



OPEN

Larvicidal and adulticidal effects of some Egyptian oils against *Culex pipiens*

Mohamed M. Baz¹, Abdelfattah Selim²✉, Ibrahim Taha Radwan³,
Abeer Mousa Alkhaibari⁴ & Hanem F. Khater⁵

Mosquitoes and mosquito-borne diseases represent an increasing global challenge. Plant extract and/or oils could serve as alternatives to synthetic insecticides. The larvicidal effects of 32 oils (1000 ppm) were screened against the early 4th larvae of *Culex pipiens* and the best oils were evaluated against adults and analyzed by gas chromatography-mass spectrometry (GC mass) and HPLC. All oils had larvicidal activity (60.0–100%, 48 h Post-treatment, and their Lethal time 50 (LT₅₀) values ranged from 9.67 (*Thymus vulgaris*) to 37.64 h (*Sesamum indicum*). Oils were classified as a highly effective group (95–100% mortalities), including *Allium sativum*, *Anethum graveolens*, *Camellia sinensis*, *Foeniculum vulgare*, *Nigella sativa*, *Salvia officinalis*, *T. vulgaris*, and *Viola odorata*. The moderately effective group (81–92% mortalities) included *Boswellia serrata*, *Cuminum cyminum*, *Curcuma aromatic*, *Allium sativum*, *Melaleuca alternifolia*, *Piper nigrum*, and *Simmondsia chinensis*. The least effective ones were *C. sativus* and *S. indicum*. *Viola odorata*, *Anethum graveolens*, *T. vulgaris*, and *N. sativa* provide 100% adult mortalities PT with 10, 25, 20, and 25%. The mortality percentages of the adults subjected to 10% of oils (H group) were 48.89%, 88.39%, 63.94%, 51.54%, 92.96%, 44.44%, 72.22%, and 100% for *A. sativum*, *An. graveolens*, *C. sinensis*, *F. vulgare*, *N. sativa*, *S. officinalis*, *T. vulgaris*, and *V. odorata*, respectively. *Camellia sinensis* and *F. vulgare* were the most potent larvicides whereas *V. odorata*, *T. vulgaris*, *An. graveolens* and *N. sativa* were the best adulticides and they could be used for integrated mosquito control.

Mosquitoes are an ancient nuisance pest and mosquito-borne diseases represent an increasing global health challenge, threatening over 40% of the world's population and it is expected that almost half of the world's population will be at risk of arbovirus transmission by 2050¹. *Culex pipiens* (Diptera: *Culicidae*) is widely distributed, transmitting dreadful diseases leading to severe morbidity and sometimes mortality to humans and animals^{2–5}.

Vector control is the primary method for reducing public concerns about mosquito-borne diseases^{6–11}. Controlling adults and larvae through repellents and insecticides^{12,13}, are the most effective approach for reducing mosquito bites. Using synthetic insecticides led to insecticide resistance, environmental pollution, and health hazards to human health and non-target organisms.

Searching for eco-friendly alternatives in botanicals such as essential oils (EOs) is a curtail need. EOs are volatile components found in many plant families like Asteraceae, Rutaceae, Myrtaceae, Lauraceae, Lamiaceae, Apiaceae, Piperaceae, Poaceae, Zingiberaceae, and Cupressaceae¹⁴. EOs contain complicated mixtures of products as phenols, sesquiterpenes, and monoterpenes¹⁵.

EOs have antibacterial, antiviral, and antifungal activities. They also possess insecticidal effect interfering with insects' physiological, metabolic, behavioral, and biochemical functions through inhalation, ingestion, or skin absorption of EOs inducing a neurotoxic action¹⁶. EOs act as adulticides, larvicides, deterrents, and repellents. They are less toxic, biodegradable, and overcome insecticidal resistance^{15,17,18}.

EOs have higher popularity with organic growers and environmentally conscious consumers and suitability for urban areas, homes, and other sensitive areas.

¹Department of Entomology, Faculty of Science, Benha University, Benha 13518, Egypt. ²Department of Animal Medicine (Infectious Diseases), Faculty of Veterinary Medicine, Benha University, Toukh 13736, Egypt. ³Supplementary General Sciences Department, Faculty of Oral and Dental Medicine, Future University in Egypt, P.O. Box 11835, Cairo, Egypt. ⁴Department of Biology, Faculty of Science, University of Tabuk, Tabuk 71491, Kingdom of Saudi Arabia. ⁵Department of Parasitology, Faculty of Veterinary Medicine, Benha University, Toukh 13736, Egypt. ✉email: Abdelfattah.selim@fvmtm.bu.edu.eg

No.	Oil name	Plant oils		
		Order	Family	English name
1	<i>Allium sativum</i> ^a	Asparagales	Amaryllidaceae	Garlic
2	<i>Anethum graveolens</i> ^a	Apiales	Apiaceae	Dill
3	<i>Argania spinosa</i> ^b	Ericales	Sapotaceae	Argan
4	<i>Boswellia serrata</i> R. ^a	Sapindales	Burseraceae	Olibanum
5	<i>Brassica carinata</i> ^a	Brassicales	Brassicaceae	Mustard
6	<i>Camellia sinensis</i> ^a	Ericales	Theaceae	Green Tea
7	<i>Cedrus libani</i> A. ^a	Pinales	Pinaceae	Cedar wood
8	<i>Citrullus colocynthis</i> L. ^b	Cucurbitales	Cucurbitaceae	Bitter apple
9	<i>Crocus sativus</i> L. ^a	Asparagales	Iridaceae	Saffron crocus
10	<i>Cucurbita maxima</i> D. ^a	Cucurbitales	Cucurbitaceae	Pumpkin
11	<i>Cuminum cyminum</i> L. ^a	Apiales	Apiaceae	Cumin
12	<i>Cupressus sempervirens</i> ^b	Pinales	Cupressaceae	Italian cypress
13	<i>Curcuma aromatica</i> S. ^a	Zingiberales	Zingiberaceae	Curcuma
14	<i>Curcuma longa</i> L. ^a	Zingiberales	Zingiberaceae	Common turmeric
15	<i>Foeniculum vulgare</i> M. ^a	Apiales	Apiaceae	Sweet fennel
16	<i>Gadus morhua</i> ^a	Gadiformes	Gadidae	Cod Liver
17	<i>Lepidium sativum</i> L. ^a	Brassicales	Brassicaceae	Garden pepperwort
18	<i>Linum usitatissimum</i> L. ^a	Malpighiales	Linaceae	Common flax
19	<i>Melaleuca alternifolia</i> ^a	Myrtales	Myrtaceae	Tea tree
20	<i>Nigella sativa</i> ^a	Ranunculales	Ranunculaceae	Black cumin
21	<i>Panax ginseng</i> ^a	Apiales	Araliaceae	Chinese ginseng
22	<i>Piper nigrum</i> L. ^a	Piperales	Piperaceae	Black pepper
23	<i>Prunus dulcis</i> ^b	Rosales	Rosaceae	Almond
24	<i>Ruta chalepensis</i> L. ^a	Sapindales	Rutaceae	Rues
25	<i>Salvia officinalis</i> L. ^a	Lamiales	Lamiaceae	Sage
26	<i>Sesamum indicum</i> ^a	Lamiales	Pedaliaceae	Sesame
27	<i>Simmondsia chinensis</i> ^b	Caryophyllales	Simmondsiaceae	Joboba
28	<i>Syzygium aromaticum</i> L.	Myrtales	Myrtaceae	Clove
29	<i>Tilia americana</i> L. ^a	Malvales	Malvales	Tilia
30	<i>Thymus vulgaris</i> L.	Lamiales	Lamiaceae	Garden
31	<i>Viola odorata</i> L. ^a	Malpighiales	Violaceae	Sweet violet
32	<i>Zingiber officinale</i> ^a	Zingiberales	Zingiberaceae	Ginger

Table 1. Plants species screened (oil No = 32) used for larvicidal activity. ^aPlant oils purchased from EL CAPTAIN company for extracting natural oils, plants and cosmetics “Cap Pharm”. ^bPlant oils purchased from Harraz for Food Industry & Natural products.

The role of EOs in mosquito control has been discussed^{15,19}. This study aimed to screen and evaluate the lethal time values of the larvicidal effects of thirty-two oils and evaluate the adulticidal effect and phytochemical analyses of the most effective ones against *Cx. pipiens*.

Materials and methods

Plant oils. Thirty- two oils were purchased from EL CAPTAIN Company for extracting natural oils, plants, and cosmetics “Cap Pharm,” El Obor, Cairo, Egypt and Harraz for Food Industry & Natrual products, Cairo, Egypt (Table 1).

Culex pipiens. *Culex pipiens* (anautozenous strain) was provided from the colony reared at the Department of Entomology, Faculty of Science, Benha University, Egypt, and maintained at 27 ± 2 °C, 75–85% RH and 14: 10 h (L/D) photoperiod.

Larvicidal efficacy. Thirty-two oils were screened for their larvicidal efficacy²⁰ against the early fourth instar larvae, *Cx. pipiens*. Oils were added to a solvent (emulsifier) consisting of dechlorinated water plus 1.0 mL 0.5% Tween-20, through a shaker plate to yield a homogenous solution. Oils were added to a solvent consisting of dechlorinated water plus 5% tween 20. For each oil, twenty larvae were placed in a 500 mL glass beaker containing 250 mL of 1000 ppm. The experiment and the control group, treated with the solvent only, were replicated three times. Larval mortalities were recorded 0.5, 2, 8, 24, and 48 h post-treatment (PT).

Adulticidal efficacy. Susceptibility tests for adult mosquitoes were performed for the promising larvicidal oils through the CDC bottle bioassays²¹ with modifications. For each concentration, three bottles were coated. Several concentrations for each oil were prepared using pure ethanol as a solvent. The bottles were coated with the desired concentrations and left overnight at 27 ± 2 °C for solvent evaporation.

Adult mosquitoes (15–10, aged 3–4 days) fed on 10% sucrose solution were released to each bottle using a hand aspirator. The exposure time was set to 30 min. The mosquitoes were removed from the bottles. Mosquito groups were added to separate transparent paper cups (10 × 9 × 6 cm) having 10% sucrose solution and mortalities were checked after 24 h. Three replicates were made for each concentration.

GC/MS analysis. A Thermo Scientific Trace GC Ultra/ISQ Single Quadrupole MS, TG-5MS fused silica capillary column was used for the GC/MS study (0.1 mm, 0.251 mm and 30 m film thickness). An electron ionisation device with a 70 eV ionisation energy was employed for GC/MS detection. At a constant flow rate of 1 mL/min, helium gas was used as the carrier gas. Temperatures were established at 280 °C for the injector and MS transfer line. The oven temperature was set at 50 °C (hold for 2 min), then increased to 150 °C at a rate of 7 °C per minute, then to 270 °C at a rate of 5 °C per minute (hold for 2 min), and finally to 310 °C at a rate of 3.5 °C per minute (hold 10 min). A percent relative peak area was used to explore the quantification of all of the discovered components. The chemicals were tentatively identified by comparing their respective retention times and mass spectra to those of the NIST, WILLY library data from the GC/MS instrument. The identification was done using mass spectra and a computer search of user-generated reference libraries. To check peak homogeneity, single-ion chromatographic reconstruction was used. When identical spectra could not be identified, only the structural type of the relevant component was provided based on its mass spectral fragmentation. When possible, reference compounds were co-chromatographed to confirm GC retention durations²².

Data analysis. Data were analyzed through one-way analysis of variance (ANOVA), Duncan's multiple range tests, and Probit analysis for calculating the lethal concentration (LC) and lethal time (LT) values using the computer program PASW Statistics 2009 (SPSS version 22). The relative efficacies (RE) were calculated¹⁸ according to the following formula:

$$RE \text{ for LC} = LC_{50}(LC_{90} \text{ or } LC_{99}) \text{ for reference oil} / LC_{50}(LC_{90} \text{ or } LC_{99}) \text{ for EO.}$$

$$RE \text{ for LT} = LT_{50}(LT_{90} \text{ or } LT_{99}) \text{ for reference oil} / LT_{50}(LT_{90} \text{ or } LT_{99}) \text{ for EO.}$$

Non-parametric, Kruskal–Wallis test was performed to compare the mean differences of more than two groups followed by the Mann–Whitney test to compare the mean differences between the effective oil groups.

Results

The larvicidal effect of 32 oils was screened against the early 4th larvae, *Cx. pipiens*. The results showed that all plant oils had larvicidal activity (60.0–100%, 48 h PT) and their Lethal time 50 (LT₅₀) values ranged from 9.67 (*Thymus vulgaris*) to 37.64 h (*Sesamum indicum*), Tables 2 and 3.

The efficacy of oils could be classified, 48 h post-treatment (PT) as the highly effective group (H group) inducing 95–100% mortalities, including eight oils: *Allium sativum*, *Anethum graveolens*, *Camellia sinensis*, *Foeniculum vulgare*, *Nigella sativa*, *Salvia officinalis*, *T. vulgaris*, and *Viola odorata*. *Camellia sinensis* and *F. vulgare* provided 100%, 24 h PT (Table 2).

