefore, the chemokines associated with bone resorption include CXCL-8, CCL-20, CCL-6, CCXL-10, CCL-3, while CCCL-4 is associated with bone formation. Among the inflammatory progression, most chemokines serve as a mediator for osteoclastogenics.

### 11. The Effect of Macrophages in Osteocytes

The role of osteocytes is important in the bone remodeling cycle and may be regulated by osteoclasts and osteoblasts. The differentiation of osteocytes is influenced by osteoblasts. Osteocytes form the major structures of cortical and calcaneus bone and have different physiological function for bone resorption or formation. Osteocytes promote the production of RANKL and decrease OPG expression. Consequently, the RANKL/OPG ratio increases, osteoclastogenesis occurs, and the enhancement of bone resorption in the unloading activity is seen. The loss of RANKL in osteocytes may increase cancellous bone mass in the osteogenesis imperfecta mouse model [80]. Activation of macrophages can induce the production of proinflammatory cytokines including IL-1, IL-6 and TNF- $\alpha$ , which may be important for the inflammatory bone loss [81]. After stimulation of IL-1 and TNF- $\alpha$ , active fibroblast growth factor-23 secretion by osteocytes may contribute to hypophosphataemia during sepsis [82]. IL-1 may promote loss of osteocyte viability through NF-kB/RANKL signaling [83]. Soluble IL-6 may promote bone formation or osteoclastogenesis in different levels among the progression of normal bone growth and remodeling [84]. The osteoclastogenesis-supporting activity is reduced in zoledronate-treated osteocyte-like MLO-Y4 cells in the presence of IL-6 neutralizing therapy [85]. Bisphosphonate-related osteonecrosis of the jaw may be progressing by this pathway. Immunohistochemical staining of TNF- $\alpha$ , IL-6, and sclerostin in osteocytes are statistically higher in the patient with spinal cord injury [86]. Macrophages may inhibit osteocyte viability through the effect of TNF- $\alpha$  and IL-6 in the model of monosodium urate crystal-induced inflammation [87]. During the stage of bone repair, expression of IL-18 from macrophages may serve an important role by increasing expression during bone formation or decreasing expression in the model of bisphosphonate-related osteonecrosis of the jaw [88,89]. Osteocytes may be regulated by macrophage secreting inflammatory cytokines (TNF- $\alpha$ , IL-6) to influence bone turnover. However, osteocytes suppress sclerostin expression with the consequent induction of Wnt signaling and enhanced bone formation in the loading activity. The physiological effect of osteocyte-intrinsic mTORC1 signaling may decrease trabecular bone mass [90]. In different conditions, osteoclastogenesis or osteoblastogenesis influence bone loss. In animal models, N-methyl pyrrolidone may prevent estrogen deficiency, induce increased expression of sclerostin, and decrease the progression of osteoporosis [91]. The stimulation of bone formation is induced after anti-sclerostin antibody treatment in ovariectomized rats [92,93]. Sclerostin from osteocytes may suppress bone formation by decreasing osteoblast-related autophagy or apoptosis in multiple myeloma-related osteolytic bone diseases [94]. In the mouse model of estrogen deficiency, the condition of autophagy in osteocytes is associated with oxidative stress [95]. Aging, estrogen deficiency, and steroid may stimulate oxidative stress and induce the abnormal apoptosis of osteocytes and osteoblasts with consequent progression of osteoporosis or bone loss [96]. IFN- $\beta$  is an anti-osteoclastogenic cytokine that activates the signaling of Toll-like receptor 5 and can be secreted by osteocytes. Adequate activation and proliferation of osteocytes are important for bone formation. Blocking and inhibiting the abnormal apoptosis of osteocytes by low-dose risedronate can prevent bone loss in ovariectomized rats [97].

### 12. The Modulation of Insulin-Like Growth Factor-1 (IGF-1) in Macrophages

During inflammation, macrophages are the major producers of cytokines. Activated macrophages are able to promote osteoclastogenesis for the progression of osteoporosis. IGF-1 can inhibit the activation of macrophages under the stimulation of IL-4 [98]. Overexpression of IGF-1 may delay the secretion of proinflammatory cytokines by macrophages in the model of skeletal muscle injury [99]. The limitation of activated macrophages by IGF-1 is through the PI3K/Akt signaling pathway [100]. IGF-1 serves as a key factor for anti-inflammation processes and macrophage modulation in models of murine skeletal muscle [101]. IGF-1 may activate the differentiation of osteoblasts and promote bone formation [102]. Therefore, IGF-1 has different effects during bone remodeling. IGF-I may stimulate both markers of bone formation and bone resorption, but a low dose of IGF-1 can promote bone formation [103]. Elevation of parathyroid hormone (PTH) is associated with the activation of bone turnover and is important to regulate bone remodeling. PTH receptor signaling can increase the RANKL/OPG ratio in osteoblasts and osteocytes. Therefore, primary and secondary hyperparathyroidism may increase the risk of osteoporosis related fracture. T lymphocytes express the PTH receptor and are essential for the development of osteoclastogenesis [104]. The direct effects of PTH on osteoblasts and osteocytes, and indirect actions on osteoclasts, promote both bone formation and bone resorption [105]. The interaction between T cells and macrophages is associated with osteoimmunology. The administration of PTH has different dose-dependent effects including bone formation or osteoclastic callus remodeling during fracture healing [106].

