



In Vitro Activity of Rifampin, Rifabutin, and Rifapentine against Enterococci and Streptococci from Periprosthetic Joint Infection

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ABSTRACT After staphylococci, streptococci and enterococci are the most frequent causes of periprosthetic joint infection (PJI). MICs and minimum biofilm bactericidal concentrations of rifampin, rifabutin, and rifapentine were determined for 67 enterococcal and 59 streptococcal PJI isolates. Eighty-eight isolates had rifampin MICs of $\leq 1 \mu g/ml$, among which rifabutin and rifapentine MICs were ≤ 8 and $\leq 4 \mu g/ml$, respectively. There was low rifamycin *in vitro* antibiofilm activity except for a subset of *Streptococcus mitis* group isolates.

Microbiology Spectrum

AMERICAN SOCIETY FOR MICROBIOLOGY

IMPORTANCE Rifampin is an antibiotic with antistaphylococcal biofilm activity used in the management of staphylococcal periprosthetic joint infection with irrigation and debridement with component retention; some patients are unable to receive rifampin due to drug interactions or intolerance. We recently showed rifabutin and rifapentine to have *in vitro* activity against planktonic and biofilm states of rifampin-susceptible periprosthetic joint infection-associated staphylococci. After staphylococci, streptococci and enterococci combined are the most common causes of periprosthetic joint infection. Here, we investigated the *in vitro* antibiofilm activity of rifampin, rifabutin, and rifapentine against 126 *Streptococcus* and *Enterococcus* periprosthetic joint infection isolates. In contrast to our prior findings with staphylococcal biofilms, there was low antibiofilm activity of rifampin, rifabutin, and rifapentine against PJI-associated streptococcus mitis group isolates.

KEYWORDS rifamycin, periprosthetic joint infection, streptococci, enterococci, biofilm, rifampin, rifabutin, rifapentine

Rifampin is an antibiotic with antibiofilm activity used in the management of staphylococcal periprosthetic joint infection (PJI) with irrigation and debridement with component retention (IDCR) (1, 2); some patients are unable to receive rifampin due to drug interactions or intolerance. We recently showed that rifabutin and rifapentine, which have more favorable drug interaction/side effect profiles than rifampin, have *in vitro* activity against planktonic and biofilm states of rifampin-susceptible PJI-associated staphylococci (3), and that these rifamycins are as active as rifampin in combination therapy regimens in experimental rat *Staphylococcus aureus* foreign body osteomyelitis (4). After staphylococci, streptococci and enterococci combined are the most common causes of PJI, accounting for up to 20% of cases (5–8). Here, we investigated the *in vitro* activity of rifampin, rifabutin, and rifapentine, alongside levofloxacin, against planktonic and biofilm states of *Streptococcus* and *Enterococcus* PJI isolates.

The *in vitro* activity of rifampin, rifabutin, rifapentine, and levofloxacin against planktonic and biofilm states of 126 *Streptococcus* and *Enterococcus* PJI isolates was tested. Isolates were collected between 1996 and 2018 from separate patients with infected Quaintance KE, Osmon DR, Oravec CP, Berry DJ, Abdel MP, Patel R. 2021. *In vitro* activity of rifampin, rifabutin, and rifapentine against enterococci and streptococci from periprosthetic joint infection. Microbiol Spectr 9:e00071-21. https://doi.org/10.1128/Spectrum .00071-21. **Editor** William Lainhart, University of Arizona/

Citation Albano M, Karau MJ, Greenwood-

Editor William Lainhart, University of Arizona/ Banner Health

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Received 22 April 2021 **Accepted** 14 June 2021 **Published** 14 July 2021

