

Review Article

Polymeric Micelles, a Promising Drug Delivery System to Enhance Bioavailability of Poorly Water-Soluble Drugs

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Oral administration is the most commonly used and readily accepted form of drug delivery; however, it is found that many drugs are difficult to attain enough bioavailability when administered via this route. Polymeric micelles (PMs) can overcome some limitations of the oral delivery acting as carriers able to enhance drug absorption, by providing (1) protection of the loaded drug from the harsh environment of the GI tract, (2) release of the drug in a controlled manner at target sites, (3) prolongation of the residence time in the gut by mucoadhesion, and (4) inhibition of efflux pumps to improve the drug accumulation. To explain the mechanisms for enhancement of oral bioavailability, we discussed the special stability of PMs, the controlled release properties of pH-sensitive PMs, the prolongation of residence time with mucoadhesive PMs, and the P-gp inhibitors commonly used in PMs, respectively. The primary purpose of this paper is to illustrate the potential of PMs for delivery of poorly water-soluble drugs with bioavailability being well maintained.

1. Introduction

Oral administration is the most commonly preferred route for drug delivery because of its simplicity, convenience, and patient acceptance, especially in the case of repeated dosing for chronic therapy [1–3]. In contrast to the intravenous administration, which probably results in toxic blood level after injection and sometimes an under concentration of the desired threshold towards the end of the dosing interval, oral chemotherapy can provide a prolonged and continuous exposure to a relatively lower and thus safer concentration [2]. Now, more than 60% of marketed drugs are used as oral products [4]. However, it is intricate to formulate a therapeutic agent for oral administration. The bioavailability of oral drugs is strongly influenced by two important parameters, solubility and permeability [3]. Based on that, the Biopharmaceutical Classification System (BCS) defines four categories of drugs [5]. Many existing and new therapeutic entities are characterized as BCS class II (low solubility and high permeability) or BCS class IV (low solubility and low permeability). Poorly water-soluble drug candidates encountered in drug

discovery cause increasing problems of poor and variable bioavailability. It is estimated that approximately 70% of new chemical entities are poorly soluble in aqueous medium and many even in organic medium. Besides, approximately 40% of currently marketed immediate-release oral drugs are considered practically insoluble (solubility less than 100 $\mu\text{g/mL}$) in water [6, 7]. Low solubility limits the drug dissolution rate, frequently resulting in low bioavailability of the oral drug [8]. To achieve the desired therapeutic concentration in the target sites, dose escalation study of the drug was often applied in clinic [9, 10]. However, it may be undesirable due to the possibility of increased toxicity and therefore decreased patient compliance. Meanwhile, the high drug loading of pharmaceutical products often makes it difficult to complete the study [11].

Nanotechnology brings some advantages to the drug delivery, particular for oral drug. It allows (1) the delivery of poorly water-soluble drugs; (2) the targeting of drugs to specific parts of the gastrointestinal tract (GI); (3) the transcytosis of drugs across the tight intestinal barrier; and (4) the intracellular and transcellular delivery of large

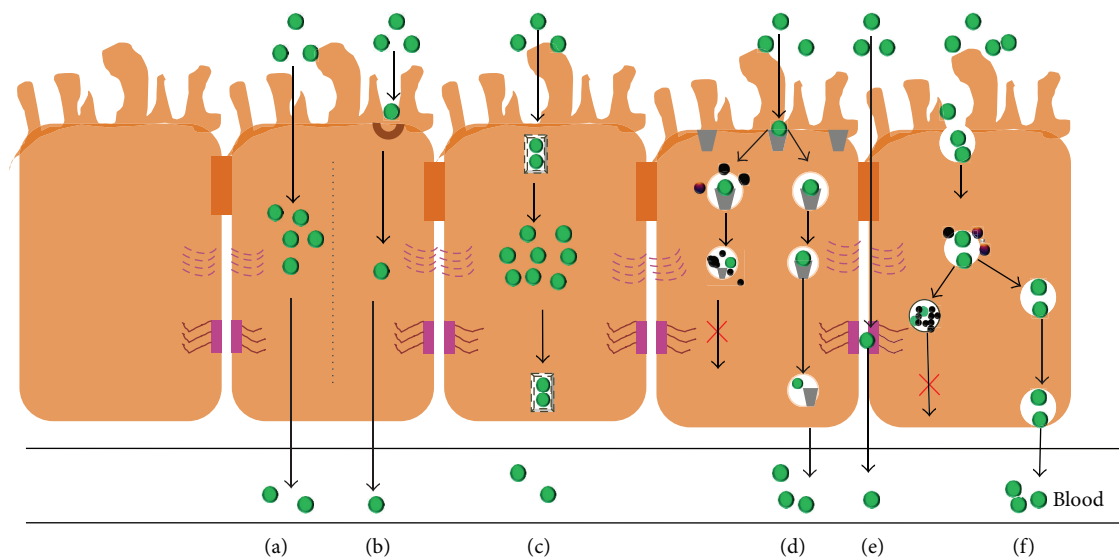


FIGURE 1: Schematic representation of the mechanisms involved in the absorption of exogenous drugs in the small intestine. (a) Transcellular transport; (b) active transport; (c) facilitated diffusion; (d) receptor-mediated endocytosis; (e) paracellular transport; (f) pinocytosis [3].

macromolecules [12, 13]. In recent years, nanotechnology has been widely focused on by numbers of researchers throughout the world for its superiority in increasing efficacy, specificity, tolerability, and therapeutic index of corresponding drugs [14]. Several strategies have been proposed such as micronization, complexation, formation of solid solutions, microemulsification, and novel drug delivery systems, including nanoparticles, lipid-based vesicles, and micelles [15–18]. Among these approaches, polymeric micelles (PMs) have gained considerable attention in the last two decades as a multifunctional nanotechnology-based delivery system for poorly water-soluble drugs. The application of PMs as drug delivery system was pioneered by the group of H. Ringsdorf in 1984 [19] and subsequently used by Kataoka in the early 1990s through the development of doxorubicin-conjugated block copolymer micelles [20]. Due to their nanoscopic size, ability to solubilize hydrophobic drugs in large amounts and achieve site-specific delivery, PMs hold promise to obtain desirable biopharmaceutical and pharmacokinetic properties of drugs [21] and enhance their bioavailability. In this review article, we will discuss the development of the PMs and focus on the mechanisms of various kinds of PMs for enhancement of oral bioavailability.

2. Absorption of Oral Drugs in the Gastrointestinal Tract

2.1. Pathways of Drug Absorption. A drug that is administered orally must survive transit through the gastrointestinal (GI) tract. Although part of the absorption process occurs in the oral cavity and stomach due to the presence of salivary amylase and gastric protease (pepsin), the small intestine remains the major site for absorption [22]. There exist many pathways for nutrient absorption in the small intestine; however, the absorption of oral drugs is restricted to either transport through the cells (transcellular pathway, see

Figure 1(a)) or between adjacent cells (paracellular pathway, see Figure 1(e)) [3]. Generally, the low-molecular weight hydrophobic entities which are able to diffuse through the membrane are absorbed by the transcellular pathway, and the absorption rate is determined by the concentration gradient across the intestinal membrane (Figure 1(c)). On the contrary, hydrophilic molecules cannot freely diffuse through the intestinal membrane, due to their low affinity for the lipidic constituents [23]. Therefore, in the absence of an appropriate membrane transporter, the paracellular pathway is the only available route for their absorption (Figure 1(e)). In some particular instances, drugs may be absorbed by fluid-phase endocytosis (pinocytosis), an energy-dependent saturable process in which the molecule travels inside membrane vesicles (Figure 1(f)).

2.2. Barriers for Absorption of Oral Drugs. Although oral administration is the preferred route for drug delivery, and the mechanisms of drug absorption have been widely studied, there still exists the serious problem of low bioavailability which has severely impeded the development of oral therapy. The bioavailability of a drug strongly depends on its intrinsic properties and physiological conditions. A drug that is administered orally must survive transit through the chemical and enzymatic GI liquids, cross the mucus layer and the epithelium before being absorbed [24, 25]. Intrinsic properties of drugs such as poor stability in the gastric environment, low mucosal permeability, and low solubility in the mucosal fluids will contribute to low absorption [26, 27]. Physiological factors such as gastrointestinal transit time, regional pH, surface area, enzymatic activity, and colonic microflora will also influence drug absorption [28].

Therefore, to achieve good absorption and bioavailability, oral drugs should be stable at the low gastric pH and have a reproducible and good pharmaceutical dissolution profile and adequate hydrophilic/lipophilic balance to cross

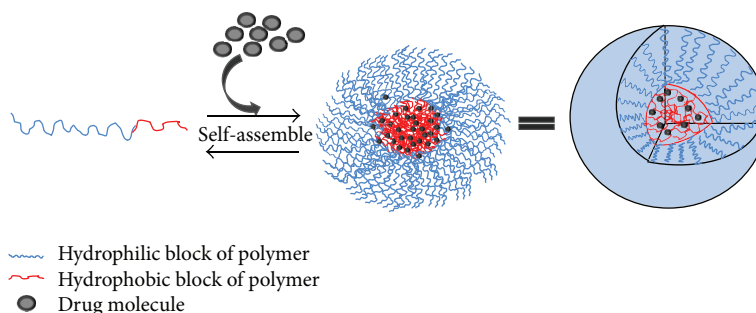


FIGURE 2: Formation and drug loading of PMs by self-assembly of amphiphilic block copolymers in aqueous solution.

the intestinal epithelial membrane. Furthermore, they should not induce significant gastrointestinal toxicities, such as nausea, vomiting, loss of appetite, or diarrhea, that would limit continued oral administration or result in poor compliance [29, 30]. To overcome these barriers and achieve above-mentioned requirements, several strategies have been proposed including the reduction of drug particle size [31], salt formation [32], or prodrug synthesis [33]. It is worth mentioning that nanosized carriers such as PMs [34] could encapsulate drugs into protective vehicles, avoiding destruction in the GI tract and releasing them in a temporally or spatially controlled manner, which could potentially enhance drug absorption and offer a promising direction for oral therapy [28].

3. Introduction of PMs

3.1. Formation of PMs. PMs are self-assembled core-shell nanostructures formed in an aqueous solution consisting of amphiphilic block copolymers (see Figure 2) [35, 36]. Formation of micelles in aqueous solution occurs when the concentration of the block copolymer increases above a certain concentration named the critical aggregation concentration (CAC) or critical micelle concentration (CMC). At the CAC or CMC, hydrophobic segments of block copolymers start to associate to minimize the contact with water molecules, leading to the formation of a vesicular or core-shell micellar structure.

Theoretically, the formation of micelles is driven by decrease of free energy. The removal of hydrophobic fragments from the aqueous environment and the reestablishing of hydrogen bond network in water decrease free energy of the system and finally form the micelles. The typical methods used for encapsulation of poorly water-soluble drugs are dialysis method, oil-in-water emulsion solvent evaporation method, and solid dispersion method [37, 38]. Other methods used are direct dissolution [39], complexation [40], chemical conjugation [41], and various solvent evaporation procedures [42].

3.2. Structure of PMs. PMs present a great potential as a drug delivery system for compounds that are hydrophobic and exhibit poor bioavailability which results from the unique core-shell structure. The inner hydrophobic core enables incorporation of poorly water-soluble drugs thus improving

their stability and bioavailability. Typically, the inner core of the PMs was formed with hydrophobic blocks of the copolymers by hydrophobic interaction. Besides, it can also be formed by electrostatic interactions, using charged block copolymers of oppositely charged macromolecules, resulting in the formation of polyion complex (PIC) micelles [43, 44]. In addition, there have been reports of PMs formed by complexation via hydrogen bonding [45–47] as well as metal-ligand coordination interactions [48], both referred to as noncovalently connected micelles. The outer shell of PMs was formed by the hydrophilic blocks of the copolymers, playing an important role in the *in vivo* behavior, particular for their steric stabilization and ability to interact with the cells [49]. Lengths of the hydrophobic and hydrophilic blocks affect the conformation of polymers in medium, as lengthier hydrophilic blocks of polymer cause it to remain monomeric in water [50].

