

# Milk exosomes-mediated miR-31-5p delivery accelerates diabetic wound healing through promoting angiogenesis

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## ABSTRACT

The refractory diabetic wound has remained a worldwide challenge as one of the major health problems. The impaired angiogenesis phase during diabetic wound healing partly contributes to the pathological process. MicroRNA (miRNA) is an essential regulator of gene expression in crucial biological processes and is a promising nucleic acid drug in therapeutic fields of the diabetic wound. However, miRNA therapies have limitations due to lacking an effective delivery system. In the present study, we found a significant reduction of miR-31-5p expression in the full-thickness wounds of diabetic mice compared to normal mice. Further, miR-31-5p has been proven to promote the proliferation, migration, and angiogenesis of endothelial cells. Thus, we conceived the idea of exogenously supplementing miR-31-5p mimics to treat the diabetic wound. We used milk-derived exosomes as a novel system for miR-31-5p delivery and successfully encapsulated miR-31-5p mimics into milk exosomes through electroporation. Then, we proved that the miR-31-5p loaded in exosomes achieved higher cell uptake and was able to resist degradation. Moreover, our miRNA-exosomal formulation demonstrated dramatically improved endothelial cell functions *in vitro*, together with the promotion of angiogenesis and enhanced diabetic wound healing *in vivo*. Collectively, our data showed the feasibility of milk exosomes as a scalable, biocompatible, and cost-effective delivery system to enhance the bioavailability and efficacy of miRNAs.

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## Introduction

Non-healing wound is one of the diabetic chronic complications, which not only causes large physiological and psychological pain to patients but also impose a tremendous burden on the entire economy and society (Boulton et al., 2005). Under normal conditions, the process of wound closure involves four overlapping and coordinated stages: hemostasis, inflammation, proliferation, and remodeling (Rodrigues et al., 2019). In patients with diabetic non-healing wounds, the wound healing process slows down or even stalls due to impairment and prolongation of the four stages (Davis et al., 2018; Zubair & Ahmad, 2019). Typically, the dysfunction of endothelial cells and microcirculation disorders occur very often in diabetic patients, which contributes to the impaired angiogenesis process, a process that happens at the beginning of the proliferation stage (Grennan, 2019).

MicroRNAs (miRNAs) are one class of short, evolutionarily conserved non-coding RNAs, which contain about 20–24 nucleotides (Ambros, 2001). A growing number of studies have reported that miRNAs have great potential as predictive, diagnostic, and even therapeutic biomarkers in many types of diseases, including diabetic wound healing (Ohtsuka et al., 2021; Zhang et al., 2017; Petkovic et al., 2020). For example, miR-126-3p promoted endothelial cell proliferation and migration, thus improving angiogenesis and wound healing in diabetic mice (Tao et al., 2017). Benefiting from its anti-inflammatory role, the application of miR-146a conjugated with cerium oxide nanoparticles enhanced diabetic wound healing (Zgheib et al., 2019). Despite the high potential of miRNA-based gene therapy, it has many obstacles that need to be overcome. Firstly, due to the negative charge of miRNAs, they are difficult to pass through cell membranes (Boca et al., 2020). In addition, miRNAs are unstable *in vitro*

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and are easy to degrade in the wound microenvironment (Meng et al., 2018). Therefore, the development of safe and efficient miRNA delivery systems is of great significance to optimize miRNA-based gene therapy.

Exosomes are one kind of cell-derived, nano-sized membranous vesicles, which originate from the endosomal system and are 30–150 nm in diameter (EL Andaloussi et al., 2013). Accumulating studies have reported that exosomes and their cargos have a therapeutic effect in various diseases ranging from cancers, cardiovascular disease, to diabetic complications (Zheng et al., 2020; Chen et al., 2021; Sorop et al., 2021). Exosomes have attracted significant attention not only because of their therapeutic effect or their function as biomarkers but also because of their potential use for next-generation drug delivery systems (Liang et al., 2021). Compared with the traditional gene carrier, the advantages of exosomes lie in the following aspects: (1) as an endogenous vesicle, exosomes have low immunogenicity; (2) their biocompatible structure makes them easy to enter cells, which enables gene delivery more efficient; (3) their robust membrane can protect therapeutic nucleic acids from degradation; (4) their ability to avoid phagocytosis and bypass engulfment by lysosomes makes them well-tolerated in body fluids; (5) their membrane can be artificially engineered for targeted delivery, which makes tissue-specific or cell-specific distribution possible (Pirisinu et al., 2020; Herrmann et al., 2021; Ullah et al., 2021; Yao et al., 2021).

However, the yield of exosomes from cell culture supernatant is so low that the application of exosomes for drug delivery is limited (Pirisinu et al., 2020). Therefore, the urgent task is to effectively obtain exosomes in mass quantities for further pharmaceutical applications. In 1973, Plantz et al. discovered the presence of exosomes in bovine milk, which is now considered a promising candidate in developing a new

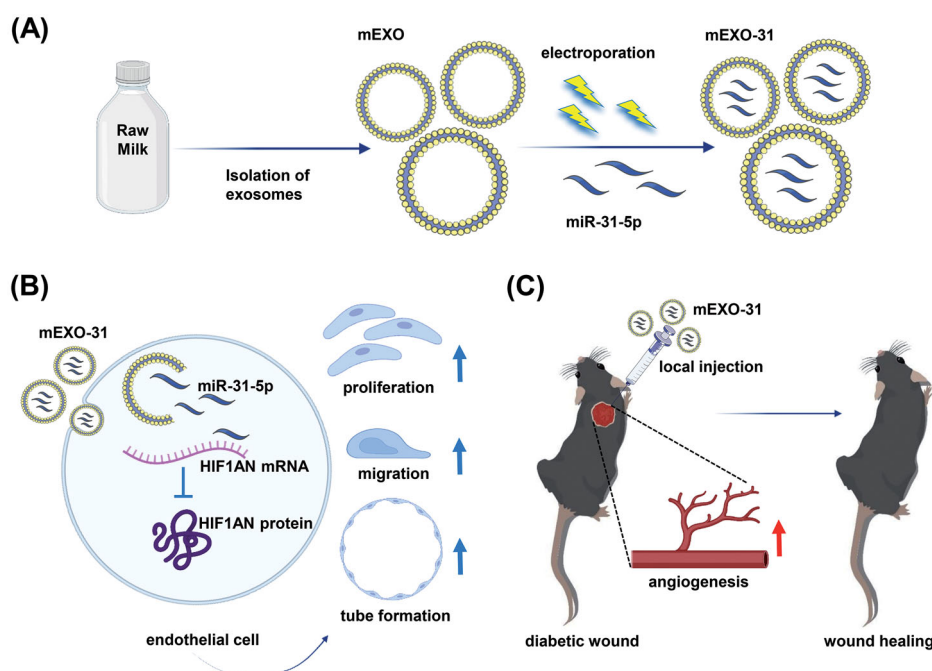
drug delivery system (Plantz et al., 1973; Munagala et al., 2016). Firstly, the quantity of exosomes isolated from bovine milk is relatively large, which is the biggest advantage that cannot be matched by any other sources of exosomes (Sedykh et al., 2020). In addition, milk-derived exosomes are highly resistant to harsh gastrointestinal tract environments and can be modified as an oral delivery vehicle (Aqil et al., 2016; Rani et al., 2017; Betker et al., 2019).

Here, we investigated the therapeutic efficacy of miR-31-5p encapsulated in milk exosomes in diabetic wound healing. We detected that miR-31-5p expression was reduced in skin wounds of diabetic mice compared with normal ones. Then, we isolated milk-derived exosomes (mEXO) by differential centrifugation and synthesized miR-31-5p-loaded exosomes (mEXO-31) by electroporation. We treated endothelial cells with mEXO-31 and found that miR-31-5p was upregulated in the recipient cells, while the target gene hypoxia-inducible factor 1 subunit alpha inhibitor (HIF1AN) was downregulated. Furthermore, the exosomes-coated miR-31-5p displayed potent pro-angiogenesis activity both *in vitro* and *in vivo* (Figure 1).

## Materials and methods

### Cell culture and transfection

Human umbilical vein endothelial cells (HUVECs) (#GDC166, CCTCC) were obtained from the China Center for Type Culture Collection (CCTCC, Wuhan, China) and grown in high-glucose Dulbecco's modified Eagle's medium (DMEM; Gibco, USA) supplemented with 10% fetal bovine serum (FBS; Gibco, USA), 100 U/ml of penicillin, and 100 µg/ml of streptomycin in a 5% CO<sub>2</sub> humidified atmosphere at 37 °C. The miR-31-5p mimic, miRNA-31-5p inhibitor, and the relevant negative controls (mimic NC and inhibitor NC) were



**Figure 1.** Schematic image. (A) Isolation of milk-derived exosomes (mEXO) and preparation of miR-31-5p-loaded exosomes (mEXO-31). (B) Action mechanism of mEXO-31 *in vitro*. (C) mEXO-31 treatment on mice model.

obtained from Ribobio company (Guangzhou, China). The sequences were presented in Table S1. HUVECs were transfected using riboFECT<sup>TM</sup>CP Reagent according to the manufacturer's instruction. The plasmids for overexpressing were purchased from Genomeditech Biotechnology (Shanghai, China). Cell transfection was performed with the NEOFECT<sup>TM</sup> DNA transfection reagent according to the manufacturer's protocol. At 48 h after transfection, the cells were processed for *in vitro* assays.

