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RESEARCH ARTICLE

# The role of environmental enteric dysfunction in the pathogenesis of *Schistosoma mansoni*-associated morbidity in school-aged children

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### **Abstract**

# **Background**

Studies have implicated schistosomiasis as a cause of intestinal barrier disruption, a salient feature of environmental enteric dysfunction (EED), as eggs translocate from the sterile bloodstream through the gut wall. We examined the longitudinal impact of praziquantel (PZQ) treatment on a) EED biomarkers and b) Insulin growth factor I (IGF-1), a key driver of childhood linear growth, since EED has been implicated in linear growth stunting.

#### Methodology

290 children infected with *S. mansoni* in Brazil were treated with PZQ at baseline. EED biomarkers lipopolysaccharide (LPS) and intestinal fatty acid binding-protein (I-FABP) were measured, as well as IGF-1 at baseline, 6 and 12-months. Multivariate regression analysis was applied to assess associations between *S. mansoni* intensity and plasma biomarkers (LPS, I-FABP, and IGF-1), controlling for potential confounding variables.

#### **Principal findings**

At baseline, *S. mansoni* infection intensities were 27.2% light, 46.9% moderate, and 25.9% heavy. LPS concentrations were significantly reduced at the 12-month visit compared to baseline (p = 0.0002). No longitudinal changes were observed for I-FABP or IGF-1 in the 6-or 12-month periods following baseline treatment. After 6-months, I-FABP concentration was significantly higher in high vs low intensity (p = 0.0017). IGF-1 concentrations were significantly lower among children with high and moderate vs low intensity infections at each study visit.

participants that data will be made publicly available even if de identified. For this reason, we request an exemption from sharing the data publicly.

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**Competing interests:** The authors have declared that no competing interests exist.

# Conclusions/significance

We report that *S. mansoni* infection impacts LPS, I-FABP and IGF-1. These findings suggest a mechanistic role for EED in schistosomiasis-related morbidities, particularly linear growth.

# **Author summary**

Schistosoma mansoni is a tropical parasitic infection that causes intestinal schistosomiasis. In infected humans, the parasite worms shed eggs that migrate across the gut barrier, which damages intestinal structure and function. In children, intestinal schistosomiasis leads to anemia, undernutrition, and linear growth stunting. The mechanistic pathways between schistosomiasis and stunting are not fully understood, but this research explores the role of environmental enteric dysfunction (EED) in schistosomiasis-related morbidity. EED is an intestinal condition that affects children living in areas of poor water, sanitation, and hygiene and also leads to impaired growth and stunting. In a longitudinal cohort of Brazilian children infected with S. mansoni, we measured blood biomarkers of EED and linear growth at three time points over 12 months. All of the children were treated for schistosomiasis at baseline, and after 12 months, we observed a significant decrease in a marker of EED, suggesting improvement in gut integrity. We also found that children who had higher parasite egg burden at the baseline visit had lower levels of insulin-like growth factor-1, a hormone that drives growth in children. Our findings suggest that EED may play a role in schistosomiasis-related stunting and furthers our understanding for S. mansoni pathogenesis in children.

# **Background**

Human schistosomiasis, a Neglected Tropical Disease (NTD), affects over 140 million individuals and remains a significant cause of morbidity and mortality in developing countries, despite available drug treatment [1,2]. Two of the three main species affecting humans, *S. japonicum* and *S. mansoni*, live in the mesenteric venules and deposit thousands of eggs daily that migrate through vessel walls, the interstitium, and ultimately penetrate the gut wall to pass into the gut lumen for excretion in feces. If untreated, higher egg burden and longer duration of infection can lead to more severe morbidity, namely liver fibrosis and hepatosplenic disease, which in some cases culminates in portal hypertension and death [3]. Much of the global burden of disease impacts children, whereby schistosomiasis can cause anemia, undernutrition, linear growth stunting, and impaired neurocognitive development [4–10]. *S. mansoni* is endemic in Brazil, one of the few nations in the Americas that has not eliminated this NTD [11,12].

Schistosomiasis is a chronic inflammatory disease, resulting in the elaboration of proinflammatory cytokines detectable in the systemic circulation [13–15]. In the context of schistosomiasis, these inflammatory responses are implicated in the pathogenesis of anemia, undernutrition, and linear growth faltering [7,10,13–20]. Prolonged systemic inflammation in children has been linked to disruptions in the insulin-like growth factor (IGF) axis, which is a main regulator of linear growth [21]. Studies in children with chronic inflammatory diseases, both infectious and non-infectious in origin, have shown that increased concentrations of pro-inflammatory biomarkers and decreased concentrations of anabolic growth factors, such as insulin-like growth factor-1 (IGF-1), are associated with linear growth faltering [22–24]. Specific to *S. mansoni* infection, children experiencing hepatic fibrosis or hepatosplenic schistosomiasis had significantly lower IGF-1 concentrations compared to uninfected children [25,26].

