Influenza Viruses: Breaking All the Rules

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ABSTRACT Influenza A viruses (IAV) are significant pathogens able to repeatedly switch hosts to infect multiple avian and mammalian species, including humans. The unpredictability of IAV evolution and interspecies movement creates continual public health challenges, such as the emergence of the 2009 pandemic H1N1 virus from swine, as well as pandemic threats from the ongoing H5N1 and the recent H7N9 epizootics. In the last decade there has been increased concern about the "dual use" nature of microbiology, and a set of guidelines covering "dual use research of concern" includes seven categories of potentially problematic scientific experiments. In this Perspective, we consider how in nature IAV continually undergo "dual use experiments" as a matter of evolution and selection, and we conclude that studying these properties of IAV is critical for mitigating and preventing future epidemics and pandemics.

nfluenza A viruses (IAV) are important pathogens of humans and animals that cause continually recurring epizootics, annual epidemics, and periodic pandemics. As single-stranded segmented RNA viruses of the family Orthomyxoviridae, IAV show antigenic diversity that reflects 17 different hemagglutinin (HA) and ten neuraminidase (NA) surface protein subtypes. Multiple HA-NA subtype combinations and genotypes are generated through mixed infection and reassortment, and an error-prone RNA-dependent RNA polymerase generates complex viral quasispecies, from which variants can be rapidly selected under such evolutionary pressures as exposure to new host species, host immunity, and antiviral drugs. IAV host barriers are relatively weak, allowing viruses to repeatedly switch hosts to infect multiple avian and mammalian species. The unpredictability of IAV evolution and interspecies movement creates continual public health challenges (1, 2).

All avian and mammalian IAV are believed to have descended from avian influenza viruses resident in an enormous global avian virus gene pool. These viruses infect hundreds of wild bird species and undergo frequent reassortment (3). Occasionally, viruses from this avian pool switch hosts to infect either domestic poultry or mammals. The mechanisms of host switching and stable adaptation to new host populations are incompletely understood (4). In 1997, highly pathogenic avian influenza A viruses (HPAI) of the H5N1 subtype in Asia escaped the wild avian gene pool and became adapted to domestic poultry. These H5N1 viruses have circulated enzootically in domestic poultry for over 15 years, spreading to the Middle East, Africa, and Europe (5, 6). Spillover from poultry to humans has caused 630 documented human infections with 375 fatalities (as of 4 June 2013; WHO) (7), without adaptation to or enhanced transmissibility between humans. (It should be noted that poultry-adapted IAV rarely infect humans and that the underlying basis and mechanisms of these rare but severe cases of human H5N1 infections are unknown). More recently (starting in February 2013), a novel H7N9 virus, presumably also from a domestic poultry source, has caused 132 documented human infections with 37 fatalities (as of 7 June 2013) (8). Whether this virus will adapt to efficient human-to-human transmissibility or infect other mammals is unknown (9-11).

Over the last decade, there has been increased concern about the "dual use" nature of microbiology, including concerns about misapplication of scientific investigation for bioterrorism or bio-

weapons development and concerns about accidental release of consequential pathogens from laboratories in which they are being studied or stored. Recent experiments with strains of HPAI H5N1 engineered to enhance ferret and guinea pig transmissibility have raised additional biosafety concerns (12-17). As a result, in 2012, the U.S. government established policies for the oversight of "dual use research of concern" (DURC) relating to fifteen "high-consequence" pathogens and toxins, two of which are IAV: HPAI H5N1 and the reconstructed 1918 pandemic influenza virus (18). DURC was defined as "life sciences research that, based on current understanding, can be reasonably anticipated to provide knowledge, information, products, or technologies that could be directly misapplied to pose a significant threat with broad potential consequences to public health and safety, agricultural crops and other plants, animals, the environment, materiel, or national security" (18). DURC policy (here referred to as "dual use" policy) is defined as including seven categories of potentially problematic scientific experiments as follows:

- (i) enhancing the harmful consequences of an agent or toxin
- (ii) disrupting immunity or the effectiveness of an immunization against an agent or toxin without clinical or agricultural justification
- (iii) conferring to an agent or toxin resistance to clinically or agriculturally useful prophylactic or therapeutic interventions against the agent or toxin, or facilitating its ability to evade detection methodologies
- (iv) increasing the stability or transmissibility of or the ability to disseminate an agent or toxin
- (v) altering the host range or tropism of an agent or toxin
- (vi) enhancing the susceptibility of a host population to an agent or toxin

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(vii) generating or reconstituting an eradicated or extinct agent or toxin

In this Perspective, we consider how in nature IAV continually undergo such "dual use experiments" as a matter of evolution and selection and how such viral evolution determines the consequent capacity of IAV to cause harm. These considerations lead us to conclude that scientifically studying such fundamental evolutionary properties of IAV is critical for mitigating and preventing the devastation these viruses cause. We do not imply that all IAV evolution in nature should be considered dual use but rather hope that consideration of the dynamic nature of IAV biology will inform policy decisions.

Naturally occurring IAV routinely break the enumerated dual use rules. (i) Enhancing the harmful consequences of an agent. The ability of different strains of IAV to cause disease varies widely between viruses and infected hosts. Numerous examples exist in which IAV strains emerge in nature with enhanced pathogenicity, transmissibility, or other altered phenotypic properties.

The emergence of HPAI strains in domestic poultry is one such example. These viruses share a class of mutation, independently arising on numerous occasions, creating an insertional element in the HA cleavage site of avian H5 or H7 subtype IAV which codes for multiple basic amino acids (19) and which allows these viruses to be activated by ubiquitous furin-like proteases. Such insertions create strains capable of causing systemic viral replication associated with severe and fatal disease in infected poultry (i.e., HPAI strains) but generally not in humans or other mammals, in which HPAI viruses as a group are unable to cause disease. In the case of HPAI H5N1, epizootic poultry disease also forms a bridge of viral access to poultry-exposed humans.