### **HUVECs proliferation, migration, and tube formation assay**

HUVECs from different treated groups were grown in 96-well culture plates (4000 cells per well) for 12 h. After 2 h incubation with the EdU, the proliferation rates of HUVECs from each group were evaluated with Cell-Light EdU Apollo *In Vitro* Kit (Ribobio, Guangzhou, China). The 24-well Transwell Chamber (8.0 μm pore size, Corning, USA) was used for assessing the migration ability of HUVECs from different treated groups. Briefly, HUVECs (16,000 cells per well) suspended in DMEM without serum were added to the upper compartment and then incubated in a complete culture medium containing 10% FBS for 24 h. Then, HUVECs migrated to the bottom surface were stained with crystal violet staining (Solarbio, Beijing, China) and counted under microscopy. For tube formation assay, HUVECs from different treated groups were seeded in 96-well culture plates (25,000 cells per well) that had been coated with 70 μl Matrigel Basement Membrane Matrix (BD Biosciences, CA, USA). Tube formation was detected under microscopy at 6 h incubation. The total tube length of the endothelial tubes was measured by the angiogenesis plugin available from ImageJ.

### **RT-PCR**

MiRNA and total RNA were isolated using miRNA Purification Kit (CW0627S, CWBIO) and Ultrapure RNA Kit (CW0581M, CWBIO), respectively according to the manufacturer's instruction. MiRNA and total RNA were reversely transcribed into cDNA using the Prime-Script<sup>®</sup> RT reagent Kit (#RR037A, TaKaRa). Real-time PCR was performed on the StepOnePlus<sup>TM</sup> platform (Applied Biosystems, USA) using TB Green<sup>®</sup> Premix Ex Taq<sup>TM</sup> II kit (#RR820A, TaKaRa). The primer sequences were presented in Tables S2, S3. The relative expression levels of targeted genes were calculated using the  $2^{-\Delta\Delta Ct}$  method and normalized to  $\beta$ -tubulin or u6.

### **Western blot**

Total proteins were extracted by RIPA lysis buffer with proteinase inhibitor (Roche, Switzerland). An equal amount of total protein (20–40 μg) was separated by SDS-PAGE (Beyotime Biotechnology, Shanghai, China), transferred into the PVDF membrane (Millipore, USA), and then incubated overnight with primary antibodies specific for HIF1AN (Abcame, USA),  $\beta$ -tubulin (Proteintech, China), CD9 (Abcame, USA), CD81 (Abcame, USA), HSP70 (Abcame, USA), TSG101

(ABclonal, China). Then, the membrane was incubated with secondary antibodies (Aspen, China) for 1 h and exposed to X-ray film (UVP, USA).

### **Dual-luciferase reporter gene assay**

The 3'UTR of HIF1AN was inserted into pmirGLO-luciferase promoter vector. For the mutant assay, the binding motif TCTTGCC was replaced with ATCGAGG. HEK-293T cells were seeded and cultured on 24-well plates. Then, cells were co-transfected with the miRNAs (mimic NC, miR-31-5p mimic) and pmirGLO-luciferase promoter vector containing the WT or MUT 3'UTR of HIF1AN. After 48 h, cells were collected and luciferase was detected using Dual-Luciferase Reporter Assay (Promega Corporation, USA). The firefly luciferase signal was normalized to that of the renilla luciferase signal.

### **Isolation of milk-derived exosomes (mEXO)**

Exosomes were isolated by differential centrifugation from raw milk as described (Aqil et al., 2019; Tao et al., 2020). Briefly, milk was centrifuged at  $13,000 \times g$  at 4 °C for 30 min to remove fat globules, cells, and cell debris. The upper-fat layer and pellet in the bottom of the tube were discarded and the supernatant was collected. The supernatant was then centrifuged at  $100,000 \times g$  at 4 °C for 60 min to remove large particles and microvesicles. Then the supernatant was centrifuged at  $145,000 \times g$  for 90 min at 4 °C and the exosome pellet was collected and washed three times with PBS and filtered through a 0.22 μm filter membrane. Exosome suspension was stored at –80 °C until used.

### **Preparation and characterization of miR-31-5p-loaded mEXO (mEXO-31)**

Milk-derived exosomes (mEXO) were loaded with miR-31-5p mimics (mEXO-31) or mimic NC (mEXO-NC) by electroporation using the CUY21EDIT II (BEX, Japan) electroporation system. The electroporation mixture was prepared by mixing mEXO and miR-31-5p in a 1:1 (wt/wt) ratio in PBS, with the final concentration of mEXO in the mixture was 0.1 mg/ml. The mixture was transferred into ice-cold 0.4-cm cuvettes and electroporated for 10 cycles with a perforation voltage of 110 V, a perforation opening time of 6 ms, a perforation interval of 10 ms, a penetration voltage of 25 V, and a capacitance of 940 μF. Post-electroporation, the mixture was transferred into a new tube at 37 °C for 30 min. Then, the unloaded mimics or mimic NC were removed through ultracentrifugation methods. The morphology and the size distribution of exosomes in each sample were detected by transmission electron microscope (TEM) and nanoparticle tracking analysis (NTA), and the protein level was quantified with Pierce BCA Protein Assay Kit (Aspen, China) as the manufacturer's instructions. To measure the stability of the miR-31-5p loaded in exosomes, free miR-31-5p mimics mixed with mEXO and mEXO-31 were incubated at 37 °C for 5 days. The miR-31-5p levels remaining in each sample were detected through RT-PCR.

### Confocal microscopy

mEXOs were incubated with a red fluorescent dye (Dil, Biotium, USA) for 30 min, and then were centrifuged to remove contaminating dye and to obtain the labeled mEXOs. For internalization assay, HUVECs were seeded in 24-well culture plates (30,000 cells per well) for 12 h, and co-cultured with Dil-labeled mEXOs for 24 h. After incubation, cells were washed twice with PBS and fixed in 4% paraformaldehyde for 10 min; thereafter, the nucleic was stained with DAPI (Solarbio, Beijing, China) and the cytoskeleton was stained with FITC phalloidin (Yeasen Biotech Co., Shanghai, China) according to the manufacturer's instructions. The mEXO uptake by cells was observed by using the laser scanning confocal microscope. For cell miR-31-5p uptake assay, FAM-labeled miR-31-5p mimics were obtained from Ribobio company (Guangzhou, China) and were encapsulated into Dil-labeled mEXOs through electroporation. HUVECs were seeded in 24-well culture plates (30,000 cells per well) for 12 h, and co-cultured with PBS, FAM-labeled miR-31-5p mimics, Dil-labeled mEXOs, and FAM-Dil-labeled mEXO-31, respectively for 24 h. After incubation, cells were washed twice with PBS and fixed in 4% paraformaldehyde for 10 min; thereafter, the nucleic was stained with DAPI (Solarbio, Beijing, China). The cell miR-31-5p uptake was observed by using the laser scanning confocal microscope.

### Diabetic wound model

All animal experiments were approved by the Animal Care Committee of Tongji Medical College. Eight-week-old male BALB/c mice were intraperitoneally injected with streptozotocin (STZ, 50 mg/kg) for 5 days, and after 2 weeks, the blood glucose was measured by a blood glucose monitor. Diabetic mice were successfully induced when the blood glucose was above 16.7 mM and maintained for another 4 weeks before full-thickness cutaneous wounds were formed. Before surgery, a total of 36 diabetic mice were anesthetized with pentobarbital sodium (Sigma–Aldrich) (1%, 50 mg/kg). After shaving and sterilization, a full-thickness excision wound at a diameter of 8 mm was performed on the back of all mice. Then the 36 mice were randomly divided into 6 groups: PBS (control) group; mimic NC group (2 nmol/wound); free miR-31-5p mimic group (2 nmol/wound); mEXO group (1.0  $\mu\text{g}/\mu\text{l}$ ); mEXO-NC group (1.0  $\mu\text{g}/\mu\text{l}$ ) and mEXO-31 group (1.0  $\mu\text{g}/\mu\text{l}$ ). Treatments were performed at day 0, 5, and 10 post-wounding. Digital photographs were taken at day 0, 5, 10, and 15, and the wound area was measured using the Image J software.