### 13. The Possible Therapy of Osteoporosis by Targeting Macrophages

Osteoporosis is treated by different traditional drugs. Calcium and vitamin D supplementation are used in the prevention and treatment of osteoporosis. Hormone replacement therapy is used in postmenopausal women with osteoporosis. Estrogen agonist-antagonists have antiresorptive effects on the bone and can reduce the risk of vertebral fractures in postmenopausal women. Bisphosphonates are most commonly used for the therapy of osteoporosis due to their effects in the suppression of osteoclast-mediated bone resorption. Strontium can compete with calcium to stimulate osteoblastic activity and decrease osteoclastic activity in the course of bone turnover. Parathyroid hormone therapy can increase bone formation and mass through parathyroid hormone-related protein receptor signaling. Denosumab is a fully human monoclonal antibody that binds RANK and inhibits the activity of RANKL. Decreasing the levels of RANKL may decrease the activity of osteoclasts.

The action of TRAF3 may prevent osteoporosis-related bone loss in inflammatory osteoporosis. RANKL can enhance TNF-induced osteoclast formation independently of TRAF6 and by degrading TRAF3 [107]. TRAF3 may be an agent to prevent bone loss. Increased expression of sclerostin is found during bone remodeling along with the elevation of RANKL, and inhibition of sclerostin expression may improve bone loss. Anti-sclerostin may serve as an agent for bone formation. Macrophages are activated to induce the progression of bone resorption during the cycle of bone remodeling. Adequate blockage of macrophage activation may suppress the progression of bone resorption and the pathogenesis of osteoporosis. Macrophages proliferate via M-CSF, and the adequate anti-M-CSF treatment may suppress the proliferation of macrophages. However, macrophages also have an important role in regulating the quality of bone repair. In patients with RA, tocilizumab adequately inhibits circulating IL-6, which leads to improved bone loss in osteoporosis [108]. Abatacept that interferes with T cell functioning, may increase bone mineral density at the femoral neck in patients with RA [109]. When the systemic inflammation is controlled by different biologics (including anti-TNF- $\alpha$ , anti-IL-6, anti-IL-1, T and B cell regulators), these anti-cytokine therapies are associated with the improvement of bone loss [110]. Macrophages were stimulated to promote osteoclastogenesis via M-CSF, TNF- $\alpha$ , IL-1 $\beta$ , IL-6, IL-17, and IL-23 [111]. When macrophage-related cytokines, which include IL-1 $\beta$ , IL-6, and TNF- $\alpha$ , are regulated, the progression of bone loss is improved. When the functions of B and T cells are regulated, the suppression of macrophage may develop through the inhibition of IFN- $\gamma$ . Therefore, osteoporosis-related bone loss is prevented or limited under the suppression of macrophages or inhibition of macrophage-related cytokines. IL-23 expression by macrophages can induce osteoclastogenesis with bone destruction during inflammation. Monotherapy against IL-23 has been used to manage post-menopausal bone loss [31]. IL-27 from macrophages can suppress the expression of RANKL and inhibit the apoptosis of osteoblasts. Treatment with IL-27 agent prevented the loss of trabecular micro-architecture and preserved cortical bone parameters [34]. However, macrophages have other effects on bone formation during fracture-related bone repair. Adequate activated macrophages are important for new bone formation.

### 14. Conclusions

Activated macrophages may secrete proinflammatory cytokine that induces bone loss by osteoclastogenesis, and are associated with activation of bone resorption. Furthermore, adequate activation of macrophages is needed during the state of bone repair and is also associated with bone formation. Different expression of activated macrophages including proinflammatory mediators in M1 and anti-inflammatory reactions in M2, can regulate the progression of bone resorption and bone formation. Osteoporosis is always associated with systemic inflammation via the abnormal activation of osteoclasts. Adequate inhibition of macrophages may improve the systemic inflammation associated with osteoporosis. Macrophages can affect osteoclasts, osteoblasts, and osteocytes during the progression of bone loss. Regulation of proinflammatory cytokine from macrophages may inhibit osteocalstogenesis. IL-4 and IL-23 may induce macrophages to form multinucleated giant cells during the phase of bone resorption. The pharmaceutical targets in macrophage polarization or macrophage related cytokine dysregulation may be an important issue to develop in the future.

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### Abbreviations

Receptor activator of NF-ĸB
Receptor activator of NF-ĸB ligand
Macrophage-colony stimulating factor
Transforming growth factor-β
Interferon- $\gamma$
Tumor necrosis factor- $\alpha$
Osteoprotegerin
Parathyroid hormone
Rheumatoid arthritis
Tumor necrosis receptor-associated factor

