RESEARCH ARTICLE

Open Access



Cytomegalovirus infection and outcome in immunocompetent patients in the intensive care unit: a systematic review and meta-analysis

Xi Li^{1,2}, Yongbo Huang^{1,2}, Zhiheng Xu^{1,2}, Rong Zhang^{1,2}, Xiaoqing Liu^{1,2}, Yimin Li^{1,2*} and Pu Mao^{1,3*}

Abstract

Background: Cytomegalovirus (CMV) infection is common in immunocompetent patients in intensive care units (ICUs). However, whether CMV infection or CMV reactivation contributes to mortality of immunocompetent patients remains unclear.

Methods: A literature search was conducted for relevant studies published before May 30, 2016. Studies reporting on CMV infection in immunocompetent patients in ICUs and containing 2×2 tables on CMV results and all-cause mortality were included.

Results: Eighteen studies involving 2398 immunocompetent patients admitted to ICUs were included in the meta-analysis. The overall rate of CMV infection was 27% (95%Cl 22–34%, $l^2 = 89\%$, n = 2398) and the CMV reactivation was 31% (95%Cl 24–39%, $l^2 = 74\%$, n = 666). The odds ratio (OR) for all-cause mortality among patients with CMV infection, compared with those without infection, was 2.16 (95%Cl 1.70–2.74, $l^2 = 10\%$, n = 2239). Moreover, upon exclusion of studies in which antiviral treatment was possibly or definitely provided to some patients, the association of mortality rate with CMV infection was also statistically significant (OR: 1.69, 95%Cl 1.01–2.83, $l^2 = 37\%$, n = 912,). For CMV seropositive patients, the OR for mortality in patients with CMV reactivation as compared with patients without CMV reactivation was 1.72 (95%Cl 1.04–2.85, $l^2 = 29\%$, n = 664). Patients with CMV infection required significantly longer mechanical ventilation (mean difference (MD): 9 days (95% Cl 5–14, $l^2 = 81\%$, n = 875)) and longer duration of ICU stay (MD: 12 days (95% Cl 7–17, $l^2 = 70\%$, n = 949)) than patients without CMV infection. When analysis was limited to detection in blood, CMV infection without antiviral drug treatment or reactivation was not significantly associated with higher mortality (OR: 1.69, 95%Cl 0.81–3.54, $l^2 = 52\%$, n = 722; OR: 1.49, $l^2 = 63\%$, n = 469).

Conclusion: Critically ill patients without immunosuppression admitted to ICUs show a high rate of CMV infection. CMV infection during the natural unaltered course or reactivation in critically ill patients is associated with increased mortality, but have no effect on mortality when CMV in blood. More studies are needed to clarify the impact of CMV infection on clinical outcomes in those patients.

Keywords: Cytomegalovirus, Immunocompetent, Meta-analysis

Medical University), Guangzhou, China Full list of author information is available at the end of the article



^{*} Correspondence: dryiminli@vip.163.com; maopu1981@126.com

¹State Key Laboratory of Respiratory Disease, National Clinical Research
Center for Respiratory Disease, Guangzhou Institute of Respiratory Disease,
the First Affiliated Hospital of Guangzhou Medical University (Guangzhou

Background

Human cytomegalovirus (CMV) is a prototypic member of the β herpes virus subfamily [1]. The prevalence of CMV seropositivity in human populations is roughly 50–95% [2–4] and highest amongst older people [5]. Cytomegalovirus infection induces innate immune responses (eg. natural killer cells) and adaptive immunity (eg. CD4+/CD8+ T cells). However, the virus can evade host detection by expressing genes that interfere with both the innate and adaptive immune systems. Eventually, CMV is able to establish latency in which either the host fails to eliminate the virus or the virus cannot replicate. However, CMV can become reactivated during periods of host immune suppression [6].

It is well known that CMV infection is common in canonical immunodeficiency patients, such as those with human immunodeficiency virus infection, solid organ or stem cell transplantation and patients undergoing chemoor radiotherapy [7–9]. With the development of more sensitive detection method, the rate of CMV detection is high in intensive care units (ICUs) [10–25]. However, so far, there is no convincing research to support the use of antiviral treatment when critically ill but immunocompetent patients present with CMV infection. Furthermore, whether CMV is a contributor or simply a bystander to the severity of illness remains under debate [26–28].

Whether CMV infection is associated with increased mortality in immunocompetent ICU patients remains controversial [13–16]. A previous meta-analysis published in 2009 demonstrated that CMV infection was associated with a higher mortality rate, nearly twice that observed in patients without CMV infection [29]. However, this study did not consider the influence of antiviral drugs on clinical outcomes. Moreover, many clinical studies about CMV have been reported in recent years. Thus, to acquire a better understanding of the potential role of CMV infection in contributing to mortality in critically ill patients, especially those not receiving antiviral agents and CMV detected in blood, we performed a meta-analysis of data available in the literature, focusing on the outcome in immunocompetent ICU patients with CMV infection.

Methods

Search strategy

A literature search for relevant publications included within the electronic databases PubMed, EMBASE and the Cochrane Library was performed using combinations of the keywords "cytomegaloviruses", "salivary gland viruses", "herpes virus", "cytomegaloviral infection", "HHV5", "intensive care", "critical care", "critical illness", "mechanical ventilation", and "pulmonary ventilator". All searches were updated on May 30, 2016. No language restriction was

enforced. We also consulted relevant reference articles and searched using Google Scholar.