The LT₅₀ values of the H group ranged from 9.67 (*T. vulgaris*) to 19.91 (*An. graveolens*) hours and those of LT₉₉ values ranged from 29.97 (*Foeniculum vulgare*) to 55.32 (*An. graveolens*). The relative effects (RE) of such oils according to LT₅₀ values were 2.7, 1.9, 2.9, 3.7, 2.4, 2.4, 3.9, and 3.6 times, respectively, times than *S. indicum*; whereas those of LT₉₉ values were 2.1, 1.8, 2.4, 3.3, 2.0, 2.0, 3.0, and 3.0 times, respectively, than *C. sativus*. The Chi-square, significance, and regression equations were provided for all teste oils (Table 3).

The moderately effective (M group) group of oils resulted in 81–92% mortalities 48 h PT, including *B. serrata*, *C. cyminum*, *C. aromaticum*, *L. sativum*, *M. alternifolia*, *P. nigrum*, and *S. chinensis*. They provided 63.33–71.67% mortalities, 24 h PT (Table 2).

The LT₅₀ values of M group ranged from 19.00 (*S. chinensis*) to 22.65 (*C. cyminum*) hours and those of LT₉₉ values ranged from 57.95 (*S. chinensis*) to 66.22 (*M. alternifolia*) (Table 3). Their RE regarding the LT₅₀ values were 1.8, 1.7, 1.8, 1.9, 1.7, 1.9, and 1.9 times than *S. indicum*, respectively, whereas those of LT₉₉ values were 1.7, 1.6, 1.6, 1.7, 1.5, 1.6, and 1.8 times than *C. sativus*, respectively (Table 3).

The least effective group (L group) included the other 17 oils, and the least effective ones were *C. sativus*, and *S. indicum*, providing 62.33 and 60.00% mortalities, 48 h PT, whereas their LT₅₀ values were 37.07 and 37.64 h and their LT₉₉ values were 96.88 and 92.89 h, respectively (Table 3).

Furthermore, the Kruskal–Wallis test was performed to compare the mean differences of more than two groups, followed by the Mann–Whitney test to compare the mean differences between groups. Whereas Kruskal–Wallis and Friedman's tests showed there are significant indications between the three groups at different times ($P=0.001$) (Tables 4 and 5).

Viola odorata, *A. graveolens*, *T. vulgaris*, and *N. sativa* provide 100% adult mortalities PT with 10, 25, 20, and 25%. The mortality percentages of the adults subjected to 10% of oils (H group) were 48.89%, 88.39, 63.94, 51.54, 92.96, 44.44, 72.22, and 100.0% for *A. sativum*, *An. graveolens*, *C. sinensis*, *F. vulgare*, *N. sativa*, *S. officinalis*, *T. vulgaris*, and *V. odorata*, respectively. Their adulticidal LC₅₀ values, 24 h PT, were 15.57, 2.42, 9.01, 15.07, 3.42, 20.46, 3.08, and 1.88%; whereas their LC₉₀ values were 38.86, 9.47, 32.18, 33.34, 5.44, 50.76, 16.08, and 7.37%, respectively. *Salvia officinalis* followed by *A. sativum* were the least effective oils against adults. According to LC₉₀, *N. sativa*, *V. odorata* and *An. graveolens* killed mosquitoes 9.3, 6.9, and 5.4 times more than *S. officinalis* (Table 6).

Oils	Mortality % (mean ± SD)/h					Grouping
	0.5	2	8	24	48	
<i>Allium sativum</i>	6.67 ± 0.58 ^{abE}	22.33 ± 1.53 ^D	46.67 ± 0.58 ^{efgC}	81.33 ± 1.53 ^{dB}	96.67 ± 0.58 ^{EA}	H
<i>Anethum graveolens</i>	8.33 ± 0.58 ^{abE}	23.33 ± 1.15 ^D	48.67 ± 1.15 ^{JC}	83.67 ± 1.53 ^{dB}	98.33 ± 0.58 ^{EA}	H
<i>Argania spinosa</i>	5.00 ± 1.00 ^{abE}	11.67 ± 0.58 ^D	21.67 ± 1.53 ^{bcdC}	43.33 ± 1.53 ^{EB}	66.67 ± 1.53 ^{dA}	L
<i>Boswellia serrata</i>	3.33 ± 0.58 ^{abE}	15.00 ± 1.00 ^D	31.67 ± 1.53 ^{bcdC}	70.00 ± 1.00 ^{dB}	90.00 ± 1.00 ^{EA}	M
<i>Brassica carinata</i>	3.33 ± 0.58 ^{abE}	13.33 ± 0.58 ^D	25.00 ± 1.00 ^{bcdC}	45.00 ± 1.53 ^{EB}	68.33 ± 2.08 ^{dA}	L
<i>Camellia sinensis</i>	8.33 ± 0.58 ^{abE}	23.33 ± 1.00 ^{EC}	61.67 ± 1.53 ^{1B}	100.00 ± 1.00 ^{dA}	100.00 ± 0.58 ^{EA}	H
<i>Cedrus libani</i>	5.00 ± 1.00 ^{abE}	15.00 ± 0.00 ^{ED}	25.00 ± 1.00 ^{EC}	56.67 ± 1.00 ^{dB}	78.33 ± 1.53 ^{EA}	L
<i>Citrullus colocynthis</i>	3.33 ± 0.58 ^{abE}	11.67 ± 0.58 ^{deD}	33.33 ± 0.58 ^{defC}	65.00 ± 1.00 ^{dfB}	75.00 ± 1.00 ^{deA}	L
<i>Crocus sativus</i>	3.33 ± 0.58 ^{abE}	10.00 ± 1.00 ^{defD}	21.67 ± 1.15 ^{hijC}	39.33 ± 1.00 ^{hiB}	62.33 ± 1.00 ^{fgA}	L
<i>Cucurbita maxima</i>	3.33 ± 0.58 ^{abE}	10.00 ± 1.00 ^{defD}	21.67 ± 1.53 ^{hijC}	48.33 ± 1.53 ^{ghB}	65.00 ± 1.35 ^{efgA}	L
<i>Cuminum cyminum</i>	3.33 ± 0.58 ^{abE}	8.33 ± 0.58 ^{ED}	33.33 ± 1.53 ^{defC}	63.33 ± 1.53 ^{dfB}	88.33 ± 1.53 ^{bca}	M
<i>Cupressus sempervirens</i>	5.00 ± 1.00 ^{abE}	8.33 ± 0.58 ^{ED}	16.67 ± 0.58 ^{HC}	41.67 ± 2.08 ^{hiB}	63.33 ± 2.00 ^{fgA}	L
<i>Curcuma aromatic</i>	5.00 ± 1.00 ^{abE}	16.67 ± 1.53 ^{abcdeD}	35.00 ± 1.73 ^{defC}	71.67 ± 1.53 ^{dB}	88.33 ± 1.53 ^{bca}	M
<i>Curcuma longa</i>	5.00 ± 1.00 ^{abE}	10.00 ± 1.00 ^{defD}	20.00 ± 1.00 ^{HC}	40.00 ± 2.08 ^{hiB}	61.67 ± 1.53 ^{fgA}	L
<i>Foeniculum vulgare</i>	8.33 ± 0.58 ^{abE}	25.00 ± 1.15 ^{EC}	63.33 ± 0.58 ^{AB}	100.00 ± 1.00 ^{EA}	100.00 ± 0.00 ^{AA}	H
<i>Gadus morhua</i>	5.00 ± 1.00 ^{abE}	13.33 ± 0.58 ^{bcdE}	31.67 ± 1.53 ^{defghC}	55.00 ± 1.00 ^{fgB}	75.00 ± 1.00 ^{deA}	L
<i>Lepidium sativum</i>	6.67 ± 0.58 ^{abE}	15.00 ± 1.00 ^{abcdeD}	36.67 ± 1.15 ^{deC}	70.00 ± 1.00 ^{cdeB}	90.00 ± 1.00 ^{abcA}	M
<i>Linum usitatissimum</i>	3.33 ± 0.58 ^{abE}	15.00 ± 1.00 ^{abcdeD}	40.00 ± 1.00 ^{cdC}	55.00 ± 1.00 ^{fgB}	75.00 ± 1.00 ^{deA}	L
<i>Melaleuca alternifolia</i>	6.67 ± 0.58 ^{abE}	10.00 ± 1.00 ^{defD}	40.00 ± 1.00 ^{cdC}	71.67 ± 1.53 ^{dB}	81.67 ± 0.58 ^{cdA}	M
<i>Nigella sativa</i>	5.00 ± 1.00 ^{abE}	20.00 ± 1.00 ^{abcdD}	50.00 ± 1.00 ^{bcC}	78.67 ± 1.53 ^{bcB}	95.00 ± 1.00 ^{abA}	H
<i>Panax ginseng</i>	5.00 ± 1.00 ^{abE}	11.67 ± 0.58 ^{deD}	30.00 ± 1.73 ^{defghC}	48.33 ± 1.53 ^{ghB}	71.67 ± 1.15 ^{defA}	L
<i>Piper nigrum</i>	5.00 ± 1.00 ^{abE}	20.00 ± 1.00 ^{abcdD}	38.33 ± 0.58 ^{dC}	70.00 ± 1.00 ^{cdeB}	88.33 ± 1.58 ^{bca}	M
<i>Prunus dulcis</i>	3.33 ± 0.57 ^{abE}	13.33 ± 0.33 ^{bcdE}	31.67 ± 0.88 ^{defghC}	50.00 ± 0.57 ^{ghB}	75.00 ± 0.57 ^{deA}	L
<i>Ruta chalepensis</i>	3.33 ± 0.58 ^{abE}	15.00 ± 1.00 ^{abcdeD}	33.33 ± 2.08 ^{defC}	60.00 ± 2.00 ^{fb}	80.00 ± 1.00 ^{cdA}	L
<i>Salvia officinalis</i>	6.67 ± 0.58 ^{abE}	21.67 ± 1.53 ^{abcd}	51.67 ± 1.53 ^{bc}	80.00 ± 1.53 ^{cbB}	97.33 ± 1.00 ^{abA}	H
<i>Sesamum indicum</i>	3.33 ± 0.58 ^{abE}	8.33 ± 1.15 ^{ED}	15.00 ± 1.00 ^{EC}	36.67 ± 1.15 ^{EB}	60.00 ± 1.15 ^{gA}	L
<i>Simmondsia chinensis</i>	5.00 ± 1.00 ^{abE}	11.67 ± 0.58 ^{deD}	36.67 ± 1.53 ^{deC}	70.00 ± 2.00 ^{cdeB}	91.67 ± 0.58 ^{abA}	M
<i>Syzygium aromaticum</i>	5.00 ± 1.00 ^{abE}	13.33 ± 0.58 ^{bcdE}	23.33 ± 1.15 ^{ghijC}	50.00 ± 1.00 ^{ghB}	76.67 ± 1.53 ^{dA}	L
<i>Tilia americana</i>	5.00 ± 0.57 ^{abE}	15.00 ± 0.00 ^{abcdeD}	25.00 ± 0.57 ^{ghijC}	56.67 ± 0.88 ^{fgB}	88.33 ± 0.88 ^{bca}	L
<i>Thymus vulgaris</i>	8.33 ± 0.58 ^{abE}	21.67 ± 0.58 ^{abcd}	58.33 ± 2.08 ^{abc}	85.00 ± 0.58 ^{bb}	100.00 ± 1.00 ^{abA}	H
<i>Viola odorata</i>	8.33 ± 0.58 ^{abE}	23.33 ± 1.00 ^{abd}	58.67 ± 1.53 ^{abc}	89.67 ± 1.53 ^{abB}	100.00 ± 0.00 ^{abA}	H
<i>Zingiber officinale</i>	5.00 ± 1.00 ^{abE}	13.33 ± 0.58 ^{bcdE}	26.67 ± 1.53 ^{efghiC}	48.33 ± 1.53 ^{ghB}	75.00 ± 1.00 ^{deA}	L
Control	0.33 ± 0.33 ^{AA}	0.33 ± 0.33 ^{IA}	0.33 ± 0.33 ^{kA}	0.33 ± 0.33 ^{IA}	0.33 ± 0.33 ^{hA}	L

Table 2. Larval mortality (%) of plant oils used at 1000 ppm through different time periods. Numbers of the same row followed by the same small letter are not significantly different (one-way ANOVA, Duncan's MRT, $P > 0.05$). H: The highly effective (95–100% mortalities), 8 oils. M: The moderately effective group (81–92% mortalities), 7 oils. L.: The moderately effective group, include the rest of oils, 17 oils.