Amphiphilic copolymers which constitute PMs are usually block copolymers [51, 52]. Block copolymers can be diblock copolymers or triblock copolymers. Generally, diblock copolymers of the A-B type, where A represents a hydrophilic block and B represents a hydrophobic block, are commonly used to design PMs, whereas triblock copolymers consist of two types of polymers (ABA) [53] or three types of polymers (ABC). Most drug carrier applications have been studied with A-B or A-B-A type block copolymers due to the close relationship between the properties of micelles and the structure of polymers [54]. The physicochemical characteristics of the building blocks influence the physical and biological properties of the PMs [55]. Hence, micelle-forming block copolymers have been the focus of several studies over the past few years. For oral drug delivery system, the block copolymers used to form micelles should (1) spontaneously self-assemble in water, (2) enhance drug solubility by several orders of magnitude and provide high loading efficiency, (3) remain stable upon dilution in the GI tract, (4) be biocompatible and nontoxic, and (5) be easy to synthesize at large scale [28, 56, 57]. The choice of core-forming polymers is the major determinant for important properties of PMs such as stability, drug loading capacity, and drug release profiles [58]. Poly(propylene oxide) (PPO) [53, 59] which belongs to Pluronics, poly(esters) such as poly(lactic acid) (PLA) [60], hydrophobic poly(amino acids) [61], copolymers of lactic acid and glycolic acids [62, 63], and poly(caprolactone) (PCL) [64], which are regarded as the

commonly used core-forming blocks of PMs, and have been studied in the past 10 years. These core-forming polymers cover a wide range of structural diversity and polarity for solubilizing numbers of poorly water-soluble drugs [51, 52]. Meanwhile, the chemical nature and molecular weight of the hydrophilic block will strongly affect the stealth properties and accordingly influence the circulation kinetics of the micellar assembly. Poly(ethylene glycol) (PEG) is most commonly used as the hydrophilic segment of the block copolymers, since it is a nontoxic polymer with FDA approval as a component of various pharmaceutical formulations. Furthermore, its unique physicochemical properties (high water solubility, high flexibility, and large exclusion volume) provide good “stealth” properties for PMs [65, 66], while poly(N-vinyl-2-pyrrolidone) (PVP) [67] and poly(acrylic acid) (PAA) [68] are frequently used as PEG alternatives.

4. PMs for Enhancement of Bioavailability

The main mechanisms involved in the enhancement of drug absorption by PMs are: (1) protection of the loaded drug from the harsh environment of the GI tract, (2) release of the loaded drug in a controlled manner at target sites, (3) prolongation of the residence time in the gut by mucoadhesion, and (4) inhibition of efflux pumps to improve drug accumulation [69]. Several physicochemical parameters seem to influence translocation of micelles across the epithelium, including surface hydrophobicity, polymer nature, and particle size [69]. There exist many characteristics of PMs that allow them to traverse across the epithelium. For example, PMs with appropriate particle size can be taken up and then cross the intestinal barrier [40, 70, 71]. Furthermore, to achieve a good bioavailability, drugs may be delivered at a specific region in the GI tract, the so-called absorption window. To reach the absorption window, PMs can be manipulated by coupling different types of polymers or by grafting various functional groups at the hydrophilic end of the copolymer, such as the pH-sensitive [72–74] and receptor sensitive groups [75].

4.1. Special Stability of PMs for Enhancement of Bioavailability. As we discussed above, GI tract is the major barrier for oral drugs. After oral administration, drugs will encounter the harsh physicochemical environment of the GI tract and be degraded due to the variation of pH levels as well as the presence of enzymes or bile salts. To ensure delivery of the carried drugs to the absorption sites, PMs must be able to resist rapid dissociation upon dilution and retain the stable core-shell structure before target sites. It is known that PMs possess two aspects of structural stability, thermodynamic and kinetic, provided by the entanglement of polymer chains in the inner core [76–78].

For a micelle to be thermodynamically stable, the copolymer concentration should be above its CMC. The CMC is influenced by the hydrophilic-lipophilic balance (HLB) of the block copolymer [79]. A reverse relationship between the CMC and hydrophobicity of the core-forming blocks has been shown in many studies: an increase in the hydrophobic block length results in a lower CMC if the hydrophilic

segment is kept constant [80]. Generally, PMs show very low CMC values in a range from 10^{-6} to 10^{-7} M. These CMC values are much smaller than those of micelles formed from low-molecular weight surfactants (10^{-3} – 10^{-4} M) [81], which allows a series of dilution and still retain the micellar structure. The second aspect, kinetic stability of PMs, comes into the picture when the concentration of the copolymer falls below the CMC. Kinetic stability may be more important than the thermodynamic stability for the nonequilibrium drug delivery conditions. Unlike micelles formed from low molecular weight surfactant molecules, the kinetic stability of PMs is high for the stiff or bulky core structure. Therefore, the disassembly of PMs at a concentration below CMC occurs at a relatively slower rate because of the relatively high kinetic stability. The slow dissociation allows PMs to retain their integrity and drug content before reaching the target sites, which is also helpful to enhance oral bioavailability.

4.2. pH-Sensitive PMs for Enhancement of Bioavailability. It is indicated that non-pH-sensitive micelles may enhance drug solubilization but probably not necessarily the drug absorption. Free (readily absorbable) form of a drug is one of the most important requirements for absorption in the GI tract. However, drug release from such PMs will occur only by diffusion when polymer concentration is well above the CMC, preventing the complete drug release [11]. Moreover, Camilleri once studied the stomach emptying time (ca. 177 min) and the small bowel transit time (ca. 168 min) in healthy human volunteers by monitoring the migration of a radio-labeled marker previously mixed in their meal [82]. Thus, there is also a possibility that the PMs might be excreted before complete drug release or drug might not be released close to its absorption window in the GI tract. Several PMs systems designed to increase the oral bioavailability of hydrophobic compounds exhibit release times which largely exceeded the transit time in the small intestine [83, 84]. This is also true for surfactant micelles which have been found in some cases to impede the absorption of hydrophobic drugs due to excessive retention in the micellar phase [85]. Hence, when developing oral formulations for poorly water-soluble drugs, it is important to adequately control the release rate in order to avoid either precipitation upon dilution or sequestration within the micellar phase which may lead to incomplete absorption.

4.2.1. Introduction of pH-Sensitive PMs. The potential disadvantage of normal PMs can be solved by application of additional stimuli that cause micelle destabilization in a specially controlled manner thus increasing the selectivity and efficiency of drug delivery to target sites. External factors such as heat [86, 87], light [88], and sound (ultrasound) [89, 90] have already been studied by many researchers. However, these external stimuli may only activate the carriers that are situated closely underneath the skin but not those deeply distributed in the body. The intracellular signals also play an important role in regulating drug release which causes a great deal of interests, and here we focus our attention on pH-responsive systems.

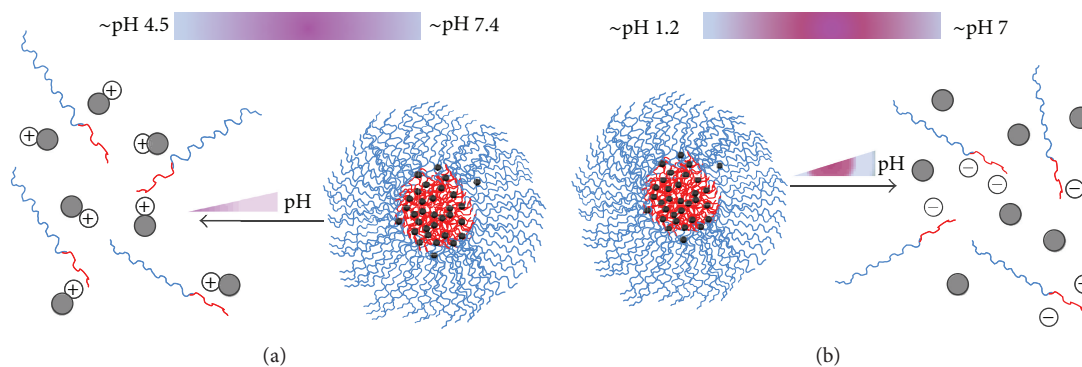


FIGURE 3: Schematic representation of the mechanisms of pH sensitivity. (a) PMs with basic core units, (b) PMs with acidic core units.

As is known, blood and normal tissues have a pH of 7.23 [91]. The mildly acidic pH encountered in a tumor (pH \sim 6.8) as well as endosomes and lysosomes (pH 5.0–5.5) provides a potential trigger to accelerate the degradation of pH-sensitive PMs and release of encapsulated drugs. Therefore, numerous pH-sensitive polymeric micellar systems have been employed for intravenous administration of anticancer drugs to tumors [92–94]. In the GI tract, the pH varies from high acidity in the stomach (pH 1.0–2.5) to a neutral or slightly alkaline pH in the small intestine (pH 5.1–7.8) [95]. Such wide variation of pH along the GI tract has been utilized for controlled drug release from carriers [2]. Strategies to prevent GI degradation or to promote absorption in the intestine by making use of the pH gradient appear promising.

4.2.2. Mechanisms of pH-Sensitive PMs for Enhancement of Bioavailability. Among the various polymers composed micelles, polyacids, or polybases may be used as building blocks that impart pH sensitivity to drug release [73, 96]. Basic core monomeric units such as amines are uncharged and thus hydrophobic at high pH condition while hydrophilic upon protonation at low pH (see Figure 3(a)). On the contrary, acidic core units such as carboxylic acids are uncharged when protonated at low pH and become negatively charged at a relatively high pH (see Figure 3(b)). Many examples of “protonation” approaches to trigger destabilization of micelles have been reported, such as incorporating L-histidine [97, 98], pyridine, and tertiary amine groups [99] in their hydrophobic segments. PMs are formed at a pH above the pKa of the protonatable group, where the hydrophobic segment essentially is uncharged. As the pH decreases below the pKa, the ionization of the polymer causes increased hydrophilicity and electrostatic repulsions of the polymers, leading to the destabilization of the micelles and controlled drug release.

4.2.3. Polymers Commonly Used in Oral pH-Sensitive PMs. Acrylic-based polymers are widely used in oral pH-sensitive drug delivery, such as poly(methacrylic acid) (PMAA). PMAA retains a collapsed state in the low pH of the stomach and swells as it transits through the intestines. Blends of this polymer with polyethylacrylate (PMAA-PEA) and polymethacrylate (PMAA-PMA) can be tailored to dissolve

in specific pH ranges corresponding to specific locations in the GI tract [100–102]. These pH-responsive micelles based on the acrylic acid core can be either multimolecular or unimolecular [103, 104]. Upon pH increase, the core of the unimolecular micelles became more polar hence promoting the release of the incorporated hydrophobic drug [103]. As these micelles do not possess a CMC, they have the advantage of being intrinsically stable upon dilution. Conversely to unimolecular micelles that maintain their integrity upon a change in pH, pH-sensitive multimolecular PMs based on ionizable polyanions disassemble following an increase in environmental pH.