arthroplasties managed at the Mayo Clinic and included 61 isolates of E. faecalis, 6 E. faecium, 23 S. agalactiae, 1 S. pyogenes, 6 S. dysgalactiae, 17 S. mitis group, 6 S. anginosus group, 4 S. salivarius group, 1 S. mutans group, and 1 S. gallolyticus. E. faecalis ATCC 29212 and S. pneumoniae ATCC 49619 were used as guality control strains. Rifampin, rifabutin, rifapentine, and levofloxacin (Sigma-Aldrich, St. Louis, MO) MICs were determined by broth microdilution by following Clinical and Laboratory Standards Institute (CLSI) guidelines (9, 10). Rifampin and levofloxacin were prepared following CLSI guidelines (10). Rifabutin and rifapentine were prepared in dimethyl sulfoxide and methanol, respectively, per the manufacturer's instructions. Current CLSI rifampin breakpoints for enterococci are $\leq 1 \ \mu g/ml$ susceptible, 2 μ g/ml intermediate, and \geq 4 μ g/ml resistant. There are no rifampin breakpoints defined by the CLSI for beta-hemolytic or viridans group streptococci. No rifabutin or rifapentine breakpoints are defined for any of the tested bacteria (10). EUCAST rifampin breakpoints for Streptococcus groups A, B, C, and G are $\leq 0.06 \ \mu$ g/ml susceptible and $>0.5 \ \mu$ g/ml resistant, and the EUCAST epidemiological cutoff (ECOFF) for viridans group streptococci is 0.125 μ g/ml (11). Levofloxacin breakpoints defined by CLSI for all organism types tested are $\leq 2 \mu g/ml$ susceptible, 4 $\mu g/ml$ intermediate, and $\geq 8 \mu g/ml$ resistant (10). Minimum biofilm bactericidal concentration (MBBC) values were determined using a pegged-lid microtiter plate assay, as previously described (3, 12).

Detailed findings for all study isolates are depicted in Table S1 in the supplemental material, which shows the aggregated MIC and MBBC values for the *E. faecalis, S. agalactiae*, and *S. mitis* group isolates. Overall, 29/61 (48%) *E. faecalis* isolates were rifampin susceptible, among which rifabutin and rifapentine MICs were ≤ 8 and $\leq 4 \mu g/ml$, respectively (Table 1). All enterococcal rifamycin MBBCs were $\geq 8 \mu g/ml$, except for *E. faecalis* IDRL-11962 (all rifamycin MBBCs, 4 $\mu g/ml$). Overall, 48/61 (79%) *E. faecalis* isolates were levofloxacin susceptible; levofloxacin MBBCs were $\geq 8 \mu g/ml$, except for *E. faecalis* IDRL-10026 (levofloxacin MBBC, 4 $\mu g/ml$) (Table 1).

All 23 *S. agalactiae* isolates tested had rifampin MICs of $\leq 0.25 \ \mu$ g/ml (among which 3 would be considered susceptible and 20 intermediate by EUCAST breakpoints), with rifabutin and rifapentine MICs of ≤ 0.25 and $\leq 1 \ \mu$ g/ml, respectively (Table 1). All 6 *S. dysgalactiae* isolates had rifampin MICs of 0.03 μ g/ml (susceptible based on EUCAST breakpoints), among which rifabutin and rifapentine MICs were 0.03 and $\leq 0.06 \ \mu$ g/ml, respectively (Table S1). *S. agalactiae* and *S. dysgalactiae* rifamycin MBBCs were all $>8 \ \mu$ g/ml (Table S1).

S. mitis group isolates had rifampin, rifabutin, and rifapentine MICs of ≤ 0.25 , ≤ 0.125 , and $\leq 0.5 \ \mu$ g/ml, respectively, except one isolate, which had MICs of 4, >8, and 4 μ g/ml, respectively; 87% of these isolates were at or below the EUCAST rifampin ECOFF (Table 1). MBBC₅₀ values for rifampin, rifabutin, and rifapentine were ≥ 8 , 1, and $\geq 8 \ \mu$ g/ml, respectively.

All six *S. anginosus* group isolates tested had rifampin MICs of $\leq 0.5 \ \mu$ g/ml, rifabutin MICs of $\leq 0.5 \ \mu$ g/ml, and rifapentine MICs of $\leq 1 \ \mu$ g/ml (Table S1). Four isolates were at or below the EUCAST rifampin ECOFF. All *S. anginosus* group rifamycin MBBCs were >8 μ g/ml, except for IDRL-12364 (rifabutin MBBC, 0.5 μ g/ml).