Oil phytochemical analysis. Phytochemical analysis of oils of *F. vulgare* Mill., *An. graveolens* L., *V. odorata* L., *T. vulgaris* L., *A. sativum*, *S. officinalis* and *C. sinensis* by GC/MS and HPLC analysis revealed their major compounds. *F. vulgare* oil contains Estragole (70.36%); Limonene (8.96%) and 1,3,3-trimethyl Bicyclo [2.2.1] heptan-2-one (2.81%) (Table 7 and Fig. 1).

Anethum graveolens showed abundance of 4-Pyridinecarbaldehyde-4-propyl-3-thiosemicarbazone (32.13%); 1,5-dimethyl-1,5-Cyclooctadiene (17.19%); Dihydrocarvone (5.98%); 3a(1H)-Azulenol,2,3,4,5,8a-hexahydro-6,8-adimethyl-3-(1-methylethyl),[3R-(3à,3aà,8aà)] (Carotol) (21.26%); and tricyclic compound Daucol (2.39%) (Table 8 and Fig. 2).

Viola odorata L. oil contains Diphenyl ether (42.04%); alpha.-Ionone(11.87%); (Z)-5-(4-tert-Butyl-1-hydroxycyclohexyl)-3-methylpent-2-en-4-yne (7.22%); 2,3,3a,4,5,5a,6,7,9a,9b-decahydro-3,5a,9-trimethyl-7,9a-peroxy Naphtho-[1,2-b]furan-2-one (6.6%); 2-hexyl-1-Decanol (4.15%); and hexadecahydro-Pyrene (2.79%) (Table 9 and Fig. 3).

Thymus vulgaris oil included 2-Ethynyl-3-hydroxypyridine (12.37%); 2-à-pinene(8.92%),2,5-Dipropoxybenzaldehyde (7.70%); 5-Amino-8-cyano-7-methoxy-3,4-dihydro-3-methyl-1,6-naphthyridin- (1H)-one (5.05%); à-terpinyl acetate (5.00%); 4-methyl-1-(1-methyl-ethyl)-3-Cyclohexen-1-ol (4.73%), 3-(6,6-Dimethyl-5-oxohept-2-enyl)-cyclo-heptanone (4.54%); 10-Methylnonadecane(4.12%); 9-methyl Nonadecane-(3.55%); n1,1'-oxybis Decane (2.36%); 7,11-Hexadecadienal (2.14%); and (2R,3R)-3-(2-Methoxy-4-methylphenyl)-2,3-dimethylcyclopentanone (2.01%) (Table 10 and Fig. 4).

Allium sativum contains many effective chemical compounds including the 9-Octadecenamide, (Z)-(29.07%), Trisulfide, di-2-propenyl (14.86%), and isochiapin B%2 < (8.63%) compounds (Table 11 and Fig. 5).

Oil name	LT ₅₀ (lower-upper)	RE (LT ₅₀)	LT ₉₀ (lower-upper)	RE (LT ₉₀)	LT ₉₉ (lower-upper)	RE (LT ₉₉)	Chi (Sig)	Regrision equation
<i>Allium sativum</i>	13.95 (3.16–54.44)	2.7	31.17 (18.49–174.49)	2.2	45.20 (26.92–276.44)	2.1	39.30 (0.000a)	y = 0.86 + 0.06*x
<i>Anethum graveolens</i>	19.90 (11.30–36.52)	1.9	39.41 (27.22–81.32)	1.8	55.31 (37.96–120.10)	1.8	23.13 (0.000a)	y = 1.23 + 0.06*x
<i>Argania spinosa</i>	33.02 (22.75–55.92)	1.1	63.55 (45.59–120.49)	1.1	88.45 (62.33–175.00)	1.1	13.91 (0.008a)	y = 1.31 + 0.04*x
<i>Boswellia serrata</i>	20.78 (12.05–37.26)	1.8	41.01 (28.56–82.20)	1.7	57.50 (39.77–121.10)	1.7	22.42 (0.000a)	y = 1.27 + 0.06*x
<i>Brassica carinata</i>	32.09 (21.04–59.25)	1.2	62.39 (43.53–132.05)	1.1	87.09 (59.69–193.58)	1.1	17.05 (0.002a)	y = 1.33 + 0.04*x
<i>Camellia sinensis</i>	13.02 (3.56–56.12)	2.9	27.65 (16.38–172.03)	2.5	39.58 (23.51–269.84)	2.4	40.31 (0.000a)	y = 0.96 + 0.07*x
<i>Cedrus libani A</i>	26.87 (17.55–44.77)	1.4	52.99 (38.06–98.01)	1.3	74.29 (52.64–143.56)	1.3	16.60 (0.002a)	y = 1.24 + 0.05*x
<i>Citrullus colocynthis</i>	26.08 (12.80–65.61)	0.0	52.72 (34.03–169.10)	0.0	74.44 (47.49–257.33)	1.3	32.23 (0.000a)	y = 1.25 + 0.05*x
<i>Crocus sativus</i>	37.07 (25.39–68.56)	1.0	70.02 (49.05–147.56)	1.0	96.88 (66.53–213.77)	1.0	14.35 (0.006a)	y = 1.41 + 0.04*x
<i>Cucurbita maxima</i>	30.90 (22.00–47.60)	1.2	57.85 (43.01–97.25)	1.2	79.81 (58.44–139.44)	1.2	12.91 (0.012a)	y = 1.44 + 0.05*x
<i>Cuminum cyminum</i>	22.65 (13.54–140.07)	1.7	43.44 (30.47–86.24)	1.6	60.39 (42.00–126.16)	1.6	22.68 (0.000a)	y = 1.39 + 0.06*x
<i>Cupressus sempervirens</i>	34.67 (26.87–47.96)	1.1	67.29 (52.45–100.54)	1.0	93.88 (71.85–144.86)	1.0	18.16 (0.66a)	y = 1.41 + 0.05*x
<i>Curcuma aromatic</i>	20.49 (10.77–39.97)	1.8	41.98 (28.40–94.24)	1.7	59.51 (40.00–141.25)	1.6	25.53 (0.000a)	y = 1.14 + 0.05*x
<i>Curcuma longa</i>	33.89 (24.46–52.94)	1.1	63.92 (47.28–109.44)	1.1	88.41 (64.29–157.09)	1.1	11.35 (0.023a)	y = 1.37 + 0.04*x
<i>Foeniculum vulgare</i>	10.22 (5.29–21.14)	3.7	20.99 (13.93–49.73)	3.3	29.77 (19.68–74.34)	3.3	21.56 (0.000a)	y = 1.06 = 0.1*x
<i>Gadus morhua</i>	27.64 (16.47–54.29)	1.4	55.69 (37.98–128.11)	1.3	78.56 (52.78–191.03)	1.2	21.54 (0.000a)	y = 1.2 + 0.04*x
<i>Lepidium sativum</i>	20.06 (11.18–36.90)	1.9	41.06 (28.31–84.97)	1.7	58.18 (39.83–126.60)	1.7	22.42 (0.000a)	y = 1.11 + 0.05*x
<i>Linum usitatissimum</i>	26.78 (12.80–77.92)	1.4	55.74 (35.22–213.81)	1.3	79.35 (49.44–328.66)	1.2	31.75 (0.000a)	y = 1.18 + 0.04*x
<i>Melaleuca alternifolia</i>	22.36 (9.11–58.90)	1.7	46.52 (29.47–159.02)	1.5	66.22 (41.73–244.98)	1.5	36.44 (0.000a)	y = 1.12 + 0.05*x
<i>Nigella sativa</i>	15.67 (5.25–46.57)	2.4	33.48 (20.57–130.64)	2.1	48.00 (29.54–202.69)	2.0	36.89 (0.000a)	y = 1.01 + 0.06*x
<i>Panax ginseng</i>	30.16 (19.05–57.39)	1.2	59.66 (41.18–131.40)	1.2	83.70 (56.80–194.15)	1.2	18.86 (0.001a)	y = 1.25 + 0.04*x
<i>Piper nigrum</i>	20.14 (9.84–41.84)	1.9	42.45 (28.17–103.75)	1.6	60.63 (40.01–157.34)	1.6	27.10 (0.000a)	y = 1.07 + 0.05*x
<i>Prunus dulcis</i>	26.75 (19.88–36.78)	2.6	58.25 (45.50–85.63)	1.4	78.56 (64.49–127.36)	1.2	21.11(0.03a)	y = 1.2 + 0.04*x
<i>Ruta chalepensis</i>	25.12 (14.06–50.27)	1.5	50.74 (34.32–119.52)	1.4	71.63 (47.88–178.94)	1.4	24.68 (0.000a)	y = 1.24 + 0.05
<i>Salvia officinalis</i>	15.42 (5.38–41.36)	2.4	34.12 (21.26–116.53)	2.1	49.37 (30.77–181.26)	2.0	32.84 (0.000a)	y = 0.89 + 0.06*x
<i>Sesamum indicum</i>	37.64 (32.87–44.04)	1.0	68.08 (58.97–81.70)	1.0	92.89 (79.68–112.98)	1.0	8.60 (0.720a)	y = 1.54 + 0.04*x
<i>Simmondsia chinensis</i>	19.00 (14.03–25.19)	1.9	40.45 (32.52–55.17)	1.8	57.95 (46.08–81.12)	1.8	4.20 (0.241a)	y = 1.23 + 0.06*x
<i>Syzygium aromaticum</i>	32.14 (21.00–44.84)	1.2	63.13 (43.91–102.50)	1.1	88.39 (60.37–19.40)	1.1	16.81 (0.031a)	y = 1.26 + 0.04*x
<i>Tilia americana</i>	26.03 (19.61–35.05)	1.4	52 (43.55–78.29)	1.3	78.62 (61.30–115.31)	1.2	16.6 (0.471a)	y = 1.24 + 0.05*x
<i>Thymus vulgaris</i>	9.67 (3.58–33.79)	3.9	21.89 (13.29–104.01)	3.2	31.86 (19.19–163.28)	3.0	33.04 (0.000a)	y = 0.88 + 0.09*x
<i>Viola odorata</i>	10.31 (3.88–28.58)	3.6	22.15 (13.76–78.00)	3.2	31.81 (19.76–120.35)	3.0	29.95 (0.000a)	y = .96 + 0.09*x
<i>Zingiber officinale</i>	29.27 (19.73–48.49)	1.3	57.30(41.31–105.43)	1.2	80.16 (56.91–153.86)	1.2	14.90 (0.005a)	y = 1.26 + 0.04*x
Reference oil	<i>Sesamum indicum</i>		<i>Crocus sativus</i>					

Table 3. Lethal time values of applied oils (1000 ppm) against *Culex pipiens* larvae. RE Relative efficacy. Significant values are in [bold].

Oil groups	Mortality % (mean ± SD)*				
	0.5 h	2 h	8 h	24 h	48 h
Low	4.2 ± 0.847	12.3 ± 2.278	25.980 ± 6.590	49.4 ± 7.838	71.6 ± 7.39
Medium	5.0 ± 1.361	13.8 ± 4.050	35.950 ± 2.864	69.5 ± 2.841	88.3 ± 3.191
High	7.5 ± 1.260	22.7 ± 1.527	54.792 ± 6.389	87.1 ± 8.533	98.3 ± 1.992
Chi-Square	16.909**	18.152**	23.037**	25.391**	25.098**
df	2	2	2	2	2
Asymp. Sig	0.001	0.001	0.001	0.001	0.001

Table 4. Kruskal–Wallis test for larval mosquito mortality (%) of plant oil groups at 1000 ppm. *Means produced by non-parametric analysis (Kruskal–Wallis, $p < 0.05$). **The X^2 value is sig. at significant level 1% H: The highly effective group (95–100% mortalities) are 8 oils (*A. sativum*, *A. graveolens*, *C. sinensis*, *F. vulgare*, *N. sativa*, *S. officinalis*, *T. vulgaris*, and *V. odorata*). M: The moderately effective group (81–92% mortalities) are 7 oils (*B. serrata*, *C. cuminum*, *C. aromaticum*, *L. sativum*, *M. alternifolia*, *P. nigrum*, and *S. chinensis*). L.: The moderately effective group are included the rest of oils, 17 oils (*A. spinosa*, *B. carinata*, *C. libani*, *C. colocynthis*, *C. sativus*, *C. maxima*, *C. sempervirens*, *C. longa*, *G. morhua*, *L. usitatissimum*, *P. ginseng*, *P. dulcis*, *R. chalepensis*, *S. indicum*, *S. aromaticum*, *T. americana*, and *Z. officinale*).