Although there is a strong correlation between pro-inflammatory cytokines and schistosomiasis-related morbidity [7,14,20], the mechanisms by which schistosome infection results in the elaboration of pro-inflammatory cytokines and consequent morbidity remain understudied. Another driver of inflammatory responses is microbial translocation (MT), whereby luminal microbes cross the intestinal wall into the normally sterile bloodstream [27]. MT is a feature of environmental enteric dysfunction (EED), an acquired subclinical condition of the small intestine associated with impaired linear growth and chronic undernutrition in children exposed to conditions of poor water, sanitation, and hygiene [28,29]. MT is typically detected by the presence of serum lipopolysaccharide (LPS) or endotoxin, LPS binding protein (LBP), or circulating antibodies against LPS [30] in serum. There is increasing evidence linking schistosomiasis with MT. During schistosome infection, the passage of eggs into the gut lumen damages the integrity of the gut wall, which can enable MT into the bloodstream. Schistosome egg-induced MT may represent an important stimulus for pro-inflammatory responses during schistosome infection. Studies have shown higher concentrations of MT biomarkers among adults with schistosomiasis compared to uninfected individuals [31,32]. Further, in adult cases of hepatosplenic schistosomiasis, MT biomarkers were associated with systemic inflammation [33]. However, a recent report among adolescents in Kenya found that LPS was not correlated with schistosome egg burden and was not associated with infection intensity prior to treatment [34].

To our knowledge, studies have yet to examine the impact of schistosomiasis treatment on both gut health markers and IGF-1, a key promotor of linear growth in children. The present study enrolled children infected with *S. mansoni* in Brazil to examine a) the relationships between infection intensity (egg burden), gut health markers, and IGF-1 and b) the longitudinal impact of *S. mansoni* treatment [praziquantel (PZQ)] on gut health markers and IGF-1. We include gut health markers assessing both MT (LPS) and intestinal epithelial damage [intestinal fatty acid binding-protein (I-FABP)], another characteristic feature of EED.

#### **Methods**

#### **Ethics statement**

This study was approved by the Brazilian National Ethics Committee (CAAE 531.282). For each participant, formal written informed consent was obtained from a parent. Additionally, assent was obtained from participants  $\geq$  7 years old.

#### Study area, population, and design

This longitudinal study was conducted in the Jequitinhonha Valley in northern Minas Gerais State (MG), Brazil. Children with *S. mansoni* infection were recruited in schools from multiple communities within five different municipalities between March-December 2014. Participant recruitment and enrollment has been described previously [35]. Students and their parents or legal guardians were invited by members of the study team to participate in the parasitological screening. Students (n = 3,661) provided stool specimens and 20.4% (n = 750) were positive for *S. mansoni*. The eligibility criteria included males and females between the ages of 6–15 years with *S. mansoni* infection. Those who were pregnant or breastfeeding or who had symptoms of diarrhea were excluded. Enrolled participants attended baseline, 4-week, 6-month,

and 12-month study visits. Demographic and socioeconomic information was collected at the baseline visit using questionnaires as previously described [36,37]. The present analysis includes a sample of n=290 participants who met the inclusion criteria for the cohort study described above and who had available plasma for biomarker measurements (LPS, I-FABP, and IGF-1).

At the baseline visit, all participants received treatment for *S. mansoni* infection: a single oral dose of PZQ (60 mg/kg). Any participants with geohelminth infections at baseline were treated with a single oral dose of albendazole (400 mg). Four weeks following PZQ treatment, participants were re-assessed for *S. mansoni* infection and retreated if necessary. Re-infection of *S. mansoni* was assessed at the 12-month visit, and any positive cases were treated with PZQ. All treatments were administered with direct observation and under medical supervision as recommended by the Brazilian Ministry of Health [38]. At baseline, 6-month, and 12-month visits, blood samples were collected in vacuum blood collection tubes and transported to the laboratory at the Instituto René Rachou, FIOCRUZ for processing, and plasma fractions were stored in -80C freezers. Biomarkers (LPS, I-FABP, and IGF-1) were measured in plasma collected at baseline, 6-month, and 12-month visits.

#### Laboratory measures

S. mansoni infection was determined by microscopy using the Kato-Katz fecal thick smear method [39]. At screening, baseline, 4-week, and 12-month visits, participants provided stool samples collected on two separate days. Slides were prepared and assessed in duplicate for each stool sample within 24 hours of collection. For each visit, the slide concentrations of eggs per gram of stool (EPG) were averaged, and any value equal to or greater than 1 EPG was considered positive. For quality control, 10% of the slides were randomly selected and examined by a senior microscopist at the Instituto René Rachou, FIOCRUZ, in Belo Horizonte, MG. Plasma LPS was quantified using the Pierce LAL Chromogenic Endotoxin Quantitation Kit (Thermo Fisher Scientific, Waltham, MA, USA). Plasma I-FABP and IGF-1 concentrations were measured by enzyme-linked immunosorbent assay (ELISA) using kits from R&D Systems (Human FABP2/I-FABP DuoSet ELISA #DY3078, Human IGF-I/IGF-1 Quantikine ELISA #DG100B). Biomarker measurements were conducted at the Instituto René Rachou, FIO-CRUZ in Brazil.

#### Statistical analysis

S. mansoni intensity is reported in eggs per gram of stool (EPG) and intensity categories are defined as light (1–99 EPG), moderate (100–399 EPG), and heavy (≥400 EPG) [39]. Weightfor-age z-score (WAZ) was calculated using the CDC growth chart reference for ages 0 to <20 years, which considers age, weight, and sex. Socioeconomic status (SES) categories were determined using methods designed by Gwatkin, et al. [40] and described previously [35]. Values are reported in median and interquartile range (IQR) unless otherwise stated. Multivariate regression analysis was applied to assess associations between plasma biomarker concentrations and baseline S. mansoni intensity category and changes in plasma biomarker concentrations across study visits. Fully adjusted models applied automated stepwise selection with entry and stay thresholds of 0.1 to consider potential confounding variables including age, sex, WAZ, SES, and hookworm infection status. Continuous variables included in regression models were natural log-transformed. P values < 0.05 were considered significant. Statistical analyses were conducted using SAS Studio 3.8 (SAS Institute Inc., Cary, NC).