### Histological analysis

At day 15 post-wounding, the whole wound bed of each mouse was obtained for histological analysis. The wounds were fixed with 4% paraformaldehyde. After being dehydrated with a series of graded ethanol, the tissues were then embedded in paraffin and cut into 8  $\mu\text{m}$  thick longitudinal sections before further staining. The rate of

re-epithelialization was analysis by using hematoxylin and eosin (H&E) staining, and the degree of collagen accumulation was evaluated by Masson staining.

### Immunofluorescence analysis

To determine angiogenesis of wound beds, the sections were incubated with CD31, and  $\alpha$ -SMA antibody (Abcam, USA) overnight at 4 °C. After being washed three times with PBS, the sections were incubated with a second antibody (Aspen, China) for 1 h at room temperature. The image was taken by a microscope and then analyzed by using ImageJ software.

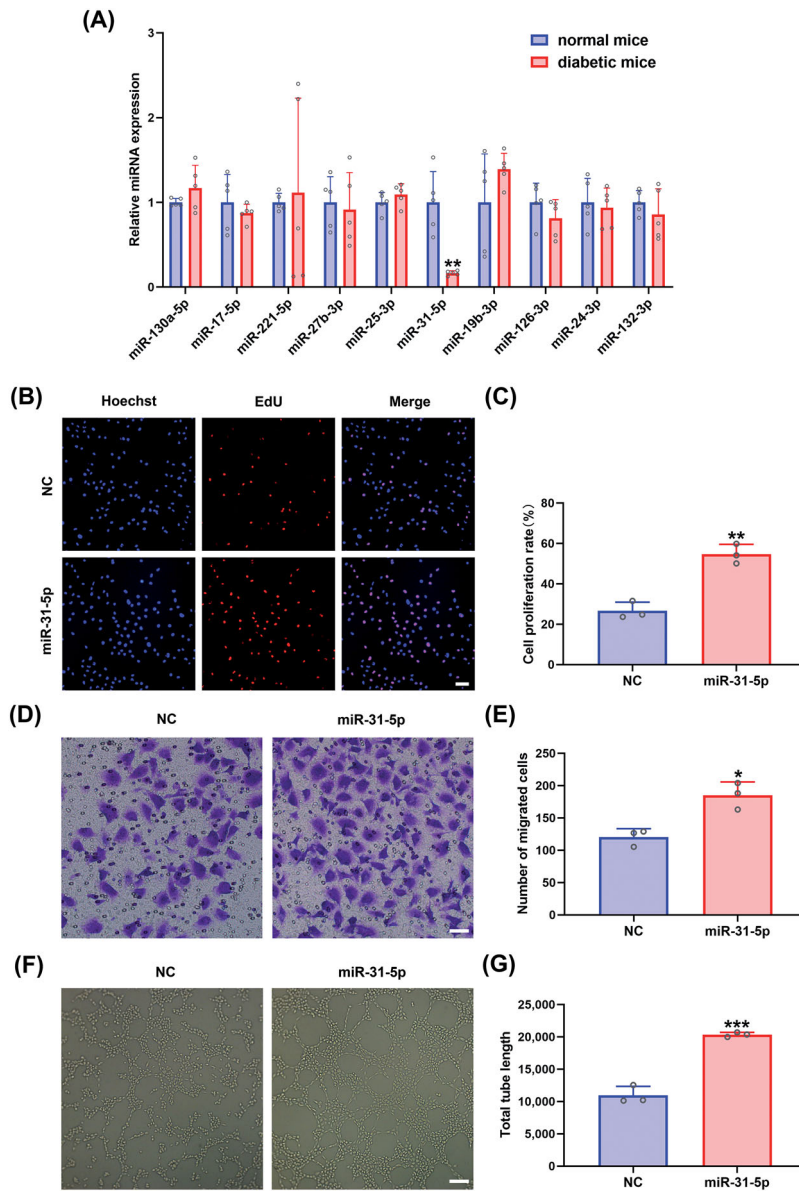
### Statistics

All statistical analyses were performed using GraphPad Prism software (version 8.0.2, La Jolla, CA, USA). For the comparison of two groups, an unpaired Student's *t*-test was applied. For group  $\geq 3$ , a one-way analysis of variance (ANOVA) with Tukey *post-hoc* test was carried out. All data are presented as mean  $\pm$  standard deviation (*SD*). Statistical significance was set at  $p < .05$ .

## Results

### Overexpression of miR-31-5p promoted endothelial cell proliferation, migration, and angiogenesis

To encapsulate a proangiogenic miRNA in milk exosomes to enhance diabetic wound healing, we compared the expression levels of ten miRNAs, including miR-130a-5p, miR-17-5p, miR-221-5p, miR-27b-3p, miR-25-3p, miR-31-5p, miR-19b-3p, miR-126-3p, miR-24-3p, and miR-132-3p, in full-thickness skin lesions between diabetic and normal mice to find the aberrantly downregulated miRNAs. Among five downregulated miRNAs in diabetes wounds (miR-17-5p, miR-27b-3p, miR-31-5p, miR-126-3p, and miR-132-3p), miR-31-5p showed statistically significant down regulation (Figure 2(A)). The significance of lower miR-31-5p was also tested in db/db T2DM murine model (Figure SF3(A)). To simulate the abnormally expressed miR-31-5p in diabetic wounds and to investigate the role of miR-31-5p, HUVECs were transfected with miR-31-5p inhibitors. We found that miR-31-5p inhibitors displayed an anti-proliferation, anti-migration, and anti-angiogenesis activity in HUVECs (Figure SF1). Thus, we hypothesized that the exogenous supplement of miR-31-5p could restore endothelial cell functions. After transfecting miR-31-5p mimics to HUVECs for 48 h, we conducted a series of assays below. Firstly, the proliferation of the cells was determined by EdU assays, and much more EdU-positive HUVECs were found when the cells were transfected with mimics (Figure 2(B,C)). Additionally, transwell assays showed that miR-31-5p significantly increased migration of HUVECs (Figure 2(D,E)). Lastly, *in vitro* tube formation assays were performed to assess the ability of miR-31-5p to promote endothelial cell angiogenesis. As we expected, there was a nearly 2-fold increase in total tube length, which indicated that miR-31-5p



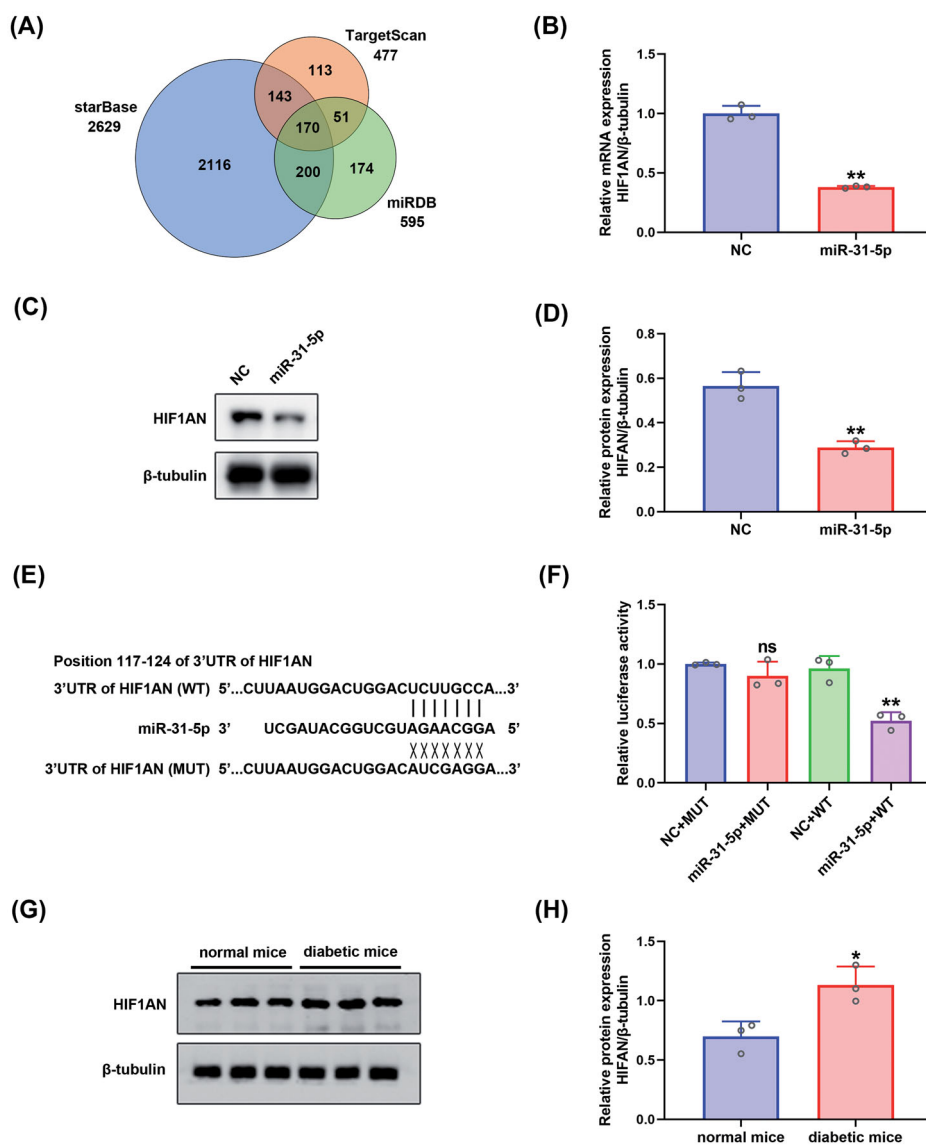
**Figure 2.** Overexpression of miR-31-5p promoted endothelial cell proliferation, migration, and angiogenesis. (A) Relative expression levels of miRNAs in wound tissue compared between normal mice and diabetic mice.  $n = 5$ ,  $^{*}p < .01$  vs. normal mice. (B,C) EdU assay analysis of the proliferation rate of HUVECs treated with mimic-NC/miR-31-5p mimics. The proliferative cells and cellular nuclei were stained with red and blue colors.  $n = 3$ ,  $^{**}p < .01$  vs. NC. Scale bar, 50  $\mu$ m. (D,E) Images of migrated HUVECs in each group.  $n = 3$ ,  $^{*}p < .05$  vs. NC. Scale bar, 50  $\mu$ m. (F,G) Images of tube formation of HUVECs in each group.  $n = 3$ ,  $^{***}p < .001$  vs. NC. Scale bar, 100  $\mu$ m. Data were presented as mean  $\pm$  SD. Unpaired Student's *t*-test was used.