All but one of the streptococcal isolates tested were levofloxacin susceptible. For *S. agalactiae*, levofloxacin MBBCs were >8 μ g/ml for 18/23 isolates (Table 1). For levofloxacin-susceptible *S. dysgalactiae*, levofloxacin MBBCs were 4 μ g/ml for 2 isolates and >8 μ g/ml for 4 isolates (Table S1). For *S. anginosus* group, levofloxacin MBBCs were 2 μ g/ml for 2 isolates and ≥8 μ g/ml for 4 isolates (Table S1). For *S. mitis* group isolates, levofloxacin MBBCs were ≥1 μ g/ml (Table 1).

In contrast to our findings with staphylococcal biofilms (3), results of this study show low *in vitro* activity of rifamycins against enterococcal biofilms. The biofilm results reported here are consistent with those of other reports. Holmberg et al. studied rifampin and ciprofloxacin alone and in combination against 15 PJI *E. faecalis* isolates (13). All except one isolate was rifampin susceptible, but MBBCs (tested for four isolates) were 64 to 128 μ g/ml (13). Likewise, for ciprofloxacin, three isolates had ciprofloxacin MICs of >16 μ g/ml, with the remaining classified as susceptible; ciprofloxacin

Volume 9	Issue 1	e00071-21

		No. of i	solates (c	umulative	No. of isolates (cumulative %) with the following value (μ g/ml):	following	value (µg	/ml):						
Parameter	Drug	0.03	0.06	0.125	0.25	0.5	۲	2	4	≥8	MIC ₅₀ (<i>μ</i> g/ml)	MIC ₉₀ (<i>µ</i> g/ml)	וו (MBBC ₅₀ (אשן MB)) MBBC ₉₀ (<i>µ</i> g/ml)
<i>E. faecalis</i> MIC	Rifampin			2 (3)	3 (8)	13 (30)	11 (48)	14 (70)	11 (89)	7 (100)	2	8		
	Rifabutin			1 (2)	3 (7)	4 (13)	11 (31)	6 (41)	15 (66)	21 (100)	4	8		
	Rifapentine				1 (2)	4 (8)	6 (18)	18 (48)	20 (80)	12 (100)	4	8 //		
	Levofloxacin				3 (5)	7 (16)	27 (61)	11 (79)	1 (80)	12 (100)	1	8 /		
MBBC	Rifampin								1 (2)	60 (100)			8	8/1
	Rifabutin								1 (2)	60 (100)			8	8/1
	Rifapentine								1 (2)	60 (100)			8	8 /l
	Levofloxacin								1 (2)	60 (100)			8 /l	8 /l
S. agalactiae														
MIC	Rifampin		3 (13)	5 (35)	15 (100)						0.25	0.25		
	Rifabutin		7 (30)	10 (74)	6 (100)						0.125	0.25		
	Rifapentine			3 (13)	5 (35)	14 (96)	1 (100)				0.5	0.5		
	Levofloxacin					10 (43)	11 (91)	2 (100)			-	1		
MBBC	Rifampin									23 (100)			8	8~1
	Rifabutin									23 (100)			8 /l	8/1
	Rifapentine									23 (100)			8 /	8/1
	Levofloxacin						1 (4)		4 (22)	18 (100)			8	8
<i>S. mitis</i> group														
MIC	Rifampin	5 (31)	8 (81)	1 (87)	1 (94)				1 (100)		0.06	0.125		
	Rifabutin	3 (19)	6 (56)	6 (94)						1 (100)	0.06	0.125		
	Rifapentine		4 (25)	6 (62)	4 (87)	1 (94)			1 (100)		0.125	0.25		
	Levofloxacin					6 (37)	9 (94)	1 (100)			1	-		
MBBC	Rifampin		2 (12)	2 (24)	1 (29)				1 (35)	11 (100)			8 /l	8/1
	Rifabutin	1 (6)	3 (24)			1 (29)	5 (59)		3 (76)	4 (100)			-	8/1
	Rifapentine			2 (12)	3 (29)	1 (35)	1 (41)			10 (100)			8	8/1