Kim and his coworkers hypothesized that the physical stability of hydrotropic polymeric (HP) micelles containing AA moieties may decrease in the intestine, releasing the loaded drugs faster in the intestine than in the stomach [105]. To examine this hypothesis, they took paclitaxel (PTX) as model drug and developed a hydrotropic polymer, PEG-b-(4-(2-vinylbenzyloxy)-N,N-(diethylnicotinamide)) (PEG-b-VBODENA), doped with AA units (\leq 50 mol%) to confer pH sensitivity to PMs, testing PTX loading/release profiles by changing the pH condition. They observed that the loading content and efficiency of PTX were governed by the pH of the loading medium, with both maxima at $\text{pH} \leq 4$. Increasing the pH above the pKa of the polymers provoked a rapid dissociation of the complexes. The self-association into well-defined micellar structure is facilitated by the hydrophobic nonionizable Al(M)A units, whereas the pH sensitivity is conferred by the carboxylic acid groups of the MAA moieties [106]. The PTX release from HPC with more than 20% AA contents was completed within 12 h in a simulated intestinal fluid (pH = 6.5) while the PMs without any AA moiety showed very slow release profiles. Therefore, the pH-dependent release of PTX from HP micelles can be used to increase the bioavailability of PTX upon oral delivery.

Some other groups have also developed the pH-sensitive oral drug delivery systems. In an earlier report, Sant et al. prepared and characterized a pH-sensitive PMs incorporating poorly water-soluble model drugs [104]. The self-assemblies were constructed from novel pH-sensitive polymers composed of poly(ethylene glycol)-block-poly(alkyl acrylate-co-methacrylic acid) (PEG-b-P(AIA-co-MAA)). Due to the presence of pendant carboxyl groups in the hydrophobic part,

these copolymers exhibit pH-dependent aggregation behavior and form supramolecular micelles below pH 4.7. Hence, these copolymers dissociate partially or completely with increase in pH owing to the ionization of carboxylic groups. Two water-insoluble model drugs, named indomethacin (IND) and fenofibrate (FNB), were incorporated in the supramolecular assemblies by dialysis or oil-in-water (O/W) emulsion methods. The pH-dependent drug release in vitro from the micelles was also confirmed in their study. To make sure whether pH-sensitive PMs could improve the bioavailability of a poorly water-soluble drug, further in vivo study was carried out [1]. For in vivo study, FNB was chosen as the poorly water-soluble model drug. The pharmacokinetics of FNB incorporated in PMs was evaluated in male Sprague-Dawley rats after oral dosing and compared with the commercial micronized formulation, Lipidil MicroR and an FNB coarse suspension. The oral bioavailability of FNB from these self-assemblies revealed 156% and 15% increases versus FNB coarse suspension and Lipidil MicroR, respectively. The results suggest that these pH-sensitive PMs could efficiently improve the bioavailability of poorly water-soluble drugs. Other types of pH-controlled release carriers such as pH-sensitive polymer-drug conjugates [107, 108] are beyond the scope of PMs and not discussed in this review.

4.3. Mucoadhesive PMs for Enhancement of Bioavailability

4.3.1. Introduction of Mucoadhesive PMs. Nanocarriers for oral administration should adhere to mucus and cross the mucus layer. Drugs delivered to mucosal surfaces are usually efficiently removed by mucus clearance mechanisms [109]. The luminal surface of mucosal tissues is protected by a highly viscoelastic layer [110], and the protective coatings rapidly remove foreign particles from the GI tract which probably lead to low bioavailability. Unlike the relatively high requirements of intravenous infusions, oral formulations could include high-molecular weight polymers as long as these components are metabolizable and cannot find their way into the systemic circulation. Hence, it may be an effective means of increasing uptake of drugs with mucoadhesive PMs [111, 112], and there have been considerable interests in the concept of mucoadhesive PMs. Firstly, mucosal retention can be used to increase the transit time in the GI tract, resulting in prolonged time window for the release of the payload. Secondly, mucoadhesive polymers swell and fill the crevices of the mucous membrane, contributing to the effective surface area in contact with the intestinal mucosa and yielding a high local concentration of the drug [113]. Thirdly, bioadhesion could also localize the PMs at a given target site and increase the drug concentration gradient for the intense contact of the particles with the mucosal surface [27].

The ability to develop mucoadhesive interactions within the gut would be one of the key factors influencing their ability to promote oral absorption of the loaded drug. It was demonstrated that there exists a direct relationship between mucoadhesion and drug absorption [114, 115]. In fact, the development of adhesive interactions (between PMs and mucosa) would probably induce the immobilization of these

carriers in intimate contact with the absorptive membrane. This fact would facilitate the establishment of a concentration gradient of the loaded drug from the PMs to the circulation, which finally results in an enhancement of absorption and bioavailability.

4.3.2. Mechanisms of Mucoadhesive PMs for Enhancement of Bioavailability. Mucoadhesion is a complex phenomenon, and several steps have been suggested in mucoadhesive bond formation [116]. The first step is the spreading, wetting, and dissolution of mucoadhesive polymer at the interface. The second step is the mechanical or physical entanglement between the polymer and the tissue surface mucus layer, resulting in an interpenetration layer. The next step is the result of chemical interactions [116]. Mucoadhesion can be obtained by the building of either nonspecific interactions with the mucosal surface, such as covalent bonds, ionic bonds, hydrogen bonding, and van der Waals' interactions [117], or specific interactions by functionalizing polymers with targeting ligands (e.g., lectins [118, 119]) or reactive groups such as thiols [120].

The fates of the mucoadhesive PMs in the GI tract include at least three different pathways: mucoadhesion, translocation through the mucosa or transit, and direct faecal elimination. Among the various factors, the surface charges of PMs seem to play an important role in particle uptake. On one hand, the negatively charged intestinal mucosa, due to the existence of glycocalyx, attracts more positively charged PMs. Therefore, a considerable number of studies have been conducted using positively charged polymers such as chitosan to increase residence time in the GI tract [121, 122]. On the other hand, the particle mobility also seems to be strongly dependent on surface charges, and it was indicated that transport rates were inversely related to particle surface potentials. Negatively charged particles display significantly higher transport rates than near neutral or positively charged particles whose transport was probably limited by particle aggregation and electrostatic adhesive interactions with mucosa [123]. Crater and Carrier demonstrated a 20–30 times faster diffusion for anionic particles in comparison with cationic ones [123], which proved the opinion discussed above. Therefore, it is crucial to control the balance between mucoadhesion and mucus penetration for an efficient oral delivery.

4.3.3. Polymers Commonly Used in Mucoadhesive PMs. Polymers such as cross-linked polyacrylic acids (PAA) [124–126], carboxypolymethylene, carboxymethyl cellulose, alginate, chitosan (CS), and their derivatives [127–129] are commonly accepted as mucoadhesive and safe polymers. Mucoadhesive polymers, especially positively charged polymers, were preferential to enhance drug absorption by prolonging the residence time at the site of absorption. Chitosan (CS), a linear amino polysaccharide composed of randomly distributed (1–4) linked d-glucosamine and N-acetyl-d-glucosamine units, is a well-known naturally occurring cationic biopolymer, which has received increasing attention owed to its biocompatibility, nontoxicity, and low immunogenicity [130, 131]. The adhesive properties of chitosan caused by the development of

electrostatic interactions with glycoproteins of mucus [132] are of primary interest for oral delivery and its cationic properties below pH 6.5 favor the mucoadhesive ability. Moreover, among the existing cationic polymers, chitosan is an ideal candidate for oral DNA and protein delivery [133] due to its mucoadhesive properties and its ability to induce a transient opening of the tight junctions [134]. Nevertheless, due to the insolubility of chitosan observed at pH values above its pKa (6.4) in water, micelles of amphiphilic chitosan rapidly precipitate in biological solution (pH 7.4). Therefore, water-soluble chitosan derivatives have often been used for development of drug delivery systems like glycol chitosan (GC) and chitosan oligosaccharide (CSO), exhibiting good solubility over a broad range of pH [135, 136].

Other synthetic mucoadhesive polymers have been currently investigated in pharmaceutical formulations including PEG, cellulose derivatives (methylcellulose) [137, 138] and hydroxypropyl cellulose (HPC) [139], and polyelectrolytes (PAA) [39]. These polymers bind to the mucus via noncovalent bonds such as hydrogen bonding, electrostatic interactions, and van der Waals forces. Mucus interpenetration and chain entanglement may also contribute to the phenomenon of mucoadhesion, particularly with regard to uncharged polymers. Another commonly used mucoadhesive polymers are Pluronic-PAA copolymers. Strong mucoadhesive properties of the Pluronic-PAA copolymers originate from both the carboxyl-mucin interactions and the ability of the polyether segments to interpenetrate into and anchor the copolymer on the mucosa [124]. Mucoadhesive parameters of several types of Pluronic-PAA copolymers have already exceeded those of Carbopol or carbomer (lightly cross-linked PAA), which is an industry standard for mucoadhesive polymers used as pharmaceutical excipients. According to previous studies, mucoadhesive PAA and thiomers increase the residence time of insulin at the site of intestinal absorptive cells, thus enhancing its intestinal absorption [140–142]. Investigators assumed that the insulin uptake can be significantly enhanced after oral administration due to the positive attributes of the thiomers PAA-Cys including mucoadhesion, permeation enhancement and shielding against enzymatic degradation.

Much stronger bioadhesion can be achieved by functionalizing polymers with targeting ligands (e.g., lectins) [118, 119] or reactive groups such as thiols [120]. Lectins are proteins or glycoproteins of nonimmunological origin which specifically recognize sugar molecules and therefore are capable of binding to glycosylated membrane components [143, 144]. Sugars are all present in glycoproteins and glycolipids of mammalian mucosa, either at the surface of epithelial cells or in mucous layers. Through strong adherence to glycoproteins and glycolipids in the membrane of enterocytes, lectins may prove useful in both prolonging the transit time of a host cargo through the small intestine as well as promoting its uptake via receptor-mediated endocytosis. Bernkop-Schnürch and coworkers have demonstrated that the thiolation of classical PMs substantially increases their mucoadhesive properties and therefore further improves the oral absorption of therapeutic proteins [145]. Surface-exposed thiols are thought to form disulfide bonds with cysteine-rich subdomains of mucus glycoproteins. Thiolated polymers also exhibit

an increased permeation-enhancing effect as well as enzyme inhibitory properties [145]. Thiol-decorated polyion complex micelles prepared through complexation between PEG-*b*-poly(2-(*N,N*-dimethylamino)ethyl methacrylate) and a 20-mer oligonucleotide have been shown to interact with mucin through the formation of disulfide bonds [146]. While these micelles were initially designed to carry nucleic acid drugs, a similar strategy may be applied to deliver hydrophobic drugs through the use of thiol-functionalized PEG-*b*-PLA or PEG-*b*-PCL PMs [147].

4.4. *P-gp* Inhibitors for Enhancement of Bioavailability

4.4.1. *Introduction of P-gp.* Besides uptake, drugs are often pumped out of enterocytes by efflux transporters on the surface of intestinal mucosa. The extent of absorption for poorly water-soluble drugs (and orally administered drugs in general) is affected by these efflux pathways [148]. Among the efflux transporters, the most well known and widely studied is the P-glycoprotein (P-gp) efflux transporters [149]. Pgp is a 170-kDa membrane transporter which is part of the ATP-binding cassette (ABC) [150]. Using ATP, the human multidrug resistance-associated protein (MDR1) and P-gp can actively transport a wide range of relatively hydrophobic, amphipathic drugs out of the cell. When drugs encapsulated in PMs, they remain mainly associated with the particles and are not likely to be substrate of the efflux pumps. However, hydrophobic drugs can be released from the micelles and are more likely to be transported by the efflux pumps [151]. Compounds transported by P-gp include important anticancer drugs like Vinca alkaloids [152], anthracyclines [153], epipodophyllotoxins, and taxanes [154]. So ABC transporters may reduce the amount of drug absorbed and limit bioavailability in a dose-dependent, inhibitable, and saturable manner [155]. Due to its ability to expel therapeutics, the presence of intestinal P-gp is associated with a decrease in oral bioavailability and is thought to be one of the most significant causes for decreased permeability and therefore oral bioavailability. Therefore, modulation of its activity is regarded as a potential means to improve drug bioavailability.