dramatically promoted angiogenesis (Figure 2(F,G)). In summary, these data suggest that miR-31-5p was down-regulated in diabetic mice, which may be one of the reasons for delayed wound healing, and the overexpression of miR-31-5p enhanced the HUVEC proliferation, migration, and angiogenesis *in vitro*.

### HIF1AN was a direct target of miR-31-5p

To seek out the downstream mechanisms governing the function of endothelial cells, three independent algorithms: starBase (Li et al., 2014), TargetScan (Agarwal et al., 2015), and miRDB (Chen & Wang, 2020) were used to predict the potential targets of miR-31-5p. 170 putative miR-31-5p targets were predicted by the three algorithms (Figure 3(A)). Then, we searched DAVID, a web-based functional

annotation tool (Jiao et al., 2012), for functional annotations of the 170 genes. We found that HIF1AN, STARD13, and SPARC were three genes related to the negative regulation of angiogenesis. We selected HIF1AN for further study because the target score of HIF1AN was the highest among the three genes. The three algorithms revealed that HIF1AN was a conserved target of miR-31-5p with the pairing position of 117-124 in the 3' untranslated region (UTR) of HIF1AN 3'UTR (Figure 3(E)). To further confirm that HIF1AN was a direct target of miR-31-5p, miR-31-5p mimics were transfected into HUVECs. We found that HIF1AN levels in miR-31-5p mimics transfected HUVECs decreased in both mRNA (Figure 3(B)) and protein (Figure 3(C,D)) levels. Then the luciferase reporters (pmirGLO-3'UTR of HIF1AN-WT and HIF1AN-MUT) were constructed and transfected into the 293T cells. It was seen that overexpression of miR-31-5p

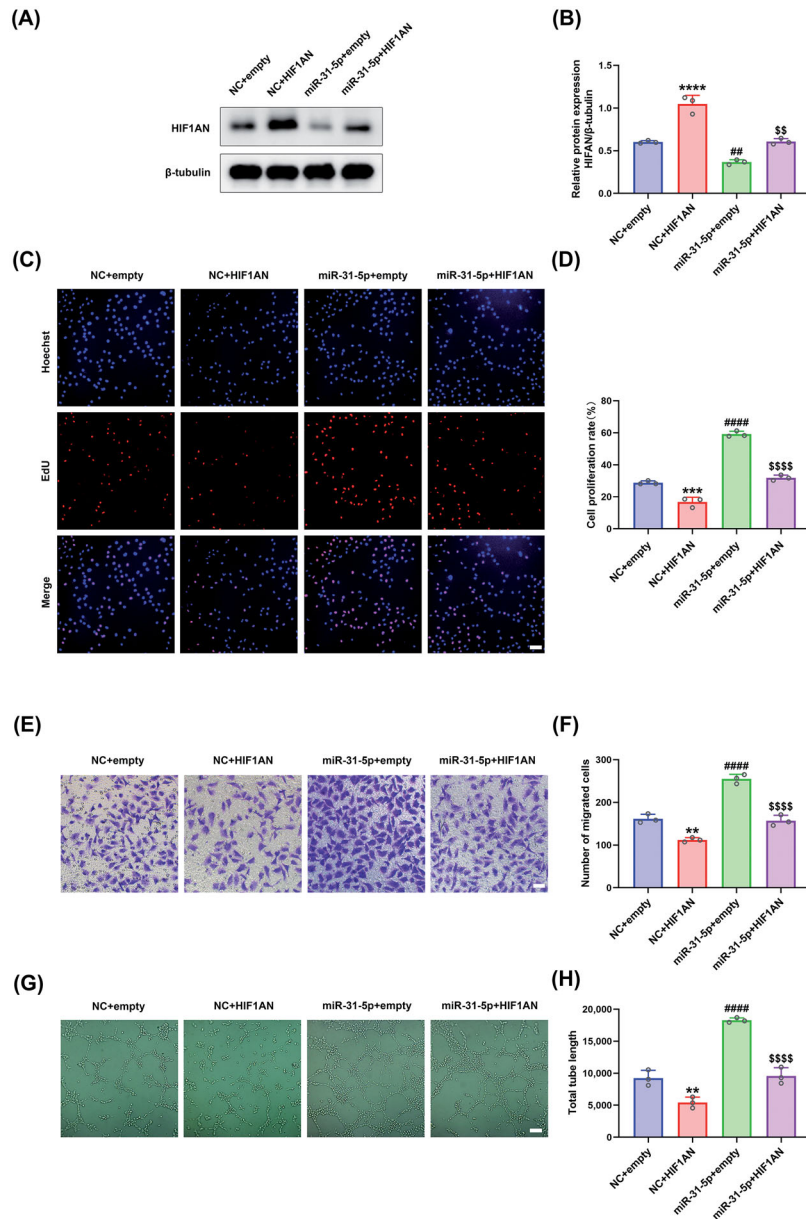


**Figure 3.** HIF1AN was a direct target of miR-31-5p. (A) Venn diagram depicting the number of potential targets of miR-31-5p predicted by three algorithms. (B) RT-PCR analysis of HIF1AN mRNA expression in HUVECs transfected with mimic-NC/miR-31-5p mimics.  $n = 3$ ,  $**p < .01$  vs. NC. (C,D) Western blotting analysis of HIF1AN protein expression in HUVECs in each group.  $n = 3$ ,  $**p < .01$  vs. NC. (E) Schematic drawing of the putative binding sites or mutations of miR-31-5p in HIF1AN mRNA 3'UTR. (F) Luciferase activity of each group was detected at 48 h post-transfection.  $n = 3$ , ns no significant vs. NC + MUT;  $**p < .01$  vs. NC + WT. (G,H) Western blotting analysis of HIF1AN protein expression in wound tissue from normal and diabetic mice.  $n = 3$ ,  $*p < .05$  vs. normal mice. Data were presented as mean  $\pm$  SD. Unpaired Student's *t*-test was used in (B,D,H). One-way ANOVA with Tukey *post-hoc* test was used in (F).

apparently suppressed the luciferase activity of pmirGLO-HIF1AN-WT compared with empty vector control, while exerted no significant effects on the luciferase activity of pmirGLO-HIF1AN-MUT in 293T cells (Figure 3(F)). In view of the association of HIF1AN with miR-31-5p, we also assessed the expression level of HIF1AN in wound tissue from diabetic mice via western blot. We then found an upregulation of HIF1AN in diabetic wounds compared to normal ones (Figure 3(G,H)), which presented the completely opposite tendency to miR-31-5p. The significance of higher HIF1AN was also tested in db/db T2DM murine model (Figures SF3(B,C)). Collectively, these results revealed that the expression of HIF1AN was upregulated in diabetic wounds and miR-31-5p could inhibit HIF1AN expression by binding to the conservative complementary sequence of the HIF1AN mRNA 3'UTR.