Study selection

Two researchers (LX and HYB) performed data extraction independently, and any discrepancies were addressed by discussion and reevaluation until consensus was achieved. Observational studies were eligible if they reported on CMV infection in immunocompetent patients in the ICU, and if a 2×2 table could be constructed based on CMV results and all-cause mortality. All patients were over 18 years of age. The systematic review included only studies in which all patients were tested for CMV. An episode of CMV infection was defined by one of the examination CMV viral culture, polymerase chain reaction (PCR), CMV antigen (pp65) in blood, tracheal aspirates, urine, or a combination of these. A case was defined by the presence of reactivation, where the patient had CMV infection and was seropositive. Immunocompetent patients were defined as those patients who did not receive a solid organ or hematopoietic stem cell transplant, did not receive immunosuppressive treatment, did not have human immunodeficiency virus infection, did not have primary immunodeficiency, and did not receive chemotherapy or radiotherapy before ICU admission.

Data extraction and quality appraisal

We obtained information on basic study characteristics (author, year of publication, country of origin, study period, setting, and study design), characteristic population, the site and detection method of sample, CMV seropositivity, CMV infection incidence, all-cause mortality, length of ICU/hospital stay, length of mechanical ventilation, and administration of antiviral drugs.

The Newcastle-Ottawa scale, developed for evaluating the quality of observational studies (Additional file 1: Table S1) [30], was used to assess the validity of included studies.

Data analysis

Continuous variables are reported as mean or median values and categorical variables are reported as n (%). Meta-analytic pooling was performed for outcome variables with a Logit transformation approach, reporting results as summary point estimates (95% confidence interval, CI). We used the Mantel–Haenszel method to obtain odds ratios (ORs) and 95% CI. When only the median, range, or interquartile range of length of mechanical ventilation and the length of ICU stay were reported, we used simple formulas to estimate the mean and standard deviation [31].

Between-study heterogeneity was examined using the I² measure of inconsistency and the chi-square test of heterogeneity.

To evaluate publication bias, we constructed a funnel plot and used the Egger test. Sensitivity analyses of the Begg's test were additionally conducted to ascertain the robustness of our findings. All meta-analyses were performed with R software (version 3.3.3 for Windows) and SPSS 18 (IBM, Armonk, NY, USA).

Results

Study selection

The initial database search identified 1846 potentially relevant studies. Following this, assessment of the full text yielded 17 studies suitable for analysis. Another publication was incorporated after examining references from the extracted articles [23]. Consequently, our meta-analysis consisted of 18 articles (Fig. 1), including one case-control [20] and 17 cohort studies [10–19, 21–25, 32].

Characteristics of included studies

Most studies were conducted in the United States and Europe, except one cohort study in Egypt [12], and were published between 1990 and 2016 (Table 1). Overall, the studies were well done, with a median score of 7 (range 6–8) on the Newcastle–Ottawa scale for appraising the quality of observational studies.

A total of 2398 patients were included, having been admitted to the ICU for a variety of reasons, with a median age of 59 years. The median period of prospective

studies was 24 months, ranging broadly from 3 to 78 months. All studies used CMV blood assays, and 6 studies also assayed sputum samples. Most studies indicated that the frequency of sample collection was once a week. In our analysis, the methods used to assess CMV infection were virus culture, pp65 antigen detection and PCR detection of CMV DNA in ten, three and two studies, respectively, and combinations of two diagnostic methods in the remaining three studies.

CMV infection and outcome in immunocompetent patients

As shown in Fig. 2, the overall detection rate of CMV was 27% (95% CI 22–34%, I^2 = 89%, n = 2398). As compared with patients without CMV infection, the all-cause mortality of patients with CMV infection was significantly higher (OR: 2.16; 95% CI 1.70–2.74, I^2 = 10%, n = 2239) (Fig. 3a). When analysis was limited to CMV detection in blood, there was still statistical significance in mortality rate between patients with CMV infection (OR: 2.15, 95% CI 1.48–3.15, I^2 = 34%, n = 1441) compared with patients without infection (Additional file 2: Figure S1).

To rule out the impact of antiviral drugs on patients with CMV infection, four studies in which patients received antiviral drugs during their ICU stay and eight studies that did not specify the use of antiviral drugs were excluded. The