### References

- Hadji, P.; Klein, S.; Gothe, H.; Haussler, B.; Kless, T.; Schmidt, T.; Steinle, T.; Verheyen, F.; Linder, R. The epidemiology of osteoporosis—bone evaluation study (best): An analysis of routine health insurance data. *Dtsch. Arztebl. Int.* 2013, *110*, 52–57. [PubMed]
- 2. Cunha-Henriques, S.; Costa-Paiva, L.; Pinto-Neto, A.M.; Fonsechi-Carvesan, G.; Nanni, L.; Morais, S.S. Postmenopausal women with osteoporosis and musculoskeletal status: A comparative cross-sectional study. *J. Clin. Med. Res.* **2011**, *3*, 168–176. [CrossRef]
- 3. Johnell, O.; Kanis, J. Epidemiology of osteoporotic fractures. *Osteoporos. Int.* **2005**, *16*, S3–S7. [CrossRef] [PubMed]
- 4. Rachner, T.D.; Khosla, S.; Hofbauer, L.C. Osteoporosis: Now and the future. *Lancet* 2011, 377, 1276–1287. [CrossRef]
- Kassem, M.; Kveiborg, M.; Eriksen, E.F. Production and action of transforming growth factor-beta in human osteoblast cultures: Dependence on cell differentiation and modulation by calcitriol. *Eur. J. Clin. Investig.* 2000, 30, 429–437. [CrossRef]
- 6. Jilka, R.L.; Weinstein, R.S.; Bellido, T.; Parfitt, A.M.; Manolagas, S.C. Osteoblast programmed cell death (apoptosis): Modulation by growth factors and cytokines. *J. Bone Miner. Res.* **1998**, *13*, 793–802. [CrossRef]
- Lucas, P.A. Chemotactic response of osteoblast-like cells to transforming growth factor beta. *Bone* 1989, 10, 459–463. [CrossRef]
- 8. Kasagi, S.; Chen, W. Tgf-beta1 on osteoimmunology and the bone component cells. *Cell Biosci.* **2013**, *3*, 4. [CrossRef] [PubMed]
- 9. Zupan, J.; Jeras, M.; Marc, J. Osteoimmunology and the influence of pro-inflammatory cytokines on osteoclasts. *Biochem. Med.* (*Zagreb*) **2013**, *23*, 43–63. [CrossRef]
- 10. Baron, R.; Rawadi, G. Targeting the wnt/beta-catenin pathway to regulate bone formation in the adult skeleton. *Endocrinology* **2007**, *148*, 2635–2643. [CrossRef]
- 11. Canalis, E.; Giustina, A.; Bilezikian, J.P. Mechanisms of anabolic therapies for osteoporosis. *N. Engl. J. Med.* **2007**, *357*, 905–916. [CrossRef]
- Poole, K.E.; van Bezooijen, R.L.; Loveridge, N.; Hamersma, H.; Papapoulos, S.E.; Lowik, C.W.; Reeve, J. Sclerostin is a delayed secreted product of osteocytes that inhibits bone formation. *FASEB J.* 2005, 19, 1842–1844. [CrossRef]
- 13. Long, C.L.; Humphrey, M.B. Osteoimmunology: The expanding role of immunoreceptors in osteoclasts and bone remodeling. *Bonekey Rep.* **2012**, *1*. [CrossRef]
- 14. Kylmaoja, E.; Nakamura, M.; Tuukkanen, J. Osteoclasts and remodeling based bone formation. *Curr. Stem. Cell Res. Ther.* **2016**, *11*, 626–633. [CrossRef]
- 15. Chen, X.; Wang, Z.; Duan, N.; Zhu, G.; Schwarz, E.M.; Xie, C. Osteoblast-osteoclast interactions. *Connect. Tissue Res.* **2018**, *59*, 99–107. [CrossRef]
- 16. Matsuo, K.; Irie, N. Osteoclast-osteoblast communication. *Arch. Biochem Biophys* **2008**, 473, 201–209. [CrossRef]
- 17. Horwood, N.J. Macrophage polarization and bone formation: A review. *Clin. Rev. Allergy Immunol.* **2016**, *51*, 79–86. [CrossRef]