Oil groups	0.5 h	2 h	8 h	24 h	48 h	Chi ² Df= 4
Low	4.2 ± 0.847	12.3 ± 2.278	25.980 ± 6.590	49.4 ± 7.838	71.6 ± 7.39	68**
Medium	5.0 ± 1.361	13.8 ± 4.050	35.950 ± 2.864	69.5 ± 2.841	88.3 ± 3.191	28**
High	7.5 ± 1.260	22.7 ± 1.527	54.792 ± 6.389	87.1 ± 8.533	98.3 ± 1.992	31.7**
total	5.21 ± 1.733	15.21 ± 5.111	35.36 ± 13.379	63.23 ± 17.613	81.93 ± 13.09	127.6**

Table 5. Friedman test for larval mosquito mortality (%) of plant oil groups at 1000 ppm. **The X^2 value is sig. at significant level 1%

Salvia officinalis oil showed abundance of Terpinen-4-ol (17.35%), Camphor (16.08%), 14- α -H-PREGNA (9.25%), and 1-CHLOROOCCTADECANE (6.82%), (Table 12 and Fig. 6). Finally, *C. sinensis* oil is dissolved in distilled water and its major components include Gallic acid (1674 $\mu\text{g/ml}$), Catechin (421 $\mu\text{g/ml}$), Methyl gallate (1076 $\mu\text{g/ml}$), Caffeic acid (678 $\mu\text{g/ml}$), Coumaric acid (566 $\mu\text{g/ml}$), Naringenin (178 $\mu\text{g/ml}$), and Kaempferol (218 $\mu\text{g/ml}$), Table 13. Essential oils and the most active ingredients of the analyzed oils were drawn (Fig. 7).

Discussion

EOs could serve as suitable alternatives to synthetic insecticides because they are relatively safe, available, and biodegradable¹⁵. In this study, 32 oils were evaluated against *Cx. pipiens*. *Thymus vulgare* and *C. sinensis* were the most effective larvicides (100% mortality 24 h PT). The larvicidal effect of the H group could be arranged according to their LT_{50} values (h) as follows: *T. vulgaris* (9.67), *F. vulgare* (10.22), *V. odorata* (10.31), *C. sinensis* (13.02), *A. sativum* (13.95), *S. officinalis* (15.42), *N. sativa* (15.67), then *An. graveolens* (19.90). On the other hand, their LT_{99} values ranged from 29.77 (*F. vulgare*) to 55.31 (*An. graveolens*).

In this study, the most effective oils against adults were *An. graveolens* and *V. odorata* followed by *T. vulgaris* then *N. sativa*. The data revealed that *F. vulgare* is a highly potent larvicide. Similarly, its oil controlled *Anopheles atroparvus*, *Culex quinquefasciatus*^{23,24}, and *Aedes aegypti*²⁵. Despite its effectiveness as larvicide in this study, *F. vulgare* was the least effective adulticide. In contrast, it induced adulticidal properties against *Cx. quinquefasciatus*²³.

Our data indicated that *C. sinensis* was a highly effective larvicide and the less effective adulticide. Comparatively, the chemical extracts of *C. sinensis* induced larvicidal and adult repellent effects against *Cx. pipiens* providing the highest protection (100%) from the bites of starved females at the dose of 6 mg/cm²⁶. Moreover, its leaf extract showed larvicidal effect against *Anopheles arabiensis* and *Anopheles gambiae* (s.s.)²⁷.

Thymus vulgare and *An. graveolens* showed potent larvicidal and adulticidal effects in this work. Likewise, *T. vulgaris* has both effects against *Cx. quinquefasciatus*²⁸ and *Ae. aegypti*²⁹. *Thymus vulgare* exhibited larvicidal properties, 100% mortality, against *Cx. pipiens* larvae, at 200 ppm, whereas the LC_{25} and LC_{50} values indicated no effect on AChE activity, activation of the detoxification system, as indicated by an increase in GST activity and a decrease in GSH rate³⁰.

Our findings agree with another study found that the most potent EOs out of 53 oils against larvae were *F. vulgare*, *T. vulgaris*, *Citrus medica* (lime), and *C. sinensis* ($LC_{50} = 27.5, 31.6, 51.3, 53.5$ ppm, respectively). *C. sinensis* was the most efficient EOs enhancing the efficacy of deltamethrin, co-toxic factor = 316.67, over than PBO, the positive control, co-toxic factor = 283.35³¹.

Some oils applied in this study showed a similar larvicidal effect against *Cx. pipiens* as *N. sativa*^{32,33} and *S. officinalis*³⁴. Some essential oils such as *T. vulgaris*, *S. officinalis*, *C. sempervirens* and *A. graveolens* had a larvicidal effect against mosquito larvae and their LC_{90} values were < 200–300 ppm. This result may be due to several

Oil name	Conc. %	Mortality% (mean ± SD)	LC ₅₀ (lower–upper limit)	RE (LC ₅₀)	LC ₉₀ (lower–upper limit)	RE (LC ₉₀)	LC ₉₅ (lower–upper limit)	RE (LC ₉₅)	Chi (Sig)	Equation
<i>Allium sativum</i>	0	0 ± 0e	15.57 (8.49–28.46)	2.4	38.86 (26.79–81.87)	1.9	45.47 (31.19–97.80)	1.9	24.40 (0.000a)	Y = 0.051 + 0.008*x
	0.5	20.00 ± 6.67d								
	2.0	24.44 ± 5.88d								
	5.0	42.22 ± 2.22c								
	10	48.89 ± 4.44c								
	20	62.22 ± 8.01b								
<i>Anethum graveolens</i>	0	6.37 ± 18.75d	2.42 (0.08–4.22)	8.05	9.47 (4.66–17.80)	5.4	23.25 (7.17–129.13)	2.6	33.254 (.000a)	Y = 0.242 + 0.130*x
	0.1	36.86 ± 15.46bc								
	0.5	41.66 ± 27.57b								
	2	46.12 ± 11.77b								
	5	75.96 ± 18.84a								
	10	88.39 ± 7.27a								
	20	91.85 ± 9.24a								
<i>Camellia sinensis</i>	0	3.57 ± 20.00c	9.01 (–17.75 to 23.09)	2.3	32.18 (19.96–170.57)	1.6	38.754 (24.052–218.98)	1.5	26.52 (0.000a)	Y = 0.644 + 0.106*x
	2	51.51 ± 2.62b								
	5	61.21 ± 6.30ab								
	10	63.94 ± 10.22ab								
	15	75.35 ± 29.22ab								
	20	78.78 ± 16.87ab								
<i>Foeniculum vulgare</i>	0	10.50 ± 25.00d	15.07 (0.10–104.60)	1.4	33.34 (21.67–789.17)	1.5	38.53 (24.63–986.39)	1.5	22.19 (0.000a)	Y = 0.331 + 0.03*x
	5	36.73 ± 16.93bc								
	10	51.54 ± 11.47ab								
	15	51.70 ± 2.27ab								
	20	59.00 ± 16.87ab								
<i>Nigella sativa</i>	0	4.95 ± 20.61e	3.42 (–53.96 to 30.15)	6.0	5.44 (–14.41 to 84.13)	9.3	29.95 (15.87–1184.48)	2.0	57.88 (0.000a)	Y = 0.261 + 0.06*x
	0.05	41.87 ± 12.75 cd								
	0.1	60.68 ± 3.73bc								
	0.5	72.91 ± 6.45ab								
	1	74.54 ± 19.78ab								
	2	78.09 ± 18.28ab								
	10	92.96 ± 9.44ab								
<i>Salvia officinalis</i>	0	0 ± 0e	20.46 (11.34–45.85)	1.0	50.76 (33.24–140.52)	1.0	59.35 (38.59–168.23)	1.0	25.35 (0.000a)	Y = 0.8022 + 0.091*x
	0.5	17.78 ± 2.22d								
	2.0	22.22 ± 2.22d								
	5.0	37.78 ± 4.45c								
	10	44.44 ± 4.44bc								
	20	53.33 ± 3.85b								
<i>Thymus vulgaris</i>	0	3.57 ± 7.15c	3.08 (–3.29 to 7.48)	6.6	16.08 (10.43–41.60)	3.2	19.76 (12.83–52.76)	3.0	34.12 (0.000a)	Y = 0.350 + 0.091*x
	0.1	38.74 ± 4.28b								
	0.5	61.66 ± 7.26ab								
	2	69.82 ± 9.85ab								
	10	72.22 ± 14.69ab								
	20	100.00 ± 0.00a								
Continued										

Oil name	Conc. %	Mortality% (mean ± SD)	LC ₅₀ (lower–upper limit)	RE (LC ₅₀)	LC ₉₀ (lower–upper limit)	RE (LC ₉₀)	LC ₉₅ (lower–upper limit)	RE (LC ₉₅)	Chi (Sig)	Equation
<i>Viola odorata</i>	0	3.57 ± 7.15d	1.88 (–1.80 to 5.29)	10.8	7.37 (4.46–29.82)	6.9	8.92 (5.43–37.58)	6.6	21.99 (0.001a)	Y = 0.190 + 0.112*x
	0.1	50.00 ± 10.00c								
	0.5	54.95 ± 15.61c								
	1	57.50 ± 19.20c								
	2	65.83 ± 13.21bc								
	6	85.05 ± 13.62ab								
10	100.00 ± 0.00a									
Reference oils			<i>Salvia officinalis</i>							

Table 6. The adulticidal effects of selected plant oils against *Culex pipiens* after 24 h post-treatments.

Peak no.	R _i (min.)	MW	MF	Area %	Probabilities of the detected compounds
1	5.03	40	C3H4	0.14	1-Propyne
2	5.22	138	C7H10N2O	0.26	2,3,3a,4,7,7a-Hexahydro-1H-benzimidazol-2-one
3	5.28	348	C19H22ClFN2O	1.06	1-Chloro-3-(3-fluorobenzoyl)-4-(2-(diethylamino)ethylamino)benzene
4	6.38	136	C10H16	0.41	Sabinene
5	6.49	262	C12H23O4P	1.01	Dimethyl[[2,2-dimethyl-3-(2'-methylprop-1'-cyclopropyl)methyl]phosphate
6	7.57	670	C44H27DN4Ni	0.15	(5,10,15,20-tetraphenyl[2-(2)H1]prophyrin-ato)zinc(II)
7	9.17	136	C10H16	8.96	Limonene
8	10.90	152	C10H16O	2.81	1,3,3-trimethyl Bicyclo[2.2.1]heptan-2-one
10	14.26	148	C10H12O	70.36	Estragole
11	14.72	818	C44H28Br2N4Ti	0.11	Tetraphenylporphyrinatodibromotitanium (IV)
12	16.70	166	C11H18O	0.47	3,7-Dimethyl-2,6-Nonadienal
13	17.28	152	C10H16O	1.41	2,4-Decadienal
14	18.07	194	C14H26	0.17	1,1'-Bicycloheptyl
15	29.40	300	C17H36O2Si	0.20	Tetradecanoic acid, trimethylsilyl ester
16	32.19	160	C10H21F	0.15	Fluoro decane
17	32.36	244	C13H24O4	0.11	Oxalic acid isohexylpentyl ester
18	33.14	328	C19H40O2Si	1.74	Hexadecanoic acid, trimethylsilyl ester
19	33.78	282	C18H34O2	0.15	(Z) 9-Octadecenoic acid
20	34.03	138	C10H18	0.25	7-Methyl-1-nonyne
21	34.12	282	C18H34O2	0.30	(Z) 9-Octadecenoic acid
22	34.58	256	C16H32O2	0.12	Hexadecanoic acid
23	35.57	280	C18H32O2	1.44	(Z,Z) 9,12-Octadecadienoic acid
24	35.64	280	C18H32O2	1.03	(Z,Z) 9,12-Octadecadienoic acid
25	35.70	356	C21H40O4	0.53	2,3-Dihydroxypropylelaidate
26	35.76	238	C16H30O	1.67	Z-7-Hexadecenal
27	36.25	280	C18H32O2	0.23	(Z,Z) 9,12-Octadecadienoic acid
28	36.38	266	C18H34O	0.43	12-Octadecenal
29	42.83	142	C9H18O	0.13	Nonanal
31	46.93	660	C20Cl12	0.13	Dodecachloroperylene
32	48.70	295	C20H25NO	0.61	(R)-1-[N-1-cyclopentylpropionylamino-1-ethyl]naphthalene
33	50.05	354	C20H18O6	0.38	Isosamin

Table 7. GC/MS analysis of the *Foeniculum vulgare* Mill.

reasons, including the percentages of their principal components compositions that are manipulated according to the origin of plant oil, quality of oil, susceptibility of the strain used, oil storage conditions, and technical conditions^{35–37}.

Likewise our findings, *An. graveolens* and *F. vulgare* act as larvicidal, pupicidal, and oviposition deterrent agents against *M. domestica*³⁸. Moreover, *Ocimum basilicum* was the most effective extract tested on *Cx. pipiens* larvae and adults^{39,40}.

Allium sativum showed high potency against larvae in this study. A similar finding was recorded for *Cx. pipiens* and *Culex restuans* (LC₅₀ = 7.5 and 2.7 ppm, respectively)⁴¹. *Argania spinosa* oil showed a low larvicidal effect in this study. A similar effect was recorded against *Cx. quinquefasciatus* larvae⁴².

