




Draft Genome Sequence of a *Pseudomonas aeruginosa* NA04 Bacterium Isolated from an Entomopathogenic Nematode

Rosalba Salgado-Morales,^a Nancy Rivera-Gómez,^a
Luis Fernando Lozano-Aguirre Beltrán,^b Armando Hernández-Mendoza,^c
 Edgar Dantán-González^a

Laboratorio de Estudios Ecogenómicos, Centro de Investigación en Biotecnología, Universidad Autónoma del Estado de Morelos, Cuernavaca, Morelos, Mexico^a; Programa de Genómica Evolutiva, Centro de Ciencias Genómicas, Universidad Nacional Autónoma de México, Cuernavaca, Morelos, Mexico^b; Centro de Investigación en Dinámica Celular, Universidad Autónoma del Estado de Morelos, Cuernavaca, Morelos, Mexico^c

ABSTRACT We report the draft genome sequence of Gram-negative bacterium *Pseudomonas aeruginosa* NA04, isolated from the entomopathogenic nematode *Heterorhabditis indica* MOR03. The draft genome consists of 54 contigs, a length of 6.37 Mb, and a G+C content 66.49%.

The gammaproteobacterium *Pseudomonas aeruginosa* is Gram negative, aerobic, and highly versatile. It is distributed in different ecological niches, including water and soil, and in association with host organisms (1). The genus *Pseudomonas* is particularly interesting because of its importance as an opportunistic pathogen in humans and its biotechnological potential (2, 3).

We isolated a bacterium from the entomopathogenic nematode *Heterorhabditis indica* MOR03, recovered from soil samples cultivated with sugarcane in Yautepec, Morelos, Mexico (18°55'16.9"N and 99°02'24.0"W). A previous analysis of 16S rRNA gene sequences showed that the bacterium has a 100% sequence identity with *Pseudomonas aeruginosa*. In addition, this strain shows potential for the biological control of pests and is interesting as a model for studying the interactions between bacteria and host nematodes.

We obtained the genome sequence of *Pseudomonas aeruginosa* NA04. The strain was grown in Luria broth (LB) and incubated for 12 h at 30°C with shaking at 250 rpm. Genomic DNA was extracted with the ZR fungal/bacterial miniprep kit (Zymo Research), and 5 µg of genomic DNA was sequenced using the Illumina HiSeq platform (2 × 300 bp paired-end) approach. We obtained a total of 9,447,636 reads that were quality trimmed and error corrected using DynamicTrim (SolexaQA++) Perl script (4). Reads were assembled using SPAdes version 3.5.0 (5). The draft genome consists of 54 contigs with a total length of 6,375,895 bp. The largest contig was 891,545 bp, the N_{50} value was 757,654 bp, and L_{50} was reached in 4 contigs. The genome sequence of *P. aeruginosa* PAO1 was used as a reference. The total percentage of aligned bases in the reference is 96.08%, which was used to order the contigs in Mauve (6). Automated annotation was made with Rapid Annotations using Subsystems Technology (RAST) (7). *P. aeruginosa* NA04 has a G+C content of 66.49% and includes 5,823 coding sequences. RNAmmer and ARAGORN obtained 7 rRNAs (5S, 16S, 23S) and 67 tRNAs (8, 9). The analysis of the concatenated sequences (*gyrB*, 16S, 23S of rRNA genes) of *P. aeruginosa* NA04 shows that it is closely related to *P. aeruginosa* M18 (NC_017548), isolated from the rhizosphere soil of sweet melon (10). The genome of *P. aeruginosa* NA04 includes

Received 15 June 2017 Accepted 18 July 2017 Published 7 September 2017

Citation Salgado-Morales R, Rivera-Gómez N, Lozano-Aguirre Beltrán LF, Hernández-Mendoza A, Dantán-González E. 2017. Draft genome sequence of a *Pseudomonas aeruginosa* NA04 bacterium isolated from an entomopathogenic nematode. Genome Announc 5:e00746-17. <https://doi.org/10.1128/genomeA.00746-17>.