4.4.2. *Polymers Commonly Used in P-gp Inhibiting PMs for Enhancement of Bioavailability.* The first P-gp inhibitors proposed were substrates that could bind to the protein and inhibit its activity. Several drugs, including cyclosporine A (cyA) and verapamil, have been studied for this purpose [156, 157]. However, these molecules may be associated with toxic side effects, and amphiphilic polymers were presented as a potential alternative [158]. Mostly, the inhibition of efflux transport with amphiphilic polymers appears to be related to a modification of the fluidity of the cellular membrane [159]. This inhibitory effect has been demonstrated with both low-molecular weight and polymeric micelles, among which D- α -tocopheryl polyethylene glycol succinate (TPGS) [160, 161] and Pluronics have been extensively studied.

Pluronic block copolymers (also known under their nonproprietary name “poloxamers”) consist of hydrophilic ethylene oxide (EO) and hydrophobic propylene oxide (PO)

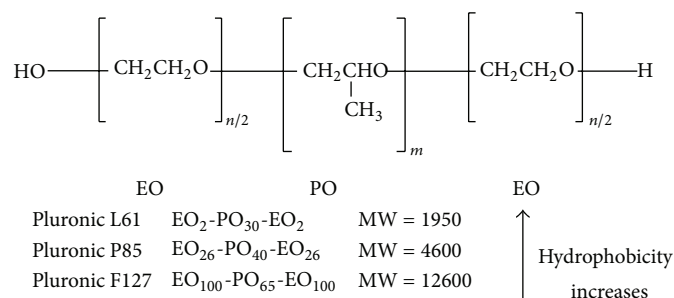


FIGURE 4: Pluronic block copolymers available from BASF (Wyandotte, MI, USA) contain two hydrophilic EO blocks and a hydrophobic PO block [167].

blocks arranged in a basic A-B-A structure: EO_{n/2}-PO_m-EO_{n/2}. The structure formula of Pluronic block copolymers is shown in Figure 4. Membrane fluidization is known to contribute to inhibition of P-gp efflux function. Pluronic block copolymers are known to induce drastic changes in the microviscosity of cell membranes, and these changes can be attributed to the alterations in the structure of the lipid bilayers as a result of absorption of the block copolymer molecules on the membranes [162]. Yoncheva et al. once prepared, characterized, and evaluated the pharmacokinetics of PTX incorporated in stabilized Pluronic micelles [49]. The stabilization of micelles was performed by cross-linking of their core, aiming to prevent disaggregation of micelles upon dilution in physiological fluids. Moreover, Pluronic copolymers may inhibit the activity of drug efflux transporters such as P-gp, MRPs, and BCRP [163, 164], which make it an adequate strategy to increase the bioavailability and promote the efficacy of PTX. Furthermore, it is believed that inhibition of P-gp ATPase activity, presumably through nonspecific changes in lipid and protein conformation and mobility, has a major contribution to the inhibition of P-gp efflux function [3]. Pluronic copolymers could inhibit drug efflux transporters, drug sequestration in acidic vesicles, and the glutathione/glutathione S-transferase detoxification system in an energy-dependent manner. Therefore, ATP depletion caused by the inhibition of the ATPase activity induced by the Pluronic copolymers has been proposed to be a reason for chemosensitization of these cells [165, 166].

D-α-tocopheryl polyethylene glycol succinate (Vitamin E TPGS or simply TPGS) (see Figure 5) is a water-soluble derivative of natural Vitamin E, which is formed by esterification of Vitamin E succinate with polyethylene glycol (PEG) [168]. Therefore, it has advantages over PEG and Vitamin E in application of various drug delivery device, including extending the half-life of the drug in plasma and enhancing the cellular uptake [169]. TPGS has amphiphilic structure of lipophilic alkyl tail and hydrophilic polar head with an HLB value of 13.2 and a low CMC value [170].

The effect of TPGS on the bioavailability of a P-gp substrate was first reported in enhancing CyA absorption. It was initially postulated that the improvement in oral availability was due solely to micelle formation and increased drug solubility. Subsequently, Chang and coworkers demonstrated an increased CyA absorption at TPGS concentrations below the CMC [171]. Since CyA is a known P-gp substrate,

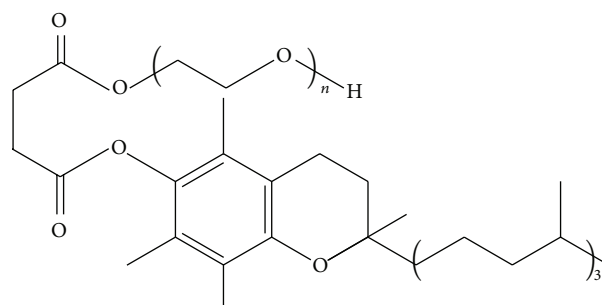


FIGURE 5: Structure of D-α-tocopheryl polyethylene glycol succinate (TPGS).

the authors hinted at a possible mechanism implicating the efflux transporter, a premise which was later confirmed. Dabholkar and his coworkers made use of PEG-PE/TPGS mixed micelles as drug carrier and investigated some properties of the efficiency in solubilizing PTX and the ability to bypass the P-gp-mediated drug efflux [172]. It was shown that PTX was efficiently solubilized in the nontoxic PEG-PE/TPGS micelles, and the entrapment was quite stable with only about 20% of the incorporated drug released from micelles after 48 h at 37°C. In addition, PTX-containing PEG-PE/TPGS micelles were stable *in vitro* under various conditions, in particular, at low pH values and in the presence of bile acids, which is especially important for oral administration. Contrary to other surfactants, TPGS seems to have only a minor effect on membrane fluidity [173], challenging earlier reports [159]. Indeed, it was speculated that the inhibition of P-gp resulted from a decrease in ATPase activity following substrate binding [173]. Further *in vitro* studies were carried out to investigate the mechanisms of P-gp inhibition using Caco-2 cells model [174]. The data suggest that TPGS is neither a P-gp substrate nor a trigger of intracellular ATP depletion. Instead, TPGS might act as an allosteric modulator not involving the Cis(Z)-flupentixol binding site.

In addition, some other amphiphilic polymers have been reported as P-gp inhibitors, such as mPEG-block-polycaprolactone [175], PEG-phosphatidylethanolamine [172], PEG-b-PLA [176], mPEG-poly(caprolactone-trimethylene carbonate) [177], and N-octyl-O-sulfate chitosan [178]. Among them, N-octyl-O-sulfate chitosan (NOSC) has been extensively studied. NOSC, synthesized by Q. Ping's group, is an amphipathic chitosan derivative, carrying sulfated groups

as hydrophilic moieties and octyl groups as hydrophobic moieties [179]. The oral bioavailability of PTX-loaded NOSC micelles and Taxol was further compared. It was suggested that NOSC, an inhibitor of P-gp, could enhance the oral absorption of PTX by a P-gp-independent micelle internalization [180]. In vivo study, the oral bioavailability of PTX loaded in NOSC micelles was increased by 6-fold in comparison with that of an orally dosed Taxol. In the Caco-2 cell uptake studies, NOSC micelles brought about a significantly higher amount of PTX accumulated via both clathrin- and caveolae-mediated endocytosis. The mechanism of P-gp inhibition by NOSC was probably related to interfering with the P-gp ATPase rather than reducing the P-gp expression.

5. Conclusion

Oral administration is the most commonly preferred route for drug delivery, especially in the case of repeated dosing for chronic therapy. To achieve good oral absorption of poorly water-soluble drugs, the loaded drug should be protected from the harsh gastrointestinal environment and release in a controlled manner at the target sites. In this review article, we aim to illustrate the potential of PMs for delivery of poorly water-soluble drugs, especially in the areas of oral delivery. It was suggested that PMs could enhance the oral drug bioavailability probably because the special stability (thermodynamic and kinetic stability) facilitating the safe transport of PMs through the GI tract, the pH-sensitivity of PMs promoting the controlled release properties of loaded drugs at target region, the mucoadhesivity of PMs prolonging the residence time in the gut, and the P-gp inhibitors contributing to drug accumulation. To make a methodical layout, we introduced various kinds of PMs separately in this article. However, a possible direction of combining two or more properties, such as pH-sensitive and mucoadhesive PMs, has gained much attention and offers a promising way to enhance the bioavailability of oral delivery.

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References

- [1] V. P. Sant, D. Smith, and J.-C. Leroux, "Enhancement of oral bioavailability of poorly water-soluble drugs by poly(ethylene glycol)-*block*-poly(alkyl acrylate-co-methacrylic acid) self-assemblies," *Journal of Controlled Release*, vol. 104, no. 2, pp. 289–300, 2005.
- [2] L. Bromberg, "Polymeric micelles in oral chemotherapy," *Journal of Controlled Release*, vol. 128, no. 2, pp. 99–112, 2008.
- [3] G. Gaucher, P. Satturwar, M.-C. Jones, A. Furtos, and J.-C. Leroux, "Polymeric micelles for oral drug delivery," *European Journal of Pharmaceutics and Biopharmaceutics*, vol. 76, no. 2, pp. 147–158, 2010.
- [4] Y. Masaoka, Y. Tanaka, M. Kataoka, S. Sakuma, and S. Yamashita, "Site of drug absorption after oral administration: assessment of membrane permeability and luminal concentration of drugs in each segment of gastrointestinal tract," *European Journal of Pharmaceutical Sciences*, vol. 29, no. 3–4, pp. 240–250, 2006.
- [5] S. Shugarts and L. Z. Benet, "The role of transporters in the pharmacokinetics of orally administered drugs," *Pharmaceutical Research*, vol. 26, no. 9, pp. 2039–2054, 2009.
- [6] C. A. Lipinski, "Drug-like properties and the causes of poor solubility and poor permeability," *Journal of Pharmacological and Toxicological Methods*, vol. 44, no. 1, pp. 235–249, 2000.
- [7] E. M. Merisko-Liversidge and G. G. Liversidge, "Drug nanoparticles: formulating poorly water-soluble compounds," *Toxicologic Pathology*, vol. 36, no. 1, pp. 43–48, 2008.
- [8] Y. K. Choi, B. K. Poudel, N. Marasini et al., "Enhanced solubility and oral bioavailability of itraconazole by combining membrane emulsification and spray drying technique," *International Journal of Pharmaceutics*, vol. 434, no. 1–2, pp. 264–271, 2012.
- [9] A. M. Monjazeb, D. Ayala, C. Jensen et al., "A phase I dose escalation study of hypofractionated IMRT field-in-field boost for newly diagnosed glioblastoma multiforme," *International Journal of Radiation Oncology Biology Physics*, vol. 82, no. 2, pp. 743–748, 2012.
- [10] S. Gillessen, U. S. Gnad-Vogt, E. Gallerani et al., "A phase I dose-escalation study of the immunocytokine EMD, 521873 (Selectikine) in patients with advanced solid tumours," *European Journal of Cancer*, vol. 49, no. 1, pp. 35–44, 2013.
- [11] Y. Lu and K. Park, "Polymeric micelles and alternative nanonized delivery vehicles for poorly soluble drugs," *International Journal of Pharmaceutics*, 2012.
- [12] O. C. Farokhzad and R. Langer, "Impact of nanotechnology on drug delivery," *ACS Nano*, vol. 3, no. 1, pp. 16–20, 2009.
- [13] A. Lavasanifar, J. Samuel, and G. S. Kwon, "Poly(ethylene oxide)-*block*-poly(L-amino acid) micelles for drug delivery," *Advanced Drug Delivery Reviews*, vol. 54, no. 2, pp. 169–190, 2002.
- [14] J. B. Hall, M. A. Dobrovolskaia, A. K. Patri, and S. E. McNeil, "Characterization of nanoparticles for therapeutics," *Nanomedicine*, vol. 2, no. 6, pp. 789–803, 2007.
- [15] R. G. Strickley, "Solubilizing excipients in oral and injectable formulations," *Pharmaceutical Research*, vol. 21, no. 2, pp. 201–230, 2004.
- [16] K. Kawakami, T. Yoshikawa, T. Hayashi, Y. Nishihara, and K. Masuda, "Microemulsion formulation for enhanced absorption of poorly soluble drugs: II. In vivo study," *Journal of Controlled Release*, vol. 81, no. 1–2, pp. 75–82, 2002.
- [17] R. H. Müller, K. Mäder, and S. Gohla, "Solid lipid nanoparticles (SLN) for controlled drug delivery—a review of the state of the art," *European Journal of Pharmaceutics and Biopharmaceutics*, vol. 50, no. 1, pp. 161–177, 2000.
- [18] C. J. H. Porter and W. N. Charman, "In vitro assessment of oral lipid based formulations," *Advanced Drug Delivery Reviews*, vol. 50, no. 1, pp. S127–S147, 2001.
- [19] J. Kalal, J. Drobnik, J. Kopecek, and J. Exner, "Water soluble polymers for medicine," *British Polymer Journal*, vol. 10, no. 2, pp. III–114, 1978.
- [20] M. Yokoyama, G. S. Kwon, T. Okano, Y. Sakurai, T. Seto, and K. Kataoka, "Preparation of micelle-forming polymer-drug conjugates," *Bioconjugate Chemistry*, vol. 3, no. 4, pp. 295–301, 1992.
- [21] E. C. Ale, B. Maggio, and M. L. Fanani, "Ordered-disordered domain coexistence in ternary lipid monolayers activates sphingomyelinase by clearing ceramide from the active phase,"