### miR-31-5p/HIF1AN axis regulated endothelial cell function

We further studied the biological function of miR-31-5p/HIF1AN axis in HUVECs. Firstly, the protein levels of HIF1AN were detected in HUVECs under different treatments, showing that transfection of HIF1AN overexpressing plasmid achieved a higher level of HIF1AN than empty control plasmid, and rescued the reduced HIF1AN content induced by miR-31-5p mimics (Figure 4(A,B)). Then, EdU assays were performed to detect the proliferation activity of HUVECs. We demonstrated that the overexpression of HIF1AN exhibited a notable anti-proliferation effect and obviously counteracted miR-31-5p-improved HUVEC proliferation (Figure 4(C,D)). Moreover, a similar trend could be observed in the transwell assays detecting the migration activity of the cells



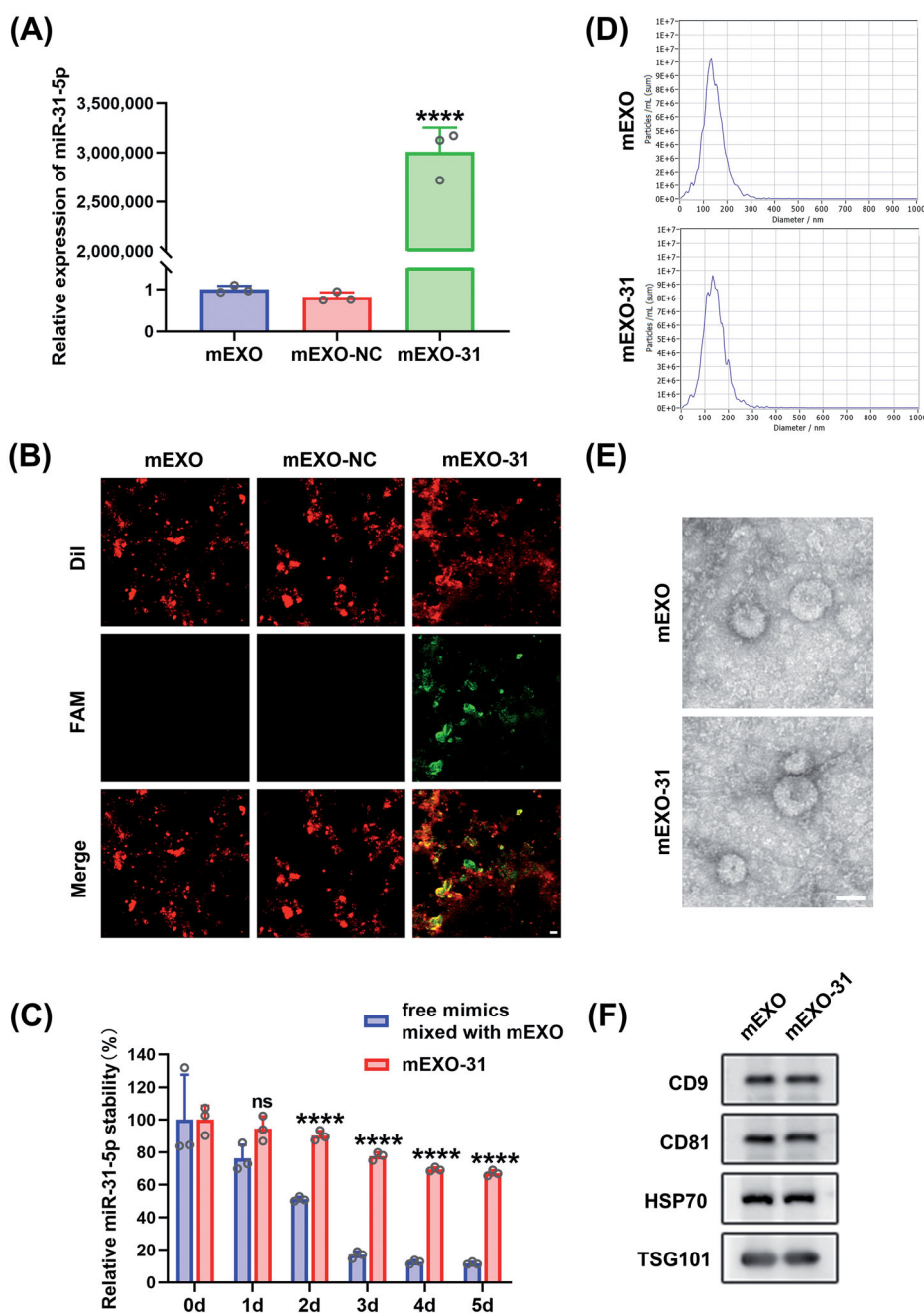
**Figure 4.** miR-31-5p/HIF1AN axis regulated endothelial cell function. (A,B) Western blotting analysis of HIF1AN protein expression in HUVECs in different treated groups.  $n = 3$ , \*\*\*\* $p < .0001$ , ## $p < .01$  vs. NC + empty;  $^{SS}p < .01$  vs. miR-31-5p + empty. (C,D) EdU assay analysis of the proliferation rate of HUVECs in each group. The proliferative cells and cellular nuclei were stained with red and blue colors. Scale bar, 50  $\mu$ m.  $n = 3$ , \*\*\* $p < .001$ , #### $p < .0001$  vs. NC + empty;  $p < .0001$  vs. miR-31-5p + empty. (E,F) Images of migrated HUVECs in each group. Scar bar, 50  $\mu$ m.  $n = 3$ , \*\* $p < .01$ , #### $p < .0001$  vs. NC + empty;  $p < .0001$  vs. miR-31-5p + empty. (G,H) Images of tube formation of HUVECs in each group. Scar bar, 100  $\mu$ m.  $n = 3$ , \*\* $p < .01$ , #### $p < .0001$  vs. NC + empty;  $p < .0001$  vs. miR-31-5p + empty. Data were presented as mean  $\pm$  SD. One-way ANOVA with Tukey *post-hoc* test was used.

(Figure 4(E,F)). We supposed that the overexpression of HIF1AN could inhibit the vascular formation of HUVECs, and the subsequent *in vitro* tube formation assays confirmed the hypothesis (Figure 4(G,H)). We also demonstrated that HIF1AN overexpression abolished the miR-31-5p mimic-induced promotion of angiogenesis (Figure 4(G,H)). In summary, our rescue experiments confirmed that miR-31-5p promoted endothelial cell functions through targeting HIF1AN.

#### Preparation and characterization of mEXO-31

To investigate whether milk-derived exosomes (mEXO) can function as an ideal cargo carrier to deliver miR-31-5p to

endothelial cells and diabetic wounds, we isolated exosomes from raw milk using a differential centrifugation approach. We demonstrated the internalization and the biocompatibility of mEXO in HUVECs (Figure SF2). Then, we used electroporation approach to load mimic-NC and miR-31-5p mimics into mEXO (named as mEXO-NC and mEXO-31, respectively), followed by removing the unloaded NC and mimics through ultracentrifugation. To verify that miR-31-5p mimics were successfully loaded into mEXO, we detected the level of miR-31-5p in mEXO, mEXO-NC, and mEXO-31 by RT-PCR. The results showed that the miR-31-5p level was dramatically higher in mEXO-31 than in mEXO or mEXO-NC (Figure 5(A)). Moreover, confocal laser-scanning microscopy visually



**Figure 5.** Preparation and characterization of mEXO-31. (A) RT-PCR analysis of relative miR-31-5p levels in mEXO, mEXO-NC, and mEXO-31.  $n = 3$ , \*\*\*\* $p < .0001$  vs. mEXO-NC. (B) Confocal images showed successful loading of miR-31-5p into mEXO. Red and green fluorescence represented mEXO and miR-31-5p mimics, respectively. Scale bar, 10  $\mu\text{m}$ . (C) RT-PCR analysis of remaining miR-31-5p in each group.  $n = 3$ , ns no significant, \*\*\*\* $p < .0001$  vs. free mimics mixed with mEXO. (D) NTA identified the size distribution of mEXO and mEXO-31. (E) TEM identified the morphology of mEXO and mEXO-31. Scale bar, 50  $\mu\text{m}$ . (F) Western blotting analysis of exosome specific markers including CD9, CD81, HSP70, and TSG101 of mEXO and mEXO-31. Data were presented as mean  $\pm$  SD. One-way ANOVA with Tukey *post-hoc* test was used in (A). Unpaired Student's *t*-test was used in (C).