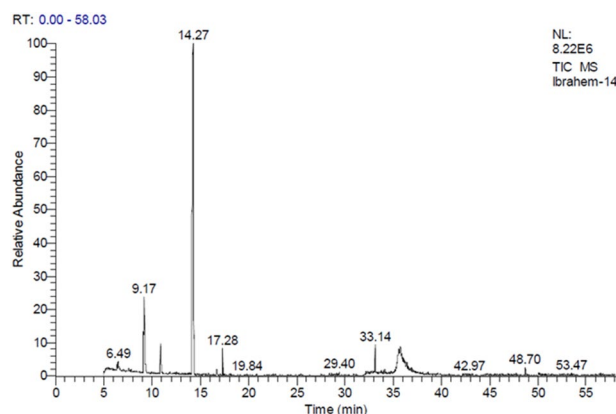


Figure 1. GC/MS analysis of the *Foeniculum vulgare* Mill.

Peak no.	R _t (min.)	MW	MF	Area %	Probabilities of the detected compounds
1	5.14	238	C13H18O4	0.49	Diethyl 3,4-bis(methylene)cyclopentane-1,1-dicarboxylate
2	5.21	600	C33H28O11	0.69	(2'S,3S,3'S,P)-hydroxyanhydrophlegmacin-9,10-quinone 8'-O-methylether
3	7.65	290	C19H30O2	0.06	2-(2'-Isopropenyldec-2'-enyl)methylcyclopentane-1,3-dione
4	9.18	136	C10H16	17.19	1,5-Dimethyl-1,5-Cyclooctadiene
5	9.35	136	C10H16	0.23	DL-Limonene
6	14.05	152	C10H16O	5.98	Dihydrocarvone
7	14.25	152	C10H16O	0.86	CIS-DIHYDROCARVONE
8	15.44	150	C10H14O	14.62	2-Methyl-5-(1-methylethenyl)2-Cyclohexen-1-one
9	15.80	733	C44H28Cl2N4V	0.07	Dichloro(5,10,15,20-tetra phenylporphyrinato)vanadium
10	16.71	692	C41H33FeO5P	0.13	Dicarbonyl(1,3-5-ü-6-phenyl-2-(phenylethynyl)cyclohept-4-ene-1,3-diyl) triphenoxyphosphaneiron
11	17.29	110	C8H14	0.47	octahydro Pentalene
12	18.89	675	C44H28CuN4	0.09	(5,10,15,20-tetraphenyl[2-(2)H1]prophyrinato)copper(II)
13	20.82	204	C15H24	0.10	à-Humulene
14	21.36	686	C37H24Cl2N6O4	0.08	2,2-Bis[4[[4-chloro-6-(3-ethynylphenoxy)-1,3,5-triazin-2-yl]oxy]phenyl]propane
15	21.92	134	C10H14	0.14	1,2,3,4-Tetramethyl-5-methylenecyclopenta-1,3-diene
16	22.07	204	C15H24	0.38	á-Bisabolene
17	22.16	648	C35H38Cl2N4O4	0.11	2,4-bis(á-chloroethyl)-6,7-bis[á-methoxycarbonylethyl]-1,3,5-trimethylporphyrin
18	22.36	640	C32H64O5Si4	0.23	OTETRAKIS(TRIMETHYLSILYL)3,5-DIHYDROXY-2-(3-HYDROXY-1-OCTENYL)CYCLOPENTANEHEP-TANOATE
19	23.34	208	C14H24O	0.18	3-Oxabicyclo[3.3.1]non-6-ene
20	24.23	222	C15H26O	21.26	3a(1H)-Azulenol,2,3,4,5,8,8a-hexahydro-6,8-adimethyl-3-(1-methylethyl),[3R-(3à,3aà,8aà)]
21	24.57	572	C23H26Br2O7	0.10	Dibromogomisin A
22	25.05	222	C10H14N4S	32.13	4-Pyridinecarbaldehyde-4-propyl-3-thiosemicarbazone
23	25.28	238	C15H26O2	2.39	Daucol
24	26.01	194	C12H18O2	0.06	3-(1-Hydroxyhexyl)phenol
25	27.54	220	C15H24O	0.06	Trans-Z-à-Bisaboleneepoxide
26	33.01	2598	N/A	0.07	YGRKKRRQRRRGPKRRLLDL/5
27	34.16	691	C51H33NO2	0.07	2,6-Bis(2,3,5-triphenyl-4-oxocyclopentadienyl)pyridine
28	35.47	733	C44H28Cl2N4V	0.08	Dichloro(5,10,15,20-tetraphenylporphyrinato)vanadium
29	40.31	739	C39H81NO4Si4	0.13	(3S,4R,1'E,2'R,3'R)-1-tertButyldimethylsilyl-4-(3'-tertbutyldimethylsilyloxy-2'-methylprop-1'-enyl)-3-(1'',3'' di(tertbutyldimethylsilyloxy)-2''-methylhex-5''-yl)-3-methylazetidin-2-one
31	43.48	114	C6H10O2	0.13	3,4-Hexanedione
32	50.56	680	C35H40O5Si5	0.06	Pentamethylpentaphenylcyclopentasiloxane
33	51.11	733	C44H28Cl2N4V	0.09	Dichloro(5,10,15,20-tetraphenylporphyrinato)vanadium

Table 8. GC/MS analysis of the *Anethum graveolens* L.

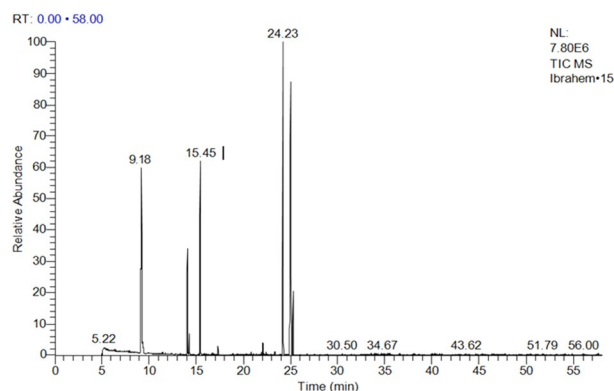


Figure 2. GC/MS analysis of the *Anethum graveolens L.*

Peak no.	R _t (min.)	MW	MF	Area %	Probabilities of the detected compounds
1	23.923	170	C12H10O	42.04	Diphenyl ether
2	24.735	192	C13H20O	11.87	.alpha.-Ionone
3	26.485	192	C13H20O	7.73	3-Buten-2-one, 4-(2,6,6-trimethyl-1-cyclohexen-1-yl)
4	28.317	236	C15H24O2	0.61	Limonen-6-ol, pivalate
5	28.58	226	C13H22O3	0.9	2-Hydroxy-1,1,10-trimethyl-6,9-epidioxydecalin
6	28.786	238	C16H30O	1.26	7-Hexadecenal, (Z)-
7	29.599	236	C16H28O	0.83	7,11-Hexadecadienal
8	29.713	296	C20H40O	1.48	Phytol
9	29.959	242	C16H34O	2.15	2-Hexyl-1-Decanol
10	30.074	378	C25H46O2	1.09	Undec-10-ynoic acid, tetradecyl ester
11	30.211	296	C20H40O	1.02	PHYTOL ISOMER
12	30.881	266	C16H26O3	0.67	2-Dodecen-1-yl(-)succinic anhydride
13	31.338	242	C16H34O	2.14	1-Decanol, 2-hexyl-
14	31.939	218	C16H26	2.79	hexadecahydroPyrene
15	32.054	240	C17H36	0.7	Tetradecane, 2,6,10-trimethyl
16	34.245	250	C16H26O2	7.22	(Z)-5-(4-tert-Butyl-1-hydroxycyclohexyl)-3-methylpent-2-en-4-yne
17	35.092	264	C15H20O4	6.6	2,3,3a,4,5,5a,6,7,9a,9b-decahydro-3,5a,9-trimethyl-7,9a-peroxy Naphtho[1,2-b]furan-2-one
18	35.269	264	C15H20O4	4.73	2,3,3a,4,5,5a,6,7,9a,9b-decahydro-3,5a,9-trimethyl-7,9a-peroxy Naphtho [1,2-b]furan-2-one
19	35.905	242	C16H34O	2.19	2-hexyl-1-Decanol
20	37.146	266	C18H34O	1.89	Z,E-2,13-Octadecadien-1-ol
21	23.923	170	C12H10O	0.78	Diphenyl ether

Table 9. GC/MS analysis of the *Viola odorata L.*

Curcuma species was less effective in this study, but its 27 components as curcuminoids and monocarbonyl curcumin derivatives were effective larvicidal agents against *Cx. pipiens* and *Ae. albopictus*⁴³ and hexane extraction of *Curcuma longa* showed 100% larvicidal activity against *Cx. pipiens* and *Aedes albopictus* at 1000 ppm after being treated 24 h⁴⁴.

Zingiber officinale and *Syzygium aromaticum* were less effective. In contrast, they were effective against *Cx. pipiens* (LC₅₀ = as 71.85 and 30.75, respectively)⁴⁵.

Sesamum indicum is one of the L group in this study. In contrast, petroleum ether extract showed larvicidal, antifeedant and repellent action against *Cx. pipiens*³³. Furthermore, EOs of *N. sativa*, *Allium cepa*, and *S. indicum*, induced larvicidal effect and their LC₅₀ values against both field and laboratory strains of *Cx. pipiens* were 247.99 and 108.63; 32.11 and 2.87; and finally, 673.22 and 143.87 ppm, respectively. They influenced the pupation and adult emergence rates besides developmental abnormalities at sublethal concentrations⁴⁶.

Boswellia serrata (M group) and *Brassica carinata* (L group) showed relative larvicide against *Cx. pipiens* in this study. A similar result was reported^{47,48}. The lethal concentration values of Fenugreek (*Trigonella foenum-grecum*), earth almond (*Cyperus esculentus*), mustard (*Brassica campestris*), olibanum (*Boswellia serrata*), rocket (*Eruca sativa*), and parsley (*Carum ptroselinum*) were 32.42, 47.17, 71.37, and 83.36, 86.06, and 152.94 ppm,

User Chromatograms

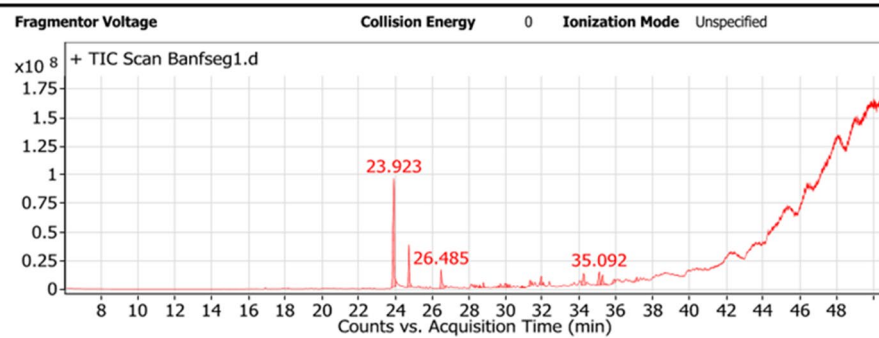


Figure 3. GC/MS analysis of the sample *Viola odorata* L.

respectively. Against *Cx. pipiens* larvae. Furthermore, increasing concentrations were directly proportional to the reduction of both pupation and adult emergences rates⁴⁸.

Some oil-resins as *Commiphora molmol*, *Araucaria heterophylla*, *Eucalyptus camaldulensis*, *Pistacia lentiscus*, and *Boswellia sacra* showed larvicidal activity against *Cx. pipiens* larvae. The larvicidal effect 24 and 48 h PT, respectively, were for acetone extracts, 1500 ppm, of *C. molmol* (83.3% and 100% and LC_{50} = 623.52 and 300.63 ppm) and *A. heterophylla* (75% and 95% and LC_{50} = 826.03 and 384.71 ppm). On the other hand, the aqueous extract of *A. heterophylla* induced higher mortalities (LC_{50} = 2819.85 ppm and 1652.50 ppm), followed by *C. molmol*, (LC_{50} = 3178.22 and 2322.53 ppm)⁴⁹.

A similar larvicidal effect was recorded for *Rosmarinus officinalis*, hexane extract (80 and 160 ppm), reduced 100% mortality against 3rd and 4th instars larvae of *Cx. pipiens* and the toxicity increased in the pupal and adult stages⁵⁰.

Out of 36 essential oils, red moor besom leaf oil has strong fumigation activity against *Cx. pipiens* pallens adults⁵¹. Similar to the adulticidal effect of the applied oils in this work, some other oils have adulticidal activities against mosquitoes as *Cedrus deodara*, *Eucalyptus citriodora*, *Cymbopogon flexuosus*, *Cymbopogon winterianus*, *Pinus roxburghii*, *S. aromaticum*, and *Tagetes minuta*⁵². The Leaf Oils of *Cinnamomum* species had adulticidal activities against *Ae. aegypti* and *Aedes albopictus*⁵³. EOs have adulticidal effects against *Musca domestica*⁵⁴ as *A. sativum*, *S. aromaticum*, and *F. vulgare*⁵⁵. Essential oils of *Melaleuca leucadendron* (L.) and *Callistemon citrinus* (Curtis) showed 100% adult mortality against *Aedes aegypti* (L.) and *Cx. quinquefasciatus* (Say), 24 h exposure⁵⁶.