- Dou, C.; Ding, N.; Zhao, C.; Hou, T.; Kang, F.; Cao, Z.; Liu, C.; Bai, Y.; Dai, Q.; Ma, Q.; et al. Estrogen deficiency-mediated m2 macrophage osteoclastogenesis contributes to m1/m2 ratio alteration in ovariectomized osteoporotic mice. *J. Bone Miner. Res.* 2018, 33, 899–908. [CrossRef]
- 19. Pereira, M.; Petretto, E.; Gordon, S.; Bassett, J.H.D.; Williams, G.R.; Behmoaras, J. Common signalling pathways in macrophage and osteoclast multinucleation. *J. Cell Sci.* **2018**, *131*. [CrossRef]
- 20. Murray, P.J. Macrophage polarization. Annu. Rev. Physiol. 2017, 79, 541-566. [CrossRef]
- 21. Dey, A.; Allen, J.; Hankey-Giblin, P.A. Ontogeny and polarization of macrophages in inflammation: Blood monocytes versus tissue macrophages. *Front. Immunol.* **2014**, *5*, 683. [CrossRef]
- 22. Silva, M.T. When two is better than one: Macrophages and neutrophils work in concert in innate immunity as complementary and cooperative partners of a myeloid phagocyte system. *J. Leukoc. Biol.* **2010**, *87*, 93–106. [CrossRef]
- 23. Srivastava, R.K.; Dar, H.Y.; Mishra, P.K. Immunoporosis: Immunology of osteoporosis-role of t cells. *Front. Immunol.* **2018**, *9*, 657. [CrossRef]
- 24. Yun, T.J.; Chaudhary, P.M.; Shu, G.L.; Frazer, J.K.; Ewings, M.K.; Schwartz, S.M.; Pascual, V.; Hood, L.E.; Clark, E.A. Opg/fdcr-1, a tnf receptor family member, is expressed in lymphoid cells and is up-regulated by ligating cd40. *J. Immunol.* **1998**, *161*, 6113–6121.
- 25. Jung, Y.K.; Kang, Y.M.; Han, S. Osteoclasts in the inflammatory arthritis: Implications for pathologic osteolysis. *Immune Netw.* **2019**, *19*, e2. [CrossRef]
- Sims, N.A.; Green, J.R.; Glatt, M.; Schlict, S.; Martin, T.J.; Gillespie, M.T.; Romas, E. Targeting osteoclasts with zoledronic acid prevents bone destruction in collagen-induced arthritis. *Arthritis Rheum.* 2004, 50, 2338–2346. [CrossRef]
- 27. Mansoori, M.N.; Shukla, P.; Kakaji, M.; Tyagi, A.M.; Srivastava, K.; Shukla, M.; Dixit, M.; Kureel, J.; Gupta, S.; Singh, D. Il-18bp is decreased in osteoporotic women: Prevents inflammasome mediated il-18 activation and reduces th17 differentiation. *Sci. Rep.* **2016**, *6*, 33680. [CrossRef]
- 28. Maugeri, D.; Mamazza, C.; Lo Giudice, F.; Puglisi, N.; Muscoso, E.G.; Rizzotto, M.; Testai, M.; Bennati, E.; Lentini, A.; Panebianco, P. Interleukin-18 (il-18) and matrix metalloproteinase-9 (mmp-9) in post-menopausal osteoporosis. *Arch. Gerontol. Geriatr.* **2005**, *40*, 299–305. [CrossRef]
- 29. Tang, M.; Tian, L.; Luo, G.; Yu, X. Interferon-gamma-mediated osteoimmunology. *Front. Immunol* **2018**, *9*, 1508. [CrossRef]
- 30. Sato, K.; Suematsu, A.; Okamoto, K.; Yamaguchi, A.; Morishita, Y.; Kadono, Y.; Tanaka, S.; Kodama, T.; Akira, S.; Iwakura, Y.; et al. Th17 functions as an osteoclastogenic helper t cell subset that links t cell activation and bone destruction. *J. Exp. Med.* **2006**, 203, 2673–2682. [CrossRef]
- 31. Shukla, P.; Mansoori, M.N.; Singh, D. Efficacy of anti-il-23 monotherapy versus combination therapy with anti-il-17 in estrogen deficiency induced bone loss conditions. *Bone* **2018**, *110*, 84–95. [CrossRef]
- 32. Khera, A.; Kanta, P.; Kalra, J.; Dumir, D.; M, T. Resveratrol restores the level of key inflammatory cytokines and rankl/opg ratio in the femur of rat osteoporosis model. *J. Women Aging* **2018**, 1–13. [CrossRef]
- 33. Woodward, J. Regulation of haematopoietic progenitor cell proliferation and survival: The involvement of the osteoblast. *Cell Adh. Migr.* **2010**, *4*, 4–6. [CrossRef]
- Shukla, P.; Mansoori, M.N.; Kakaji, M.; Shukla, M.; Gupta, S.K.; Singh, D. Interleukin 27 (il-27) alleviates bone loss in estrogen-deficient conditions by induction of early growth response-2 gene. *J. Biol. Chem.* 2017, 292, 4686–4699. [CrossRef]
- 35. Boyce, B.F.; Xing, L. Functions of rankl/rank/opg in bone modeling and remodeling. *Arch. Biochem. Biophys.* **2008**, 473, 139–146. [CrossRef]
- 36. Udagawa, N.; Takahashi, N.; Akatsu, T.; Tanaka, H.; Sasaki, T.; Nishihara, T.; Koga, T.; Martin, T.J.; Suda, T. Origin of osteoclasts: Mature monocytes and macrophages are capable of differentiating into osteoclasts under a suitable microenvironment prepared by bone marrow-derived stromal cells. *Proc. Natl. Acad. Sci. USA* 1990, *87*, 7260–7264. [CrossRef]
- 37. Kao, W.J.; McNally, A.K.; Hiltner, A.; Anderson, J.M. Role for interleukin-4 in foreign-body giant cell formation on a poly(etherurethane urea) in vivo. *J. Biomed. Mater. Res.* **1995**, *29*, 1267–1275. [CrossRef]
- DeFife, K.M.; Jenney, C.R.; McNally, A.K.; Colton, E.; Anderson, J.M. Interleukin-13 induces human monocyte/macrophage fusion and macrophage mannose receptor expression. *J. Immunol.* 1997, 158, 3385–3390.