Other noteworthy examples of IAV breaking this first dual use rule include the emergence of pandemic IAV and the continual evolution of seasonal IAV strains. IAV pandemics have occurred sporadically since at least 1510 (20) and four times in the last 100 years-1918 H1N1, 1957 H2N2, 1968 H3N2, and 2009 H1N1 (21). Each pandemic emergence has featured an antigenically novel HA associated with enhanced morbidity and mortality (22), including those in younger age groups (23). The epidemiological impact of seasonal influenza varies from year to year (24), with morbidity and mortality being in some years on par with those of pandemic years (21). For example, the moderately severe A/Fujian/411/2002-like epidemic of the 2003-to-2004 season (25) was caused by an intrasubtypic reassortment between different clades of human H3N2 viruses (26, 27). It is sobering to realize that virtually all known deaths from influenza-23,000 to 36,000 deaths in the United States in a typical year and as many as 50 million deaths globally during a pandemic-result from the natural ability of IAV to break this first dual use rule by undergoing genetic alterations in nature that enable efficient infection, severe disease, and sometimes death.

(ii) Disrupting immunity or the effectiveness of an immunization. Like other RNA viruses, IAV disrupt host immunity by a variety of mechanisms, including down-regulating host type I interferon (IFN) responses (28). One key component of this is the IAV nonstructural protein NS1, which acts pleiotropically to attenuate the host interferon responses. Sequence variations, mutations, and in-frame deletions within this gene among different IAV strains may account for altered pathogenicity (29). For example, the NS1 protein from the 1918 influenza virus is a very potent inhibitor of antiviral and type I IFN responses in cultured human lung cells (30). Other NS1 mutations have been correlated with enhanced pathogenicity of HPAI H5N1 viruses (31–33). The alternatively spliced PB1-F2 may independently alter host IFN signaling via a mitochondrial component of the pathway (34), and mutations in the PB1-F2 protein of the 1918 virus and some HPAI H5N1 viruses have been linked to enhanced pathogenicity (35) and enhanced risk of secondary bacterial coinfections (36, 37). The newly described frameshifted PA-X protein may also play a role in host response modulation: reduced PA-X expression augments the pathogenicity of the 1918 influenza virus in mice (38).

One of the greatest hurdles to mitigating the public health impact of influenza is the rapid and constant evolution of seasonal IAV (39), necessitating the annual production of vaccines containing new strains that keep pace with the antigenic drift of circulating strains (40). Regular emergence of antigenic escape mutants in the HA and also in the neuraminidase (NA) genes is a defining feature of IAV biology in humans (41, 42) and an important example of dual use rule-breaking in nature. Antigenic escape occurs through acquisition of point mutations in the HA and NA genes, by intrasubtypic reassortment, or, dramatically, by the de novo emergence of pandemic virus strains with novel HA subtypes. If human IAV were unable to undergo continual and successful antigenic drift, growing population immunity would soon drive them to complete extinction. Breaking the second dual use rule is thus a fundamental mechanism of human influenza disease pathogenesis.

(iii) Conferring resistance to prophylactic or therapeutic interventions. Four approved antiviral drugs in two categories—the adamantane ion channel blockers and the neuraminidase inhibitors-are available for IAV treatment. Unfortunately, human and animal origin IAV often develop mutations conferring resistance to each of these drugs (43). Circulating human seasonal H3N2 viruses developed adamantane resistance after 2004 (44), severely limiting drug utility in prevention and treatment. Similarly, circulating human seasonal H1N1 viruses developed resistance to the most commonly used neuraminidase inhibitor, oseltamivir (45, 46). Upon its introduction in humans, 2009 pandemic H1N1 (H1N1pdm) viruses were already resistant to the adamantanes by virtue of a mutation in the matrix M2 gene derived from one of the parental swine IAV (47) involved in the reassortment event that generated it (48). Isolates with resistance to oseltamivir were also commonly observed (49). Resistance to other neuraminidase inhibitors, including zanamavir and the newer peramivir, has also been described with different subtypes of human IAV (50–53), and oseltamivir resistance has been described after treatment of patients infected with HPAI H5N1 viruses (54, 55). The recent H7N9 viruses possess an M2 mutation conferring adamantine resistance (56), and one of the human isolates has an NA mutation that likely confers reduced sensitivity to oseltamivir (57). Nature therefore easily and repeatedly breaks the third dual use rule by creating drug-resistant mutant viruses, reducing IAV treatment options.

(iv) Increasing the transmissibility of the agent. Pandemic IAV strains are zoonotically derived, either through reassortment with human- or mammal-adapted influenza virus or through *de novo* adaptation. The 2009 pandemic H1N1 virus was derived by reassortment of two preexisting swine IAV (48), which had both circulated in pigs and had caused only limited dead-end infections in humans (58). Why then did the 2009 pandemic reassortant

virus transmit between humans? Recent studies have shown correlations between transmission in ferrets and the unique constellation of genes associated with the 2009 pandemic virus (the Eurasian avian-like swine H1N1 IAV parental virus contributed its NA and matrix segments), in which both NA activity and viral morphology played a role in enhanced respiratory droplet transmission (59) as well as HA-NA functional balance (60). Since all known pandemic influenza viruses have been derived from precursor zoonotic IAVs or their gene segments and yet are necessarily highly transmissible between humans, new pandemic viruses must acquire genetic changes that break the fourth dual use rule of enhancing transmissibility; moreover, because pandemic viruses give rise to subsequent seasonal IAV, all human IAV result directly from the continual breaking of this rule.