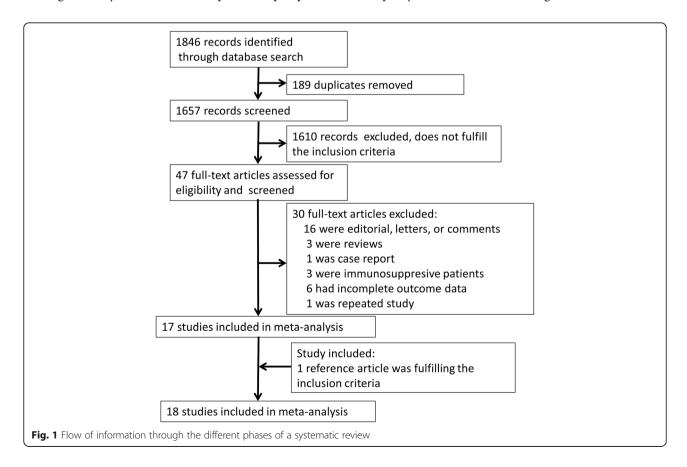


Table 1 Characteristics of studies included in the meta-analysis

Study	Year	Location	Research type	ICU type	NOS	Age, y (media)	Male/ Femal	Period, month	Selected population	Sample	Detective way
Domart [25]	1990	France	CP	S	7	53	97/18	66	Mediastinistis after cardiac surgery	Blood, urine	Culture
Cook [24]	1998	USA	CR	S	7	NA	NA	14	Sepsis without bacteria or fungus	Blood, sputum	Culture
Kutza [23]	1998	Germany	CP	S	6	58	27/7	NA	Sepsis	Blood	PCR, pp65
Heininger [22]	2001	Germany	CP	S	7	68	35/21	12	Seropositive patients after major surgery and trauma	Blood, sputum	PCR, Culture
Cook [21]	2003	USA	CP	S	8	59	62/42	15	Sepsis, SICU stay>7d	Blood, sputum	Culture
Jaber [20]	2005	France	CC	M-S	6	NA	138/62	78	fever>72 h, without bacteria or fungus	Blood	pp65
von Muller [19]	2006	Germany	CP	М	7	NA	15/10	9	septic shock, ICU stay>7d	Blood	pp65
Limaye [18]	2008	USA	CP	M-S	8	52	73/47	24	Seropositive	Blood	PCR
Ziemann [17]	2008	Germany	CR	S	7	NA	33/66	33	ICU stay>13d	Blood	PCR
Chiche [16]	2009	France	CP	М	7	63	159/83	24	Mechanical ventilation16	Blood, sputum	PP65, Culture
Chilet [15]	2010	Spanish	CP	S	7	NA	37/16	13	Seropositive, ICU stay>5d	Blood, sputum	PCR
Bordes [14]	2011	France	CP	S	7	52	22/7	20	Seropositive, burn patients,	Blood	PCR
Heininger [13]	2011	Germany	CP	M-S	7	68	67/19	30	Seropositive, severe sepsis	Blood, sputum	PCR
Osman [12]	2014	Egypt	CP	М	7	59	29/22	3	Mechanical ventilation	Blood	PCR
Walton [11]	2014	France	CP	M-S	7	NA	305/255	48	Sepsis	Blood	PCR
Ong [10]	2015	New Zealand	CP	М	8	NA	196/110	24	ARDS, Mechanical ventilation>4d	Blood	PCR
Frantzeska [32]	2015	Greece	CP	М	7	63	51/29	24	Seropositive, mechanical ventilation	Blood	PCR
Ong [33]	2016	Netherlands	CP	М	7	NA	164/107	36	ARDS, Mechanical ventilation>4d	Blood	PCR

CP cohort prospective research, CR cohort and retrospective research, CC case-control research, ARDS acute respiratory distress syndrome, PCR polymerase chain reaction, NA not available, M-S medical-surgical, S surgical, M medical

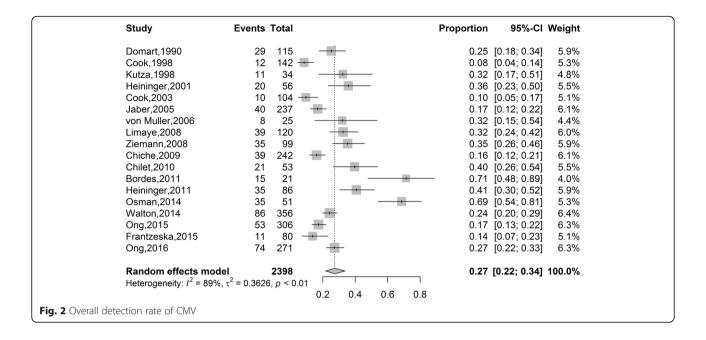
remaining six studies of patients without antiviral treatment during the course of ICU stay were analyzed [10, 13, 18, 19, 21, 33]. The difference in mortality rates between patients with CMV infection remained significant (OR: 1.69, 95% CI 1.01-2.83, $I^2=37\%$, n=912) compared with patients without infection (Fig. 3b). When analysis was limited to CMV detection in blood, there was no statistical significance in mortality rate between patients with CMV infection (OR: 1.69, 95% CI 0.81-3.54, $I^2=52\%$, n=722) as compared with patients without infection (Additional file 3: Figure S2).