- 39. DeFife, K.M.; Jenney, C.R.; Colton, E.; Anderson, J.M. Cytoskeletal and adhesive structural polarizations accompany il-13-induced human macrophage fusion. *J. Histochem. Cytochem.* **1999**, 47, 65–74. [CrossRef]
- Kong, Y.Y.; Yoshida, H.; Sarosi, I.; Tan, H.L.; Timms, E.; Capparelli, C.; Morony, S.; Oliveira-dos-Santos, A.J.; Van, G.; Itie, A.; et al. Opgl is a key regulator of osteoclastogenesis, lymphocyte development and lymph-node organogenesis. *Nature* 1999, 397, 315–323. [CrossRef]
- 41. Khosla, S. Minireview: The opg/rankl/rank system. Endocrinology 2001, 142, 5050–5055. [CrossRef]
- 42. Garnero, P.; Darte, C.; Delmas, P.D. A model to monitor the efficacy of alendronate treatment in women with osteoporosis using a biochemical marker of bone turnover. *Bone* **1999**, *24*, 603–609. [CrossRef]
- 43. Greenblatt, M.B.; Shim, J.H. Osteoimmunology: A brief introduction. *Immune Netw.* **2013**, *13*, 111–115. [CrossRef] [PubMed]
- Mann, G.N.; Jacobs, T.W.; Buchinsky, F.J.; Armstrong, E.C.; Li, M.; Ke, H.Z.; Ma, Y.F.; Jee, W.S.; Epstein, S. Interferon-gamma causes loss of bone volume in vivo and fails to ameliorate cyclosporin a-induced osteopenia. *Endocrinology* 1994, 135, 1077–1083. [CrossRef]
- 45. Amarasekara, D.S.; Yun, H.; Kim, S.; Lee, N.; Kim, H.; Rho, J. Regulation of osteoclast differentiation by cytokine networks. *Immune Netw.* **2018**, *18*, e8. [CrossRef]
- 46. Kong, Y.Y.; Feige, U.; Sarosi, I.; Bolon, B.; Tafuri, A.; Morony, S.; Capparelli, C.; Li, J.; Elliott, R.; McCabe, S.; et al. Activated t cells regulate bone loss and joint destruction in adjuvant arthritis through osteoprotegerin ligand. *Nature* **1999**, *402*, 304–309. [CrossRef]
- 47. Geusens, P.; Lems, W.F. Osteoimmunology and osteoporosis. Arthritis Res. Ther. 2011, 13, 242. [CrossRef]
- Manabe, N.; Kawaguchi, H.; Chikuda, H.; Miyaura, C.; Inada, M.; Nagai, R.; Nabeshima, Y.; Nakamura, K.; Sinclair, A.M.; Scheuermann, R.H.; et al. Connection between b lymphocyte and osteoclast differentiation pathways. J. Immunol. 2001, 167, 2625–2631. [CrossRef]
- 49. Nemeth, K.; Schoppet, M.; Al-Fakhri, N.; Helas, S.; Jessberger, R.; Hofbauer, L.C.; Goettsch, C. The role of osteoclast-associated receptor in osteoimmunology. *J. Immunol.* **2011**, *186*, 13–18. [CrossRef]
- 50. Kameda, T.; Mano, H.; Yuasa, T.; Mori, Y.; Miyazawa, K.; Shiokawa, M.; Nakamaru, Y.; Hiroi, E.; Hiura, K.; Kameda, A.; et al. Estrogen inhibits bone resorption by directly inducing apoptosis of the bone-resorbing osteoclasts. *J. Exp. Med.* **1997**, *186*, 489–495. [CrossRef]
- 51. Kim, H.; Tabata, A.; Tomoyasu, T.; Ueno, T.; Uchiyama, S.; Yuasa, K.; Tsuji, A.; Nagamune, H. Estrogen stimuli promote osteoblastic differentiation via the subtilisin-like proprotein convertase pace4 in mc3t3-e1 cells. *J. Bone Miner. Metab.* **2014**. [CrossRef]
- 52. Stepan, J.J.; Alenfeld, F.; Boivin, G.; Feyen, J.H.; Lakatos, P. Mechanisms of action of antiresorptive therapies of postmenopausal osteoporosis. *Endocr. Regul.* **2003**, *37*, 225–238.
- 53. Gao, J.; Liu, Q.; Liu, X.; Ji, C.; Qu, S.; Wang, S.; Luo, Y. Cyclin g2 suppresses estrogen-mediated osteogenesis through inhibition of wnt/beta-catenin signaling. *PLoS ONE* **2014**, *9*, e89884.
- 54. Cenci, S.; Toraldo, G.; Weitzmann, M.N.; Roggia, C.; Gao, Y.; Qian, W.P.; Sierra, O.; Pacifici, R. Estrogen deficiency induces bone loss by increasing t cell proliferation and lifespan through ifn-gamma-induced class ii transactivator. *Proc. Natl. Acad. Sci. USA* **2003**, *100*, 10405–10410. [CrossRef]
- 55. D'Amelio, P.; Isaia, G.C. Immune system and postmenopausal bone loss. *Clinic Rev. Bone Miner. Metab* 2009, 7, 262–268. [CrossRef]
- 56. Zhao, R. Immune regulation of osteoclast function in postmenopausal osteoporosis: A critical interdisciplinary perspective. *Int. J. Med. Sci.* 2012, *9*, 825–832. [CrossRef]
- 57. Chin, K.Y. The relationship between follicle-stimulating hormone and bone health: Alternative explanation for bone loss beyond oestrogen? *Int. J. Med. Sci.* **2018**, *15*, 1373–1383. [CrossRef]
- 58. Suzuki, R.; Fukami, S.; Tomomura, M.; Tomomura, A.; Shirataki, Y. Screening for natural medicines effective for the treatment of osteoporosis. *J. Nat. Med.* **2019**, *73*, 331–337. [CrossRef]
- Jin, H.; Yao, L.; Chen, K.; Liu, Y.; Wang, Q.; Wang, Z.; Liu, Q.; Cao, Z.; Kenny, J.; Tickner, J.; et al. Evodiamine inhibits rankl-induced osteoclastogenesis and prevents ovariectomy-induced bone loss in mice. *J. Cell Mol. Med.* 2019, 23, 522–534. [CrossRef]
- 60. Kim, B.; Lee, K.Y.; Park, B. Icariin abrogates osteoclast formation through the regulation of the rankl-mediated traf6/nf-kappab/erk signaling pathway in raw264.7 cells. *Phytomedicine* **2018**, *51*, 181–190. [CrossRef]
- Hwang, Y.H.; Ha, H.; Kim, R.; Cho, C.W.; Song, Y.R.; Hong, H.D.; Kim, T. Protective effects of a polysaccharide ble0 isolated from barley leaf on bone loss in ovariectomized mice. *Int. J. Biol. Macromol.* 2019, 123, 314–321. [CrossRef] [PubMed]