- Biochimica Et Biophysica Acta-Biomembranes*, vol. 1818, no. 11, pp. 2767–2776, 2012.
- [22] H. Suzuki and Y. Sugiyama, “Role of metabolic enzymes and efflux transporters in the absorption of drugs from the small intestine,” *European Journal of Pharmaceutical Sciences*, vol. 12, no. 1, pp. 3–12, 2000.
- [23] A. L. Daugherty and R. J. Mersny, “Regulation of the intestinal epithelial paracellular barrier,” *Pharmaceutical Science and Technology Today*, vol. 2, no. 7, pp. 281–287, 1999.
- [24] M. A. Deli, “Potential use of tight junction modulators to reversibly open membranous barriers and improve drug delivery,” *Biochimica et Biophysica Acta*, vol. 1788, no. 4, pp. 892–910, 2009.
- [25] M. Schenk and C. Mueller, “The mucosal immune system at the gastrointestinal barrier,” *Best Practice and Research*, vol. 22, no. 3, pp. 391–409, 2008.
- [26] L. M. Ensign, R. Cone, and J. Hanes, “Oral drug delivery with polymeric nanoparticles: the gastrointestinal mucus barriers,” *Advanced Drug Delivery Reviews*, vol. 64, no. 6, pp. 557–570, 2012.
- [27] G. Ponchel and J.-M. Irache, “Specific and non-specific bioadhesive particulate systems for oral delivery to the gastrointestinal tract,” *Advanced Drug Delivery Reviews*, vol. 34, no. 2-3, pp. 191–219, 1998.
- [28] L. Plapied, N. Duhem, A. des Rieux, and V. Pr at, “Fate of polymeric nanocarriers for oral drug delivery,” *Current Opinion in Colloid and Interface Science*, vol. 16, no. 3, pp. 228–237, 2011.
- [29] P. Bytzer, S. J. Connolly, S. Yang et al., “Analysis of upper gastrointestinal adverse events among patients given dabigatran in the RE-LY trial,” *Clinical Gastroenterology and Hepatology*, vol. 11, no. 3, pp. 246.e5–252.e5, 2013.
- [30] Z. Harel, S. Harel, P. S. Shah, R. Wald, J. Perl, and C. M. Bell, “Gastrointestinal adverse events with sodium polystyrene sulfonate (kayexalate) use: a systematic review,” *The American Journal of Medicine*, vol. 126, no. 3, pp. 264.e9–264.e24, 2013.
- [31] J. C. Chaumeil, “Micronization: a method of improving the bioavailability of poorly soluble drugs,” *Methods and Findings in Experimental and Clinical Pharmacology*, vol. 20, no. 3, pp. 211–215, 1998.
- [32] A. T. M. Serajuddin, “Salt formation to improve drug solubility,” *Advanced Drug Delivery Reviews*, vol. 59, no. 7, pp. 603–616, 2007.
- [33] V. J. Stella and K. W. Nti-Addae, “Prodrug strategies to overcome poor water solubility,” *Advanced Drug Delivery Reviews*, vol. 59, no. 7, pp. 677–694, 2007.
- [34] S. Alonso-Romanowski, N. S. Chiamoni, V. S. Liroy, R. A. Gargini, L. I. Viera, and M. C. Taira, “Characterization of diacetylenic liposomes as carriers for oral vaccines,” *Chemistry and Physics of Lipids*, vol. 122, no. 1-2, pp. 191–203, 2003.
- [35] G. Riess, “Micellization of block copolymers,” *Progress in Polymer Science*, vol. 28, no. 7, pp. 1107–1170, 2003.
- [36] M.-C. Jones and J.-C. Leroux, “Polymeric micelles—a new generation of colloidal drug carriers,” *European Journal of Pharmaceutics and Biopharmaceutics*, vol. 48, no. 2, pp. 101–111, 1999.
- [37] K. Van Butsele, P. Sibret, C. A. Fustin et al., “Synthesis and pH-dependent micellization of diblock copolymer mixtures,” *Journal of Colloid and Interface Science*, vol. 329, no. 2, pp. 235–243, 2009.
- [38] J. Taillefer, M. Jones, N. Bresseur, J. Van Lier, and J. Leroux, “Preparation and characterization of pH-responsive polymeric micelles for the delivery of photosensitizing anticancer drugs,” *Journal of Pharmaceutical Sciences*, vol. 89, no. 1, pp. 52–62, 2000.
- [39] V. Y. Alakhov, E. Y. Moskaleva, E. V. Batrakova, and A. V. Kabanov, “Hypersensitization of multidrug resistant human ovarian carcinoma cells by pluronic P85 block copolymer,” *Bioconjugate Chemistry*, vol. 7, no. 2, pp. 209–216, 1996.
- [40] N. Nishiyama and K. Kataoka, “Preparation and characterization of size-controlled polymeric micelle containing cis-dichlorodiammineplatinum(II) in the core,” *Journal of Controlled Release*, vol. 74, no. 1-3, pp. 83–94, 2001.
- [41] Y. Li and G. S. Kwon, “Methotrexate esters of poly(ethylene oxide)-*block*-poly(2-hydroxyethyl-L-aspartamide). Part I: effects of the level of methotrexate conjugation on the stability of micelles and on drug release,” *Pharmaceutical Research*, vol. 17, no. 5, pp. 607–611, 2000.
- [42] A. Lavasanifar, J. Samuel, and G. S. Kwon, “Micelles self-assembled from poly(ethylene oxide)-*block*-poly(*N*-hexyl stearate-L-aspartamide) by a solvent evaporation method: effect on the solubilization and haemolytic activity of amphotericin B,” *Journal of Controlled Release*, vol. 77, no. 1-2, pp. 155–160, 2001.
- [43] Y. Luo, X. Yao, J. Yuan, T. Ding, and Q. Gao, “Preparation and drug controlled-release of polyion complex micelles as drug delivery systems,” *Colloids and Surfaces B*, vol. 68, no. 2, pp. 218–224, 2009.
- [44] I. K. Voets, A. De Keizer, M. A. Cohen Stuart, J. Justynska, and H. Schlaad, “Irreversible structural transitions in mixed micelles of oppositely charged diblock copolymers in aqueous solution,” *Macromolecules*, vol. 40, no. 6, pp. 2158–2164, 2007.
- [45] C.-H. Hsu, S.-W. Kuo, J.-K. Chen, F.-H. Ko, C.-S. Liao, and F.-C. Chang, “Self-assembly behavior of A-B diblock and C-D random copolymer mixtures in the solution state through mediated hydrogen bonding,” *Langmuir*, vol. 24, no. 15, pp. 7727–7734, 2008.
- [46] W.-P. Gao, Y. Bai, E.-Q. Chen et al., “Controlling vesicle formation via interpolymer hydrogen-bonding complexation between poly(ethylene oxide)-*block*-polybutadiene and poly(acrylic acid) in solution,” *Macromolecules*, vol. 39, no. 14, pp. 4894–4898, 2006.
- [47] S.-W. Kuo, P.-H. Tung, C.-L. Lai, K.-U. Jeong, and F.-C. Chang, “Supramolecular micellization of diblock copolymer mixtures mediated by hydrogen bonding for the observation of separated coil and chain aggregation in common solvents,” *Macromolecular Rapid Communications*, vol. 29, no. 3, pp. 229–233, 2008.
- [48] R. Dobrawa and F. W urthner, “Metallo-supramolecular approach toward functional coordination polymers,” *Journal of Polymer Science, Part A*, vol. 43, no. 21, pp. 4981–4995, 2005.
- [49] K. Yoncheva, P. Calleja, M. Agueros et al., “Stabilized micelles as delivery vehicles for paclitaxel,” *International Journal of Pharmaceutics*, vol. 436, no. 1-2, pp. 258–264, 2012.
- [50] M. Yokoyama, “Polymeric micelles for the targeting of hydrophobic drugs,” *Polymeric Drug Delivery Systems*, vol. 148, pp. 533–576, 2005.
- [51] M. F. Francis, L. Lavoie, F. M. Winnik, and J.-C. Leroux, “Solubilization of cyclosporin a in dextran-g-polyethyleneglycolalkyl ether polymeric micelles,” *European Journal of Pharmaceutics and Biopharmaceutics*, vol. 56, no. 3, pp. 337–346, 2003.
- [52] Y. Kakizawa and K. Kataoka, “Block copolymer micelles for delivery of gene and related compounds,” *Advanced Drug Delivery Reviews*, vol. 54, no. 2, pp. 203–222, 2002.