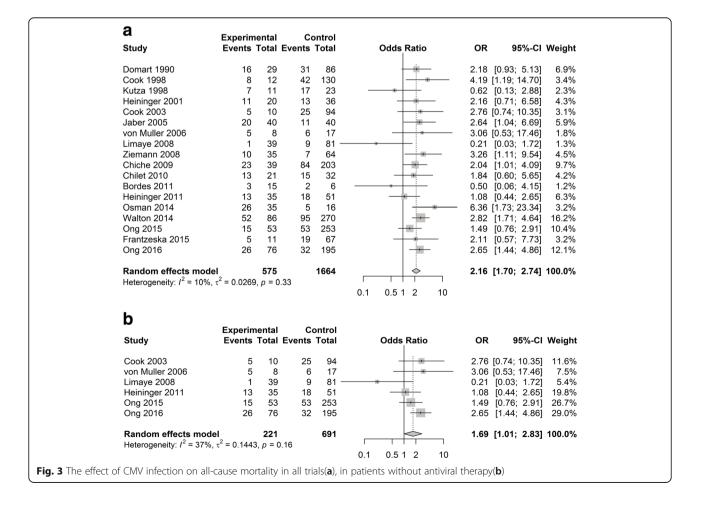
The mean difference in mechanical ventilation days and duration of ICU stay was an increase of 9 days (95% CI 5–14, $I^2 = 81\%$, n = 875) and 12 days (95% CI 7–17, $I^2 = 70\%$, n = 949), respectively, between patients with

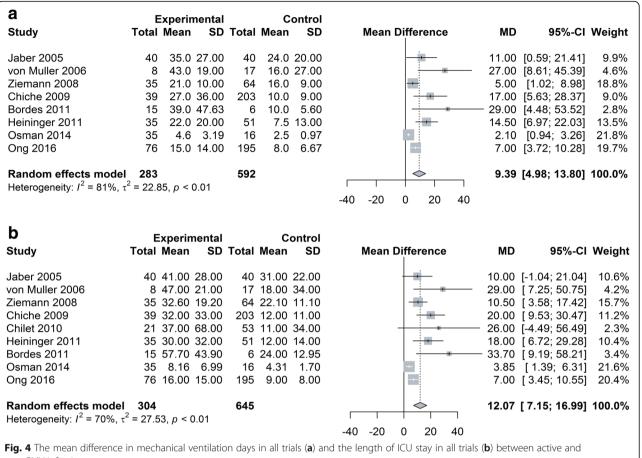
and without CMV infection (Fig. 4a and b). When analysis was limited to CMV detection in blood, there was still a statistically significant difference in length of mechanical ventilation and ICU stay between patients with CMV infection as compared with patients without infection (MD: 7 days (95% CI 3–11, $I^2 = 77\%$, n = 547) and MD: 9 days (95% CI 4–13, $I^2 = 66\%$, n = 547)), respectively (Additional file 4: Figure s3 and Additional file 5: Figure s4).

CMV reactivation and outcome in immunocompetent patients

The CMV seropositivity rate, which represents previous infection, was 71% (95% CI 68–75%, $I^2 = 35\%$, n = 1242) in immunocompetent ICU patients (Fig. 5a). Patients







non-CMV infection

with CMV reactivation, which represents CMV detected among seropositive patients, was 31% (95% CI 24-39%, $I^2 = 72\%$, n = 666) (Fig. 5b).

The OR for mortality in patients with CMV reactivation as compared with patients without CMV reactivation was 1.72 (95% CI 1.04–2.85, $I^2 = 29\%$, n = 664) (Fig. 5c). But for patients of CMV infection in blood, the reactivation was not associated with higher mortality (OR: 1.49, 95% CI 0.46–4.28, $I^2 = 63\%$, n = 469) (Additional file 6: Figure S5).

We also analyzed the rate of CMV and mortality thought categorized by the detection methods (Additional file 7: Figure S6, Additional file 8: Figure S7: Additional file 9: Figure S8 and Additional file 10: Figure S9).

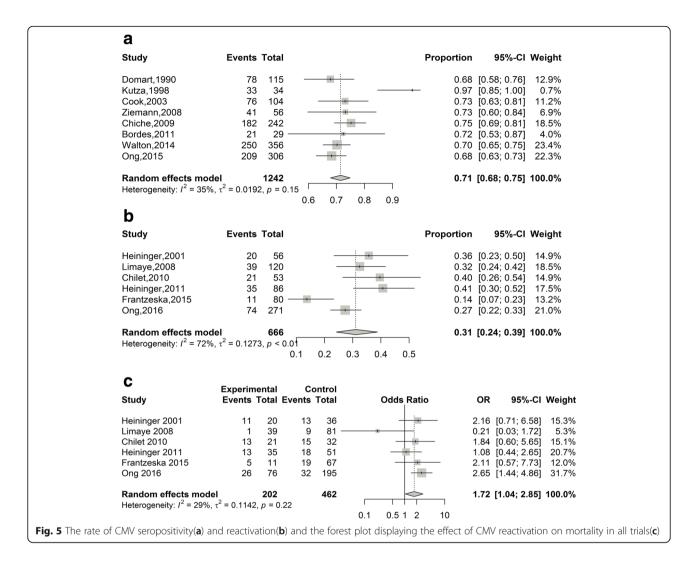
Publication bias and sensitivity analysis

We used the Egger test to detect publication bias. There was no publication bias either in the overall CMV prevalence analysis (t = 1.1264, p = 0.2766) or in the all-cause CMV mortality analysis (t = -1.3418, p = 0.1984). We also used Begg's test to detect sensitivity analysis, and the results showed that the analyses were robust.