- 62. Boyce, B.F.; Li, J.; Xing, L.; Yao, Z. Bone remodeling and the role of traf3 in osteoclastic bone resorption. *Front. Immunol.* **2018**, *9*, 2263. [CrossRef]
- 63. Bishop, G.A.; Stunz, L.L.; Hostager, B.S. Traf3 as a multifaceted regulator of b lymphocyte survival and activation. *Front. Immunol.* **2018**, *9*, 2161. [CrossRef] [PubMed]
- 64. Kaneko, J.; Okinaga, T.; Hikiji, H.; Ariyoshi, W.; Yoshiga, D.; Habu, M.; Tominaga, K.; Nishihara, T. Zoledronic acid exacerbates inflammation through m1 macrophage polarization. *Inflamm. Regen.* **2018**, *38*, 16. [CrossRef] [PubMed]
- 65. Ono, T.; Nakashima, T. Recent advances in osteoclast biology. *Histochem. Cell Biol.* **2018**, *149*, 325–341. [CrossRef] [PubMed]
- 66. Kim, K.J.; Lee, Y.; Hwang, H.G.; Sung, S.H.; Lee, M.; Son, Y.J. Betulin suppresses osteoclast formation via down-regulating nfatc1. *J. Clin. Med.* **2018**, *7*, 154. [CrossRef]
- 67. Vi, L.; Baht, G.S.; Soderblom, E.J.; Whetstone, H.; Wei, Q.; Furman, B.; Puviindran, V.; Nadesan, P.; Foster, M.; Poon, R.; et al. Macrophage cells secrete factors including lrp1 that orchestrate the rejuvenation of bone repair in mice. *Nat. Commun.* **2018**, *9*, 5191. [CrossRef]
- 68. Lind, M.; Deleuran, B.; Yssel, H.; Fink-Eriksen, E.; Thestrup-Pedersen, K. Il-4 and il-13, but not il-10, are chemotactic factors for human osteoblasts. *Cytokine* **1995**, *7*, 78–82. [CrossRef]
- 69. Silfversward, C.J.; Frost, A.; Brandstrom, H.; Nilsson, O.; Ljunggren, O. Interleukin-4 and interleukin-13 potentiate interleukin-1 induced secretion of interleukin-6 in human osteoblast-like cells. *J. Orthop. Res.* **2004**, 22, 1058–1062. [CrossRef]
- 70. Palmqvist, P.; Lundberg, P.; Persson, E.; Johansson, A.; Lundgren, I.; Lie, A.; Conaway, H.H.; Lerner, U.H. Inhibition of hormone and cytokine-stimulated osteoclastogenesis and bone resorption by interleukin-4 and interleukin-13 is associated with increased osteoprotegerin and decreased rankl and rank in a stat6-dependent pathway. *J. Biol. Chem.* **2006**, *281*, 2414–2429. [CrossRef]
- 71. Panagakos, F.S. Transforming growth factor–alpha stimulates chemotaxis of osteoblasts and osteoblast-like cells in vitro. *Biochem. Mol. Biol. Int.* **1994**, *33*, 643–650. [PubMed]
- 72. Harmer, D.; Falank, C.; Reagan, M.R. Interleukin-6 interweaves the bone marrow microenvironment, bone loss, and multiple myeloma. *Front. Endocrinol. (Lausanne)* **2018**, *9*, 788. [CrossRef] [PubMed]
- 73. Sun, W.; Meednu, N.; Rosenberg, A.; Rangel-Moreno, J.; Wang, V.; Glanzman, J.; Owen, T.; Zhou, X.; Zhang, H.; Boyce, B.F.; et al. B cells inhibit bone formation in rheumatoid arthritis by suppressing osteoblast differentiation. *Nat. Commun.* **2018**, *9*, 5127. [CrossRef] [PubMed]
- 74. Pathak, J.L.; Bakker, A.D.; Verschueren, P.; Lems, W.F.; Luyten, F.P.; Klein-Nulend, J.; Bravenboer, N. Cxcl8 and ccl20 enhance osteoclastogenesis via modulation of cytokine production by human primary osteoblasts. *PLoS ONE* 2015, 10, e0131041. [CrossRef]
- 75. Collins, F.L.; Williams, J.O.; Bloom, A.C.; Singh, R.K.; Jordan, L.; Stone, M.D.; McCabe, L.R.; Wang, E.C.Y.; Williams, A.S. Ccl3 and mmp-9 are induced by tl1a during death receptor 3 (tnfrsf25)-dependent osteoclast function and systemic bone loss. *Bone* 2017, *97*, 94–104. [CrossRef]
- Xuan, W.; Feng, X.; Qian, C.; Peng, L.; Shi, Y.; Xu, L.; Wang, F.; Tan, W. Osteoclast differentiation gene expression profiling reveals chemokine ccl4 mediates rankl-induced osteoclast migration and invasion via pi3k pathway. *Cell Biochem. Funct.* 2017, 35, 171–177. [CrossRef] [PubMed]
- 77. Dong, Y.; Song, C.; Wang, Y.; Lei, Z.; Xu, F.; Guan, H.; Chen, A.; Li, F. Inhibition of prmt5 suppresses osteoclast differentiation and partially protects against ovariectomy-induced bone loss through downregulation of cxcl10 and rsad2. *Cell Signal.* 2017, 34, 55–65. [CrossRef] [PubMed]
- 78. Liu, P.; Lee, S.; Knoll, J.; Rauch, A.; Ostermay, S.; Luther, J.; Malkusch, N.; Lerner, U.H.; Zaiss, M.M.; Neven, M.; et al. Loss of menin in osteoblast lineage affects osteocyte-osteoclast crosstalk causing osteoporosis. *Cell Death Differ.* 2017, 24, 672–682. [CrossRef]
- 79. Koizumi, K.; Saitoh, Y.; Minami, T.; Takeno, N.; Tsuneyama, K.; Miyahara, T.; Nakayama, T.; Sakurai, H.; Takano, Y.; Nishimura, M.; et al. Role of cx3cl1/fractalkine in osteoclast differentiation and bone resorption. *J. Immunol.* 2009, *183*, 7825–7831. [CrossRef]
- 80. Zimmerman, S.M.; Heard-Lipsmeyer, M.E.; Dimori, M.; Thostenson, J.D.; Mannen, E.M.; O'Brien, C.A.; Morello, R. Loss of rankl in osteocytes dramatically increases cancellous bone mass in the osteogenesis imperfecta mouse (oim). *Bone Rep.* **2018**, *9*, 61–73. [CrossRef]
- 81. Komori, T. Mouse models for the evaluation of osteocyte functions. *J. Bone Metab.* **2014**, *21*, 55–60. [CrossRef] [PubMed]