(v) Altering the host range of the agent. One of the most important features of IAV ecobiology is the ability to undergo stable host switching, including bird-to-mammal switches (2, 61). The host range of IAV is very broad, including many species of wild birds, domestic anseriform and gallinaceous poultry, humans, swine, horses, dogs, cats, seals, and other mammals (2). All pandemic IAV infections are likely zoonotic, involving adaptations to humans of zoonotically derived viruses either *in toto* (e.g., 2009 pH1N1 [48] and possibly 1918 H1N1 [62]) or by reassortment that incorporates avian influenza virus gene segments into an existing human-adapted virus (e.g., 1957 H2N2 and 1968 H3N2 [63, 64]). Numerous other alterations of host range have been documented, some of which are noted here.

In 1979, an avian H1N1 virus adapted *in toto* to northern European swine (65, 66), creating a new swine lineage genetically and antigenically distinct from the North American classical swine H1N1 lineage thought to be derived from the 1918 pandemic influenza virus (67, 68). This new H1N1 lineage then replaced classical swine H1N1 viruses in Europe (69). Other avian IAV have also infected swine, including an independently emerging avian H1N1 virus in China (70). H4N6, H3N3, H1N1, and H2N3 avian viruses have all caused swine epizootics in North America (71–73).

Reassortment between swine IAV strains resulted in the 2009 pandemic H1N1 emergence, a particularly unfortunate example of many such swine host-switching events that have been occurring with increasing frequency (74). One of the parental viruses of the 2009 pandemic was derived from a North American lineage of swine-adapted IAV, itself a product of complex reassortment events between swine, human, and avian IAV (75). These triplereassortant swine viruses, containing an epizootically active "triple-reassortant internal gene," or TRIG, cassette, have been evolving dynamically at the swine-human interface (76–78), and human infections with a novel variant H3N2 swine IAV have recently been noted as well (79, 80). The recent development of complex global hyperevolution of IAV within swine is an ominous occurrence that may greatly increase the risk of future pandemics.

Equine influenza has been recognized clinically for centuries (81) and since the 1950s has been linked separately to H7N7 and H3N8 viruses. In 1989, an independent avian H3N8 host-switch event caused a serious new equine epizootic in China (82). Interestingly, equine H3N8 viruses have undergone further host-switching events to become stably adapted to and transmitted between dogs (83). Clinically documented for centuries, canine influenza has long been considered a dead-end infection acquired from horses which never led to stable canine adaptation. Yet an

avian H3N2 virus recently jumped into dog populations in North Korea and China, creating a second new stably adapted canine virus (84). Curiously, these two canine-adapted IAV strains, as well as a group of new swine H3N2 viruses with the TRIG cassette and the related 2009 pandemic H1N1 virus, all have the same peculiar genetic marker in the PA-X open reading frame that may indicate an unappreciated property of host switching or new-host adaptation (85). Thus, IAV exist in a large complex ecosystem that includes not only hundreds of species of wild birds but also poultry and numerous mammals and is characterized by numerous naturally occurring host-switching events that break the fifth dual use rule. Understanding how these host switches occur is an important and growing area of research likely to have a bearing on understanding and controlling pandemic emergence.

(vi) Enhancing the susceptibility of a host population. Enhancing the susceptibility of a host population can theoretically occur via a number of different mechanisms, including several of those already discussed, such as disrupting immunity or eluding immune response to immunizations, conferring resistance to prophylactic agents, improving transmissibility, and altering host range. In addition, influenza viruses may break the susceptibility rule by enhancing susceptibility to copathogenic infectious diseases. It is clearly established that secondary bacterial pneumonias following primary influenza virus infection play a key role in enhanced IAV morbidity and mortality (86, 87). Nasopharyngeal carriage of potential respiratory bacterial pathogens is common (88–91) and may predispose to coinfection (86). The pathogenetic mechanisms by which influenza virus and bacterial coinfection induce more-severe disease are complex and multifactorial (86, 92, 93) and include not only poorly understood viral properties but also respiratory epithelial dysfunction, impaired mucociliary clearance, enhanced bacterial adhesion, epithelial cell death, and apoptosis. Some IAV (e.g., the 1918 pandemic virus) are more capable of enhancing susceptibility to secondary bacterial pneumonia, albeit by mechanisms not fully understood. That nature alters the susceptibility of humans to severe and fatal pneumonias caused by viruses that interact with other microorganisms in the production of copathogenetic disease is another cogent example of breaking this sixth dual use rule.

(vii) Generating or reconstituting an eradicated or extinct agent. The 1918 pandemic virus was sequenced and reconstructed using an archaevirologic approach (94); study of this virus has yielded many insights into IAV biology (95). Epidemiological data, however, also support the idea of cyclic reemergence of human pandemic IAV with antigenically related HAs (i.e., antigenic recycling), leading to relative protection of persons of certain age groups in different pandemics (20). For example, elderly populations in 1918 are thought to have experienced immunoprotection derived from prior exposure to an antigenically related virus which may have emerged in the 1830s (20, 96); serologic and epidemiological data from the 1968 pandemic suggest analogous immunoprotection in persons born before 1893 (97), supporting the idea that the 1889 pandemic could have been caused by an H3 subtype virus. The 2009 pandemic virus has an H1 HA antigenically related to, and in fact directly derived from, the 1918 virus via the classical swine H1N1 viral lineage, explaining why persons born before 1950 showed evidence of immunoprotection in 2009 (98). Since HA subtypes circulating in the wild bird reservoir undergo little directed antigenic drift (3), independently emerging pandemic viruses bearing the same HA subtype are likely to be antigenically similar. Thus, nature continues to repeatedly break this final dual use rule by recycling viruses or viral antigens from the past.