Copyright © 2017 Salgado-Morales et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to Edgar Dantán-González, edantan@uaem.mx.

genes that code for chitinase and a chitin binding protein. Because the exoskeletons of the insects are constituted with chitin, these could influence the insecticide activity. Also, the genome contains the gene coding for a potent toxin (exotoxin A) that affects the protein synthesis of the host cells and genes encoding the RTX-like toxins found in the entomopathogenic bacterium *Photorhabdus luminescens* (11). These genes are necessary for the production of siderophores involved in the efficient uptake of iron, an important factor for the colonization of the host (12).

Accession number(s). This whole-genome shotgun project has been deposited at DDBJ/ENA/GenBank under the accession no. [MYFK00000000](#). The version described in this paper is version MYFK01000000.

ACKNOWLEDGMENTS

We are thankful for CONAcYT scholarship 296934, national postdoctoral fellowship CONAcYT 2015–2017 (to N.R.-G.), and CONAcYT project PDCPN 0247780 (to E.D.-G.).

REFERENCES

1. Spiers AJ, Buckling A, Rainey PB. 2000. The causes of *Pseudomonas* diversity. *Microbiology* 146:2345–2350. <https://doi.org/10.1099/00221287-146-10-2345>.
2. Streeter K, Mohammad K. 2016. *Pseudomonas aeruginosa*: a review of their pathogenesis and prevalence in clinical settings and the environment. *Infect Epidemiol Med* 2:25–32. <https://doi.org/10.18869/modares.iem.2.1.25>.
3. Bano N, Musarrat J. 2003. Characterization of a new *Pseudomonas aeruginosa* strain NJ-15 as a potential biocontrol agent. *Curr Microbiol* 46:324–328. <https://doi.org/10.1007/s00284-002-3857-8>.
4. Cox MP, Peterson DA, Biggs PJ. 2010. SolexaQA: At-a-glance quality assessment of Illumina second-generation sequencing data. *BMC Bioinformatics* 11:485. <https://doi.org/10.1186/1471-2105-11-485>.
5. Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Pribelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. *J Comput Biol* 19:455–477. <https://doi.org/10.1089/cmb.2012.0021>.
6. Darling AE, Mau B, Perna NT. 2010. progressiveMauve: multiple genome alignment with gene gain, loss and rearrangement. *PLoS One* 5:e11147. <https://doi.org/10.1371/journal.pone.0011147>.
7. Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O. 2008. The RAST server: Rapid Annotations using Subsystems Technology. *BMC Genomics* 9:75. <https://doi.org/10.1186/1471-2164-9-75>.
8. Lagesen K, Hallin P, Rødland EA, Staerfeldt HH, Rognes T, Ussery DW. 2007. RNAmmer: consistent and rapid annotation of ribosomal RNA genes. *Nucleic Acids Res* 35:3100–3108. <https://doi.org/10.1093/nar/gkm160>.
9. Laslett D, Canback B. 2004. ARAGORN, a program to detect tRNA genes and tmRNA genes in nucleotide sequences. *Nucleic Acids Res* 32:11–16. <https://doi.org/10.1093/nar/gkh152>.
10. Hu HB, Xu YQ, Chen F, Zhang XH, Hur BK. 2005. Isolation and characterization of a new fluorescent *Pseudomonas* strain that produces both phenazine 1-carboxylic acid and pyoluteorin. *J Microbiol Biotechnol* 15:86–90.
11. Linhartová I, Bumba L, Mašín J, Basler M, Osička R, Kamanová J, Procházková K, Adkins I, Hejnová-Holubová J, Sadílková L, Morová J, Šebo P. 2010. RTX proteins: a highly diverse family secreted by a common mechanism. *FEMS Microbiol Rev* 34:1076–1112. <https://doi.org/10.1111/j.1574-6976.2010.00231.x>.
12. Cornelis P, Dingemans J. 2013. *Pseudomonas aeruginosa* adapts its iron uptake strategies in function of the type of infections. *Front Cell Infect Microbiol* 3:75. <https://doi.org/10.3389/fcimb.2013.00075>.