Discussion

In this meta-analysis, we have demonstrated that CMV infection frequently present in critically ill immunocompetent patients at ICU admission. The overall rate of CMV infection was 27%, which was higher than the 17% presented in a previous meta-analysis [29], because eight recent studies detecting CMV infection by PCR assay were included in our meta-analysis [10–15, 32, 33]. Polymerase chain reaction has been demonstrated to be the most sensitive method of CMV detection [34], but even so, the CMV infection rate may still be underestimated because we chose only the studies containing 2×2 tables on CMV results and all-cause mortality. We excluded studies where either the rate of CMV infection or mortality was zero and we also excluded some studies with a 0% infection rate that used early monitoring of CMV, often fewer than 7 days after admission to the ICU [26, 35-37]. We believe this could have led to underestimation of the CMV infection rate because the transition to CMV infection requires time for the complete lytic virus cycle to develop from the latent phase [38].

We found that the detection rate of CMV by culture, pp65 and PCR was 13, 22 and 34%, respectively. Desachy



et al. demonstrated that positive results for CMV infection were obtained in a median of 4 days by PCR compared with 11 days by pp65 antigen detection after onset of sepsis [36]. Therefore, PCR facilitates earlier diagnosis of an episode of CMV infection than any other method. We then analyzed the association between CMV positivity and mortality, stratified by detection method. We also found that patients with CMV infection detected by PCR had higher mortality than patients without CMV infection (OR: 2.07, 95% CI 1.59–2.70, $I^2 = 40\%$, n =1441). However, when compared with other methods, the association with mortality was marginally less strong using PCR. We may think that viral burden of CMV is determinant of pathogenesis, and higher CMV loads is correlated with progression of some CMV infection disease [39, 40].

The presence of CMV seropositivity, representing previous infection, was found in 71% of immunocompetent ICU patients and the incidence of CMV reactivation was high, observed in 31% of seropositive patients in our

meta-analysis. There are several factors that might explain the high prevalence. First of all, the rate of CMV seropositivity increases with advancing age [5] and in our analysis, the median age was 59 years. Second, to inhibit the reactivation of CMV, as many as 10% of all peripheral CD4+ and CD8+ T cells are constantly required for immune surveillance to maintain functional latency [41]. Sepsis is associated with immunoparalysis, as apoptosis of CD4+ and CD8+ T cells is increased [42, 43]. Furthermore, some patients in the ICU may be immunosuppressed after trauma and major surgery [44]. In addition, treatments commonly received in the ICU, such as massive transfusion, corticosteroids, or catecholamines may transiently compromise host immunity [45]. It has also been reported that the use of heart-lung machines can lead to temporary systemic immunosuppression [46]. Therefore, patients in the ICU may show transient immunoparalysis [47], potentially resulting in the observed CMV reactivation. Third, some inflammatory cytokines including tumor necrosis factor alpha and interleukin-1 β , can stimulate reactivation of latent CMV [48]. Thus, significant numbers of immunocompetent patients harboring latent virus are susceptible to CMV reactivation during critical illness.

When the mortality analysis was limited to CMV detection in blood, CMV infection without antiviral drug treatment or reactivation was not significantly associated with higher mortality. This maybe explained that the presence of high peripheral levels of functional CMV-specific CD4+ and CD8+ T cells in immunocompetent patients, which can suppress CMV during episodes of reactivation [26]. It was observed that CMV infection was not associated with mortality in CMV colitis. In steroid-refractory patients with ulcerative colitis, CMV was found in the colon by histopathology, which is also not associated with adverse clinical outcomes [49]. Indeed, there has been no research to demonstrate that immunocompetent critically patients with CMV infection could benefit from antivirus therapy. And there are a number of side effects of antiviral drugs, such as hematologic complications (neutropenia, anemia and thrombocytopenia), renal dysfunction, mental disorders [50]. Therefore, giving antiviral drugs to critically ill patients should be considered cautiously in terms of advantage-disadvantage ratio. To address this issue, there are two ongoing, blinded, randomized placebo-controlled clinical trials of an antiviral drug with activity against CMV in critically ill patients in the ICU (NCT 01335932, NCT 02152358).

Patients with sepsis have the highest incidence of CMV infection [22]. Early in 1990's, bacterial sepsis was considered to trigger CMV reactivation [26]. The reactivation associated with sepsis was consequence of inflammatory stimulation, transient immune compromise, and maybe involving some component of epigenetic regulation of viral DNA [26].

There are five limitations in this study. First, we observed large heterogeneity in many of our analyses. However, little or no heterogeneity was observed in the meta-analysis of mortality outcome. Second, most studies were not blind, thus reducing the reliability of the results. Third, lack of sufficient data on clinical parameters (eg: severity of illness, cause of ICU admission, comorbidity) meant that stratified analyses based on such clinical characteristics were not possible. Fourth, the definition of the state of CMV infection was inconsistent and maybe restrictive to capture the dynamics of CMV infection. As such, we could not conduct meta-analysis with outcome data and this is a major limitation of our meta-analysis. Finally, one [10] cannot discount the effect of unmeasured confounders given the observational nature of the body of evidence comprising this meta-analysis.

Conclusions

Our findings suggests that there is a high incidence of CMV seropositivity and CMV infection in critically ill patients without immunosuppression. This study suggest that CMV infection without antiviral drug treatment or reactivation in critically ill patients is associated with increased mortality, and is not associated with mortality when CMV infection is detected in blood. Further research is necessary to determine the full role of CMV in this vulnerable patient demographic.