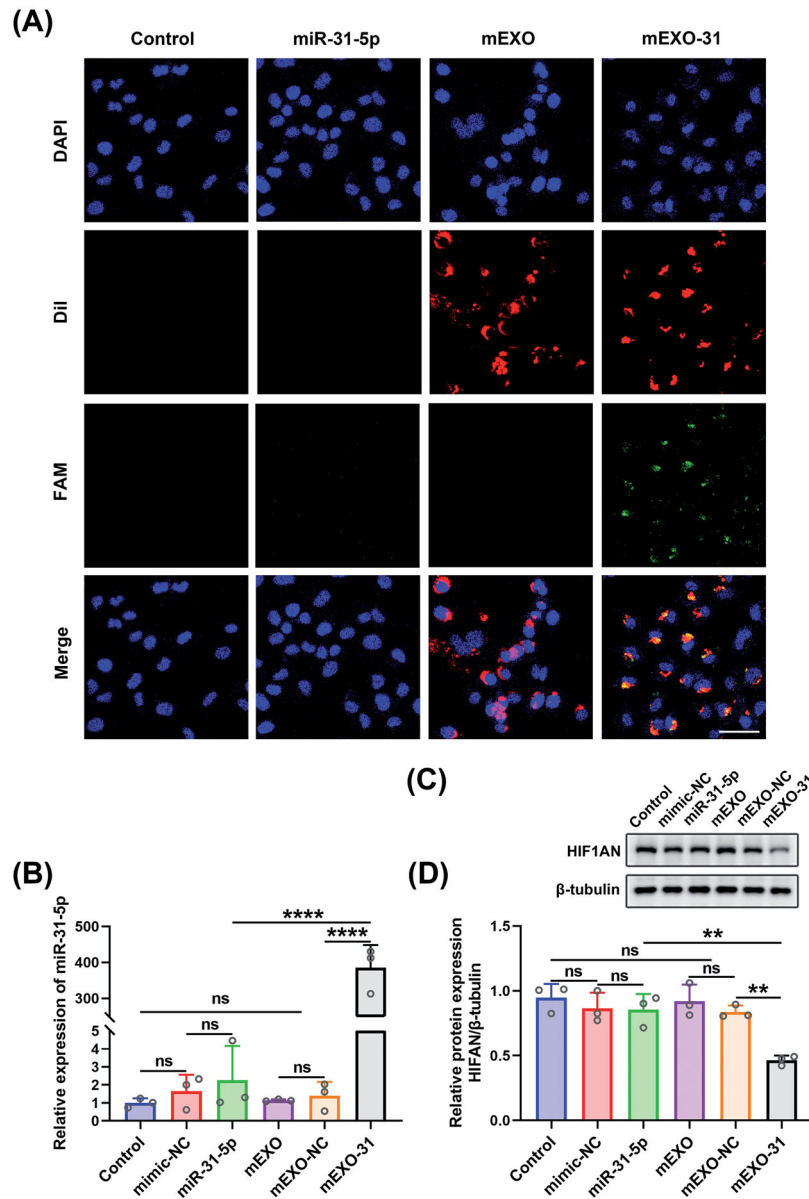
observed that FAM-labeled miR-31-5p mimics were successfully loaded into mEXO (Figure 5(B)).

The mean size of mEXO and mEXO-31 was 131.1 and 133.6 nm in diameter, respectively as measured by NTA (Figure 5(D)). Inwardly concave cuplike structure was found in the two exosomes as observed by TEM (Figure 5(E)). Western blotting analysis revealed that there was high expression of exosome-specific markers in the two exosomes including CD9, CD81, HSP70, TSG101 (Figure 5(F)). These data indicated that loading miRNA by electroporation into mEXO had almost no influence on

the particle size, morphology, or surface markers of the mEXO.

Further, we explored the extent to which the mEXO protected the miRNA. Free miR-31-5p mimics mixed with mEXO and mEXO-31 were exposed to 37  $^{\circ}\text{C}$  for 5 days, and then the miR-31-5p level was quantified from each sample. The results showed that free miR-31-5p mimics almost totally degraded at day 5, on the contrary, more than 60% of the miR-31-5p loaded in mEXO remained stable (Figure 5(C)), indicating that mEXO had the ability to protect miR-31-5p mimics from degradation.





**Figure 6.** mEXO-31 delivered miR-31-5p into cells more efficiently. (A) Confocal images showed the delivery of miR-31-5p into HUVECs in each group. Blue, red, and green fluorescence represented the cellular nuclei, mEXO, and miR-31-5p, respectively. Scale bar, 50  $\mu$ m. (B) RT-PCR analysis of relative miR-31-5p levels in HUVECs in different treated groups. (C,D) Western blotting analysis of HIF1AN protein expression in HUVECs in different treated groups.  $n = 3$ , ns: no significant, \*\* $p < .01$ , \*\*\*\* $p < .0001$ . Data were presented as mean  $\pm$  SD. One-way ANOVA with Tukey *post-hoc* test was used.

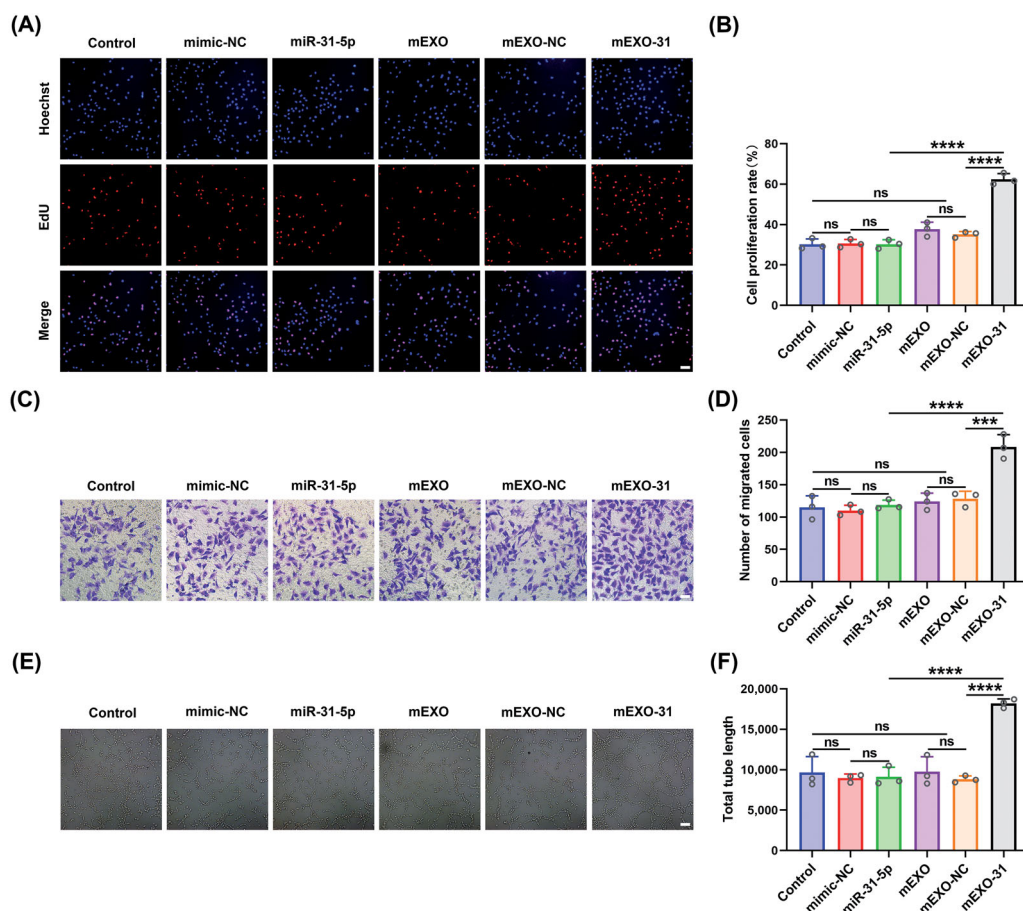
### **mEXO-31 delivered miR-31-5p into cells more efficiently**

For the sake of detecting the transfection efficiency of mEXO-31, we transfected HUVECs with PBS, free miR-31-5p mimics, mEXO, and mEXO-31, respectively, and determined the efficiency using confocal laser-scanning microscopy. As shown in Figure 6(A), almost all HUVECs were successfully transfected with miR-31-5p when the cells were treated with mEXO-31, while nearly no miR-31-5p was detected in the cells when treated with free mimics. We then respectively treated HUVECs with PBS, free mimic-NC, free miR-31-5p mimics, mEXO, mEXO-NC, and mEXO-31. RT-PCR was used to detect the expression level of miR-31-5p in HUVECs, showing that there was a remarkable increase of miR-31-5p level in mEXO-31 group than in mEXO-NC and free mimics group (Figure 6(B)). In addition, the protein level of HIF1AN was

significantly downregulated in HUVECs treated with mEXO-31, while the level of HIF1AN remained unchanged in other groups (Figure 6(C,D)). These results collectively indicated that naked free miRNA was difficult to be taken up by cells, and the dilemma of miRNA-based therapeutics could be broken by the milk exosome delivery system. More than that, the biological function of miR-31-5p encapsulated in mEXO was preserved, and it could reduce the expression of the target gene HIF1AN.