MBBCs (tested for four isolates) were 256 μ g/ml (13). This is similar to our findings with levofloxacin. Holmberg et al. also reported rifampin MICs of 1 to 2 μ g/ml and MBBCs of 64 to 128 μ g/ml for three *E. faecium* PJI isolates (14). Minardi et al. reported planktonic MICs of 2 μ g/ml for *E. faecalis* ATCC 29212 and ATCC 51299, with adherent biofilm concentrations of 16 and 32 μ g/ml, respectively, using a crystal violet assay (15). They evaluated tigecycline and rifampin alone or combined for prevention of ure-teral stent infection in an experimental rat model, showing more activity of combination therapy than either drug alone (13, 15). Oliva et al. showed that rifampin alone had no activity against enterococcal biofilms, either *in vitro* or *in vivo*, but did demonstrate activity when administered as a combination therapy (16).

Data on rifampin treatment of enterococcal PJI is sparse. Thompson et al. reported a tendency toward better outcome with rifampin-combination therapy for enterococcal PJI; however, most cases were given combination therapy directed toward coinfections with staphylococci (17). Tornero et al. reviewed characteristics and outcomes of 203 patients with enterococcal PJI at 18 hospitals in 6 European countries. For those with infection within 30 days of implantation, rifampin in combination with another active antibiotic was associated with a higher remission rate than alternatives without rifampin (18).

Fiaux et al. reported results of a retrospective multicenter cohort study of 95 streptococcal PJIs from 2001 through 2009 (19). All isolates tested were rifampin susceptible. Fifty-five cases were treated with IDCR with rifampin combinations, including with levofloxacin, used in 52 and 28 cases, respectively; the overall remission rate was 71%. Antibiotic treatment regimens other than rifampin combinations were associated with worse outcome by univariate analysis (19). Rifampin combinations, including with levofloxacin, were associated with improved remission rates. Andronic et al. found no difference in failure rates with or without rifampin in a retrospective analysis of 22 streptococcal PJIs from a single institution, five of which were treated with rifampin combination regimens (20). In a study by Loubet et al. that included six S. agalactiae PJI cases, two were treated with combinations with rifampin, one with a good outcome; however, only 57% of tested S. agalactiae strains were susceptible to rifampin (21). Lora-Tamayo et al. recently published results of a retrospective, observational, multicenter, international study of 462 streptococcal PJI cases managed with IDCR, 37% of which were managed with rifampin. Failure occurred in 42% (187/444) of evaluable patients. Early use of rifampin and treatment for ≥ 21 days with a β -lactam as monotherapy or in combination with rifampin was associated with successful outcomes (22). The relevance of in vitro biofilm susceptibility testing and its relationship with clinical success with combination rifamycin therapies is incompletely defined.

This study is one of the largest evaluating the *in vitro* planktonic and biofilm activity of rifampin against PJI-associated streptococci and enterococci and, to our knowledge, the only study evaluating rifabutin and rifapentine against PJI isolates. Overall, there was low antibiofilm activity of rifamycins against PJI-associated streptococci and enterococci, with the exception of some *S. mitis* group isolates. Whether the study findings correlate with *in vivo* efficacy or *in vitro* efficacy in combination with other agents remains to be determined.

SUPPLEMENTAL MATERIAL

Supplemental material is available online only. **SUPPLEMENTAL FILE 1**, DOCX file, 0.04 MB.

ACKNOWLEDGMENTS

R.P. reports grants from ContraFect, TenNor Therapeutics Limited, Hylomorph, Paratek, BioFire, and Shionogi. R.P. is a consultant to Curetis, Specific Technologies, Next Gen Diagnostics, PathoQuest, Selux Diagnostics, 1928 Diagnostics, PhAST, and Qvella; monies are paid to Mayo Clinic. R.P. is also a consultant to Netflix. In addition, R.P. has a patent on *Bordetella pertussis*/parapertussis PCR issued, a patent on a device/ method for sonication with royalties paid by Samsung to Mayo Clinic, and a patent on an antibiofilm substance issued. R.P. receives an editor's stipend from IDSA and honoraria from the NBME, Up-to-Date, and the Infectious Diseases Board Review Course. M.P.A. received royalties from Stryker.