The results showed that *A. sativum*, and *S. officinalis* oils were effective against mosquito larvae, maybe due to the presence of a number of active secondary compounds such as ISOCHIAPIN B%2 < (sesquiterpene lactone) and 9-Octadecenamamide, (Z)-that are anti-inflammatory activity⁵⁷, also, Terpinen-4-ol and Camphor in Sage oil that these are excellent natural insecticide⁵⁸, but these oils garlic and Sage did not show the required efficacy against adult mosquitoes.

The phytochemical analysis of this study revealed the major activated compounds of the analyzed oils. Green tea oil is a highly effective larvicide in this study contains a high amount of polyphenols that have antioxidant activity. A similar finding was reported⁵⁹. Our data indicated that green tea oil also contains polyphenols as Gallic acid, Catechin, Methyl gallate, Coffeic acid, Coumaric acid, Naringenin, and Kaempferol which might aid in its insecticidal effect.

This study indicated that *F. vulgare* contains Estragole (70.36%) and Limonene (8.96%). Similarly, Limonene as a cyclic monoterpene has a viable insecticidal effect⁶⁰. Besides, Estragole induced toxicity to adult fruit flies, *Ceratitis capitata*⁶¹. Moreover, *An. graveolens* contains thiosemicarbazone (32.13%) in this study. Likewise, thiosemicarbazide is a major component *An. graveolens* with insecticidal effect⁶². Also, Dauco and carotol are essential oils documented for *An. graveolens* in this work have repellent activity against adult *Ae. aegypti*, *Ae. albopictus*, and *Anopheles quadrimaculatus* Say⁶³. Furthermore, *V. odorata* in the present analysis contains alpha-ionone, which revealed anti-inflammatory and analgesic effects⁶⁴. *Thymus vulgaris* showed good alpha-pinene and pyridine derivatives that play an important role as larvicidal and adulticidal effects against *Ae. aegypti* and growth regulator, respectively^{65,66}. In addition, the combination of all constituents may promote their individual larvicidal and adulticidal effects.

The biochemical compositions showed that *T. vulgaris* oil affected the energy reserves with a marked effect on proteins and lipids³⁰. The differences between our findings and those of the others could be attributed to the biological activities and the chemical composition for EOs, which could vary between plant age, tissues, geographical origin, the part used in the distillation process, distillation type, and the species. Therefore, types and levels of active constituents in each oil may be responsible for the variability in their potential against pests¹⁶.

Conclusions

Diseases transmitted by mosquitoes represent global concerns. Our findings demonstrate the potential of *F. vulgare* and *C. sinensis* as the most potent larvicides and *N. sativa*, *V. odorata*, and *An. graveolens* as the most effective adulticides as they contain good command of different essential oils. EOs could be used for integrated mosquito control programs as larvicides or synergists for enhancing the efficacy of current adulticides³¹. Further

Peak no.	R _i (min.)	MW	MF	Area %	Probabilities of the detected compounds
1	5.1	208	C13H20O2	0.86	TRANS- α -IONON-5,6-EPOXIDE
2	5.23	122	C8H15B	0.79	1-Borabicyclo[4.3.0]nonane
3	6.46	136	C10H16	1.85	Tricyclene
4	6.86	136	C10H16	0.69	Camphene
5	7.64	136	C10H16	8.92	2- α -pinene
6	9.07	119	C7H5NO	12.37	2-Ethynyl-3-hydroxypyridine
7	11.32	196	C12H20O2	0.68	Linalyl acetate
8	12.50	152	C10H16O	1.27	(1S) Bicyclo[2.2.1]heptan-2-one, 1,7,7-trimethyl
9	13.39	156	C10H20O	0.78	1-Methyl-4-(1-methylethyl)Cyclohexanol
10	13.51	154	C10H18O	4.73	4-Methyl-1-(1-methylethyl)-3-Cyclohexen-1-ol
11	13.91	154	C10H18O	1.13	$\alpha,\alpha,4$ -trimethyl (S) 3-Cyclohexene-1-methanol
12	15.67	182	C11H18O2	0.63	linalyl formate
13	16.48	196	C12H20O2	1.76	EXOBORNYL ACETATE
14	18.17	196	C12H20O2	5.00	α -terpinyl acetate
15	20.52	142	C9H18O	0.56	3-Ethylheptanal
16	21.94	268	C19H40	0.58	Nonadecane
17	22.84	199	C9H13NO4	1.87	2S,7S Methyl-2-Hydroxy-3-oxotetrahydro-1-Hpyrrolizine-7a-(5H)-carboxylate
18	22.97	226	C16H34	0.92	Pentadecane-5-methyl
19	23.10	212	C15H32	0.75	3-ethyl Tridecane
20	23.22	348	C19H40O3S	0.84	hexyltridecyl ester Sulfurous acid
21	23.39	226	C16H34	1.09	3-methyl Pentadecane
22	24.06	168	C8H12N2O2	1.52	1,6-diisocyanato Hexane
23	24.24	298	C20H42O	2.36	1,1'-oxybis Decane,
24	24.40	282	C20H42	0.81	Eicosane
25	24.65	334	C18H38O3S	0.57	Sulfurous acid, butyltetradecyl ester
26	25.10	282	C20H42	4.12	10-Methylnonadecane
27	25.24	268	C19H40	1.00	7-hexyl Tridecane
28	25.37	334	C18H38O3S	1.10	6-Tetradecanesulfonic acid, butyl ester
29	25.49	334	C18H38O3S	1.44	6-Tetradecanesulfonic acid, butyl ester
31	25.68	250	C16H26O2	4.54	3-(6,6-Dimethyl-5-oxohept-2-enyl)-cycloheptanone
32	25.98	222	C13H18O3	7.70	2,5-Dipropoxybenzaldehyde
33	26.30	352	C25H52	1.33	Pentacosane
34	26.44	282	C20H42	3.55	9-methyl, Nonadecane
35	26.62	224	C16H32	1.08	1-Hexadecene
36	26.84	236	C16H28O	2.14	7,11-Hexadecadienal
37	27.25	232	C11H12N4O2	5.05	5-Amino-8-cyano-7-methoxy-3,4-dihydro-3-methyl-1,6-naphthyridin-2(1H)-one
38	27.32	232	C15H20O2	2.01	(2R,3R)-3-(2-Methoxy-4-methylphenyl)-2,3-dimethylcyclopentanone
39	27.42	282	C20H42	0.87	2,6-dimethyl Octadecane
40	27.54	310	C22H46	0.77	8-heptyl Pentadecane
41	27.65	376	C21H44O3S	0.61	Sulfurous acid, hexyl pentadecyl ester
42	27.82	226	C16H34	0.88	Hexadecane
43	28.42	164	C5H9BrO	0.62	1-Bromo-2-methyl-3-Buten-2-ol
44	28.54	242	C16H34O	1.25	2-Hexyl-1-decanol
45	28.69	111	C7H13N	1.08	1-isocyano Hexane
46	29.32	116	C7H16O	1.94	2-ethyl 1-Pentanol
47	30.70	200	C13H28O	0.82	2-Propyldecane-1-ol
48	31.33	197	C11H19NO2	0.98	2-Ethylhexyl cyanoacetate
49	33.27	592	C41H84O	0.70	1-Hentetracontanol
50	36.28	324	C23H48	0.57	9-hexyl Heptadecane
51	37.92	366	C26H54	0.58	5,14-dibutyl Octadecane

Table 10. GC/MS analysis of *Thymus vulgaris* L.

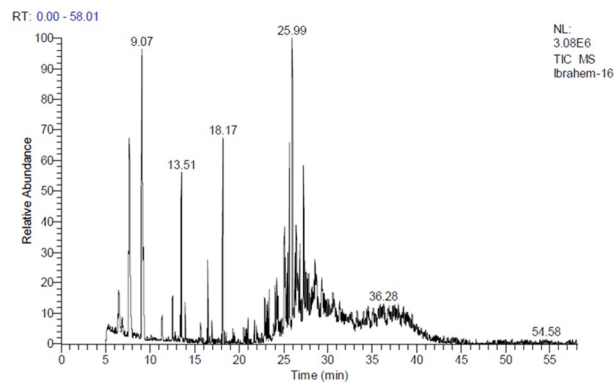


Figure 4. GC/MS analysis of *Thymus vulgaris* L.

Peak no.	R _t (min.)	MW	MF	Area %	Probabilities of the detected compounds
1	6.27	146	C6H10S2	4.54	Diallyl disulphide
2	7.49	152	C4H8S3	9.68	Trisulfide, methyl 2-propenyl
3	9.35	178	C6H10S3	14.86	Trisulfide, di-2-propenyl
4	12.22	350	C19H26O6	8.63	ISOCHIAPIN B %2 <
5	14.97	334	C20H30O4	3.54	1,2-Benzenedicarboxylic acid, butyl octyl ester
6	16.05	346	C19H22O6	3.11	ISOCHIAPIN B
7	17.67	387	C17H37N7O3	7.84	9-OCTADECENAMIDE
8	19.61	281	C18H35NO	29.07	9-Octadecenamide, (Z)-
10	21.40	208	C11H12O2S	4.25	3-(Benzylthio)acrylic acid, methyl ester
11	23.27	300	C19H24O3	5.86	3,17-DIOXO-11-à-HYDROXYANDROSTANE-1,4-DIENE
12	23.54	436	C26H44O5	1.82	3 Ethyl iso-allocholate
13	23.62	490	C34H50O2	6.81	CHOLEST-5-EN-3-YL BENZOATE

Table 11. GC/MS analysis of the *Allium sativum*. 9-Octadecenamide, (Z)- (29.07), Trisulfide, di-2-propenyl (14.86), and ISOCHIAPIN B %2 < (8.63).

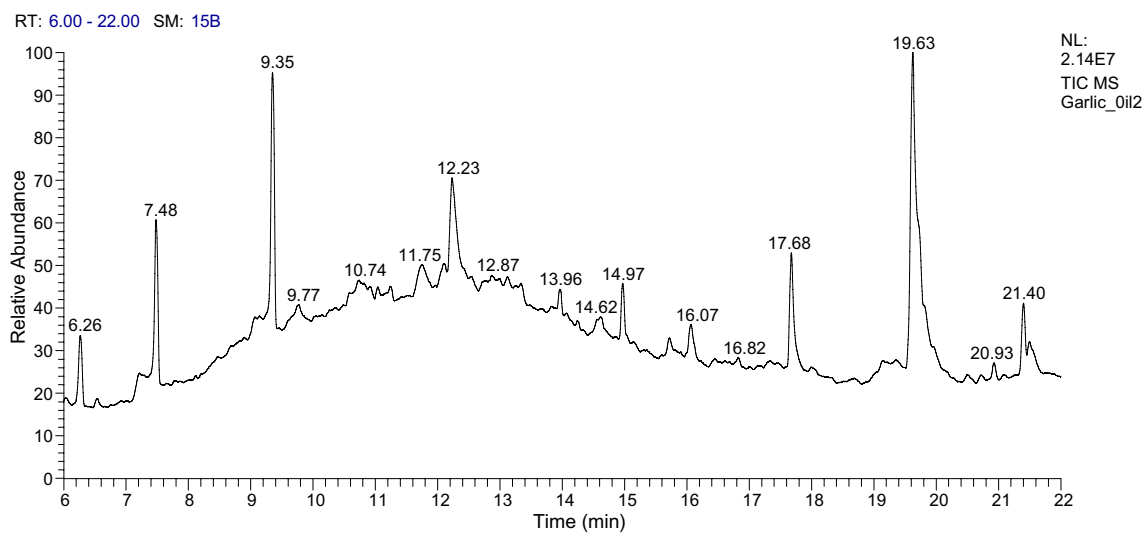


Figure 5. GC/MS analysis of *Allium sativum*.