CONCLUSION

Influenza A viruses are important human and animal pathogens and are among the leading infectious causes of human deaths globally. Influenza pandemics have been emerging for over a millennium, and we will undoubtedly see novel IAV continue to evolve for efficient human adaptation and pandemic spread in the future. Whether currently circulating avian H5N1, H9N2, or the more recent H7N9 viruses can adapt to efficient transmissibility in humans remains unknown, but the consequences of such a host switch event might be devastating (14). It is of vital importance to continue to study, utilizing appropriate biosafety and biosecurity oversight, how IAV can switch hosts, develop high transmissibility in human populations, escape immunity, develop antiviral resistance, and achieve other phenotypic properties that have a bearing upon their disease-producing and epidemic/pandemic potential.

The DURC framework to guide decisions about the funding of H5N1 experiments is a significant step in addressing biosafety and biosecurity concerns (99). But while scientists and policy makers continue to grapple with safety issues, IAV continue to evolve in complex and unpredictable ways by breaking, as a matter of their adaptable ecobiology, all of the very same dual use rules. It is an irony that influenza viruses have survived to cause millions of cumulative deaths precisely because they have learned how to break our self-imposed safety "rules" and that these rules might someday have the unintended consequence of limiting the very research that can potentially decipher the most devastating of IAV's secrets and that allows us to develop critically needed preventive and therapeutic modalities.

The issue naturally arises as to whether, in our efforts to save lives and prevent disease, we are better off avoiding research designed to elucidate the mechanisms of IAV-induced disease and host switching or whether we should proceed in conducting this type of research in as safe a manner as is possible. Like so much else in science and public health, it comes down to weighing the risks and benefits. In this case, the risks of inaction are predictable: influenza pandemics, epidemics, and epizootics will continue to wreak havoc for the foreseeable future at the cost of countless lives. In considering the relative merits of supporting or thwarting safe research aimed at better preventing and controlling influenza by elucidating fundamental viral mechanisms, IAV's own actions should surely be borne in mind.

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REFERENCES

- Palese P, Shaw ML. 2007. Orthomyxoviridae: the viruses and their replication, p 1647–1690. *In* Knipe DM, Howley PM (ed), Fields virology, vol 2, 5th ed. Lippincott Williams & Wilkins, Philadelphia, PA.
- Wright PF, Neumann G, Kawaoka Y. 2007. Orthomyxoviruses, p 1691–1740. *In* Knipe DM, Howley PM (ed), Fields virology, vol 2, 5th ed. Lippincott Williams & Wilkins, Philadelphia, PA.
- 3. Dugan VG, Chen R, Spiro DJ, Sengamalay N, Zaborsky J, Ghedin E, Nolting J, Swayne DE, Runstadler JA, Happ GM, Senne DA, Wang R,

Slemons RD, Holmes EC, Taubenberger JK. 2008. The evolutionary genetics and emergence of avian influenza viruses in wild birds. PLoS Pathog. 4:e1000076. doi: 10.1371/journal.ppat.1000076.