### **mEXO-31 promoted endothelial cell proliferation, migration, and angiogenesis in vitro**

To further investigate the functional effects of mEXO-31 on HUVECs, we treated HUVECs with PBS, free mimic-NC, free



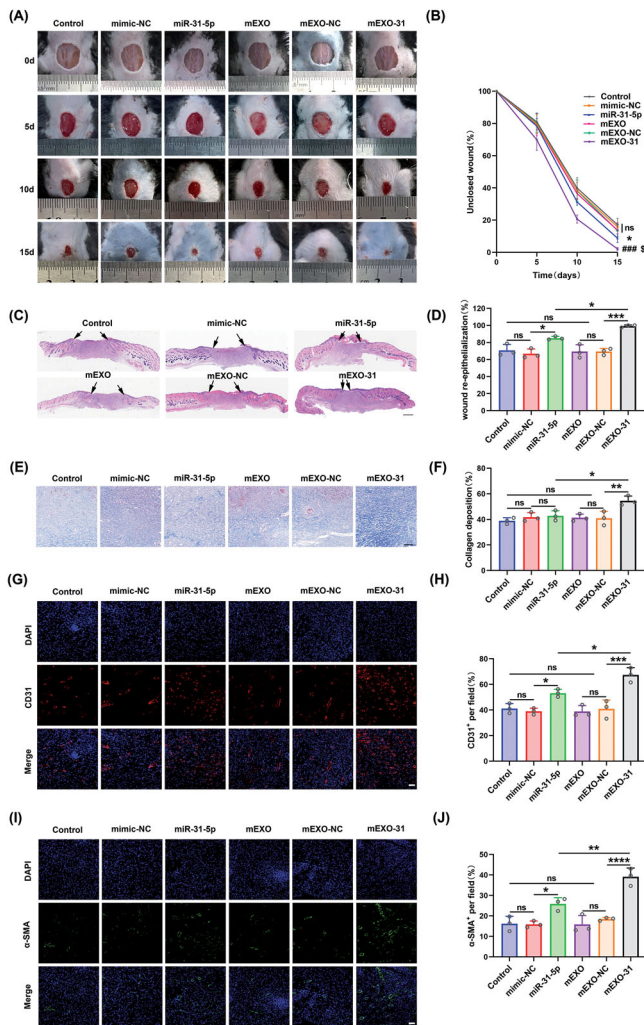
**Figure 7.** mEXO-31 promoted endothelial cell proliferation, migration, and angiogenesis *in vitro*. (A,B) EdU assay analysis of the proliferation rate of HUVECs in different treated groups. The proliferative cells and cellular nuclei were stained with red and blue colors. Scale bar, 50  $\mu\text{m}$ . (C,D) Images of tube formation of HUVECs in each group. Scale bar, 100  $\mu\text{m}$ .  $n = 3$ , ns: no significant,  $***p < .001$ ,  $****p < .0001$ . Data were presented as mean  $\pm$  SD. One-way ANOVA with Tukey *post-hoc* test was used.

miR-31-5p mimics, mEXO, mEXO-NC, and mEXO-31, respectively. Then, we evaluated the cell behavior by using EdU assays, transwell assays, and *in vitro* tube formation assays. Firstly, EdU assays were performed to assess the proliferation ability of endothelial cells. Consistent with our expectations, mEXO-31 treatment greatly promoted cell proliferation: the rate of EdU-positive HUVECs was increased more than 2-fold in mEXO-31 group compared with other groups (Figure 7(A,B)). Next, we evaluated the migration ability of HUVECs by conducting transwell assays. After being treated for 2 days, the cells were seeded in a transwell system. As the results showed, significant differences were observed between the mEXO-31 group and other groups, with migration increasing about 1.5-fold when HUVECs were treated with mEXO-31 (Figure 7(C,D)). Furthermore, the angiogenic tube-forming ability of endothelial cells was detected by vascular network formation assays in matrigel. The results revealed that stimulation of HUVECs with mEXO-31 notably promoted cell network tube formation *in vitro*, exhibiting a nearly 2-fold increase in total tube length in mEXO-31 group than in other groups (Figure 7(E,F)). The above evidence strongly suggested that neither mEXO nor free miR-31-5p mimics could promote endothelial cell functions, while delivery miR-31-5p by mEXO notably enhanced the bioavailability

of miR-31-5p, thus improving the proliferation, migration, and tube formation of HUVECs.

### mEXO-31 accelerated diabetic wound healing *in vivo*

To explore the potential of mEXO-31 for clinical treatment, we established a full-thickness wound model at the dorsum of diabetic mice, and used PBS, free mimic-NC, free miR-31-5p mimics, mEXO, mEXO-NC, and mEXO-31 to treat the mice. As we expected, the unclosed wound rate in mice treated with mEXO-31 was the smallest in these different treated groups at days 5, 10, and 15 post-wounding (Figure 8(A,B)). Interestingly, in the free miR-31-5p mimics treated diabetic wounds, the unclosed wound rate was smaller than that in control, NC, mEXO, and mEXO-NC groups (Figure 8(A,B)). This phenomenon may be attributed to our added treatment in each group at days 5 and 10 post-wounding. However, the therapeutic effect of free miR-31-5p mimics was far from satisfactory compared to mEXO-31. Next, histological analysis was performed to investigate the wound repair efficiency in different treatment groups at day 15 post-operation. H&E staining analysis showed that mEXO-31 treated group exhibited the highest re-epithelialization rate among other groups



**Figure 8.** mEXO-31 accelerated diabetic wound healing *in vivo*. (A,B) Representative images of wound closure in a wound model at the dorsum of diabetic mice at day 0, 5, 10, and 15 post-wounding.  $n = 5$ , ns: no significant,  $*p < .05$  vs. mimic-NC;  $###p < .001$  vs. mEXO-NC;  $§p < .05$  vs. miR-31-5p. (C,D) H&E staining analysis of wound sections at day 15 post-wounding. The single-headed arrows indicate the un-epithelialized areas. Scar bar, 500  $\mu\text{m}$ . (E,F) Masson staining evaluated the collagen deposition at day 15 post-wounding. Scar bar, 100  $\mu\text{m}$ . (G–J) Immunofluorescence staining of cellular nuclei (DAPI, blue), CD31 (red), and  $\alpha$ -SMA (green) of wound bed at day 15 post-wounding. Scar bar, 50  $\mu\text{m}$ .  $n = 3$ , ns: no significant,  $*p < .05$ ,  $**p < .01$ ,  $***p < .001$ ,  $****p < .0001$ . Data were presented as mean  $\pm$  SD. One-way ANOVA with Tukey *post-hoc* test was used.

(Figure 8(C,D)). We also observed an enhanced re-epithelialization rate in the free mimics group (Figure 8(C,D)). In addition, the Masson staining analysis demonstrated that the collagen deposition in mEXO-31 group was better than that of other groups (Figure 8(E,F)). We observed no difference in collagen deposition between the mimics group and control, NC, mEXO, or mEXO-NC group (Figure 8(E,F)). CD31 and  $\alpha$ -SMA are two indicators of blood vessels and were used to evaluate the angiogenesis in the diabetic wound bed. As shown in Figure 8(G–J), very few blood vessels were observed in the wound tissues of the control, NC, mEXO, and mEXO-NC group, while the treatment of free miR-31-5p mimics improved vascular network formation. Notably, the highest vascular density was achieved in mEXO-31 group. These data indicated that mEXO-31 significantly accelerated

diabetic wound healing by promoting wound re-epithelialization, collagen deposition, and angiogenesis.

## Discussion

In the present study, miR-31-5p was selected from ten miRNA candidates to encapsulate in milk exosomes to accelerate diabetic wound healing. Comprehensive literature demonstrated that these ten miRNAs have been reported to be involved in the process of angiogenesis in diabetic complications and other diseases (Chen & Gorski, 2008; Rawal et al., 2017; Zhang et al., 2017). For instance, bone marrow-derived angiogenic cells (BMACs) exhibit impaired function in diabetes. While overexpression of miR-27b-3p could alleviate impaired BMAC function by improving cell proliferation, tube formation, adhesion, and delaying apoptosis. Moreover, cell therapy of BMACs overexpressing miR-27b-3p on diabetic wounds augmented wound perfusion and capillary formation (Wang et al., 2014). Among the 10 miRNAs, miR-31-5p was the only miRNA showing statistically significant downregulation in skin wounds of diabetic mice compared with normal ones.