REFERENCES

- Zheng Z, Stewart PS. 2002. Penetration of rifampin through *Staphylococcus* epidermidis biofilms. Antimicrob Agents Chemother 46:900–903. https://doi .org/10.1128/AAC.46.3.900-903.2002.
- Zimmerli W, Sendi P. 2019. Role of rifampin against staphylococcal biofilm infections in vitro, in animal models, and in orthopedic-device-related infections. Antimicrob Agents Chemother 63:e01746-18. https://doi.org/ 10.1128/AAC.01746-18.
- Albano M, Karau MJ, Greenwood-Quaintance KE, Osmon DR, Oravec CP, Berry DJ, Abdel MP, Patel R. 2019. *In vitro* activity of rifampin, rifabutin, rifapentine, and rifaximin against planktonic and biofilm states of staphylococci isolated from periprosthetic joint infection. Antimicrob Agents Chemother 63:e00959-19. https://doi.org/10.1128/AAC.00959-19.
- Karau MJ, Schmidt-Malan SM, Albano M, Mandrekar JN, Rivera CG, Osmon DR, Oravec CP, Berry DJ, Abdel MP, Patel R. 2020. Novel use of rifabutin and rifapentine to treat methicillin-resistant *Staphylococcus aureus* in a rat model of foreign body osteomyelitis. J Infect Dis 222:1498–1504. https:// doi.org/10.1093/infdis/jiaa401.
- Aggarwal VK, Bakhshi H, Ecker NU, Parvizi J, Gehrke T, Kendoff D. 2014. Organism profile in periprosthetic joint infection: pathogens differ at two arthroplasty infection referral centers in Europe and in the United States. J Knee Surg 27:399–406. https://doi.org/10.1055/s-0033-1364102.
- Rosteius T, Jansen O, Fehmer T, Baecker H, Citak M, Schildhauer TA, Geßmann J. 2018. Evaluating the microbial pattern of periprosthetic joint infections of the hip and knee. J Med Microbiol 67:1608–1613. https://doi .org/10.1099/jmm.0.000835.
- Lam A, Rasmussen M, Thompson O. 2018. Successful outcome for patients with streptococcal prosthetic joint infections–a retrospective populationbased study. Infect Dis 50:593–600. https://doi.org/10.1080/23744235.2018 .1449961.
- Benito N, Franco M, Ribera A, Soriano A, Rodriguez-Pardo D, Sorlí L, Fresco G, Fernández-Sampedro MD, Toro MD, Guío L. 2016. Time trends in the aetiology of prosthetic joint infections: a multicentre cohort study. Clin Microbiol Infect 22:732.
- CLSI. 2018. Methods for dilution antimicrobial susceptibility tests for bacteria that grow aerobically, CLSI standard M07, 11th ed. CLSI, Wayne, PA.
- 10. CLSI. 2021. Performance standards for antimicrobial susceptibility testing, CLSI supplement 2020, 31st ed. CLSI supplement M100. CLSI, Wayne, PA.
- 11. EUCAST. 2021. Breakpoint tables for interpretation of MICs and zone diameters, version 11.0. http://www.eucast.org.
- Schmidt-Malan SM, Quaintance KEG, Karau MJ, Patel R. 2016. *In vitro* activity of tedizolid against staphylococci isolated from prosthetic joint infections. Diagn Microbiol Infect Dis 85:77–79. https://doi.org/10.1016/j.diagmicrobio .2016.01.008.
- Holmberg A, Mörgelin M, Rasmussen M. 2012. Effectiveness of ciprofloxacin or linezolid in combination with rifampicin against *Enterococcus faecalis* in biofilms. J Antimicrob Chemother 67:433–439. https://doi.org/10 .1093/jac/dkr477.
- Holmberg A, Rasmussen M. 2014. Antibiotic regimens with rifampicin for treatment of *Enterococcus faecium* in biofilms. Int J Antimicrob Agents 44:78–80. https://doi.org/10.1016/j.ijantimicag.2014.03.008.