- Taubenberger JK, Kash JC. 2010. Influenza virus evolution, host adaptation, and pandemic formation. Cell Host Microbe 7:440–451.
- Guan Y, Poon LL, Cheung CY, Ellis TM, Lim W, Lipatov AS, Chan KH, Sturm-Ramirez KM, Cheung CL, Leung YH, Yuen KY, Webster RG, Peiris JS. 2004. H5N1 influenza: a protean pandemic threat. Proc. Natl. Acad. Sci. U. S. A. 101:8156–8161.
- 6. Peiris JS, de Jong MD, Guan Y. 2007. Avian influenza virus (H5N1): a threat to human health. Clin. Microbiol. Rev. 20:243–267.
- WHO. 2013. Cumulative number of confirmed human cases for avian influenza A(H5N1) reported to WHO, 2003–2013. http://www.who.int/influenza /human_animal_interface/EN_GIP_20130604CumulativeNumberH5N1 cases.pdf.
- WHO. 2013. Number of confirmed human cases of avian influenza A(H7N9) reported to WHO. http://www.who.int/influenza/human _animal_interface/influenza_h7n9/08_ReportWebH7N9Number.pdf.
- Kageyama T, Fujisaki S, Takashita E, Xu H, Yamada S, Uchida Y, Neumann G, Saito T, Kawaoka Y, Tashiro M. 2013. Genetic analysis of novel avian A(H7N9) influenza viruses isolated from patients in China, February to April 2013. Euro Surveill. 18:20453.
- 10. Chen Y, Liang W, Yang S, Wu N, Gao H, Sheng J, Yao H, Wo J, Fang Q, Cui D, Li Y, Yao X, Zhang Y, Wu H, Zheng S, Diao H, Xia S, Zhang Y, Chan KH, Tsoi HW, Teng JL, Song W, Wang P, Lau SY, Zheng M, Chan JF, To KK, Chen H, Li L, Yuen KY. 2013. Human infections with the emerging avian influenza A H7N9 virus from wet market poultry: clinical analysis and characterisation of viral genome. Lancet 381: 1916–1925.
- Morens DM, Taubenberger JK, Fauci AS. 2013. Pandemic influenza viruses—hoping for the road not taken. N. Engl. J. Med. 368:2345–2348.
- 12. Herfst S, Schrauwen EJ, Linster M, Chutinimitkul S, de Wit E, Munster VJ, Sorrell EM, Bestebroer TM, Burke DF, Smith DJ, Rimmelzwaan GF, Osterhaus AD, Fouchier RA. 2012. Airborne transmission of influenza A/H5N1 virus between ferrets. Science 336:1534–1541.
- 13. Imai M, Watanabe T, Hatta M, Das SC, Ozawa M, Shinya K, Zhong G, Hanson A, Katsura H, Watanabe S, Li C, Kawakami E, Yamada S, Kiso M, Suzuki Y, Maher EA, Neumann G, Kawaoka Y. 2012. Experimental adaptation of an influenza H5 HA confers respiratory droplet transmission to a reassortant H5 HA/H1N1 virus in ferrets. Nature 486:420-428.
- Morens DM, Subbarao K, Taubenberger JK. 2012. Engineering H5N1 avian influenza viruses to study human adaptation. Nature 486:335–340.
- Fouchier R, Osterhaus AB, Steinbruner J, Yuen KY, Henderson DA, Klotz L, Sylvester E, Taubenberger JK, Ebright RH, Heymann DL. 2012. Preventing pandemics: the fight over flu. Nature 481:257–259.
- Zhang Y, Zhang Q, Kong H, Jiang Y, Gao Y, Deng G, Shi J, Tian G, Liu L, Liu J. 2013. H5N1 hybrid viruses bearing 2009/H1N1 virs genes transmit in guinea pigs by respiratory droplet. Science 340:1459–1463.
- 17. Perez DR. 2012. Public health and biosecurity. H5N1 debates: hung up on the wrong questions. Science 335:799–801.
- Office of Biotechnology Activities NIH. 2012. U.S. government policy for oversight of life sciences dual use research of concern. http://www.phe .gov/s3/dualuse/Documents/us-policy-durc-032812.pdf.
- Alexander DJ, Brown IH. 2009. History of highly pathogenic avian influenza. Rev. Sci. Tech. 28:19–38.
- Morens DM, Taubenberger JK. 2011. Pandemic influenza: certain uncertainties. Rev. Med. Virol. 21:262–284.
- Morens DM, Taubenberger JK, Fauci AS. 2009. The persistent legacy of the 1918 influenza virus. N. Engl. J. Med. 361:225–229.
- Miller MA, Viboud C, Balinska M, Simonsen L. 2009. The signature features of influenza pandemics—implications for policy. N. Engl. J. Med. 360:2595–2598.
- Simonsen L, Clarke MJ, Schonberger LB, Arden NH, Cox NJ, Fukuda K. 1998. Pandemic versus epidemic influenza mortality: a pattern of changing age distribution. J. Infect. Dis. 178:53–60.
- Thompson MG, Shay DK, Zhou H, Bridges CB, Cheng PY, Burns E, Bresee JS, Cox NJ. 2010. Estimates of deaths associated with seasonal influenza—United States, 1976–2007. MMWR Morb. Mortal. Wkly. Rep. 59:1057–1062.
- CDC. 2004. 2003–2004 U.S. influenza season summary. http://www.cdc .gov/flu/weekly/weeklyarchives2003-2004/03-04summary.htm.
- Holmes EC, Ghedin E, Miller N, Taylor J, Bao Y, St George K, Grenfell BT, Salzberg SL, Fraser CM, Lipman DJ, Taubenberger JK. 2005.

Whole-genome analysis of human influenza A virus reveals multiple persistent lineages and reassortment among recent H3N2 viruses. PLoS Biol. 3:e300. doi: 10.1371/journal.pbio.0030300.