#### Results

## **Population**

Characteristics for the 290 participants are described in Table 1. The median age was 12.2 years (10.0–13.8 IQR), and 57.2% were male. The median *S. mansoni* intensity was 192 EPG (90–516 IQR), and the proportions in light, moderate, and heavy intensity categories were 27.2%, 46.9%, and 25.9%, respectively. Of the 290 participants treated for *S. mansoni* infection at baseline, 46 (16.2%) were re-infected at the 12-month visit. Analyses pertaining to the 12-month visit were also run separately excluding those 46 participants who were re-infected, and none of the significant findings were impacted.

#### LPS

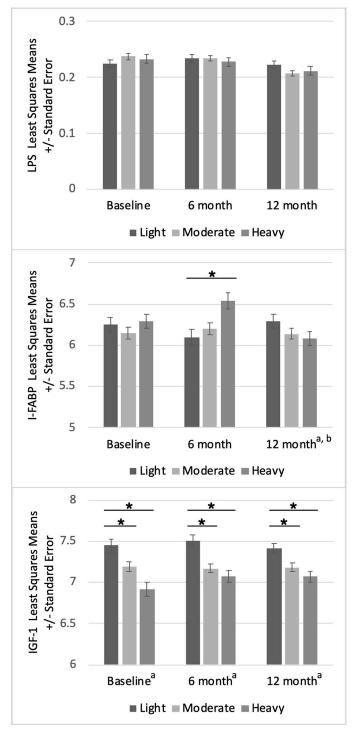
LPS concentration did not differ by *S. mansoni* intensity category at baseline nor at the 6- and 12-month follow-up visits (Fig 1, S1 Table). Following PZQ treatment administered during

Table 1. Participant characteristics at baseline, N = 290

12.2 (10.0–13.8) 166 (57.2) 36.0 (29.0–47.9) -0.55 (-1.27, 0.21) 83 (29.3) 47 (16.6) 70 (24.7)	
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70 (24.7)	
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18 (6.2)	
30 (10.3)	
40 (13.8)	
38 (13.1)	
21 (7.2)	
39 (13.4)	
26 (9.0)	
78 (26.9)	
192 (90–516)	
79 (27.2)	
136 (46.9)	
75 (25.9)	
32 (11.3)	
0.26 (0.20-0.31)	
524.7 (283.3–865.5)	
1,674.6 (969.4–2,259.7)	

 $EPG, eggs\ per\ gram\ of\ stool; LPS, lipopolysaccharide; I-FABP, intestinal\ fatty\ acid\ binding\ protein; IGF-1, insulin-like\ growth\ factor\ 1$ 

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**Fig 1. Biomarkers by baseline** *S. mansoni* **intensity.** Least squares means and standard errors for lipopolysaccharide (LPS, EU/mL), intestinal fatty acid binding protein (I-FABP, pg/mL), and insulin-like growth factor 1 (IGF-1, pg/mL) concentrations at each visit (baseline, 6-months, and 12-months) by baseline *S. mansoni* intensity category. Asterisks represent significant differences determined by multivariate regression with light infection intensity as the reference category. Stepwise selection retained variables of a) age and b) WAZ. Continuous variables were natural log-transformed.

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the baseline visit, LPS concentrations were significantly reduced at the 12-month visit compared to the baseline (Fig 2, S2 Table).

#### **I-FABP**

There were no differences in I-FABP concentrations by infection intensity category at the baseline or 12-month visits. At the 6-month visit, the I-FABP concentration was significantly higher for those in the heavy intensity category compared to the light intensity category (Fig 1, S1 Table). I-FABP concentrations measured at the 6 and 12 month follow-up visits were not significantly different from the baseline measures (Fig 2, S2 Table).

#### IGF-1

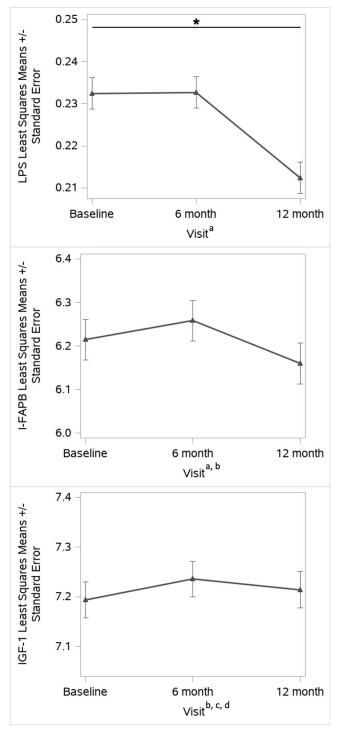
IGF-1 concentrations were significantly lower for those with moderate and heavy infection intensity compared to light infection intensity at baseline, 6-month, and 12-month visits (Fig 1, S1 Table). There were no significant differences in IGF-1 concentrations measured at the 6 and 12-month visits compared to the baseline visit (Fig 2, S2 Table). In multivariate regression models adjusting for age, IGF-1 concentrations were positively correlated with LPS concentrations at the 6-month (p = 0.0218) and 12- month visits (p = 0.0445), but not at baseline (p = 0.1105). In similar regression models adjusting for age, IGF-1 concentrations were not correlated with I-FABP concentrations at any visit (Table 2).