- Minardi D, Cirioni O, Ghiselli R, Silvestri C, Mocchegiani F, Gabrielli E, d'Anzeo G, Conti A, Orlando F, Rimini M, Brescini L, Guerrieri M, Giacometti A, Muzzonigro G. 2012. Efficacy of tigecycline and rifampin alone and in combination against *Enterococcus faecalis* biofilm infection in a rat model of ureteral stent. J Surg Res 176:1–6. https://doi.org/10.1016/j.jss.2011.05.002.
- Oliva A, Tafin UF, Maiolo EM, Jeddari S, Bétrisey B, Trampuz A. 2014. Activities of fosfomycin and rifampin on planktonic and adherent *Enterococcus faecalis* strains in an experimental foreign-body infection model. Antimicrob Agents Chemother 58:1284–1293. https://doi.org/10.1128/AAC.02583-12.
- Thompson O, Rasmussen M, Stefánsdóttir A, Christensson B, Åkesson P. 2019. A population-based study on the treatment and outcome of enterococcal prosthetic joint infections. A consecutive series of 55 cases. J Bone Jt Infect 4:285–291. https://doi.org/10.7150/jbji.35683.
- Tornero E, Senneville E, Euba G, Petersdorf S, Rodriguez-Pardo D, Lakatos B, Ferrari MC, Pilares M, Bahamonde A, Trebse R, Benito N, Sorli L, del Toro MD, Baraiaetxaburu JM, Ramos A, Riera M, Jover-Sáenz A, Palomino J, Ariza J, Soriano A, European Society Group of Infections on Artificial Implants (ESGIAI). 2014. Characteristics of prosthetic joint infections due to *Enterococcus* sp. and predictors of failure: a multi-national study. Clin Microbiol Infect 20:1219–1224. https://doi.org/10.1111/1469-0691.12721.
- Fiaux E, Titecat M, Robineau O, Lora-Tamayo J, El Samad Y, Etienne M, Frebourg N, Blondiaux N, Brunschweiler B, Dujardin F, Beltrand E, Loiez C, Cattoir V, Canarelli JP, Hulet C, Valette M, Nguyen S, Caron F, Migaud H, Senneville E, on behalf of the G4 Bone and Joint Infection Study Group (G4BJIS). 2016. Outcome of patients with streptococcal prosthetic joint infections with special reference to rifampicin combinations. BMC Infect Dis 16:1–9. https://doi.org/10.1186/s12879-016-1889-0.
- Andronic O, Achermann Y, Jentzsch T, Bearth F, Schweizer A, Wieser K, Fucentese SF, Rahm S, Zinkernagel AS, Zingg PO. 2021. Factors affecting outcome in the treatment of streptococcal periprosthetic joint infections: results from a single-centre retrospective cohort study. Int Orthop 45:57–63. https://doi.org/10.1007/s00264-020-04722-7.
- Loubet P, Koumar Y, Lechiche C, Cellier N, Schuldiner S, Kouyoumdjian P, Lavigne J-P, Sotto A. 2021. Clinical features and outcome of *Streptococcus agalactiae* bone and joint infections over a 6-year period in a French university hospital. PLoS One 16:e0248231. https://doi.org/10.1371/journal .pone.0248231.
- 22. Lora-Tamayo J, Senneville É, Ribera A, Bernard L, Dupon M, Zeller V, Li HK, Arvieux C, Clauss M, Uçkay I, Vigante D, Ferry T, Iribarren JA, Peel TN, Sendi P, Miksic NG, Rodríguez-Pardo D, Del Toro MD, Fernández-Sampedro M, Dapunt U, Huotari K, Davis JS, Palomino J, Neut D, Clark BM, Gottlieb T, Trebše R, Soriano A, Bahamonde A, Guío L, Rico A, Salles MJC, Pais MJG, Benito N, Riera M, Gómez L, Aboltins CA, Esteban J, Horcajada JP, O'Connell K, Ferrari M, Skaliczki G, Juan RS, Cobo J, Sánchez-Somolinos M, Ramos A, Giannitsioti E, Jover-Sáenz A, Baraia-Etxaburu JM, Barbero JM, Group of Investigators for Streptococcal Prosthetic Joint Infection, et al. 2017. The not-so-good prognosis of streptococcal periprosthetic joint infection managed by implant retention: the results of a large multicenter study. Clin Infect Dis 64:1742–1752. https://doi.org/10.1093/cid/cix227.