- Memoli MJ, Jagger BW, Dugan VG, Qi L, Jackson JP, Taubenberger JK. 2009. Recent human influenza A/H3N2 virus evolution driven by novel selection factors in addition to antigenic drift. J. Infect. Dis. 200: 1232–1241.
- García-Sastre A. 2011. Induction and evasion of type I interferon responses by influenza viruses. Virus Res. 162:12–18.
- Hale BG, Randall RE, Ortín J, Jackson D. 2008. The multifunctional NS1 protein of influenza A viruses. J. Gen. Virol. 89:2359–2376.
- 30. Geiss GK, Salvatore M, Tumpey TM, Carter VS, Wang X, Basler CF, Taubenberger JK, Bumgarner RE, Palese P, Katze MG, García-Sastre A. 2002. Cellular transcriptional profiling in influenza A virus-infected lung epithelial cells: the role of the nonstructural NS1 protein in the evasion of the host innate defense and its potential contribution to pandemic influenza. Proc. Natl. Acad. Sci. U. S. A. 99:10736–10741.
- 31. Seo SH, Hoffmann E, Webster RG. 2002. Lethal H5N1 influenza viruses escape host anti-viral cytokine responses. Nat. Med. 8:950–954.
- 32. Lipatov AS, Andreansky S, Webby RJ, Hulse DJ, Rehg JE, Krauss S, Perez DR, Doherty PC, Webster RG, Sangster MY. 2005. Pathogenesis of Hong Kong H5N1 influenza virus NS gene reassortants in mice: the role of cytokines and B- and T-cell responses. J. Gen. Virol. 86:1121–1130.
- 33. Jiao P, Tian G, Li Y, Deng G, Jiang Y, Liu C, Liu W, Bu Z, Kawaoka Y, Chen H. 2008. A single-amino-acid substitution in the NS1 protein changes the pathogenicity of H5N1 avian influenza viruses in mice. J. Virol. 82:1146–1154.
- McAuley JL, Chipuk JE, Boyd KL, Van De Velde N, Green DR, Mc-Cullers JA. 2010. PB1-F2 proteins from H5N1 and 20 century pandemic influenza viruses cause immunopathology. PLoS Pathog. 6:e1001014. doi: 10.1371/journal.ppat.1001014.
- 35. Conenello GM, Zamarin D, Perrone LA, Tumpey T, Palese P. 2007. A single mutation in the PB1-F2 of H5N1 (HK/97) and 1918 influenza A viruses contributes to increased virulence. PLoS Pathog. 3:1414–1421. doi: 10.1371/journal.ppat.0030141.
- McAuley JL, Hornung F, Boyd KL, Smith AM, McKeon R, Bennink J, Yewdell JW, McCullers JA. 2007. Expression of the 1918 influenza A virus PB1-F2 enhances the pathogenesis of viral and secondary bacterial pneumonia. Cell Host Microbe 2:240–249.
- Smith AM, McCullers JA. 14 March 2013. Molecular signatures of virulence in the PB1-F2 proteins of H5N1 influenza viruses. Virus Res. doi: 10.1016/j.virusres.2013.02.012. [Epub ahead of print.]
- 38. Jagger BW, Wise HM, Kash JC, Walters KA, Wills NM, Xiao YL, Dunfee RL, Schwartzman LM, Ozinsky A, Bell GL, Dalton RM, Lo A, Efstathiou S, Atkins JF, Firth AE, Taubenberger JK, Digard P. 2012. An overlapping protein-coding region in influenza A virus segment 3 modulates the host response. Science 337:199–204.
- Yewdell JW. 2011. Viva la revolucion: rethinking influenza A virus antigenic drift. Curr. Opin. Virol. 1:177–183.
- Morens DM, Taubenberger JK, Fauci AS. 2010. The 2009 H1N1 pandemic influenza virus: what next? mBio 1:e00211-10. doi: 10.1128/ mBio.00211-10.
- Fitch WM, Leiter JM, Li XQ, Palese P. 1991. Positive Darwinian evolution in human influenza A viruses. Proc. Natl. Acad. Sci. U. S. A. 88: 4270–4274.
- Smith DJ, Lapedes AS, de Jong JC, Bestebroer TM, Rimmelzwaan GF, Osterhaus AD, Fouchier RA. 2004. Mapping the antigenic and genetic evolution of influenza virus. Science 305:371–376.
- Hayden FG. 2006. Antivirals for influenza: historical perspectives and lessons learned. Antiviral Res. 71:372–378.
- 44. Bright RA, Shay DK, Shu B, Cox NJ, Klimov AI. 2006. Adamantane resistance among influenza A viruses isolated early during the 2005–2006 influenza season in the United States. JAMA 295:891–894.
- Moscona A. 2009. Global transmission of oseltamivir-resistant influenza. N. Engl. J. Med. 360:953–956.
- Bloom JD, Gong LI, Baltimore D. 2010. Permissive secondary mutations enable the evolution of influenza oseltamivir resistance. Science 328: 1272–1275.
- Krumbholz A, Schmidtke M, Bergmann S, Motzke S, Bauer K, Stech J, Dürrwald R, Wutzler P, Zell R. 2009. High prevalence of amantadine resistance among circulating European porcine influenza A viruses. J. Gen. Virol. 90:900–908.
- 48. Garten RJ, Davis CT, Russell CA, Shu B, Lindstrom S, Balish A,

Sessions WM, Xu X, Skepner E, Deyde V, Okomo-Adhiambo M, Gubareva L, Barnes J, Smith CB, Emery SL, Hillman MJ, Rivailler P, Smagala J, de Graaf M, Burke DF, Fouchier RA, Pappas C, Alpuche-Aranda CM, López-Gatell H, Olivera H, López I, Myers CA, Faix D, Blair PJ, Yu C, Keene KM, Dotson PD, Boxrud D, Sambol AR, Abid SH, St George K, Bannerman T, Moore AL, Stringer DJ, Blevins P, Demmler-Harrison GJ, Ginsberg M, Kriner P, Waterman S, Smole S, Guevara HF, Belongia EA, Clark PA, Beatrice ST, et al. 2009. Antigenic and genetic characteristics of swine-origin 2009 A(H1N1) influenza viruses circulating in humans. Science 325:197–201.