- 82. Ito, N.; Wijenayaka, A.R.; Prideaux, M.; Kogawa, M.; Ormsby, R.T.; Evdokiou, A.; Bonewald, L.F.; Findlay, D.M.; Atkins, G.J. Regulation of fgf23 expression in idg-sw3 osteocytes and human bone by pro-inflammatory stimuli. *Mol. Cell Endocrinol.* **2015**, *399*, 208–218. [CrossRef] [PubMed]
- Qu, X.; Mei, J.; Yu, Z.; Zhai, Z.; Qiao, H.; Dai, K. Lenalidomide regulates osteocytes fate and related osteoclastogenesis via il-1beta/nf-kappab/rankl signaling. *Biochem. Biophys. Res. Commun.* 2018, 501, 547–555. [CrossRef]
- 84. McGregor, N.E.; Murat, M.; Elango, J.; Poulton, I.J.; Walker, E.C.; Crimeen-Irwin, B.; Ho, P.W.M.; Gooi, J.H.; Martin, T.J.; Sims, N.A. Il-6 exhibits both cis and trans signaling in osteocytes and osteoblasts, but only trans signaling promotes bone formation and osteoclastogenesis. *J. Biol. Chem.* **2019**. [CrossRef] [PubMed]
- 85. Kim, H.J.; Choi, Y.; Bae, M.K.; Hwang, D.S.; Shin, S.H.; Lee, J.Y. Zoledronate enhances osteocyte-mediated osteoclast differentiation by il-6/rankl axis. *Int. J. Mol. Sci.* **2019**, *20*, 1467. [CrossRef]
- 86. Metzger, C.E.; Gong, S.; Aceves, M.; Bloomfield, S.A.; Hook, M.A. Osteocytes reflect a pro-inflammatory state following spinal cord injury in a rodent model. *Bone* **2019**, *120*, 465–475. [CrossRef] [PubMed]
- 87. Chhana, A.; Pool, B.; Callon, K.E.; Tay, M.L.; Musson, D.; Naot, D.; McCarthy, G.; McGlashan, S.; Cornish, J.; Dalbeth, N. Monosodium urate crystals reduce osteocyte viability and indirectly promote a shift in osteocyte function towards a proinflammatory and proresorptive state. *Arthritis Res. Ther.* 2018, 20, 208. [CrossRef] [PubMed]
- Park, M.J.; Lee, J.; Byeon, J.S.; Jeong, D.U.; Gu, N.Y.; Cho, I.S.; Cha, S.H. Effects of three-dimensional spheroid culture on equine mesenchymal stem cell plasticity. *Vet. Res. Commun.* 2018, 42, 171–181. [CrossRef] [PubMed]
- 89. de Barros Silva, P.G.; de Oliveira, C.C.; Brizeno, L.; Wong, D.; Lima Junior, R.; Goncalves, R.P.; Sousa, F.B.; Mota, M.; de Albuquerque Ribeiro, R.; Alves, A. Immune cellular profile of bisphosphonate-related osteonecrosis of the jaw. *Oral Dis.* **2016**, *22*, 649–657. [CrossRef]
- 90. Liu, Q.; Liu, C.; Yang, Y.; Yang, H.; Chen, J. Osteocyte-intrinsic mtorc1 signaling restrains trabecular bone accrual in mice. *J. Cell Biochem.* **2018**, *119*, 8743–8749. [CrossRef]
- Siegenthaler, B.; Ghayor, C.; Gjoksi-Cosandey, B.; Ruangsawasdi, N.; Weber, F.E. The bromodomain inhibitor n-methyl pyrrolidone prevents osteoporosis and bmp-triggered sclerostin expression in osteocytes. *Int. J. Mol. Sci.* 2018, *19*, 3332. [CrossRef]
- 92. Taylor, S.; Hu, R.; Pacheco, E.; Locher, K.; Pyrah, I.; Ominsky, M.S.; Boyce, R.W. Differential time-dependent transcriptional changes in the osteoblast lineage in cortical bone associated with sclerostin antibody treatment in ovariectomized rats. *Bone Rep.* **2018**, *8*, 95–103. [CrossRef]
- Boyce, R.W.; Brown, D.; Felx, M.; Mellal, N.; Locher, K.; Pyrah, I.; Ominsky, M.S.; Taylor, S. Decreased osteoprogenitor proliferation precedes attenuation of cancellous bone formation in ovariectomized rats treated with sclerostin antibody. *Bone Rep.* 2018, *8*, 90–94. [CrossRef]
- 94. Toscani, D.; Bolzoni, M.; Ferretti, M.; Palumbo, C.; Giuliani, N. Role of osteocytes in myeloma bone disease: Anti-sclerostin antibody as new therapeutic strategy. *Front. Immunol.* **2018**, *9*, 2467. [CrossRef]
- 95. Yang, Y.; Zheng, X.; Li, B.; Jiang, S.; Jiang, L. Increased activity of osteocyte autophagy in ovariectomized rats and its correlation with oxidative stress status and bone loss. *Biochem. Biophys. Res. Commun.* **2014**, 451, 86–92. [CrossRef]
- 96. Florencio-Silva, R.; Sasso, G.R.; Simoes, M.J.; Simoes, R.S.; Baracat, M.C.; Sasso-Cerri, E.; Cerri, P.S. Osteoporosis and autophagy: What is the relationship? *Rev. Assoc. Med. Bras.* **2017**, *63*, 173–179. [CrossRef]
- 97. Ye, T.; Cao, P.; Qi, J.; Zhou, Q.; Rao, D.S.; Qiu, S. Protective effect of low-dose risedronate against osteocyte apoptosis and bone loss in ovariectomized rats. *PLoS ONE* **2017**, *12*, e0186012. [CrossRef]
- Reis, L.C.; Ramos-Sanchez, E.M.; Petitto-Assis, F.; Nerland, A.H.; Hernandez-Valladares, M.; Selheim, F.; Floeter-Winter, L.M.; Goto, H. Insulin-like growth factor-i as an effector element of the cytokine il-4 in the development of a leishmania major infection. *Mediators Inflamm.* 2018, 2018, 9787128. [CrossRef]
- 99. Sun, K.T.; Cheung, K.K.; Au, S.W.N.; Yeung, S.S.; Yeung, E.W. Overexpression of mechano-growth factor modulates inflammatory cytokine expression and macrophage resolution in skeletal muscle injury. *Front. Physiol.* **2018**, *9*, 999. [CrossRef]
- 100. Wang, J.; Xie, L.; Wang, S.; Lin, J.; Liang, J.; Xu, J. Azithromycin promotes alternatively activated macrophage phenotype in systematic lupus erythematosus via pi3k/akt signaling pathway. *Cell Death Dis.* 2018, *9*, 1080. [CrossRef]