- [53] A. V. Kabanov, E. V. Batrakova, and V. Y. Alakhov, "Pluronic block copolymers as novel polymer therapeutics for drug and gene delivery," *Journal of Controlled Release*, vol. 82, no. 2-3, pp. 189-212, 2002.
- [54] M. Yokoyama, "Clinical applications of polymeric micelle carrier systems in chemotherapy and image diagnosis of solid tumors," *Journal of Experimental and Clinical Medicine*, vol. 3, no. 4, pp. 151-158, 2011.
- [55] C. J. F. Rijcken, O. Soga, W. E. Hennink, and C. F. V. Nostrum, "Triggered destabilisation of polymeric micelles and vesicles by changing polymers polarity: an attractive tool for drug delivery," *Journal of Controlled Release*, vol. 120, no. 3, pp. 131-148, 2007.
- [56] M. Yokoyama, "Block copolymers as drug carriers," *Critical Reviews in Therapeutic Drug Carrier Systems*, vol. 9, no. 3-4, pp. 213-248, 1992.
- [57] C. Allen, D. Maysinger, and A. Eisenberg, "Nano-engineering block copolymer aggregates for drug delivery," *Colloids and Surfaces B*, vol. 16, no. 1-4, pp. 3-27, 1999.
- [58] M. G. Carstens, C. J. Rijcken, C. F. Nostrum, and W. E. Hennink, "Pharmaceutical micelles: combining longevity, stability, and stimuli sensitivity," *Multifunctional Pharmaceutical Nanocarriers*, vol. 4, pp. 263-308, 2008.
- [59] N. Rapoport, "Combined cancer therapy by micellar-encapsulated drug and ultrasound," *International Journal of Pharmaceutics*, vol. 277, no. 1-2, pp. 155-162, 2004.
- [60] G. Ruan and S.-S. Feng, "Preparation and characterization of poly(lactic acid)-poly(ethylene glycol)-poly(lactic acid) (PLA-PEG-PLA) microspheres for controlled release of paclitaxel," *Biomaterials*, vol. 24, no. 27, pp. 5037-5044, 2003.
- [61] Y. Bae and K. Kataoka, "Intelligent polymeric micelles from functional poly(ethylene glycol)-poly(amino acid) block copolymers," *Advanced Drug Delivery Reviews*, vol. 61, no. 10, pp. 768-784, 2009.
- [62] G. K. Taek, H. Lee, Y. Jang, and G. P. Tae, "Controlled release of paclitaxel from heparinized metal stent fabricated by layer-by-layer assembly of polylysine and hyaluronic acid-g-poly(lactic-co-glycolic acid) micelles encapsulating paclitaxel," *Biomacromolecules*, vol. 10, no. 6, pp. 1532-1539, 2009.
- [63] H. Lee, C.-H. Ahn, and T. G. Park, "Poly[lactic-co-(glycolic acid)]-grafted hyaluronic acid copolymer micelle nanoparticles for target-specific delivery of doxorubicin," *Macromolecular Bioscience*, vol. 9, no. 4, pp. 336-342, 2009.
- [64] M. A. R. Meier, S. N. H. Aerts, B. B. P. Staal, M. Rasa, and U. S. Schubert, "PEO-b-PCL block copolymers: synthesis, detailed characterization, and selected micellar drug encapsulation behavior," *Macromolecular Rapid Communications*, vol. 26, no. 24, pp. 1918-1924, 2005.
- [65] G. S. Kwon, "Polymeric micelles for delivery of poorly water-soluble compounds," *Critical Reviews in Therapeutic Drug Carrier Systems*, vol. 20, no. 5, pp. 357-403, 2003.
- [66] G. Molineux, "Pegylation: engineering improved pharmaceuticals for enhanced therapy," *Cancer Treatment Reviews*, vol. 28, pp. 13-16, 2002.
- [67] A. Benahmed, M. Ranger, and J.-C. Leroux, "Novel polymeric micelles based on the amphiphilic diblock copolymer poly(*N*-vinyl-2-pyrrolidone)-*block*-poly(D,L-lactide)," *Pharmaceutical Research*, vol. 18, no. 3, pp. 323-328, 2001.
- [68] T. Inoue, G. Chen, K. Nakamae, and A. S. Hoffman, "An AB block copolymer of oligo(methyl methacrylate) and poly(acrylic acid) for micellar delivery of hydrophobic drugs," *Journal of Controlled Release*, vol. 51, no. 2-3, pp. 221-229, 1998.
- [69] A. des Rieux, V. Fievez, M. Garinot, Y.-J. Schneider, and V. Pr at, "Nanoparticles as potential oral delivery systems of proteins and vaccines: a mechanistic approach," *Journal of Controlled Release*, vol. 116, no. 1, pp. 1-27, 2006.
- [70] A. Vila, A. S anchez, M. Tob io, P. Calvo, and M. J. Alonso, "Design of biodegradable particles for protein delivery," *Journal of Controlled Release*, vol. 78, no. 1-3, pp. 15-24, 2002.
- [71] Y. J. Yamanaka and K. W. Leong, "Engineering strategies to enhance nanoparticle-mediated oral delivery," *Journal of Biomaterials Science, Polymer Edition*, vol. 19, no. 12, pp. 1549-1570, 2008.
- [72] Z. G. Gao, D. H. Lee, D. I. Kim, and Y. H. Bae, "Doxorubicin loaded pH-sensitive micelle targeting acidic extracellular pH of human ovarian A2780 tumor in mice," *Journal of Drug Targeting*, vol. 13, no. 7, pp. 391-397, 2005.
- [73] E. S. Lee, H. J. Shin, K. Na, and Y. H. Bae, "Poly(L-histidine)-PEG block copolymer micelles and pH-induced destabilization," *Journal of Controlled Release*, vol. 90, no. 3, pp. 363-374, 2003.
- [74] Y. Bae, W.-D. Jang, N. Nishiyama, S. Fukushima, and K. Kataoka, "Multifunctional polymeric micelles with folate-mediated cancer cell targeting and pH-triggered drug releasing properties for active intracellular drug delivery," *Molecular BioSystems*, vol. 1, no. 3, pp. 242-250, 2005.
- [75] X. Zhao, H. Li, and R. J. Lee, "Targeted drug delivery via folate receptors," *Expert Opinion on Drug Delivery*, vol. 5, no. 3, pp. 309-319, 2008.
- [76] N. Rapoport, "Physical stimuli-responsive polymeric micelles for anti-cancer drug delivery," *Progress in Polymer Science*, vol. 32, no. 8-9, pp. 962-990, 2007.
- [77] F. Cald erara, Z. Hruska, G. Hurtrez, J.-P. Lerch, T. Nugay, and G. Riess, "Investigation of polystyrene-poly(ethylene oxide) block copolymer micelle formation in organic and aqueous solutions by nonradiative energy transfer experiments," *Macromolecules*, vol. 27, no. 5, pp. 1210-1215, 1994.
- [78] M. Wilhelm, C.-L. Zhao, Y. Wang et al., "Poly(styrene-ethylene oxide) block copolymer micelle formation in water: a fluorescence probe study," *Macromolecules*, vol. 24, no. 5, pp. 1033-1040, 1991.
- [79] N. Wiradharma, Y. Zhang, S. Venkataraman, J. L. Hedrick, and Y. Y. Yang, "Self-assembled polymer nanostructures for delivery of anticancer therapeutics," *Nano Today*, vol. 4, no. 4, pp. 302-317, 2009.
- [80] A. B. Ebrahim, Z. Y. Ong, J. L. Hedrick et al., "Mixed micelles self-assembled from block copolymers for drug delivery," *Current Opinion in Colloid and Interface Science*, vol. 16, no. 3, pp. 182-194, 2011.
- [81] M. L. Adams, A. Lavasanifar, and G. S. Kwon, "Amphiphilic block copolymers for drug delivery," *Journal of Pharmaceutical Sciences*, vol. 92, no. 7, pp. 1343-1355, 2003.
- [82] M. Camilleri, L. J. Colemont, S. F. Phillips et al., "Human gastric emptying and colonic filling of solids characterized by a new method," *American Journal of Physiology*, vol. 257, no. 2, pp. G284-G290, 1989.
- [83] E. Pierri and K. Avgoustakis, "Poly(lactide)-poly(ethylene glycol) micelles as a carrier for griseofulvin," *Journal of Biomedical Materials Research A*, vol. 75, no. 3, pp. 639-647, 2005.
- [84] L. Ould-Ouali, M. Noppe, X. Langlois et al., "Self-assembling PEG-p(CL-co-TMC) copolymers for oral delivery of poorly water-soluble drugs: a case study with risperidone," *Journal of Controlled Release*, vol. 102, no. 3, pp. 657-668, 2005.