- Renaud C, Kuypers J, Englund JA. 2011. Emerging oseltamivir resistance in seasonal and pandemic influenza A/H1N1. J. Clin. Virol. 52:70–78.
- Samson M, Pizzorno A, Abed Y, Boivin G. 2013. Influenza virus resistance to neuraminidase inhibitors. Antiviral Res. 98:174–185.
- Memoli MJ, Davis AS, Proudfoot K, Chertow DS, Hrabal RJ, Bristol T, Taubenberger JK. 2011. Multidrug-resistant 2009 pandemic influenza A(H1N1) viruses maintain fitness and transmissibility in ferrets. J. Infect. Dis. 203:348–357.
- Memoli MJ, Hrabal RJ, Hassantoufighi A, Eichelberger MC, Taubenberger JK. 2010. Rapid selection of oseltamivir- and peramivir-resistant pandemic H1N1 virus during therapy in 2 immunocompromised hosts. Clin. Infect. Dis. 50:1252–1255.
- 53. Memoli MJ, Hrabal RJ, Hassantoufighi A, Jagger BW, Sheng ZM, Eichelberger MC, Taubenberger JK. 2010. Rapid selection of a transmissible multidrug-resistant influenza A/H3N2 virus in an immunocompromised host. J. Infect. Dis. 201:1397–1403.
- 54. de Jong MD, Tran TT, Truong HK, Vo MH, Smith GJ, Nguyen VC, Bach VC, Phan TQ, Do QH, Guan Y, Peiris JS, Tran TH, Farrar J. 2005. Oseltamivir resistance during treatment of influenza A (H5N1) infection. N. Engl. J. Med. 353:2667–2672.
- 55. Le QM, Kiso M, Someya K, Sakai YT, Nguyen TH, Nguyen KH, Pham ND, Ngyen HH, Yamada S, Muramoto Y, Horimoto T, Takada A, Goto H, Suzuki T, Suzuki Y, Kawaoka Y. 2005. Avian flu: isolation of drugresistant H5N1 virus. Nature 437:1108.
- Liu Q, Lu L, Sun Z, Chen GW, Wen Y, Jiang S. 2013. Genomic signature and protein sequence analysis of a novel influenza A (H7N9) virus that causes an outbreak in humans in China. Microbes Infect. 15:432–439.
- 57. Liu D, Shi W, Shi Y, Wang D, Xiao H, Li W, Bi Y, Wu Y, Li X, Yan J, Liu W, Zhao G, Yang W, Wang Y, Ma J, Shu Y, Lei F, Gao GF. 2013. Origin and diversity of novel avian influenza A H7N9 viruses causing human infection: phylogenetic, structural, and coalescent analyses. Lancet 381:1926–1932.
- 58. Myers KP, Olsen CW, Gray GC. 2007. Cases of swine influenza in humans: a review of the literature. Clin. Infect. Dis. 44:1084–1088.
- 59. Lakdawala SS, Lamirande EW, Suguitan AL, Jr, Wang W, Santos CP, Vogel L, Matsuoka Y, Lindsley WG, Jin H, Subbarao K. 2011. Eurasianorigin gene segments contribute to the transmissibility, aerosol release, and morphology of the 2009 pandemic H1N1 influenza virus. PLoS Pathog. 7:e1002443. doi: 10.1371/journal.ppat.1002443.
- 60. Yen HL, Liang CH, Wu CY, Forrest HL, Ferguson A, Choy KT, Jones J, Wong DD, Cheung PP, Hsu CH, Li OT, Yuen KM, Chan RW, Poon LL, Chan MC, Nicholls JM, Krauss S, Wong CH, Guan Y, Webster RG, Webby RJ, Peiris M. 2011. Hemagglutinin-neuraminidase balance confers respiratory-droplet transmissibility of the pandemic H1N1 influenza virus in ferrets. Proc. Natl. Acad. Sci. U. S. A. 108:14264–14269.
- Parrish CR, Holmes EC, Morens DM, Park EC, Burke DS, Calisher CH, Laughlin CA, Saif LJ, Daszak P. 2008. Cross-species virus transmission and the emergence of new epidemic diseases. Microbiol. Mol. Biol. Rev. 72:457–470.
- Taubenberger JK, Reid AH, Lourens RM, Wang R, Jin G, Fanning TG. 2005. Characterization of the 1918 influenza virus polymerase genes. Nature 437:889–893.
- Scholtissek C, Rohde W, Von Hoyningen V, Rott R. 1978. On the origin of the human influenza virus subtypes H2N2 and H3N2. Virology 87: 13–20.
- 64. Kawaoka Y, Krauss S, Webster RG. 1989. Avian-to-human transmission of the PB1 gene of influenza A viruses in the 1957 and 1968 pandemics. J. Virol. 63:4603–4608.
- Dunham EJ, Dugan VG, Kaser EK, Perkins SE, Brown IH, Holmes EC, Taubenberger JK. 2009. Different evolutionary trajectories of European avian-like and classical swine H1N1 influenza A viruses. J. Virol. 83: 5485–5494.
- 66. Pensaert M, Ottis K, Vandeputte J, Kaplan MM, Bachmann PA. 1981.

Evidence for the natural transmission of influenza A virus from wild ducks to swine and its potential importance for man. Bull. World Health Organ. **59**:75–78.

- 67. Taubenberger JK, Reid AH, Janczewski TA, Fanning TG. 2001. Integrating historical, clinical and molecular genetic data in order to explain the origin and virulence of the 1918 Spanish influenza virus. Philos. Trans. R. Soc. Lond. B Biol. Sci. 356:1829–1839.
- Shope RE. 1936. The incidence of neutralizing antibodies for swine influenza virus in the sera of human beings of different ages. J. Exp. Med. 63:669–684.
- 69. Brown IH. 2000. The epidemiology and evolution of influenza viruses in pigs. Vet. Microbiol. 74:29–46.
- Guan Y, Shortridge KF, Krauss S, Li PH, Kawaoka Y, Webster RG. 1996. Emergence of avian H1N1 influenza viruses in pigs in China. J. Virol. 70:8041–8046.
- Karasin AI, Brown IH, Carman S, Olsen CW. 2000. Isolation and characterization of H4N6 avian influenza viruses from pigs with pneumonia in Canada. J. Virol. 74:9322–9327.
- Karasin AI, West K, Carman S, Olsen CW. 2004. Characterization of avian H3N3 and H1N1 influenza A viruses isolated from pigs in Canada. J. Clin. Microbiol. 42:4349–4354.
- 73. Ma W, Vincent AL, Gramer MR, Brockwell CB, Lager KM, Janke BH, Gauger PC, Patnayak DP, Webby RJ, Richt JA. 2007. Identification of H2N3 influenza A viruses from swine in the United States. Proc. Natl. Acad. Sci. U. S. A. 104:20949–20954.
- 74. Vincent A, Awada L, Brown I, Chen H, Claes F, Dauphin G, Donis R, Culhane M, Hamilton K, Lewis N, Mumford E, Nguyen T, Parchariyanon S, Pasick J, Pavade G, Pereda A, Peiris M, Saito T, Swenson S, Van Reeth K, Webby R, Wong F, Ciacci-Zanella J. 5 April 2013. Review of influenza A virus in swine worldwide: a call for increased surveillance and research. Zoonoses Public Health doi: 10.1111/zph.12049. [Epub ahead of print].
- Zhou NN, Senne DA, Landgraf JS, Swenson SL, Erickson G, Rossow K, Liu L, Yoon Kj, Krauss S, Webster RG. 1999. Genetic reassortment of avian, swine, and human influenza A viruses in American pigs. J. Virol. 73:8851–8856.
- Barman S, Krylov PS, Fabrizio TP, Franks J, Turner JC, Seiler P, Wang D, Rehg JE, Erickson GA, Gramer M, Webster RG, Webby RJ. 2012. Pathogenicity and transmissibility of North American triple reassortant swine influenza A viruses in ferrets. PLoS Pathog. 8:e1002791. doi: 10.1371/journal.ppat.1002791.
- 77. Nelson MI, Detmer SE, Wentworth DE, Tan Y, Schwartzbard A, Halpin RA, Stockwell TB, Lin X, Vincent AL, Gramer MR, Holmes EC. 2012. Genomic reassortment of influenza A virus in North American swine, 1998–2011. J. Gen. Virol. 93:2584–2589.
- Nelson MI, Gramer MR, Vincent AL, Holmes EC. 2012. Global transmission of influenza viruses from humans to swine. J. Gen. Virol. 93: 2195–2203.
- CDC. 2012. Notes from the field: outbreak of influenza A (H3N2) virus among persons and swine at a county fair—Indiana, July 2012. MMWR Morb. Mortal. Wkly. Rep. 61:561.
- Nelson MI, Vincent AL, Kitikoon P, Holmes EC, Gramer MR. 2012. Evolution of novel reassortant A/H3N2 influenza viruses in North American swine and humans, 2009–2011. J. Virol. 86:8872–8878.
- Morens DM, Taubenberger JK. 2010. Historical thoughts on influenza viral ecosystems, or behold a pale horse, dead dogs, failing fowl, and sick swine. Influenza Other Resp. Viruses 4:327–337.