#### **Discussion**

To better understand the mechanistic pathway between childhood schistosomiasis and associated morbidities, such as impaired linear growth, this longitudinal cohort study sought to examine the relationships between *S. mansoni* infection and markers of gut health and the IGF axis as one potential pathway. The results demonstrate that *S. mansoni* infection impacts EED biomarkers, LPS and I-FABP, as well as IGF-1.

Compared to light infection intensity at baseline, participants with heavy or moderate infection intensity had significantly lower concentrations of IGF-1 measured at baseline, 6-month, and 12-month visits. These age-adjusted findings contribute evidence that schistosomiasis egg burden is associated with disruptions in the IGF axis, with long-term impacts during the 12 months following PZQ treatment. While the evidence is limited, previous case-control studies have demonstrated associations between severe forms of disease–hepatic fibrosis or hepatosplenic schistosomiasis—and decreased IGF-1 concentrations compared to control groups [25,26]. However, this is the first study to demonstrate an impact of schistosome egg burden on IGF-1 in the absence of overt clinical disease. These findings suggest that intensity of schistosomiasis infection leads to disruption in the IGF axis, which contributes to our understanding of at least one mechanism underlying schistosomiasis-related morbidities, specifically impaired linear growth in children.

There is strong evidence to support that chronic pro-inflammatory responses result in impaired linear growth for children experiencing schistosomiasis or EED [14,21,23]. Cytokines characteristic of a pro-inflammatory or T helper type-1 (Th1) mediated response disrupt the IGF axis and culminate in impaired growth. This has been demonstrated in chronic inflammatory childhood diseases, such as inflammatory bowel disease and chronic kidney disease, whereby increased inflammatory biomarkers and decreased anabolic growth factors like IGF-1 are associated with linear growth faltering, as well as in insults of bacteremia from MT [21,24]. Previous studies have shown that the MT biomarker LPS negatively impacts IGF-1 [41,42], however, in this cohort, LPS concentrations were positively correlated with IGF-1 six and 12 months after PZQ treatment, and not at baseline during active infection. In other



**Fig 2. Biomarkers across visits.** Least squares means and standard errors for lipopolysaccharide (LPS, EU/mL), intestinal fatty acid binding protein (I-FABP, pg/mL), and insulin-like growth factor 1 (IGF-1, pg/mL) concentrations across visits (baseline, 6-months, and 12-months). Asterisks represent significant differences determined by multivariate regression with the baseline visit as the reference category. Stepwise selection retained variables of a) WAZ, b) age, c) sex, and d) S. *mansoni* infection intensity category. Continuous variables were natural log-transformed.

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Table 2. Linear regression of IGF-1 concentrations by EED biomarkers

	Univariate		Multivariate	
	ß	P value	ß	P value <sup>a</sup>
LPS (EU/mL)				
Baseline	1.0032	0.1399	1.0526	0.1105
6-month	1.4388	0.0248	1.4454	0.0218
12-month	1.0588	0.0497	1.0650	0.0445
I-FABP (pg/mL)				
Baseline	-0.0544	0.3552	-0.0196	0.7331
6-month	-0.0914	0.0387	-0.0685	0.1200
12-month	-0.0272	0.5464	-0.0078	0.8620

LPS, lipopolysaccharide; I-FABP, intestinal fatty acid binding protein; IGF-1, insulin-like growth factor 1. Continuous variables were natural log-transformed. All continuous variables natural log-transformed.

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studies, a negative relationship between LPS and IGF-1 has been shown to be mediated by polarized Th1/Th2 responses [43,44], and additional measures of associated cytokines would offer a more comprehensive understanding of the MT and IGF axis associations in this population. Given that schistosomiasis is a chronic inflammatory disease affecting children, future research ought to target the mechanistic roles of inflammation and the IGF axis as they relate to childhood morbidities, such as growth faltering.

Our findings demonstrate the impact of *S. mansoni* infection on LPS. In adjusted regression models, LPS concentrations were significantly lower 12 months following PZQ treatment compared to baseline. Similar studies evaluating MT biomarkers following PZQ treatment for schistosomiasis showed no significant impact on LPS 6 months or 9 months after treatment [45,46]. Our results suggest an even longer timeframe required for gut barrier healing following the cessation of schistosome eggs traversing the gut wall.

The EED biomarker I-FABP is an intracellular enterocyte protein released into circulation following injury to the gut epithelia [47,48]. To our knowledge, only one previous study has examined I-FABP in the context of schistosomiasis [49]. In that study, adult fishermen in Kenya with HIV infection, and some of whom were infected with *S. mansoni*, were treated with antiretroviral therapy and PZQ at baseline as needed. I-FABP was measured at baseline, 2 weeks, 1 month, and 3 months post treatment. They found that I-FABP concentrations among participants with *S. mansoni* infection did not differ between visits and were not associated with *S. mansoni* EPG at any visit. However, those cases had relatively low infection intensity compared to the current analysis. We observed associations between infection intensity category and I-FABP concentrations 6 months following PZQ treatment. Concentrations of I-FABP were significantly higher for participants with high egg burden compared to low egg burden. Higher I-FABP concentrations related to higher egg burden at the 6-month visit may be explained by residual eggs lodged in the tissue between the mesentery and the luminal barrier and associated granulomatous inflammation, requiring time to more fully resolve [50,51].