- [85] W. Chen, P. Zhong, F. Meng et al., "Redox and pH-responsive degradable micelles for dually activated intracellular anticancer drug release," *Journal of Controlled Release*, vol. 169, no. 3, pp. 171–179, 2013.
- [86] R. Tan, Z. She, M. Wang, Z. Fang, Y. Liu, and Q. Feng, "Thermo-sensitive alginate-based injectable hydrogel for tissue engineering," *Carbohydrate Polymers*, vol. 87, no. 2, pp. 1515–1521, 2012.
- [87] G. Liu, X. Li, S. Xiong et al., "Fluorine-containing thermo-sensitive core/shell microgel particles: preparation, characterization, and their applications in controlled drug release," *Journal of Fluorine Chemistry*, vol. 135, pp. 75–82, 2012.
- [88] C. J. Chen, Q. Jin, G. Y. Liu, D. D. Li, J. L. Wang, and J. Ji, "Reversibly light-responsive micelles constructed via a simple modification of hyperbranched polymers with chromophores," *Polymer*, vol. 53, no. 17, pp. 3695–3703, 2012.
- [89] G. A. Hussein, D. Velluto, L. Kherbeck, W. G. Pitt, J. A. Hubbell, and D. A. Christensen, "Investigating the acoustic release of doxorubicin from targeted micelles," *Colloids and Surfaces B*, vol. 101, pp. 153–155, 2013.
- [90] T. Yin, P. Wang, J. Li et al., "Ultrasound-sensitive siRNA-loaded nanobubbles formed by hetero-assembly of polymeric micelles and liposomes and their therapeutic effect in gliomas," *Biomaterials*, vol. 34, no. 18, pp. 4532–4543, 2013.
- [91] G. R. Martin and R. K. Jain, "Noninvasive measurement of interstitial pH profiles in normal and neoplastic tissue using fluorescence ratio imaging microscopy," *Cancer Research*, vol. 54, no. 21, pp. 5670–5674, 1994.
- [92] I. Lee, M. Park, Y. Kim, O. Hwang, G. Khang, and D. Lee, "Ketal containing amphiphilic block copolymer micelles as pH-sensitive drug carriers," *International Journal of Pharmaceutics*, vol. 448, no. 1, pp. 256–266, 2013.
- [93] Z. Xu, M. Guo, H. Yan, and K. Liu, "Enhanced loading of doxorubicin into polymeric micelles by a combination of ionic bonding and hydrophobic effect, and the pH-sensitive and ligand-mediated delivery of loaded drug," *Reactive and Functional Polymers*, vol. 73, no. 3, pp. 564–572, 2013.
- [94] E. Ravazzolo, S. Salmaso, F. Mastrotto, S. Bersani, E. Gallon, and P. Caliceti, "pH-responsive lipid core micelles for tumour targeting," *European Journal of Pharmaceutics and Biopharmaceutics*, vol. 83, no. 3, pp. 346–357, 2013.
- [95] A. McDowell and B. J. McLeod, "Physiology and pharmacology of the brushtail possum gastrointestinal tract: relationship to the human gastrointestinal tract," *Advanced Drug Delivery Reviews*, vol. 59, no. 11, pp. 1121–1132, 2007.
- [96] O. V. Borisov and E. B. Zhulina, "Reentrant morphological transitions in copolymer micelles with pH-sensitive corona," *Langmuir*, vol. 21, no. 8, pp. 3229–3231, 2005.
- [97] E. S. Lee, H. J. Shin, K. Na, and Y. H. Bae, "Poly(L-histidine)-PEG block copolymer micelles and pH-induced destabilization," *Journal of Controlled Release*, vol. 90, no. 3, pp. 363–374, 2003.
- [98] E. S. Lee, K. Na, and Y. H. Bae, "Polymeric micelle for tumor pH and folate-mediated targeting," *Journal of Controlled Release*, vol. 91, no. 1-2, pp. 103–113, 2003.
- [99] Y. Tang, S. Y. Liu, S. P. Armes, and N. C. Billingham, "Solubilization and controlled release of a hydrophobic drug using novel micelle-forming ABC triblock copolymers," *Biomacromolecules*, vol. 4, no. 6, pp. 1636–1645, 2003.
- [100] S. Dai, K. C. Tam, and R. D. Jenkins, "Aggregation behavior of methacrylic acid/ethyl acrylate copolymer in dilute solutions," *European Polymer Journal*, vol. 36, no. 12, pp. 2671–2677, 2000.
- [101] S. Ganta, H. Devalapally, A. Shahiwala, and M. Amiji, "A review of stimuli-responsive nanocarriers for drug and gene delivery," *Journal of Controlled Release*, vol. 126, no. 3, pp. 187–204, 2008.
- [102] A. K. Bajpai, S. K. Shukla, S. Bhanu, and S. Kankane, "Responsive polymers in controlled drug delivery," *Progress in Polymer Science*, vol. 33, no. 11, pp. 1088–1118, 2008.
- [103] M.-C. Jones, M. Ranger, and J.-C. Leroux, "pH-sensitive unimolecular polymeric micelles: synthesis of a novel drug carrier," *Bioconjugate Chemistry*, vol. 14, no. 4, pp. 774–781, 2003.
- [104] V. P. Sant, D. Smith, and J.-C. Leroux, "Novel pH-sensitive supramolecular assemblies for oral delivery of poorly water soluble drugs: preparation and characterization," *Journal of Controlled Release*, vol. 97, no. 2, pp. 301–312, 2004.
- [105] S. Kim, J. Y. Kim, K. M. Huh, G. Acharya, and K. Park, "Hydrotropic polymer micelles containing acrylic acid moieties for oral delivery of paclitaxel," *Journal of Controlled Release*, vol. 132, no. 3, pp. 222–229, 2008.
- [106] P. Satturwar, M. N. Eddine, F. Ravenelle, and J.-C. Leroux, "pH-responsive polymeric micelles of poly(ethylene glycol)-b-poly(alkyl(meth)acrylate-co-methacrylic acid): influence of the copolymer composition on self-assembling properties and release of candesartan cilexetil," *European Journal of Pharmaceutics and Biopharmaceutics*, vol. 65, no. 3, pp. 379–387, 2007.
- [107] J. Kopecek, "Polymer-drug conjugates: origins, progress to date and future directions," *Advanced Drug Delivery Reviews*, vol. 65, no. 1, pp. 49–59, 2013.
- [108] H. P. Chang, J. Y. Chen, P. S. Zhong, Y. H. Chang, and M. Liang, "Synthesis and characterization of a new polymer-drug conjugate with pH-induced activity," *Polymer*, vol. 53, no. 16, pp. 3498–3507, 2012.
- [109] B. C. Tang, M. Dawson, S. K. Lai et al., "Biodegradable polymer nanoparticles that rapidly penetrate the human mucus barrier," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 106, no. 46, pp. 19268–19273, 2009.
- [110] R. A. Cone, "Barrier properties of mucus," *Advanced Drug Delivery Reviews*, vol. 61, no. 2, pp. 75–85, 2009.
- [111] D. Rahmat, C. Muller, J. Barthelme, G. Shahnaz, R. Martien, and A. B. Schnurch, "Thiolated hydroxyethyl cellulose: design and in vitro evaluation of mucoadhesive and permeation enhancing nanoparticles," *European Journal of Pharmaceutics and Biopharmaceutics*, vol. 83, no. 2, pp. 149–155, 2013.
- [112] K. M. R. Srivalli, P. K. Lakshmi, and J. Balasubramaniam, "Design of a novel bilayered gastric mucoadhesive system for localized and unidirectional release of lamotrigine," *Saudi Pharmaceutical Journal*, vol. 21, no. 1, pp. 45–52, 2013.
- [113] J. D. Smart, "The basics and underlying mechanisms of mucoadhesion," *Advanced Drug Delivery Reviews*, vol. 57, no. 11, pp. 1556–1568, 2005.
- [114] M. A. Arango, G. Ponchel, A. M. Orecchioni, M. J. Renedo, D. Duchêne, and J. M. Irache, "Bioadhesive potential of gliadin nanoparticulate systems," *European Journal of Pharmaceutical Sciences*, vol. 11, no. 4, pp. 333–341, 2000.
- [115] M. A. Arango, M. A. Campanero, M. J. Renedo, G. Ponchel, and J. M. Irache, "Gliadin nanoparticles as carriers for the oral administration of lipophilic drugs. Relationships between bioadhesion and pharmacokinetics," *Pharmaceutical Research*, vol. 18, no. 11, pp. 1521–1527, 2001.
- [116] A. Ludwig, "The use of mucoadhesive polymers in ocular drug delivery," *Advanced Drug Delivery Reviews*, vol. 57, no. 11, pp. 1595–1639, 2005.

- [117] S. K. Lai, Y.-Y. Wang, and J. Hanes, "Mucus-penetrating nanoparticles for drug and gene delivery to mucosal tissues," *Advanced Drug Delivery Reviews*, vol. 61, no. 2, pp. 158–171, 2009.
- [118] L.-L. Fu, C.-C. Zhou, S. Yao, J.-Y. Yu, B. Liu, and J.-K. Bao, "Plant lectins: targeting programmed cell death pathways as antitumor agents," *International Journal of Biochemistry and Cell Biology*, vol. 43, no. 10, pp. 1442–1449, 2011.
- [119] N. Yamazaki, S. Kojima, N. V. Bovin, S. André, S. Gabius, and H.-J. Gabius, "Endogenous lectins as targets for drug delivery," *Advanced Drug Delivery Reviews*, vol. 43, no. 2-3, pp. 225–244, 2000.
- [120] H. Kausar, R. Munagala, S. S. Bansal, F. Aqil, M. V. Vadhanam, and R. C. Gupta, "Cucurbitacin B potently suppresses non-small-cell lung cancer growth: identification of intracellular thiols as critical targets," *Cancer Letters*, vol. 332, no. 1, pp. 35–45, 2013.
- [121] J. H. Park, G. Saravanakumar, K. Kim, and I. C. Kwon, "Targeted delivery of low molecular drugs using chitosan and its derivatives," *Advanced Drug Delivery Reviews*, vol. 62, no. 1, pp. 28–41, 2010.
- [122] H. Takeuchi, J. Thongborisute, Y. Matsui, H. Sugihara, H. Yamamoto, and Y. Kawashima, "Novel mucoadhesion tests for polymers and polymer-coated particles to design optimal mucoadhesive drug delivery systems," *Advanced Drug Delivery Reviews*, vol. 57, no. 11, pp. 1583–1594, 2005.
- [123] J. S. Crater and R. L. Carrier, "Barrier properties of gastrointestinal mucus to nanoparticle transport," *Macromolecular Bioscience*, vol. 10, no. 12, pp. 1473–1483, 2010.
- [124] L. Bromberg, M. Temchenko, V. Alakhov, and T. A. Hatton, "Bioadhesive properties and rheology of polyether-modified poly(acrylic acid) hydrogels," *International Journal of Pharmaceutics*, vol. 282, no. 1-2, pp. 45–60, 2004.
- [125] J. Blanchette and N. A. Peppas, "Cellular evaluation of oral chemotherapy carriers," *Journal of Biomedical Materials Research A*, vol. 72, no. 4, pp. 381–388, 2005.
- [126] J. Blanchette and N. A. Peppas, "Oral chemotherapeutic delivery: design and cellular response," *Annals of Biomedical Engineering*, vol. 33, no. 2, pp. 142–149, 2005.
- [127] S. Dünnhaupt, J. Barthelmes, J. Hombach, D. Sakloetsakun, V. Arkhipova, and A. Bernkop-Schnürch, "Distribution of thiolated mucoadhesive nanoparticles on intestinal mucosa," *International Journal of Pharmaceutics*, vol. 408, no. 1-2, pp. 191–199, 2011.
- [128] A. Makhlof, M. Werle, Y. Tozuka, and H. Takeuchi, "A mucoadhesive nanoparticulate system for the simultaneous delivery of macromolecules and permeation enhancers to the intestinal mucosa," *Journal of Controlled Release*, vol. 149, no. 1, pp. 81–88, 2011.
- [129] V. V. Khutoryanskiy, "Advances in mucoadhesion and mucoadhesive polymers," *Macromolecular Bioscience*, vol. 11, no. 6, pp. 748–764, 2011.
- [130] D. Vllasaliu, L. Casertari, R. Fowler et al., "Absorption-promoting effects of chitosan in airway and intestinal cell lines: a comparative study," *International Journal of Pharmaceutics*, vol. 430, no. 1-2, pp. 151–160, 2012.
- [131] T. K. Giri, A. Thakur, A. Alexander, Ajazuddin, H. Badwaik, and D. K. Tripathi, "Modified chitosan hydrogels as drug delivery and tissue engineering systems: present status and applications," *Acta Pharmaceutica Sinica B*, vol. 2, no. 5, pp. 439–449, 2012.
- [132] I. Bertholon, G. Ponchel, D. Labarre, P. Couvreur, and C. Vauthier, "Bioadhesive properties of poly(alkylcyanoacrylate) nanoparticles coated with polysaccharide," *Journal of Nanoscience and Nanotechnology*, vol. 6, no. 9-10, pp. 3102–3109, 2006.
- [133] W. Guang Liu and K. De Yao, "Chitosan and its derivatives—a promising non-viral vector for gene transfection," *Journal of Controlled Release*, vol. 83, no. 1, pp. 1–11, 2002.
- [134] I. M. Van Der Lubben, J. C. Verhoef, A. C. Van Aelst, G. Borchard, and H. E. Junginger, "Chitosan microparticles for oral vaccination: preparation, characterization and preliminary in vivo uptake studies in murine Peyer's patches," *Biomaterials*, vol. 22, no. 7, pp. 687–694, 2001.
- [135] J. H. Na, S. Y. Lee, S. Lee et al., "Effect of the stability and deformability of self-assembled glycol chitosan nanoparticles on tumor-targeting efficiency," *Journal of Controlled Release*, vol. 163, no. 1, pp. 2–9, 2012.
- [136] A. Trapani, S. Di Gioia, N. Ditaranto et al., "Systemic heparin delivery by the pulmonary route using chitosan and glycol chitosan nanoparticles," *International Journal of Pharmaceutics*, vol. 447, no. 1-2, pp. 115–123, 2013.
- [137] O. Honkanen, J. Marvola, H. Kanerva et al., "Gamma scintigraphic evaluation of the fate of hydroxypropyl methylcellulose capsules in the human gastrointestinal tract," *European Journal of Pharmaceutical Sciences*, vol. 21, no. 5, pp. 671–678, 2004.
- [138] T. R. Thrimawithana, S. A. Young, C. R. Bunt, C. R. Green, and R. G. Alany, "In-vitro and in-vivo evaluation of carageenan/methylcellulose polymeric systems for transscleral delivery of macromolecules," *European Journal of Pharmaceutical Sciences*, vol. 44, no. 3, pp. 399–409, 2011.
- [139] M. A. Repka and J. W. McGinity, "Bioadhesive properties of hydroxypropylcellulose topical films produced by hot-melt extrusion," *Journal of Controlled Release*, vol. 70, no. 3, pp. 341–351, 2001.
- [140] A. H. Krauland, D. Guggi, and A. Bernkop-Schnürch, "Oral insulin delivery: the potential of thiolated chitosan-insulin tablets on non-diabetic rats," *Journal of Controlled Release*, vol. 95, no. 3, pp. 547–555, 2004.
- [141] A. H. Krauland and A. Bernkop-Schnürch, "Thiomers: development and in vitro evaluation of a peroral microparticulate peptide delivery system," *European Journal of Pharmaceutics and Biopharmaceutics*, vol. 57, no. 2, pp. 181–187, 2004.
- [142] J. Ding, R. He, G. Zhou, C. Tang, and C. Yin, "Multilayered mucoadhesive hydrogel films based on thiolated hyaluronic acid and polyvinylalcohol for insulin delivery," *Acta Biomaterialia*, vol. 8, no. 10, pp. 3643–3651, 2012.
- [143] V. Gajbhiye and S. Gong, "Lectin functionalized nanocarriers for gene delivery," *Biotechnology Advances*, 2013.
- [144] C.-M. Lehr, "Lectin-mediated drug delivery: the second generation of bioadhesives," *Journal of Controlled Release*, vol. 65, no. 1-2, pp. 19–29, 2000.
- [145] A. Bernkop-Schnürch, Y. Pinter, D. Guggi et al., "The use of thiolated polymers as carrier matrix in oral peptide delivery—proof of concept," *Journal of Controlled Release*, vol. 106, no. 1-2, pp. 26–33, 2005.
- [146] M.-H. Dufresne, M. A. Gauthier, and J.-C. Leroux, "Thiol-functionalized polymeric micelles: from molecular recognition to improved mucoadhesion," *Bioconjugate Chemistry*, vol. 16, no. 4, pp. 1027–1033, 2005.
- [147] N. C. Kalarickal, S. Rimmer, P. Sarker, and J.-C. Leroux, "Thiol-functionalized poly(ethylene glycol)-b-polyesters: synthesis and characterization," *Macromolecules*, vol. 40, no. 6, pp. 1874–1880, 2007.