- Guo Y, Wang M, Kawaoka Y, Gorman O, Ito T, Saito T, Webster RG. 1992. Characterization of a new avian-like influenza A virus from horses in China. Virology 188:245–255.
- 83. Crawford PC, Dubovi EJ, Castleman WL, Stephenson I, Gibbs EP, Chen L, Smith C, Hill RC, Ferro P, Pompey J, Bright RA, Medina MJ, Johnson CM, Olsen CW, Cox NJ, Klimov AI, Katz JM, Donis RO. 2005. Transmission of equine influenza virus to dogs. Science 310:482–485.
- Song D, Kang B, Lee C, Jung K, Ha G, Kang D, Park S, Park B, Oh J. 2008. Transmission of avian influenza virus (H3N2) to dogs. Emerg. Infect. Dis. 14:741–746.
- Shi M, Jagger BW, Wise HM, Digard P, Holmes EC, Taubenberger JK. 2012. Evolutionary conservation of the PA-X open reading frame in segment 3 of influenza A virus. J. Virol. 86:12411–12413.
- Chertow DS. 2012. Contribution of bacterial coinfection to severe influenza infection. Crit. Care Med. 40:1664–1665.
- Morens DM, Taubenberger JK, Fauci AS. 2008. Predominant role of bacterial pneumonia as a cause of death in pandemic influenza: implications for pandemic influenza preparedness. J. Infect. Dis. 198:962–970.
- García-Rodríguez JA, Fresnadillo Martínez MJ. 2002. Dynamics of nasopharyngeal colonization by potential respiratory pathogens. J. Antimicrob. Chemother. 50(Suppl. S2): 59–73.
- Kluytmans JA, Wertheim HF. 2005. Nasal carriage of Staphylococcus aureus and prevention of nosocomial infections. Infection 33:3–8.
- Simell B, Auranen K, Käyhty H, Goldblatt D, Dagan R, O'Brien KL, Pneumococcal Carriage Group. 2012. The fundamental link between pneumococcal carriage and disease. Expert Rev. Vaccines 11:841–855.
- Wertheim HF, Melles DC, Vos MC, van Leeuwen W, van Belkum A, Verbrugh HA, Nouwen JL. 2005. The role of nasal carriage in Staphylococcus aureus infections. Lancet Infect. Dis. 5:751–762.
- 92. Kash JC, Walters KA, Davis AS, Sandouk A, Schwartzman LM, Jagger BW, Chertow DS, Li Q, Kuestner RE, Ozinsky A, Taubenberger JK. 2011. Lethal synergism of 2009 pandemic H1N1 influenza virus and Streptococcus pneumoniae coinfection is associated with loss of murine lung repair responses. mBio 2:e00172-11. doi: 10.1128/mBio.00172-11.
- McCullers JA. 2006. Insights into the interaction between influenza virus and pneumococcus. Clin. Microbiol. Rev. 19:571–582.
- Taubenberger JK, Hultin JV, Morens DM. 2007. Discovery and characterization of the 1918 pandemic influenza virus in historical context. Antivir. Ther. 12:581–591.
- Taubenberger JK, Baltimore D, Doherty PC, Markel H, Morens DM, Webster RG, Wilson IA. 2012. Reconstruction of the 1918 influenza virus: unexpected rewards from the past. mBio 3:e00201-12. doi: 10.1128/ mBio.00201-12.
- Viboud C, Eisenstein J, Reid AH, Janczewski TA, Morens DM, Taubenberger JK. 2013. Age- and sex-specific mortality associated with the 1918–1919 influenza pandemic in Kentucky. J. Infect. Dis. 207:721–729.
- 97. Dowdle WR. 2006. Influenza pandemic periodicity, virus recycling, and the art of risk assessment. Emerg. Infect. Dis. 12:34–39.
- Hancock K, Veguilla V, Lu X, Zhong W, Butler EN, Sun H, Liu F, Dong L, DeVos JR, Gargiullo PM, Brammer TL, Cox NJ, Tumpey TM, Katz JM. 2009. Cross-reactive antibody responses to the 2009 pandemic H1N1 influenza virus. N. Engl. J. Med. 361:1945–1952.
- Patterson AP, Tabak LA, Fauci AS, Collins FS, Howard S. 2013. Research funding. A framework for decisions about research with HPAI H5N1 viruses. Science 339:1036–1037.

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