Because 16.2% of participants were found to be re-infected with *S. mansoni* infection at the 12-month follow-up visit, we re-ran the regression models pertaining to the 12-month visit and excluded any participants who had become re-infected. Excluding these re-infected

a) Stepwise selection considered variables of age, sex, WAZ, SES, and hookworm infection status. Of these, only age was retained and included in all multivariate regression models.

participants had no impact on the significant findings reported in the results section, and no additional significant associations emerged.

The present findings offer evidence to support relationships between *S. mansoni* infection and both EED and the IGF axis and further our understanding of the mechanistic pathways between schistosomiasis and associated morbidities. The analysis of these mechanisms would be strengthened by measuring circulating immunologic response markers (Th1 and Th2 cytokines) at each visit as well as collecting measures on schistosomiasis-specific morbidities (anemia, linear growth, and hepatic fibrosis). Additionally, the findings would benefit from enrolling participants without baseline *S. mansoni* infection who represent similar geographic and demographic characteristics. Given the significant overlapping global burden of schistosomiasis and EED, it is essential that we elucidate the immunopathology of these illnesses in order to prevent long-term morbidities in affected populations.

# **Supporting information**

**S1 File. STROBE Statement.** Checklist of items that should be included in reports of cohort studies.

(DOCX)

(DOCX)

**S1 Table. Linear regression of biomarkers by baseline** *S. mansoni* **infection intensity category.** These findings are reflected in Fig 1. All continuous variables natural log-transformed. Stepwise selection variables for multivariate regression included a) age, b) weight-for-age z-score. LPS, lipopolysaccharide; I-FABP, intestinal fatty acid binding protein; IGF-1, insulinlike growth factor 1.

**S2 Table. Linear regression of biomarkers by study visit.** These findings are reflected in Fig 2. All continuous variables natural log-transformed. Stepwise selection variables for multivariate regression included a) weight-for-age z-score, b) age, c) sex, d) *S. mansoni* intensity category. LPS, lipopolysaccharide; I-FABP, intestinal fatty acid binding protein; IGF-1, insulin-like growth factor 1.

(DOCX)

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#### **Author Contributions**

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**Methodology:** Jacqueline Araújo Fiuza, Letícia Gambogi de Ornellas, Leonardo Ferreira Matoso, Andrea Gazzinelli.

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Visualization: Susannah Colt.

Writing - original draft: Jacqueline Araújo Fiuza, Susannah Colt.

Writing – review & editing: Leonardo Ferreira Matoso, Andrea Gazzinelli, Jennifer F. Friedman, Rodrigo Corrêa-Oliveira.