- [148] Z. Rong, Y. Xu, C. Zhang, D. Xiang, X. Li, and D. Liu, "Evaluation of intestinal absorption of amtolmetin guacyl in rats: breast cancer resistant protein as a primary barrier of oral bioavailability," *Life Sciences*, vol. 92, no. 3, pp. 245–251, 2013.
- [149] R. Nieto Montesinos, A. Beduneau, Y. Pellequer, and A. Lamprecht, "Delivery of P-glycoprotein substrates using chemosensitizers and nanotechnology for selective and efficient therapeutic outcomes," *Journal of Controlled Release*, vol. 161, no. 1, pp. 50–61, 2012.
- [150] C. F. Higgins, "ABC transporters: from microorganisms to man," *Annual Review of Cell Biology*, vol. 8, pp. 67–113, 1992.
- [151] M. L. Cuestas, A. I. Castillo, A. Sosnik, and V. L. Mathet, "Downregulation of *mdr1* and *abcg2* genes is a mechanism of inhibition of efflux pumps mediated by polymeric amphiphiles," *Bioorganic & Medicinal Chemistry Letters*, vol. 22, no. 21, pp. 6577–6579, 2012.
- [152] Y. Lei, J. Tan, M. Wink, Y. Ma, N. Li, and G. Su, "An isoquinoline alkaloid from the Chinese herbal plant *Corydalis yanhusuo* W.T. Wang inhibits P-glycoprotein and multidrug resistance-associate protein 1," *Food Chemistry*, vol. 136, no. 3–4, pp. 1117–1121, 2013.
- [153] H. S. Mülder, H. Dekker, H. M. Pinedo, and J. Lankelma, "The P-glycoprotein-mediated relative decrease in cytosolic free drug concentration is similar for several anthracyclines with varying lipophilicity," *Biochemical Pharmacology*, vol. 50, no. 7, pp. 967–974, 1995.
- [154] E. Roger, F. Lagarce, E. Garcion, and J.-P. Benoit, "Reciprocal competition between lipid nanocapsules and P-gp for paclitaxel transport across Caco-2 cells," *European Journal of Pharmaceutical Sciences*, vol. 40, no. 5, pp. 422–429, 2010.
- [155] E. V. Batrakova, S. Li, W. F. Elmquist, D. W. Miller, V. Y. Alakhov, and A. V. Kabanov, "Mechanism of sensitization of MDR cancer cells by Pluronic block copolymers: selective energy depletion," *British Journal of Cancer*, vol. 85, no. 12, pp. 1987–1997, 2001.
- [156] M. Jin, T. Shimada, K. Yokogawa et al., "Site-dependent contributions of P-glycoprotein and CYP3A to cyclosporin A absorption, and effect of dexamethasone in small intestine of mice," *Biochemical Pharmacology*, vol. 72, no. 8, pp. 1042–1050, 2006.
- [157] F. Wang, D. Zhang, Q. Zhang et al., "Synergistic effect of folate-mediated targeting and verapamil-mediated P-gp inhibition with paclitaxel-polymer micelles to overcome multi-drug resistance," *Biomaterials*, vol. 32, no. 35, pp. 9444–9456, 2011.
- [158] Y.-L. Lo, "Relationships between the hydrophilic-lipophilic balance values of pharmaceutical excipients and their multidrug resistance modulating effect in Caco-2 cells and rat intestines," *Journal of Controlled Release*, vol. 90, no. 1, pp. 37–48, 2003.
- [159] B. D. Rege, J. P. Y. Kao, and J. E. Polli, "Effects of nonionic surfactants on membrane transporters in Caco-2 cell monolayers," *European Journal of Pharmaceutical Sciences*, vol. 16, no. 4–5, pp. 237–246, 2002.
- [160] H.-J. Yao, R.-J. Ju, X.-X. Wang et al., "The antitumor efficacy of functional paclitaxel nanomicelles in treating resistant breast cancers by oral delivery," *Biomaterials*, vol. 32, no. 12, pp. 3285–3302, 2011.
- [161] Z. Zhang, S. H. Lee, C. W. Gan, and S.-S. Feng, "In vitro and in vivo investigation on PLA-TPGS nanoparticles for controlled and sustained small molecule chemotherapy," *Pharmaceutical Research*, vol. 25, no. 8, pp. 1925–1935, 2008.
- [162] Z. Wei, S. Yuan, J. Hao, and X. Fang, "Mechanism of inhibition of P-glycoprotein mediated efflux by Pluronic P123/F127 block copolymers: relationship between copolymer concentration and inhibitory activity," *European Journal of Pharmaceutics and Biopharmaceutics*, vol. 83, no. 2, pp. 266–274, 2013.
- [163] J. Huang, L. Si, L. Jiang, Z. Fan, J. Qiu, and G. Li, "Effect of pluronic F68 block copolymer on P-glycoprotein transport and CYP3A4 metabolism," *International Journal of Pharmaceutics*, vol. 356, no. 1–2, pp. 351–353, 2008.
- [164] E. M. Leslie, "Arsenic-glutathione conjugate transport by the human multidrug resistance proteins (MRPs/ABCCs)," *Journal of Inorganic Biochemistry*, vol. 108, pp. 141–149, 2012.
- [165] Y. Wang, J. Hao, Y. Li et al., "Poly(caprolactone)-modified Pluronic P105 micelles for reversal of paclitaxel-resistance in SKOV-3 tumors," *Biomaterials*, vol. 33, no. 18, pp. 4741–4751, 2012.
- [166] G. Li, D. H. Lin, X. X. Xie, L. F. Qin, J. T. Wang, and K. Liu, "Uptake and transport of furanodiene in Caco-2 cell monolayers: a comparison study between furanodiene and furanodiene loaded PLGA nanoparticles," *Chinese Journal of Natural Medicines*, vol. 11, no. 1, pp. 49–55, 2013.
- [167] A. V. Kabanov, E. V. Batrakova, and V. Y. Alakhov, "Pluronic block copolymers for overcoming drug resistance in cancer," *Advanced Drug Delivery Reviews*, vol. 54, no. 5, pp. 759–779, 2002.
- [168] Z. Zhang, S. Tan, and S.-S. Feng, "Vitamin E TPGS as a molecular biomaterial for drug delivery," *Biomaterials*, vol. 33, no. 19, pp. 4889–4906, 2012.
- [169] Y. Guo, J. Luo, S. Tan, B. O. Otieno, and Z. Zhang, "The applications of Vitamin E TPGS in drug delivery," *European Journal of Pharmaceutical Sciences*, vol. 49, no. 2, pp. 175–186, 2013.
- [170] S. H.-W. Wu and W. K. Hopkins, "Characteristics of D- α -tocopheryl PEG 1000 succinate for applications as an absorption enhancer in drug delivery systems," *Pharmaceutical Technology*, vol. 23, no. 10, pp. 52–68, 1999.
- [171] T. Chang, L. Z. Benet, and M. F. Hebert, "The effect of water-soluble vitamin E on cyclosporine pharmacokinetics in healthy volunteers," *Clinical Pharmacology and Therapeutics*, vol. 59, no. 3, pp. 297–303, 1996.
- [172] R. D. Dabholkar, R. M. Sawant, D. A. Mongayt, P. V. Devarajan, and V. P. Torchilin, "Polyethylene glycol-phosphatidylethanolamine conjugate (PEG-PE)-based mixed micelles: some properties, loading with paclitaxel, and modulation of P-glycoprotein-mediated efflux," *International Journal of Pharmaceutics*, vol. 315, no. 1–2, pp. 148–157, 2006.
- [173] E.-M. Collnot, C. Baldes, M. F. Wempe et al., "Mechanism of inhibition of P-glycoprotein mediated efflux by vitamin E TPGS: influence on ATPase activity and membrane fluidity," *Molecular Pharmaceutics*, vol. 4, no. 3, pp. 465–474, 2007.
- [174] E.-M. Collnot, C. Baldes, U. F. Schaefer, K. J. Edgar, M. F. Wempe, and C.-M. Lehr, "Vitamin E TPGS P-glycoprotein inhibition mechanism: influence on conformational flexibility, intracellular ATP levels, and role of time and site of access," *Molecular Pharmaceutics*, vol. 7, no. 3, pp. 642–651, 2010.
- [175] J. Zastre, J. Jackson, M. Bajwa, R. Liggins, F. Iqbal, and H. Burt, "Enhanced cellular accumulation of a P-glycoprotein substrate, rhodamine-123, by caco-2 cells using low molecular weight methoxypolyethylene glycol-block-polycaprolactone diblock copolymers," *European Journal of Pharmaceutics and Biopharmaceutics*, vol. 54, no. 3, pp. 299–309, 2002.
- [176] L. Xiao, X. Xiong, X. Sun et al., "Role of cellular uptake in the reversal of multidrug resistance by PEG-b-PLA polymeric micelles," *Biomaterials*, vol. 32, no. 22, pp. 5148–5157, 2011.

- [177] F. Mathot, L. Van Beijsterveldt, V. Pr at, M. Brewster, and A. Ari en, "Intestinal uptake and biodistribution of novel polymeric micelles after oral administration," *Journal of Controlled Release*, vol. 111, no. 1-2, pp. 47-55, 2006.
- [178] R. Mo, X. Jin, N. Li et al., "The mechanism of enhancement on oral absorption of paclitaxel by N-octyl-O-sulfate chitosan micelles," *Biomaterials*, vol. 32, no. 20, pp. 4609-4620, 2011.
- [179] C. Zhang, G. Qu, Y. Sun et al., "Pharmacokinetics, biodistribution, efficacy and safety of N-octyl-O-sulfate chitosan micelles loaded with paclitaxel," *Biomaterials*, vol. 29, no. 9, pp. 1233-1241, 2008.
- [180] R. Mo, X. Jin, N. Li et al., "The mechanism of enhancement on oral absorption of paclitaxel by N-octyl-O-sulfate chitosan micelles," *Biomaterials*, vol. 32, no. 20, pp. 4609-4620, 2011.