Additional files

Additional file 1: Table S1. The Newcastle–Ottawa scale (PDF 17 kb)

Additional file 2: Figure S1. The effect of CMV infection on all-cause mortality in blood (TIFF 276 kb)

Additional file 3: Figure S2. The effect of CMV infection on all-cause mortality in patients without antiviral therapy in blood (TIFF 150 kb)

Additional file 4: Figure S3. The mean difference in mechanical ventilation days in blood between active and non-CMV infection (TIFF 217 kb)

Additional file 5: Figure S4. The mean difference in the length of ICU stay in blood between active and non-CMV infection (TIFF 222 kb)

Additional file 6: Figure S5. The effect of CMV reactivation on mortality in blood (TIFF 133 kb)

Additional file 7: Figure S6. Subgroup analysis of CMV detection rate according to detection method in all trials (TIFE 552 kb)

Additional file 8: Figure S7. Subgroup analysis of CMV detection rate according to detection method in blood (TIFF 393 kb)

Additional file 9: Figure S8. The effect of CMV infection on all-cause mortality in subgroup analysis according to detection method in all trials (TIFF 582 kb)

Additional file 10: Figure S9. The effect of CMV infection on all-cause mortality in subgroup analysis according to detection method in blood (TIFF 420 kb)

Abbreviations

CD: Cluster of differentiation; CI: Confidence interval; CMV: Cytomegalovirus; DNA: Deoxyribonucleic acid; HHV: Human herpes virus; ICU: Intensive care unit; OR: Odds ratio; PCR: Polymerase chain reaction

Funding

This work was funded by grants from the Science and Technology Project of Guangzhou (No. 201400000002 and No. 2014Y2–00065). The founders of the study had no role in study design, data collection, data analysis, and data interpretation.

Availability of data and materials

All data generated or analyzed during this study are included in this published article.

Authors' contributions

LX conducted the literature search, extracted data, performed statistical analysis, and drafted the manuscript. HYB conducted the search, extracted the data, and revised the manuscript. XZH performed the statistical analysis and edited the manuscript. ZR conducted the literature search and extracted data. LXQ interpreted the data. MP designed the study, interpreted data, and revised the manuscript. LYM conceived and designed the study and revised the manuscript. All authors have read and approved the final manuscript.

Ethics approval and consent to participate

This review evaluated data from published studies and was exempt from ethics committee approval. This review did not involve any direct research on patients, and no informed consent was required.

Consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Author details

¹State Key Laboratory of Respiratory Disease, National Clinical Research Center for Respiratory Disease, Guangzhou Institute of Respiratory Disease, the First Affiliated Hospital of Guangzhou Medical University, Guangzhou Medical University), Guangzhou, China. ²Intensive Care Unit, The First Affiliated Hospital of Guangzhou Medical University, Guangzhou, Guangdong, China. ³Department of Infection Control, The First Affiliated Hospital of Guangzhou Medical University, Guangzhou, Guangdong, China.

Received: 26 August 2016 Accepted: 18 June 2018 Published online: 28 June 2018

References

- Boeckh M, Geballe AP. Cytomegalovirus: pathogen, paradigm, and puzzle. J Clin Investig. 2011;121(5):1673–80.
- Stadler LP, Bernstein DI, Callahan ST, Ferreira J, Gorgone Simone GA, Edwards KM, Stanberry LR, Rosenthal SL. Seroprevalence of cytomegalovirus (CMV) and risk factors for infection in adolescent males. Clin Infect Dis. 2010;51(10):e76–81.
- Hyde TB, Schmid DS, Cannon MJ. Cytomegalovirus seroconversion rates and risk factors: implications for congenital CMV. Rev Med Virol. 2010; 20(5):311–26.
- Bate SL, Dollard SC, Cannon MJ. Cytomegalovirus seroprevalence in the United States: the national health and nutrition examination surveys, 1988-2004. Clin Infect Dis. 2010;50(11):1439–47.
- Musiani M, Zerbini M, Zauli D, Cometti G, La Placa M. Impairment of cytomegalovirus and host balance in elderly subjects. J Clin Pathol. 1988;41(7):722–5.
- Jackson SE, Mason GM, Wills MR. Human cytomegalovirus immunity and immune evasion. Virus Res. 2011;157(2):151–60.
- Owers DS, Webster AC, Strippoli GF, Kable K, Hodson EM. Pre-emptive treatment for cytomegalovirus viraemia to prevent cytomegalovirus disease in solid organ transplant recipients. Cochrane Database Syst Rev. 2013;2:CD005133.
- Atabani SF, Smith C, Atkinson C, Aldridge RW, Rodriguez-Peralvarez M, Rolando N, Harber M, Jones G, O'Riordan A, Burroughs AK, et al. Cytomegalovirus replication kinetics in solid organ transplant recipients managed by preemptive therapy. Am J Transplant. 2012;12(9):2457–64.
- Deayton JR, Sabin CA, Johnson MA, Emery VC, Wilson P, Griffiths PD. Importance of cytomegalovirus viraemia in risk of disease progression and death in HIV-infected patients receiving highly active antiretroviral therapy. Lancet. 2004;363(9427):2116–21.
- Ong DS, Klein Klouwenberg PM, Verduyn Lunel FM, Spitoni C, Frencken JF, Dekker HA, Schultz MJ, Bonten MJ, Cremer OL. Cytomegalovirus seroprevalence as a risk factor for poor outcome in acute respiratory distress syndrome*. Crit Care Med. 2015;43(2):394–400.
- Walton AH, Muenzer JT, Rasche D, Boomer JS, Sato B, Brownstein BH, Pachot A, Brooks TL, Deych E, Shannon WD, et al. Reactivation of multiple viruses in patients with sepsis. PLoS One. 2014;9(2):e98819.
- NM O, NM S, SM A-R, SA H, Aaa AA. The impact of cytomegalovirus infection on mechanically ventilated patients in the respiratory and geriatric intensive care units. Egypt J Chest Dis Tuberc. 2014;63:239–45.
- Heininger A, Haeberle H, Fischer I, Beck R, Riessen R, Rohde F, Meisner C, Jahn G, Koenigsrainer A, Unertl K, et al. Cytomegalovirus reactivation and associated outcome of critically ill patients with severe sepsis. Crit Care. 2011;15(2):R77.