#### References

- GBD Disease Injury Incidence Prevalence Collaborators. Global, regional, and national incidence, prevalence, and years lived with disability for 354 diseases and injuries for 195 countries and territories, 1990–2017: a systematic analysis for the Global Burden of Disease Study 2017. Lancet. 2018; 392 (10159):1789–858. Epub 2018/11/30. <a href="https://doi.org/10.1016/S0140-6736(18)32279-7">https://doi.org/10.1016/S0140-6736(18)32279-7</a> PMID: 30496104.
- Friedman JF. Optimizing Delivery of Mass Drug Administration for Schistosomiasis. Am J Trop Med Hyg. 2019; 101(6):1191–2. Epub 2019/11/02. https://doi.org/10.4269/ajtmh.19-0762 PMID: 31674294.
- Colley DG, Bustinduy AL, Secor WE, King CH. Human schistosomiasis. The Lancet. 2014; 383 (9936):2253–64. https://doi.org/10.1016/s0140-6736(13)61949-2 PMID: 24698483
- King CH, Dickman K, Tisch DJ. Reassessment of the cost of chronic helmintic infection: a meta-analysis
  of disability-related outcomes in endemic schistosomiasis. Lancet. 2005; 365(9470):1561–9. Epub
  2005/05/04. https://doi.org/10.1016/S0140-6736(05)66457-4 PMID: 15866310.
- Bustinduy AL, Parraga IM, Thomas CL, Mungai PL, Mutuku F, Muchiri EM, et al. Impact of polyparasitic infections on anemia and undernutrition among Kenyan children living in a Schistosoma haematobiumendemic area. Am J Trop Med Hyg. 2013; 88(3):433–40. Epub 2013/01/18. https://doi.org/10.4269/ ajtmh.12-0552 PMID: 23324217.
- Coutinho HM, Acosta LP, McGarvey ST, Jarilla B, Jiz M, Pablo A, et al. Nutritional status improves after treatment of schistosoma japonicum-infected children and adolescents. J Nutr. 2006; 136(1):183–8.
   Epub 2005/12/21. https://doi.org/10.1093/jn/136.1.183 PMID: 16365080.
- Coutinho HM, McGarvey ST, Acosta LP, Manalo DL, Langdon GC, Leenstra T, et al. Nutritional status and serum cytokine profiles in children, adolescents, and young adults with Schistosoma japonicumassociated hepatic fibrosis, in Leyte, Philippines. The Journal of infectious diseases. 2005; 192(3):528– 36. https://doi.org/10.1086/430929 PMID: 15995969.
- 8. Ezeamama AE, Friedman JF, Acosta LP, Bellinger DC, Langdon GC, Manalo DL, et al. Helminth infection and cognitive impairment among Filipino children. Am J Trop Med Hyg. 2005; 72(5):540–8. Epub 2005/05/14. PMID: 15891127.
- Friedman JF, Kanzaria HK, McGarvey ST. Human schistosomiasis and anemia: the relationship and potential mechanisms. Trends Parasitol. 2005; 21(8):386–92. https://doi.org/10.1016/j.pt.2005.06.006 PMID: 15967725.
- Leenstra T, Acosta LP, Langdon GC, Manalo DL, Su L, Olveda RM, et al. Schistosomiasis japonica, anemia, and iron status in children, adolescents, and young adults in Leyte, Philippines 1. The American journal of clinical nutrition. 2006; 83(2):371–9. https://doi.org/10.1093/ajcn/83.2.371 PMID: 16469997.
- Bezerra DVF, Queiroz JW, Camara VAV, Maciel BLL, Nascimento ELT, Jeronimo SMB. Factors Associated with Schistosoma mansoni Infestation in Northeast Brazil: A Need to Revisit Individual and Community Risk Factors. Am J Trop Med Hyg. 2021. Epub 2021/02/17. https://doi.org/10.4269/ajtmh.19-0513 PMID: 33591939.
- Nascimento GL, Pegado HM, Domingues ALC, Ximenes RAA, Itria A, Cruz LN, et al. The cost of a disease targeted for elimination in Brazil: the case of schistosomiasis mansoni. Mem Inst Oswaldo Cruz. 2019; 114:e180347. Epub 2019/01/18. https://doi.org/10.1590/0074-02760180347 PMID: 30652735.
- **13.** Abdel-Aaty HE, Selim MM, Abdel-Rehim HA. Study of gamma-interferon in schistosomiasis mansoni, autoimmune diseases and schistosomal arthropathy. J Egypt Soc Parasitol. 1999; 29(3):721–34. Epub 2003/02/04. PMID: 12561913.
- Coutinho HM, Leenstra T, Acosta LP, Su L, Jarilla B, Jiz MA, et al. Pro-inflammatory cytokines and C-reactive protein are associated with undernutrition in the context of Schistosoma japonicum infection.
   Am J Trop Med Hyg. 2006; 75(4):720–6. PMID: 17038701.
- 15. Mwatha JK, Kimani G, Kamau T, Mbugua GG, Ouma JH, Mumo J, et al. High levels of TNF, soluble TNF receptors, soluble ICAM-1, and IFN-gamma, but low levels of IL-5, are associated with hepatosple-nic disease in human schistosomiasis mansoni. Journal of immunology (Baltimore, Md: 1950). 1998; 160(4):1992–9. Epub 1998/02/20. 9469463.