Several studies have also investigated the aberrantly expressed miR-31-5p in diabetic complications. For example, the level of miR-31-5p was found to be downregulated in skin wounds of diabetic rats (Liu et al., 2015). Serum miR-31-5p level was reduced in T2D patients with diabetic nephropathy (Rovira-Llopis et al., 2018). During the wound healing process, previous studies have indicated that miR-31-5p played a crucial role in promoting keratinocyte function (Li et al., 2015; Shi et al., 2018). One study found that miR-31-5p was up-regulated in wound edge keratinocytes, and it promoted the proliferation and migration of keratinocytes by suppressing epithelial membrane protein 1 (EMP-1) (Li et al., 2015). Another study attributed the improved keratinocyte function to the miR-31-5p-mediated activation of the Ras/mitogen-activated protein kinase signaling (Shi et al., 2018). We showed here that miR-31-5p can also accelerate diabetic wound healing by improving endothelial cell function and promoting angiogenesis.

Consistent with our findings, miR-31-5p has been shown to be a proangiogenic miRNA in cancer and other diseases (Wu et al., 2011; Wang et al., 2014; Wong et al., 2015). It has been reported that miR-31-5p could induce endothelial motility and angiogenesis by targeting FAT atypical cadherin 4 (FAT4) (Wu et al., 2011). Similarly, overexpression of miR-31-5p or knockdown of FAT4 promoted endothelial progenitor cell (EPC) migration and tube formation in coronary artery disease (CAD) (Wang et al., 2014). The adhesion molecule E-selectin has been proven to be another target gene of miR-31-5p, and inhibition of E-selectin by miR-31-5p benefited HUVEC migration and angiogenesis (Suárez et al., 2010). In the current study, we demonstrated that miR-31-5p can positively regulate the proliferation, migration, and tube formation in HUVECs by targeting HIF1AN.

HIF1AN is an asparaginyl hydroxylase enzyme and it has been proven to be a negative regulator of HIF-1 transactivation domain function (Mahon et al., 2001; Lando et al., 2002).

The active form of HIF-1 is important to induce a beneficial proangiogenic response in patients with diabetic ulcers or non-healing wounds (Rey & Semenza, 2010; Hong et al., 2014). Several studies have observed that targeting HIF1AN contributed to improved angiogenesis in endothelial cells (Huang et al., 2011; Umezu et al., 2014). We validated that HIF1AN is a direct target of miR-31-5p in HUVECs through bioinformatics analysis, PCR analysis, western blotting, and luciferase reporter assay. This regulatory relationship between miR-31-5p and HIF1AN agrees with previous studies (Liu et al., 2010; Peng et al., 2012; Cheung et al., 2014; Hu et al., 2015; Zhu et al., 2019). Then, we found that the expression of HIF1AN was upregulated in diabetic wounds. Interestingly, a marked increase of HIF1AN expression was also observed in corneal epithelium from patients with diabetic keratopathies (Peng et al., 2012). The function of HIF1AN and the relationship between miR-31-5p and HIF1AN was further clarified in our rescue experiments, where the overexpression of HIF1AN suppressed cell function and counteracted the promoting effects of miR-31-5p in HUVECs.

In view of the dramatically downregulated miR-31-5p in diabetic wounds and its essential role in angiogenesis, we wondered whether miR-31-5p replacement technology could be used to restore its expression and promote diabetic wound healing. As mentioned above, miRNAs are extensively researched for their gene regulation activity, and miRNA-based therapeutics present an attractive proposition for the development of genetic therapeutics during wound healing (Chouhan et al., 2019; Nie et al., 2020). However, the utilization of miRNAs as a solution for wound healing often meets with several limitations. Firstly, naked free miRNAs are susceptible to degradation in the external environment. Not only that, the high molecular weight of miRNAs and the repulsion between negatively charged miRNAs and cell membrane hinders the successful cell uptake. As shown in our research, free miR-31-5p mimics placed in the 37 °C incubators almost totally degraded in 5 days. And nearly no miR-31-5p was detected to be taken up by cells when they were treated with naked free mimics. Hence, strong efforts must be undertaken to exploit a delivery medium to carry the miRNAs so that the enhanced retention and permeability of miRNAs in the wound environment can be achieved.

Over the last decade, several synthetic materials-based delivery systems have been reported to carry miRNAs, including polymeric nanoparticles, liposomes, peptides, and synthetic dendrimers (Liu et al., 2017; Ben-Shushan et al., 2014; Zhou et al., 2016). Despite great achievements, the use of synthetic materials-based delivery systems suffers from their poor stability, low loading efficiency, lack of scalability, and difficulty in synthesizing them in sufficient quantities (Feng & Mumper, 2013; Lim et al., 2013). Moreover, immune response, non-specific targeting, and toxicity concerns with the long-term use limit their clinical application (Sadasivam et al., 2013). Thus, we emphasized exosomes as an ideal miRNAs delivery system for the next generation of genetic therapeutics. Compared to traditional synthetic nanoparticles, a natural exosomal drug delivery system is biocompatible, safe, and efficacious (Ha et al., 2016; Rani & Ritter, 2016). In

recent years, researchers have established several miRNA delivery systems using exosomes from numerous cell types (Liang et al., 2018; Ma et al., 2018; Jeong et al., 2020; Kim et al., 2020; Liang et al., 2020). For example, human embryonic kidney 293T (HEK293T) cell-derived exosomes were used as delivery vehicles for miR-497, and the miR-497-loaded exosomes exhibited anti-tumor and anti-angiogenic effects on non-small cell lung cancer (Jeong et al., 2020).

Although some promising results have been reported to isolate exosomes from cells for delivering hydrophilic and hydrophobic drugs, the workload required to scale up the production of these cells combined with low exosome yield hamper the delivery system for clinical application (Smyth et al., 2015). In view of the current limitations of cell-derived exosomes and scalability issues, researchers have recognized milk as a potential source of exosomes, since large quantities of exosomes can be harvested from ordinary milk on an industrial scale. Arntz et al isolated exosomes from commercial semi-skimmed milk with a reported yield of 200 mg protein per liter of milk (Arntz et al., 2015). Another two researches collected exosomes from raw milk with a higher yield, about 300 mg exosomes per liter of milk (Munagala et al., 2016; Tao et al., 2020). Our exosome yield was around 200 mg per liter of raw milk. Thus, compared to very low yields of exosomes isolated from the medium of cultured cells, about 0.5–2.0 mg exosomal protein one liter of cell culture supernatant, the exosome yield from milk is over 200-fold higher, indicating that milk is a scalable source of exosomes for mass production (Sheng et al., 2011; Munagala et al., 2016; Faruqu et al., 2018).

Milk exosomes can be potentially loaded with gene therapeutic drugs (Aqil et al., 2019; Matsuda et al., 2020; Tao et al., 2020; Del Pozo-Acebo et al., 2021). Recent studies have employed passive strategies, such as incubation and active cargo-loading methods, such as sonication, extrusion, freeze and thaw cycles, electroporation, and saponin treatment to load drugs into milk exosomes (Luan et al., 2017; Adriano et al., 2021; Kandimalla et al., 2021). Here, we demonstrated successful loading of miR-31-5p mimics into milk exosomes by electroporation. Accumulating evidence have verified that the robust membrane of exosomes can protect therapeutic nucleic acid drugs from degradation (Aqil et al., 2019; Jeong et al., 2020; Tao et al., 2020; Del Pozo-Acebo et al., 2021). For instance, one study found that almost all naked free siRNA molecules were degraded after incubating at 37 °C for 8 h, while 67.1 ± 8.3% of the exosome-encapsulated siRNA retained the integrity after 24 h (Tao et al., 2020). Our present study showed that free miR-31-5p almost totally degraded incubated at 37 °C in 5 days, on the contrary, more than 60% of the miR-31-5p loaded in milk exosomes remained stable. In agreement with other studies, the effectiveness of our miRNA loaded exosomes has been proven *in vitro* and *in vivo*, which identified milk-derived exosomes as a biocompatible, safe, and cost-effective delivery system to enhance bioavailability and efficacy of drugs (Aqil et al., 2016; Munagala et al., 2016; Munagala et al., 2017; Betker et al., 2019; Del Pozo-Acebo et al., 2021; Luo et al., 2021).

## Conclusion

In conclusion, we developed a new strategy for accelerating diabetic wound healing by loading miR-31-5p into milk-derived exosomes. We showed here that raw milk can serve as a biocompatible and economical source for harvesting large quantities of exosomes, and that milk exosomes have tremendous potential as a miRNA delivery system, which can enhance the stability and cell uptake of miRNA. Our exosomal formulation of miR-31-5p significantly improved endothelial cell functions and enhanced the healing process of the diabetic wound by downregulating the expression of HIF1AN. Such effects of miR-31-5p exosomes shed light on its role in treating a diabetic wound, and we foresee a great potential of milk exosome utilization in nucleic acid drug delivery.

## Acknowledgments

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## Ethical approval

Ethical approval to conduct the study was obtained from the Ethical Committee on Animal Experiments of Huazhong University of Science and Technology.

## Disclosure statement

The authors declare that they have no competing interests.

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## Data availability statement

The authors confirm that the data supporting the findings of this study are available within the article and its [supplementary materials](#).

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