- Bordes J, Maslin J, Prunet B, d'Aranda E, Lacroix G, Goutorbe P, Dantzer E, Meaudre E. Cytomegalovirus infection in severe burn patients monitoring by real-time polymerase chain reaction: a prospective study. Burns. 2011;37(3):434–9.
- Chilet M, Aguilar G, Benet I, Belda J, Tormo N, Carbonell JA, Clari MA, Costa E, Navarro D. Virological and immunological features of active cytomegalovirus infection in nonimmunosuppressed patients in a surgical and trauma intensive care unit. J Med Virol. 2010;82(8):1384–91.
- Chiche L, Forel J, Roch A, Guervilly C, Pauly V, Allardet-Servent J, Gainnier M, Zandotti C, Papazian L. Active cytomegalovirus infection is common in mechanically ventilated medical intensive care unit patients. Crit Care Med. 2009;37(6):1850–7.
- Ziemann M, Sedemund-Adib B, Reiland P, Schmucker P, Hennig H. Increased mortality in long-term intensive care patients with active cytomegalovirus infection. Crit Care Med. 2008;36(12):3145–50.
- Limaye AP, Kirby KA, Rubenfeld GD, Leisenring WM, Bulger EM, Neff MJ, Gibran NS, Huang ML, Santo Hayes TK, Corey L, et al. Cytomegalovirus reactivation in critically ill immunocompetent patients. Jama. 2008;300(4):413–22.
- von Muller L, Klemm A, Weiss M, Schneider M, Suger-Wiedeck H, Durmus N, Hampl W, Mertens T. Active cytomegalovirus infection in patients with septic shock. Emerg Infect Dis. 2006;12(10):1517–22.
- 20. Jaber S, Chanques G, Borry J, Souche B, Verdier R, Perrigault PF, Eledjam JJ. Cytomegalovirus infection in critically ill patients: associated factors and consequences. Chest. 2005;127(1):233–41.
- Cook CH, Martin LC, Yenchar JK, Lahm MC, McGuinness B, Davies EA, Ferguson RM. Occult herpes family viral infections are endemic in critically ill surgical patients. Crit Care Med. 2003;31(7):1923–9.
- 22. Heininger A, Jahn G, Engel C, Notheisen T, Unertl K, Hamprecht K. Human cytomegalovirus infections in nonimmunosuppressed critically ill patients. Crit Care Med. 2001;29(3):541–7.
- Kutza AS, Muhl E, Hackstein H, Kirchner H, Bein G. High incidence of active cytomegalovirus infection among septic patients. Clin Infect Dis. 1998;26:1076–82.
- Cook C, Yenchar J, Kraner T, Davies E, Ferguson R. Occult herpes family viruses may increase mortality in critically ill surgical patients. Am J Surg. 1998;176(4):357–60.
- Domart Y, Trouillet JL, Fagon JY, Chastre J, Brun-Vezinet F, Gibert C. Incidence and morbidity of cytomegaloviral infection in patients with mediastinitis following cardiac surgery. Chest. 1990;97(1):18–22.
- Mansfield S, Griessl M, Gutknecht M, Cook CH. Sepsis and cytomegalovirus: foes or conspirators? Med Microbiol Immunol. 2015; 204(3):431–7.
- Kalil A. Cytomegalovirus and mortality in critical care patients: another piece of the puzzle. Crit Care Med. 2012;40:303–5.
- Cohen JI. Cytomegalovirus in the intensive care unit: pathogen or passenger? Crit Care Med. 2009;37(6):2095–6.
- Kalil AC, Florescu DF. Prevalence and mortality associated with cytomegalovirus infection in nonimmunosuppressed patients in the intensive care unit. Crit Care Med. 2009;37(8):2350–8.
- Stang A. Critical evaluation of the Newcastle-Ottawa scale for the assessment of the quality of nonrandomized studies in meta-analyses. Eur J Epidemiol. 2010;25(9):603–5.
- 31. Hozo SP, Djulbegovic B, Hozo I. Estimating the mean and variance from the median, range, and the size of a sample. BMC Med Res Methodol. 2005;5:13.
- Frantzeskaki FG, Karampi ES, Kottaridi C, Alepaki M, Routsi C, Tzanela M, Vassiliadi DA, Douka E, Tsaousi S, Gennimata V, et al. Cytomegalovirus reactivation in a general, nonimmunosuppressed intensive care unit population: incidence, risk factors, associations with organ dysfunction, and inflammatory biomarkers. J Crit Care. 2015;30(2):276–81.
- Ong DS, Spitoni C, Klein Klouwenberg PM, Verduyn Lunel FM, Frencken JF, Schultz MJ, van der Poll T, Kesecioglu J, Bonten MJ, Cremer OL. Cytomegalovirus reactivation and mortality in patients with acute respiratory distress syndrome. Intensive Care Med. 2016;42(3):333–41.
- Hardie DR, Korsman SN, Hsiao NY. Cytomegalovirus load in whole blood is more reliable for predicting and assessing CMV disease than pp65 antigenaemia. J Virol Methods. 2013;193(1):166–8.
- 35. Razonable RR, Fanning C, Brown RA, Espy MJ, Rivero A, Wilson J, Kremers W, Smith TF, Paya CV. Selective reactivation of human herpesvirus 6 variant a occurs in critically ill immunocompetent hosts. J Infect Dis. 2002;185(1):110–3.