- Zwingenberger K, Irschick E, Vergetti Sigueira J, Correia Dacal A, Feldmeier H. Tumor nucrosis factor in hepatosplenic schistosomiasis. Scandinavian Journal of Immunology. 1990; 31:205–11.
- Abdel Azim A, Sedky HA, el-Tahawy MA, Fikry AA, Mostafa H. Serum levels of tumor necrosis factor in different stages of schistosomal infection. J Egypt Soc Parasitol. 1995; 25(1):279–87. Epub 1995/04/ 01. PMID: 7602170.
- de Jesus AR, Silva A, Santana LB, Magalhaes A, de Jesus AA, de Almeida RP, et al. Clinical and immunologic evaluation of 31 patients with acute schistosomiasis mansoni. The Journal of infectious diseases. 2002; 185(1):98–105. Epub 2002/01/05. https://doi.org/10.1086/324668 PMID: 11756987.
- Khalil HM, el-Missiry AG, Abdalla HM, Khalil NM, Sabry NM, Abdel-Atty HE, et al. Serum levels of tumour necrosis factor- alpha in schistosomiasis mansoni and their analogous changes in collagen diseases and schistosomal arthropathy. J Egypt Soc Parasitol. 1995; 25(2):427–36. Epub 1995/08/01. PMID: 7665938.
- Leenstra T, Coutinho HM, Acosta LP, Langdon GC, Su L, Olveda RM, et al. Schistosoma japonicum reinfection after praziquantel treatment causes anemia associated with inflammation. Infection and immunity. 2006; 74(11):6398–407. https://doi.org/10.1128/IAI.00757-06 PMID: 16923790.
- Wong SC, Dobie R, Altowati MA, Werther GA, Farquharson C, Ahmed SF. Growth and the Growth Hormone-Insulin Like Growth Factor 1 Axis in Children With Chronic Inflammation: Current Evidence, Gaps in Knowledge, and Future Directions. Endocr Rev. 2016; 37(1):62–110. Epub 2016/01/01. https://doi.org/10.1210/er.2015-1026 PMID: 26720129.
- Cirillo F, Lazzeroni P, Sartori C, Street ME. Inflammatory Diseases and Growth: Effects on the GH-IGF Axis and on Growth Plate. Int J Mol Sci. 2017; 18(9). Epub 2017/09/01. <a href="https://doi.org/10.3390/ijms18091878">https://doi.org/10.3390/ijms18091878</a> PMID: 28858208.
- DeBoer MD, Scharf RJ, Leite AM, Férrer A, Havt A, Pinkerton R, et al. Systemic inflammation, growth factors, and linear growth in the setting of infection and malnutrition. Nutrition. 2017; 33:248–53. Epub 2016/10/08. https://doi.org/10.1016/j.nut.2016.06.013 PMID: 27712965.
- Syed S, Manji KP, McDonald CM, Kisenge R, Aboud S, Sudfeld C, et al. Biomarkers of Systemic Inflammation and Growth in Early Infancy are Associated with Stunting in Young Tanzanian Children. Nutrients. 2018; 10(9). Epub 2018/08/29. https://doi.org/10.3390/nu10091158 PMID: 30149537.
- 25. Hassan AH, Abd el Moneim MA, Abd el Aal AA, Abou Aly SA, Ahmed SH, Soliman AT, et al. Circulating growth hormone, insulin-like growth factor I, cortisol and free thyroxine in children with schistosomiasis with and without hepatic fibrosis. J Trop Pediatr. 1991; 37(1):25–30. Epub 1991/02/01. https://doi.org/10.1093/tropej/37.1.25 PMID: 2023299.
- Orsini M, Rocha RS, Disch J, Katz N, Rabello A. The role of nutritional status and insulin-like growth factor in reduced physical growth in hepatosplenic Schistosoma mansoni infection. Transactions of the Royal Society of Tropical Medicine and Hygiene. 2001; 95(4):453–6. Epub 2001/10/03. https://doi.org/10.1016/s0035-9203(01)90213-5 PMID: 11579895.
- Prendergast A, Kelly P. Enteropathies in the developing world: neglected effects on global health. Am J Trop Med Hyg. 2012; 86(5):756–63. Epub 2012/05/05. https://doi.org/10.4269/ajtmh.2012.11-0743 PMID: 22556071.
- Keusch GT, Denno DM, Black RE, Duggan C, Guerrant RL, Lavery JV, et al. Environmental enteric dysfunction: pathogenesis, diagnosis, and clinical consequences. Clinical infectious diseases: an official publication of the Infectious Diseases Society of America. 2014; 59 Suppl 4(Suppl 4):S207–12. Epub 2014/10/12. https://doi.org/10.1093/cid/ciu485 PMID: 25305288.
- Korpe PS, Petri WA Environmental enteropathy: critical implications of a poorly understood condition. Trends Mol Med. 2012; 18(6):328–36. Epub 2012/05/29. https://doi.org/10.1016/j.molmed.2012.04.007 PMID: 22633998.
- Brenchley JM, Douek DC. Microbial translocation across the GI tract. Annual review of immunology. 2012; 30:149–73. Epub 2012/01/10. <a href="https://doi.org/10.1146/annurev-immunol-020711-075001">https://doi.org/10.1146/annurev-immunol-020711-075001</a> PMID: 22224779.
- Onguru D, Liang Y, Griffith Q, Nikolajczyk B, Mwinzi P, Ganley-Leal L. Human schistosomiasis is associated with endotoxemia and Toll-like receptor 2- and 4-bearing B cells. Am J Trop Med Hyg. 2011; 84(2):321–4. Epub 2011/02/05. https://doi.org/10.4269/ajtmh.2011.10-0397 PMID: 21292908.
- Sinkala E, Kapulu MC, Besa E, Zyambo K, Chisoso NJ, Foster GR, et al. Hepatosplenic schistosomiasis is characterised by high blood markers of translocation, inflammation and fibrosis. Liver Int. 2016; 36(1):145–50. Epub 2015/06/11. https://doi.org/10.1111/liv.12891 PMID: 26058680.
- Kaonga P, Kaimoyo E, Besa E, Zyambo K, Sinkala E, Kelly P. Direct Biomarkers of Microbial Translocation Correlate with Immune Activation in Adult Zambians with Environmental Enteropathy and Hepatosplenic Schistosomiasis. Am J Trop Med Hyg. 2017; 97(5):1603–10. Epub 2017/11/16. https://doi.org/10.4269/ajtmh.17-0365 PMID: 29140241.