Peak no.	R _i (min.)	MW	MF	Area %	Probabilities of the detected compounds
1	10.22	152	C10H16O	16.08	Camphor
2	10.90	156	C10H20O	5.24	Cyclohexanol, 1-methyl-4-(1-methylethyl)-
3	11.47	154	C10H18O	17.35	Terpinen-4-ol
4	13.86	254	C13H24O2	2.47	Tridecanedial
5	14.50	280	C18H32O2	3.43	17-Octadecynoic acid
6	15.70	400	C28H48O	0.90	Cholestan-3-ol, 2-methylene-, (3 α ,5 α)-
7	16.68	268	C17H32O2	1.80	7-Methyl-Z-tetradecen-1-ol acetate
8	17.50	280	C19H36O	1.63	12-Methyl-E,E-2,13-octadecadien-1-ol
10	17.99	288	C21H36	2.03	14- \acute{a} -H-PREGNA
11	19.18	288	C18H37Cl	5.13	1-CHLOROCTADECANE
12	19.51	288	C21H36	1.77	14- \acute{a} -H-PREGNA
13	19.86	450	C32H66	4.33	DOTRIACONTANE
14	20.18	536	C37H76O	1.41	1-Heptatriacotanol
15	20.32	268	C16H28O3	1.15	Z-(13,14-Epoxy)tetradec-11-en-1-ol acetate
16	20.55	258	C16H34S	1.58	tert-Hexadecanethiol
17	20.80	312	C20H40O2	3.17	Ethanol, 2-(9-octadecenyloxy)-, (Z)-
18	20.90	288	C21H36	2.18	14- \acute{a} -H-PREGNA
19	21.26	350	C19H26O6	0.73	ISOCHIAPIN B %2<
20	21.61	288	C18H37Cl	6.82	1-CHLOROCTADECANE
21	21.84	294	C21H36	3.7	14- \acute{a} -H-PREGNA
22	22.39	288	C21H36	0.82	1-Heptatriacotanol
23	22.47	346	C19H22O6	2.74	ISOCHIAPIN B
24	22.73	288	C21H36	9.25	14- \acute{a} -H-PREGNA
25	23.09	280	C19H36O	2.20	12-Methyl-E,E-2,13-octadecadien-1-ol
26	23.23	350	C19H26O6	2.05	ISOCHIAPIN B %2<

Table 12. GC/MS analysis of the *Salvia officinalis*.

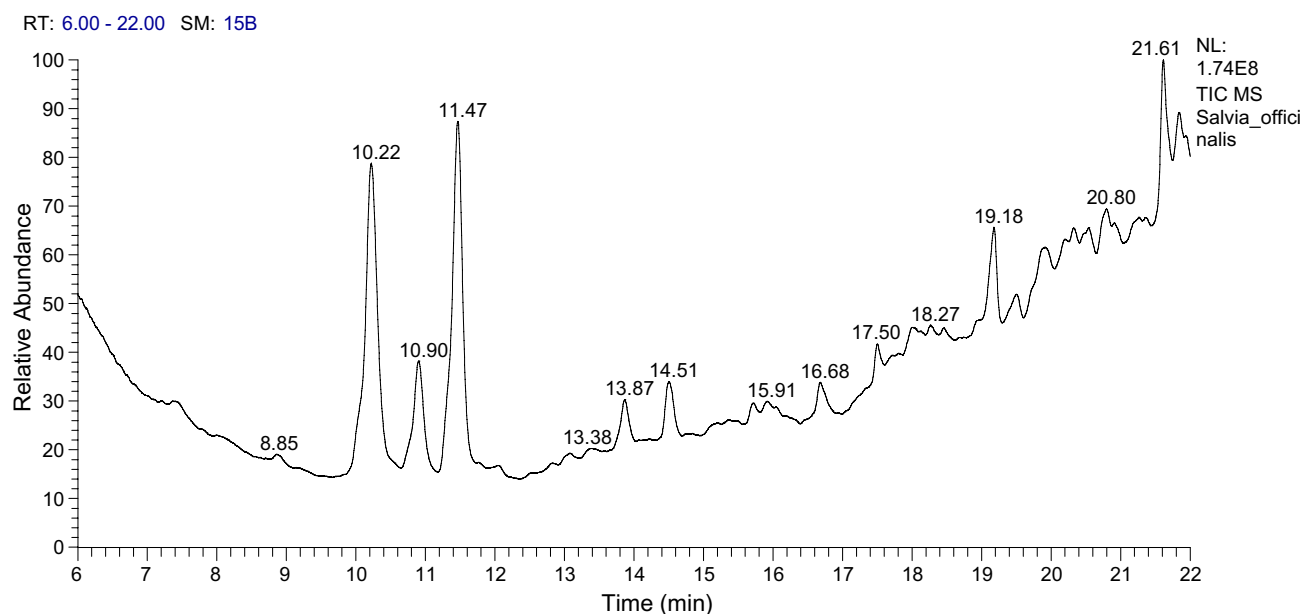


Figure 6. GC/MS analysis of *Salvia officinalis*.

Standard			Sample green tea		
St. compound	Conc. (µg/ml)	Area	Compound	Area	Conc. (µg/ml = µg/g)
allic acid	16.8	179.72	Gallic acid	895.77	1674.71
Chlorogenic acid	28	335.23	Chlorogenic acid	75.30	125.79
Catechin	67.5	584.16	Catechin	182.42	421.56
Methyl gallate	10.2	789.05	Methyl gallate	4163.86	1076.52
Coffeic acid	18	469.51	Coffeic acid	895.98	687.01
Syringic acid	17.2	389.86	Syringic acid	30.41	26.83
Pyro catechol	29.2	451.95	Pyro catechol	0.00	0.00
Rutin	61	457.55	Rutin	71.83	191.53
Ellagic acid	34.3	495.60	Ellagic acid	37.52	51.93
Coumaric acid	13.2	729.56	Coumaric acid	1566.70	566.93
Vanillin	12.9	543.81	Vanillin	0.00	0.00
Ferulic acid	12.4	353.45	Ferulic acid	71.09	49.88
Naringenin	15	266.56	Naringenin	158.25	178.11
Taxifolin	13.2	189.35	Taxifolin	16.08	22.42
Cinnamic acid	5.8	573.08	Cinnamic acid	0.00	0.00
Kaempferol	12	289.35	Kaempferol	263.99	218.97

Table 13. HPLC analysis for *Camellia sinensis*.

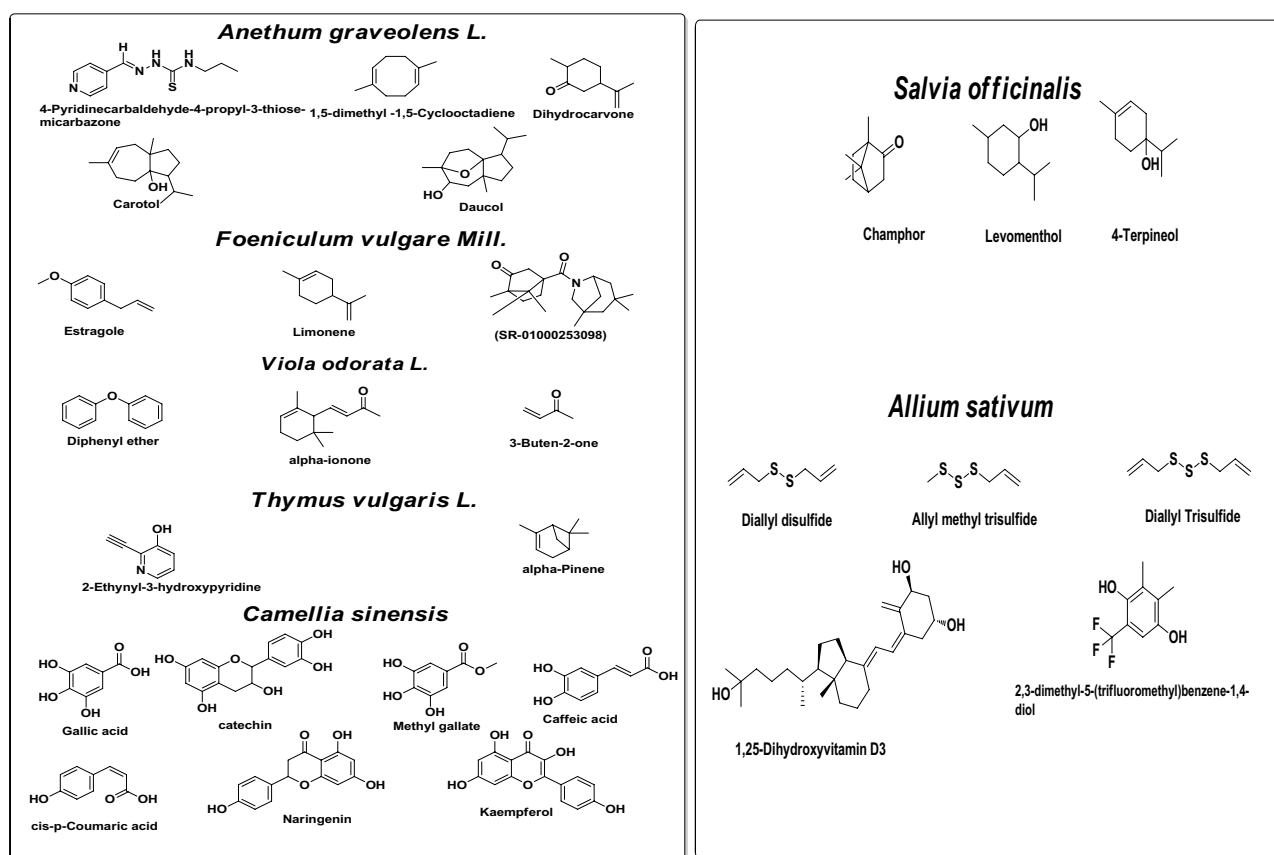


Figure 7. Essential oils and their most active ingredients.

studies are needed to develop nanoformulations that improve the efficacy and minimize applications after revealing their ecotoxicological side views.