- 101. Tonkin, J.; Temmerman, L.; Sampson, R.D.; Gallego-Colon, E.; Barberi, L.; Bilbao, D.; Schneider, M.D.; Musaro, A.; Rosenthal, N. Monocyte/macrophage-derived igf-1 orchestrates murine skeletal muscle regeneration and modulates autocrine polarization. *Mol. Ther.* 2015, 23, 1189–1200. [CrossRef]
- 102. Zhao, G.; Monier-Faugere, M.C.; Langub, M.C.; Geng, Z.; Nakayama, T.; Pike, J.W.; Chernausek, S.D.; Rosen, C.J.; Donahue, L.R.; Malluche, H.H.; et al. Targeted overexpression of insulin-like growth factor i to osteoblasts of transgenic mice: Increased trabecular bone volume without increased osteoblast proliferation. *Endocrinology* **2000**, *141*, 2674–2682. [CrossRef]
- 103. Grinspoon, S.; Baum, H.; Lee, K.; Anderson, E.; Herzog, D.; Klibanski, A. Effects of short-term recombinant human insulin-like growth factor i administration on bone turnover in osteopenic women with anorexia nervosa. *J. Clin. Endocrinol. Metab.* **1996**, *81*, 3864–3870. [PubMed]
- 104. Gao, Y.; Wu, X.; Terauchi, M.; Li, J.Y.; Grassi, F.; Galley, S.; Yang, X.; Weitzmann, M.N.; Pacifici, R. T cells potentiate pth-induced cortical bone loss through cd40l signaling. *Cell Metab.* 2008, *8*, 132–145. [CrossRef] [PubMed]
- 105. Silva, B.C.; Bilezikian, J.P. Parathyroid hormone: Anabolic and catabolic actions on the skeleton. *Curr. Opin. Pharmacol.* **2015**, *22*, 41–50. [CrossRef] [PubMed]
- 106. Milstrey, A.; Wieskoetter, B.; Hinze, D.; Grueneweller, N.; Stange, R.; Pap, T.; Raschke, M.; Garcia, P. Dose-dependent effect of parathyroid hormone on fracture healing and bone formation in mice. *J. Surg. Res.* 2017, 220, 327–335. [CrossRef] [PubMed]
- 107. Yao, Z.; Lei, W.; Duan, R.; Li, Y.; Luo, L.; Boyce, B.F. Rankl cytokine enhances tnf-induced osteoclastogenesis independently of tnf receptor associated factor (traf) 6 by degrading traf3 in osteoclast precursors. *J. Biol. Chem.* 2017, 292, 10169–10179. [CrossRef]
- 108. Suzuki, T.; Nakamura, Y.; Kato, H. Effects of denosumab on bone metabolism and bone mineral density with anti-tnf inhibitors, tocilizumab, or abatacept in osteoporosis with rheumatoid arthritis. *Ther. Clin. Risk Manag.* 2018, 14, 453–459. [CrossRef] [PubMed]
- Tada, M.; Inui, K.; Sugioka, Y.; Mamoto, K.; Okano, T.; Koike, T. Abatacept might increase bone mineral density at femoral neck for patients with rheumatoid arthritis in clinical practice: Airtight study. *Rheumatol. Int.* 2018, *38*, 777–784. [CrossRef]
- Zerbini, C.A.F.; Clark, P.; Mendez-Sanchez, L.; Pereira, R.M.R.; Messina, O.D.; Una, C.R.; Adachi, J.D.; Lems, W.F.; Cooper, C.; Lane, N.E. Biologic therapies and bone loss in rheumatoid arthritis. *Osteoporos. Int.* 2017, 28, 429–446. [CrossRef]
- 111. Moon, S.J.; Ahn, I.E.; Jung, H.; Yi, H.; Kim, J.; Kim, Y.; Kwok, S.K.; Park, K.S.; Min, J.K.; Park, S.H.; et al. Temporal differential effects of proinflammatory cytokines on osteoclastogenesis. *Int. J. Mol. Med.* 2013, *31*, 769–777. [CrossRef] [PubMed]



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