- Ondigo BN, Hamilton RE, Magomere EO, Onkanga IO, Mwinzi PN, Odiere MR, et al. Potential Utility of Systemic Plasma Biomarkers for Evaluation of Pediatric Schistosomiasis in Western Kenya. Frontiers in immunology. 2022; 13:887213. Epub 2022/05/24. <a href="https://doi.org/10.3389/fimmu.2022.887213">https://doi.org/10.3389/fimmu.2022.887213</a> PMID: 35603171.
- Gazzinelli A, Oliveira-Prado R, Matoso LF, Veloso BM, Andrade G, Kloos H, et al. Schistosoma mansoni reinfection: Analysis of risk factors by classification and regression tree (CART) modeling. PLoS One. 2017; 12(8):e0182197. Epub 2017/08/17. <a href="https://doi.org/10.1371/journal.pone.0182197">https://doi.org/10.1371/journal.pone.0182197</a> PMID: 28813451.
- Gazzinelli A, Bethony J, Fraga LA, LoVerde PT, Correa-Oliveira R, Kloos H. Exposure to Schistosoma mansoni infection in a rural area of Brazil. I: water contact. Trop Med Int Health. 2001; 6(2):126–35. Epub 2001/03/17. https://doi.org/10.1046/j.1365-3156.2001.00684.x PMID: 11251909.
- Gazzinelli A, Velasquez-Melendez G, Crawford SB, LoVerde PT, Correa-Oliveira R, Kloos H. Socioeconomic determinants of schistosomiasis in a poor rural area in Brazil. Acta Trop. 2006; 99(2–3):260–71.
   Epub 2006/10/19. https://doi.org/10.1016/j.actatropica.2006.09.001 PMID: 17045559.
- 38. Brasil Ministério da Saúde. In Vigilância em Saúde: Dengue, Esquistossomose, Hanseníase, Malária, Tracoma e Tuberculose. Cadernos de Atenção Básica n° 21. 2nd ed 2008. p. 48–65.
- World Health Organization. Prevention and control of schistosomiasis and soil-transmitted helminthiasis: report of a WHO expert committee. Geneva: 2002 Contract No.: 912.
- Gwatkin DR, Rustein S, Johnson K, Pande RP, Wagstaff A. Socio-economic differences in health, nutrition, and population in Brazil. Washington, D. C.: World Bank Group, 2000
- Fan J, Char D, Kolasa AJ, Pan W, Maitra SR, Patlak CS, et al. Alterations in hepatic production and peripheral clearance of IGF-I after endotoxin. Am J Physiol. 1995; 269(1 Pt 1):E33–42. Epub 1995/07/ 01. https://doi.org/10.1152/ajpendo.1995.269.1.E33 PMID: 7543247.
- Lang CH, Fan J, Wojnar MM, Vary TC, Cooney R. Role of central IL-1 in regulating peripheral IGF-I during endotoxemia and sepsis. Am J Physiol. 1998; 274(4):R956–62. Epub 1998/05/12. <a href="https://doi.org/10.1152/ajpregu.1998.274.4.R956">https://doi.org/10.1152/ajpregu.1998.274.4.R956</a> PMID: 9575956.
- Faim F, Passaglia P, Batalhao M, Lacchini R, Stabile AM, Carnio EC. Role of ghrelin on growth hormone/insulin-like growth factor-1 axis during endotoxemia. Growth Horm IGF Res. 2019; 48–49:36–44. Epub 2019/09/09. https://doi.org/10.1016/j.qhir.2019.08.004 PMID: 31494533.
- 44. Priego T, Ibáñez de Cáceres I, Martín AI, Villanúa MA, López-Calderón A. NO plays a role in LPS-induced decreases in circulating IGF-I and IGFBP-3 and their gene expression in the liver. Am J Physiol Endocrinol Metab. 2004; 286(1):E50–6. Epub 2003/09/18. <a href="https://doi.org/10.1152/ajpendo.00149.2003">https://doi.org/10.1152/ajpendo.00149.2003</a> PMID: 13129855.
- 45. Klemperer KM, Reust MJ, Lee MH, Corstjens P, van Dam GJ, Mazigo HD, et al. Plasma Endotoxin Levels Are Not Increased in Schistosoma mansoni-Infected Women without Signs or Symptoms of Hepatosplenic Disease. Am J Trop Med Hyg. 2020; 102(6):1382–5. Epub 2020/03/04. https://doi.org/10.4269/ajtmh.19-0875 PMID: 32124718.
- 46. McDonald EA, Stuart R, Joshi A, Wu HW, Olveda RM, Acosta LP, et al. Endotoxin at the Maternal-Fetal Interface in a Resource-Constrained Setting: Risk Factors and Associated Birth Outcomes. Am J Trop Med Hyg. 2018; 99(2):495–501. https://doi.org/10.4269/ajtmh.17-0949 PMID: 29968554.
- 47. Adriaanse MP, Tack GJ, Passos VL, Damoiseaux JG, Schreurs MW, van Wijck K, et al. Serum I-FABP as marker for enterocyte damage in coeliac disease and its relation to villous atrophy and circulating autoantibodies. Aliment Pharmacol Ther. 2013; 37(4):482–90. Epub 2013/01/08. https://doi.org/10.1111/apt.12194 PMID: 23289539.
- Vreugdenhil AC, Wolters VM, Adriaanse MP, Van den Neucker AM, van Bijnen AA, Houwen R, et al. Additional value of serum I-FABP levels for evaluating celiac disease activity in children. Scand J Gastroenterol. 2011; 46(12):1435–41. Epub 2011/10/28. <a href="https://doi.org/10.3109/00365521.2011.627447">https://doi.org/10.3109/00365521.2011.627447</a> PMID: 22029621.
- Goovaerts O, Mwinzi PNM, Muok EMO, Ceulemans A, Colebunders R, Kestens L. Aberrant plasma MMP and TIMP dynamics in Schistosoma—Immune reconstitution inflammatory syndrome (IRIS). PLoS neglected tropical diseases. 2018; 12(8):e0006710. Epub 2018/08/09. <a href="https://doi.org/10.1371/journal.pntd.0006710">https://doi.org/10.1371/journal.pntd.0006710</a> PMID: 30089120.
- Gryseels B, Polman K, Clerinx J, Kestens L. Human schistosomiasis. The Lancet. 2006; 368 (9541):1106–18. https://doi.org/10.1016/S0140-6736(06)69440-3 PMID: 16997665
- Costain AH, MacDonald AS, Smits HH. Schistosome Egg Migration: Mechanisms, Pathogenesis and Host Immune Responses. Frontiers in immunology. 2018; 9:3042. Epub 2019/01/09. <a href="https://doi.org/10.3389/fimmu.2018.03042">https://doi.org/10.3389/fimmu.2018.03042</a> PMID: 30619372.