- Desachy A, Ranger-Rogez S, Francois B, Venot C, Traccard I, Gastinne H, Denis F, Vignon P. Reactivation of human herpesvirus type 6 in multiple organ failure syndrome. Clin Infect Dis. 2001;32(2):197–203.
- Stephan F, Meharzi D, Ricci S, Fajac A, Clergue F, Bernaudin JF. Evaluation by polymerase chain reaction of cytomegalovirus reactivation in intensive care patients under mechanical ventilation. Intensive Care Med. 1996;22(11):1244–9.
- Ho M. Cytomegalovirus infection in patients with bacterial sepsis. Clin Infect Dis. 1998;26(5):1083–4.
- Griffiths PD, Cope AV, Hassan-Walker AF, Emery VC. Diagnostic approaches to cytomegalovirus infection in bone marrow and organ transplantation. Transpl Infect Dis. 1999;1(3):179–86.
- Yun Z, Lewensohn-Fuchs I, Ljungman P, Ringholm L, Jonsson J, Albert J. A real-time TaqMan PCR for routine quantitation of cytomegalovirus DNA in crude leukocyte lysates from stem cell transplant patients. J Virol Methods. 2003:110(1):73–9.
- Sylwester AW, Mitchell BL, Edgar JB, Taormina C, Pelte C, Ruchti F, Sleath PR, Grabstein KH, Hosken NA, Kern F, et al. Broadly targeted human cytomegalovirus-specific CD4+ and CD8+ T cells dominate the memory compartments of exposed subjects. J Exp Med. 2005;202(5):673–85.
- Hotchkiss RS, Monneret G, Payen D. Sepsis-induced immunosuppression: from cellular dysfunctions to immunotherapy. Nat Rev Immunol. 2013; 13(12):862–74.
- 43. Cohen J, Vincent JL, Adhikari NK, Machado FR, Angus DC, Calandra T, Jaton K, Giulieri S, Delaloye J, Opal S, et al. Sepsis: a roadmap for future research. Lancet Infect Dis. 2015;15(5):581–614.
- 44. Howard RJ, Simmons RL. Acquired immunologic deficiencies after trauma and surgical procedures. Surg Gynecol Obstet. 1974;139(5):771–82.
- Gascon P, Zoumbos NC, Young NS. Immunologic abnormalities in patients receiving multiple blood transfusions. Ann Intern Med. 1984;100(2):173–7.
- Tarnok A, Schneider P. Pediatric cardiac surgery with cardiopulmonary bypass: pathways contributing to transient systemic immune suppression. Shock. 2001;16(Suppl 1):24–32.
- Bone RC. Sir Isaac Newton, sepsis, SIRS, and CARS. Crit Care Med. 1996; 24(7):1125–8.
- Cook CH, Trgovcich J, Zimmerman PD, Zhang Y, Sedmak DD.
 Lipopolysaccharide, tumor necrosis factor alpha, or interleukin-1beta triggers reactivation of latent cytomegalovirus in immunocompetent mice. J Virol. 2006;80(18):9151–8.
- Lawlor G, Moss AC. Cytomegalovirus in inflammatory bowel disease: pathogen or innocent bystander? Inflamm Bowel Dis. 2010;16(9):1620–7.
- Chanques G, Jaber S. Treating HSV and CMV reactivations in critically ill
 patients who are not immunocompromised: con. Intensive Care Med. 2014;
 40(12):1950–3.

Ready to submit your research? Choose BMC and benefit from:

- fast, convenient online submission
- thorough peer review by experienced researchers in your field
- rapid publication on acceptance
- support for research data, including large and complex data types
- gold Open Access which fosters wider collaboration and increased citations
- maximum visibility for your research: over 100M website views per year

At BMC, research is always in progress.

Learn more biomedcentral.com/submissions