Received: 23 September 2021; Accepted: 24 February 2022

Published online: 15 March 2022

References

- Jones, R. T., Ant, T. H., Cameron, M. M. & Logan, J. G. Vol. 376 (The Royal Society, 2021).
- Abdel-Shafi, I. R. *et al.* Mosquito identification and molecular xenomonitoring of lymphatic filariasis in selected endemic areas in Giza and Qualioubiya Governorates, Egypt. *J. Egypt. Soc. Parasitol.* **46**, 93–100 (2016).
- Selim, A., Radwan, A., Arnaout, F. & Khater, H. The recent update of the situation of west Nile fever among equids in Egypt after three decades of missing information. *Pakistan Veterinary J.* **40** (2020).
- Selim, A., Megahed, A., Kandeel, S., Alouffi, A. & Almutairi, M. M. West Nile virus seroprevalence and associated risk factors among horses in Egypt. *Sci. Rep.* **11**, 1–9 (2021).
- Selim, A. & Radwan, A. Seroprevalence and molecular characterization of West Nile Virus in Egypt. *Compar. Immunol. Microbiol. Infectious Diseases.* **71**, 101473 (2020).
- Jones, R. T., Ant, T. H., Cameron, M. M. & Logan, J. G. (The Royal Society, 2021).
- Selim, A., Manaa, E., Abdelhady, A., Ben Said, M. & Sazmand, A. Serological and molecular surveys of Anaplasma spp. in Egyptian cattle reveal high A. marginale infection prevalence.
- Selim, A. *et al.* Seroprevalence and risk factors associated with Canine Leishmaniasis in Egypt. *Veterinary Sci.* **8**, 236 (2021).
- Selim, A., Megahed, A. A., Kandeel, S. & Abdelhady, A. Risk factor analysis of bovine leukemia virus infection in dairy cattle in Egypt. *Compar. Immunol. Microbiol. Infectious Diseases.* **72**, 101517 (2020).
- Selim, A. & Abdelhady, A. The first detection of anti-West Nile virus antibody in domestic ruminants in Egypt. *Trop. Anim. Health Prod.* **52**, 3147–3151 (2020).
- Selim, A., Abdelhady, A. & Alahadeb, J. Prevalence and first molecular characterization of Ehrlichia canis in Egyptian dogs. *Pak. Vet. J.* (2020).
- Khater, H. F. *et al.* *Malaria* (IntechOpen, 2019).
- Baz, M. M. Strategies for mosquito control. *PhD thesis, faculty of Science, Benha University, Egypt* (2013).
- Khater, H. F. Prospects of botanical biopesticides in insect pest management. *Pharmacologia* **3**, 641–656 (2012).
- Khater, H. F. Bioactivity of essential oils as green biopesticides: Recent global scenario. *Recent Progress Med. Plants* **37**, 151–218 (2013).
- Khan, N. & Mukhtar, H. Tea and health: Studies in humans. *Curr. Pharm. Des.* **19**, 6141–6147 (2013).
- Govindarajan, M., Rajeswary, M., Hoti, S., Bhattacharyya, A. & Benelli, G. Eugenol, α -pinene and β -caryophyllene from *Plectranthus barbatus* essential oil as eco-friendly larvicides against malaria, dengue and Japanese encephalitis mosquito vectors. *Parasitol. Res.* **115**, 807–815 (2016).
- Khater, H. & Geden, C. Potential of essential oils to prevent fly strike by *Lucilia sericata*, and effects of oils on longevity of adult flies. *J. Vector Ecol.* **43**, 261–270 (2018).
- Noutcha, M. A., Edwin-Wosu, N. I., Ogali, R. E. & Okiwelu, S. N. The role of plant essential oils in mosquito (Diptera: Culicidae) control. *Annu. Res. Rev. Biol.* 1–9 (2016).
- WHO. Larval source management: A supplementary malaria vector control measure: An operational manual. (2013).
- Vatandoost, H. *et al.* Comparison of CDC bottle bioassay with WHO standard method for assessment susceptibility level of malaria vector, *Anopheles stephensi* to three imagicides. *J. Arthropod. Borne Dis.* **13**, 17 (2019).
- Shafia, E., Aramideh, S., Valizadegan, O., Safaralizadeh, M. H. & Pesyan, N. N. GC/MS analysis of the essential oils of *Cupressus arizonica* Greene, *Juniperus communis* L. and *Mentha longifolia* L. *Bull. Chem. Soc. Ethiopia.* **33**, 389–400 (2019).
- Modise, S. A. & Ashafa, A. O. T. Larvicidal, pupicidal and insecticidal activities of *Cosmos bipinnatus*, *Foeniculum vulgare* and *Tagetes minuta* against *Culex quinquefasciatus* mosquitoes. *Trop. J. Pharm. Res.* **15**, 965–972 (2016).
- Pavela, R., Žabka, M., Bednář, J., Triska, J. & Vrhotová, N. New knowledge for yield, composition and insecticidal activity of essential oils obtained from the aerial parts or seeds of fennel (*Foeniculum vulgare* Mill.). *Ind. Crops Products.* **83**, 275–282 (2016).
- Rocha, D. K. *et al.* Larvicidal activity against *Aedes aegypti* of *Foeniculum vulgare* essential oils from Portugal and Cape Verde. *Nat. Product Commun.* **10**, 1934578X1501000438 (2015).
- Hassan, M. I., Atwa, W. A., Moselhy, W. A. & Mahmoud, D. A. Efficacy of the green tea, *Camellia sinensis* leaves extract on some biological activities of *Culex pipiens* and the detection of its phytochemical constituents. *Egypt. Acad. J. Biol. Sci. F. Toxicol. Pest Control.* **12**, 59–70 (2020).
- Muema, J. M., Bargul, J. L., Nyanjom, S. G., Mutunga, J. M. & Njeru, S. N. Potential of *Camellia sinensis* proanthocyanidins-rich fraction for controlling malaria mosquito populations through disruption of larval development. *Parasit. Vectors* **9**, 1–10 (2016).
- Pavela, R. Larvicidal property of essential oils against *Culex quinquefasciatus* Say (Diptera: Culicidae). *Ind. Crops Prod.* **30**, 311–315 (2009).
- de Oliveira, A. A. *et al.* Larvicidal, adulticidal and repellent activities against *Aedes aegypti* L. of two commonly used spices, *Origanum vulgare* L. and *Thymus vulgaris* L. *S. Afr. J. Bot.* **140**, 17–24 (2021).
- Bouguerra, N., Tine-Djebbar, F. & Soltani, N. Effect of *Thymus vulgaris* L. (Lamiales: Lamiaceae) essential oil on energy reserves and biomarkers in *Culex pipiens* L. (Diptera: Culicidae) from Tebessa (Algeria). *J. Essential Oil Bearing Plants.* **21**, 1082–1095 (2018).
- Sheng, Z. *et al.* Screening of larvicidal activity of 53 essential oils and their synergistic effect for the improvement of deltamethrin efficacy against *Aedes albopictus*. *Ind. Crops Products.* **145**, 112131 (2020).
- Alkenani, N. A. *et al.* Molecular identification and bio-control of mosquitoes using black seeds extract in Jeddah. *Pak. Vet. J.* <https://doi.org/10.29261/pakvetj/2021.025> (2021).
- Farag, M. Larvicidal and repellent potential of *Sesamum indicum* hull peels extracts against *Culex pipiens* L. (Diptera: Culicidae). *Egypt. J. Aquat. Biol. Fisheries.* **25**, 995–1011 (2021).
- Abd El Meguid, A. D., Mahmoud, S. H. & Baz, M. M. Toxicological activity of four plant oils against *Aedes caspius* and *Culex pipiens* (Diptera: Culicidae). *Int. J. Mosq. Res.* **6**, 86–94 (2019).
- El Ouali Lalami, A., El-Akhal, F., Ez Zoubi, Y. & Taghzouti, K. Study of phytochemical screening and larvicidal efficacy of ethanolic extract of *Salvia officinalis* (Lamiaceae) from North Center of Morocco against *Culex pipiens* (Diptera: Culicidae) vector of serious human diseases. *Int. J. Pharmacog. Phytochem. Res.* **8**, 1663–1668 (2016).
- Hayouni, E. A. *et al.* Tunisian *Salvia officinalis* L. and *Schinus molle* L. essential oils: Their chemical compositions and their preservative effects against Salmonella inoculated in minced beef meat. *Int. J. Food Microbiol.* **125**, 242–251 (2008).
- Nabti, I. & Bounechada, M. Larvicidal activities of essential oils extracted from five Algerian medicinal plants against *Culiseta longiareolata* Macquart. Larvae (Diptera: Culicidae). *Eur. J. Biol.* **78**, 133–138 (2019).
- Chantawee, A. & Soonwera, M. Larvicidal, pupicidal and oviposition deterrent activities of essential oils from Umbelliferae plants against house fly *Musca domestica*. *Asian Pac. J. Trop. Med.* **11**, 621 (2018).
- Belong, P., Ntonga, P. A., Fils, E., Dadji, G. A. F. & Tamesse, J. L. Chemical composition and residue activities of *Ocimum canum* Sims and *Ocimum basilicum* L. essential oils on adult female *Anopheles funestus*. *J. Anim. Plant Sci.* **19**, 2854–2863 (2013).
- El Zayyat, E. A., Soliman, M. I., Elleboudy, N. A. & Ofaa, S. E. Bioefficacy of some Egyptian aromatic plants on *Culex pipiens* (Diptera: Culicidae) adults and larvae. *J. Arthropod. Borne Dis.* **11**, 147 (2017).
- Muturi, E. J., Ramirez, J. L., Zilkowski, B., Flor-Weiler, L. B. & Rooney, A. P. Ovicidal and larvicidal effects of garlic and asafetida essential oils against West Nile virus vectors. *J. Insect Sci.* **18**, 43 (2018).

42. Alerwi, S. T. *et al.* Molecular identification and bio-control of *Culex quinquefasciatus* from Yanbu region. *J. Entomol. Zool. Stud.* **7**, 1081–1086 (2019).
43. Matiadis, D. *et al.* Curcumin derivatives as potential mosquito larvicidal agents against two mosquito vectors, *Culex pipiens* and *Aedes albopictus*. *Int. J. Mol. Sci.* **22**, 8915 (2021).
44. Prak, J.-W., Yoo, D.-H., Kim, H. K., Koo, H.-N. & Kim, G.-H. in 2014 Larvicidal and repellent activities of 33 plant extracts against two mosquitoes as *Culex pipiens* and *Aedes albopictus*. 181–181.
45. Jabbar, A., Tariq, M., Gulzar, A., Mukhtar, T. & Zainab, T. Lethal and sub lethal effects of plant extracts and green silver nanoparticles against *Culex pipiens*. (2021).
46. Khater, H. F. *Biocontrol of Some Insects* (Benha University, 2003).
47. Baz, M. M., Hegazy, M. M., Khater, H. F. & El-Sayed, Y. A. Comparative evaluation of five oil-resin plant extracts against the mosquito larvae, *Culex pipiens* Say (Diptera: Culicidae). *Pak. Vet. J.* <https://doi.org/10.29261/pakvetj> (2021).
48. Khater, H. F. & Shalaby, A.A.-S. Potential of biologically active plant oils to control mosquito larvae (*Culex pipiens*, Diptera: Culicidae) from an Egyptian locality. *Rev. Inst. Med. Trop. Sao Paulo* **50**, 107–112 (2008).
49. Baz, M. M., Hegazy, M. M., Khater, H. F. & El-Sayed, Y. A. Comparative evaluation of five oil-resin plant extracts against the mosquito larvae, *Culex pipiens* Say (Diptera: Culicidae). *Pak. Vet. J.* **41**, 191–196 (2021).
50. Shalaby, A. & Khater, H. Toxicity of certain solvent extracts of *Rosmarinus officinalis* against *Culex pipiens* larvae. *J. Egypt. German Soc. Zool. E.* **48**, 69–80 (2005).
51. Chen, W., Wu, H., Ma, Z., Feng, J. & Zhang, X. Evaluation of fumigation activity of thirty-six essential oils against *Culex pipiens pallens* (Diptera: Culicidae). *Acta Entomol. Sin.* **61**, 86–93 (2018).
52. Makhaik, M., Naik, S. N. & Tewary, D. K. Evaluation of anti-mosquito properties of essential oils. (2005).
53. Jantan, I. B., Yalvema, M. F., Ahmad, N. W. & Jamal, J. A. Insecticidal activities of the leaf oils of eight cinnamomum species against *Aedes aegypti* and *Aedes albopictus*. *Pharm. Biol.* **43**, 526–532 (2005).
54. Khater, H. F. & Geden, C. J. Efficacy and repellency of some essential oils and their blends against larval and adult house flies, *Musca domestica* L. (Diptera: Muscidae). *J. Vector Ecol.* **44**, 256–263 (2019).
55. Levchenko, M. A., Silivanova, E. A., Khodakov, P. E. & Gholizadeh, S. Insecticidal efficacy of some essential oils against adults of *Musca domestica* L. (Diptera: Muscidae). *Int. J. Trop. Insect Sci.* 1–9 (2021).
56. Pushpalatha, E. & Viswan, K. A. Adulticidal and repellent activities of *Melaleuca leucadendron* (L.) and *Callistemon citrinus* (Curtis) against filarial and dengue vectors. *Assoc. Advancement Entomol.* **38**, 149–154 (2013).
57. Sahi, N. M. Evaluation of insecticidal activity of bioactive compounds from *Eucalyptus citriodora* against *Tribolium castaneum*. *Int. J. Pharm. Phytochem. Res.* **8**, 1256–1270 (2016).
58. Fu, J. *et al.* Fumigant toxicity and repellence activity of camphor essential oil from *Cinnamomum camphora* Siebold against *Solenopsis invicta* workers (Hymenoptera: Formicidae). *J. Insect Sci.* **15**, 129 (2015).
59. Zuhussnain, M. *et al.* Insecticidal and Genotoxic effects of some indigenous plant extracts in *Culex quinquefasciatus* Say Mosquitoes. *Sci. Rep.* **10**, 1–13 (2020).
60. Sutthanont, N. *et al.* Chemical composition and larvicidal activity of edible plant-derived essential oils against the pyrethroid-susceptible and-resistant strains of *Aedes aegypti* (Diptera: Culicidae). *J. Vector Ecol.* **35**, 106–115 (2010).
61. Ling Chang, C., Kyu Cho, I. & Li, Q. X. Insecticidal activity of basil oil, trans-anethole, estragole, and linalool to adult fruit flies of *Ceratitis capitata*, *Bactrocera dorsalis*, and *Bactrocera cucurbitae*. *J. Econ. Entomol.* **102**, 203–209 (2009).
62. da Silva, J. B. P. *et al.* Thiosemicarbazones as *Aedes aegypti* larvicidal. *Eur. J. Med. Chem.* **100**, 162–175 (2015).
63. Ali, A., Radwan, M. M., Wanas, A. S. & Khan, I. A. Repellent activity of carrot seed essential oil and its pure compound, carotol, against mosquitoes. *J. Am. Mosq. Control Assoc.* **34**, 272–280 (2018).
64. Branquinho, L. S. *et al.* Anti-inflammatory and toxicological evaluation of essential oil from *Piper glabratum* leaves. *J. Ethnopharmacol.* **198**, 372–378 (2017).
65. Sarma, R., Adhikari, K., Mahanta, S. & Khanikor, B. Combinations of plant essential oil based terpene compounds as larvicidal and adulticidal agent against *Aedes aegypti* (Diptera: Culicidae). *Sci. Rep.* **9**, 1–12 (2019).
66. Gad, M., Aref, S., Abdelhamid, A., Elwassimy, M. & Abdel-Raheem, S. Biologically active organic compounds as insect growth regulators (IGRs): Introduction, mode of action, and some synthetic methods. *Curr. Chem. Lett.* **10**, 393–412 (2021).

Acknowledgements

This work was funded by the Science, Technology, Innovation Funding Authority, Egypt, entitled: “Lumpy Skin Disease in Cattle and Development of Sustainable Pest Management Tools”, Project ID: 37024.

Author contributions

Conceptualization, A.A., A.M. and M.B.; methodology, H.K., M.B., I.R.; validation, M.B., I.R. and A.A.; formal analysis, A.A. and H.K.; resources, A.A.; writing—original draft preparation, M.B., I.R., H.K. and A.A.; writing—review and editing, H.K., A.A., A.M. and A.S.; supervision, H.K.; project administration, A.S.; funding acquisition, A.S. All authors have read and agreed to the published version of the manuscript.

Competing interests

The authors declare no competing interests.

Additional information

Correspondence and requests for materials should be addressed to A.S.

Reprints and permissions information is available at www.nature.com/reprints.

Publisher’s note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>.

© The Author(s) 2022